Effects of Bariatric Surgery on Human Small Artery Function

Evidence for Reduction in Perivascular Adipocyte Inflammation, and the Restoration of Normal Anticontractile Activity Despite Persistent Obesity

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Objectives	The aim of this study was to investigate the effects of bariatric surgery on small artery function and the mechanisms underlying this.
Background	In lean healthy humans, perivascular adipose tissue (PVAT) exerts an anticontractile effect on adjacent small arteries, but this is lost in obesity-associated conditions such as the metabolic syndrome and type II diabetes where there is evidence of adipocyte inflammation and increased oxidative stress.
Methods	Segments of small subcutaneous artery and perivascular fat were harvested from severely obese individuals before $(n = 20)$ and 6 months after bariatric surgery $(n = 15)$. Small artery contractile function was examined in vitro with wire myography, and perivascular adipose tissue (PVAT) morphology was assessed with immunohistochemistry.
Results	The anticontractile activity of PVAT was lost in obese patients before surgery when compared with healthy volunteers and was restored 6 months after bariatric surgery. In vitro protocols with superoxide dismutase and catalase rescued PVAT anticontractile function in tissue from obese individuals before surgery. The improvement in anticontractile function after surgery was accompanied by improvements in insulin sensitivity, serum glycemic indexes, inflammatory cytokines, adipokine profile, and systolic blood pressure together with increased PVAT adiponectin and nitric oxide bioavailability and reduced macrophage infiltration and inflammation. These changes were observed despite the patients remaining severely obese.
Conclusions	Bariatric surgery and its attendant improvements in weight, blood pressure, inflammation, and metabolism collectively reverse the obesity-induced alteration to PVAT anticontractile function. This reversal is attributable to reductions in local adipose inflammation and oxidative stress with improved adiponectin and nitric oxide bioavailability. (J Am Coll Cardiol 2013;62:128-35) © 2013 by the American College of Cardiology Foundation

Obesity has become a global public heath challenge affecting almost one-half billion adults (1) and an estimated 40 million children (2). Large artery disease is very common in obese patients and manifests clinically as myocardial infarction, stroke, and hypertension (3). Small artery disease is also common in obesity and contributes to the development of hypertension and microvascular disease due to changes in peripheral resistance and local autoregulation (4). Historically, the small artery dysfunction in obesity has been attributed to damage to the endothelium (5), most notably to generation and release of nitric oxide (NO) (6). More

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recently, however, appreciation has grown for an additional mechanism by which vascular damage occurs in obesity; the function of fat surrounding arteries, or perivascular adipose tissue (PVAT). The PVAT surrounds the majority of blood

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vessels in the body and, in addition to adipocytes, contains inflammatory cells, stem cells, and microvasculature. In health, PVAT modulates the contractile tone of adjacent small arteries by secreting vasodilatory molecules. These adiposederived vasodilators act independently of the endothelium and include adiponectin (7), NO (7), hydrogen sulfide (8), and palmitic acid methyl ester (9). In 2009, we performed the first human small artery study of PVAT and showed that subcutaneous gluteal PVAT from lean healthy individuals reduced adrenergic constriction in adjacent arteries, an "anticontractile" effect. However, in patients with metabolic syndrome the vasodilatory effect of the PVAT was entirely lost, due to dual processes of adipose tissue hypoxia and inflammation, both of which are established sequelae of obesity in fat depots (7). More recently, we have seen that macrophage activation in adipose tissue contributes to the attenuation in PVAT anticontractile effect (10).

Bariatric surgery has been performed for nearly 60 years and is established as the most effective clinical intervention to achieve both significant and sustained weight-loss in severely obese individuals. There are 3 types of bariatric surgical procedures: restrictive, malabsorptive, and combined operations (11). Gastric bypass surgery is a combination of restriction and malabsorption and has been shown to achieve a significantly higher degree of weight loss than restrictive bariatric surgery (12).

Bariatric surgery also dramatically improves cardiovascular risk profiles in obese patients and reduces overall mortality (13,14). The mechanisms that underlie these cardiovascular improvements remain unclear, however. In the present study we investigated the effects of bariatric surgery achieved by the gastric bypass method on the vasodilatory properties of PVAT. We report amelioration of inflammation in PVAT with complete restoration of anticontractile activity as a consequence of improved adiponectin and NO bioavailability, despite persisting obesity.

Methods

Study population. Patients with severe obesity (body mass index [BMI] >35 kg/m²) who were awaiting gastric bypass surgery (n = 15) and lean healthy volunteers (n = 7) were recruited after full informed written consent in accord with local research ethics committee approval. All participants provided a fasting venous blood sample for the measurement of inflammatory markers and adipokines.

The study participants also provided gluteal subcutaneous fat samples $(1.5 \times 1.5 \times 1.5 \text{ cm})$ by undergoing a surgical biopsy under local anesthesia (7). The sample was immediately processed in 3 sections. One part was stored for immunohistology, the second was snap-frozen for estimation of NO levels, and the remainder was used to harvest small subcutaneous arteries with micro-dissection.

Blood pressure was recorded as a mean of 3 recordings measured with a semi-automated machine (OMRON

705CP, White Medical, Clifton-Upon-Dunsmore, United Kingdom), whereas the participants were seated at rest for 15 min.

Further measurements including BMI and waist circumference were also recorded.

The obese patients were invited to return for a follow-up assessment, including a biopsy, 6 months after bariatric surgery. **Biochemical analyses.** High-sensitivity C-reactive protein (hs-

Abbreviations and Acronyms
AdipoR1 = adiponectin receptor 1
BMI = body mass index
hsCRP = high-sensitivity C-reactive protein
NO = nitric oxide
PVAT = perivascular adipose tissue
TNF = tumor necrosis factor

CRP) was measured in serum by an in-house, antibody sandwich enzyme-linked immunoadsorbent assay technique with anti-human CRP antibodies, calibrators, and controls from Abcam (Cambridge, United Kingdom). Interleukin-6, tumor necrosis factor (TNF)- α , adiponectin, leptin, and resistin were measured in serum, and E-selectin was measured in plasma, all with DuoSet ELISA development kits from R&D Systems (Abingdon, United Kingdom).

Wire myography. One section of the gluteal fat biopsy was placed in chilled physiological saline solution (composition in mmol/l: sodium chloride 118, potassium chloride 3.4, magnesium sulphate 1.2, calcium chloride 1, sodium bicarbonate 25, glucose 11, potassium orthophosphate 1.2) and oxygenated. The dissection dish was placed on ice during microdissection to preserve the integrity of the tissue, whereas arterial segments 250 to 350 μ m in diameter were harvested, 1 segment with PVAT intact, and the adjacent segment devoid of PVAT. Both segments came from the same artery.

Endothelium denuded vessels were mounted on 40-µm wires and studied with wire myography (Danish MyoTech, Aarhus, Denmark).

The mounted vessel segments were oxygenated and maintained at a temperature of 37° C before vessel diameter and wall tension was normalized as previously described (7,15). Vessels were challenged with a 60-mmol/l highpotassium physiological saline solution to establish viability and baseline constriction.

Each vessel segment was stimulated with cumulative doses of norepinephrine (Sigma-Aldrich, Dorset, United Kingdom) at the following doses: 10^{-9} ; 10^{-8} ; 3×10^{-7} ; 5×10^{-7} ; 10^{-6} ; 2×10^{-6} ; 3×10^{-6} ; 5×10^{-6} ; 10^{-5} ; 2×10^{-5} ; $and 3 \times 10^{-5}$ mol/l. Contractile responses to norepinephrine are presented as a percentage of high-potassium physiological saline solution constriction, consistent with published data (7,16–18).

Pharmacological assessment. Pharmacological protocols were applied to study the effect of PVAT on adjacent small arteries. In each case 2 segments of the same artery were prepared with and without PVAT attached as previously described (7).

The role of oxidative stress was evaluated by incubation of samples from pre-operative patients with superoxide dismutase and catalase (superoxide dismutase, Sigma-Aldrich, 100- μ /ml incubation period 45 min; and catalase, Sigma-Aldrich, 100- μ /ml incubation period 45 min; n = 4).

Further protocols assessed the contribution of NO and adiponectin to PVAT function in samples taken from patients after weight loss. Arteries with PVAT were incubated with blocking peptide for adiponectin receptor 1 (AdipoR1) (5 μ g/ml, 1.6 × 10⁻⁴ mol/l, Enzo Life Sciences; 45 min; n = 7) and N5-[imino(methylamino)methyl]-Lornithine, citrate (5 \times 10⁻⁵ mol/l, Sigma; 45 min; n = 4). NO assay. Five paired pre-operative and post-operative frozen PVAT samples were homogenized by suspending the tissue in 400 µl of lysis buffer (50 mmol/l Tris base, 150 mmol/l sodium chloride, 2 mmol/l ethylenediaminetetraacetic acid, 2 mmol/l ethyleneglycoltetraacetic acid, 40 mmol/l ß-glycerophosphate, 50 mmol/l sodium fluoride, 10 mmol/l sodium pyrophosphate, 10% glycerol, 1% Triton X-100 [pH 7.4]) and protease inhibitor (Complete Mini EDTA-Free; Roche Diagnostics, Basel, Switzerland) on ice and using a Dounce (glass/glass) tissue grinder set (Sigma-Aldrich). Upon complete homogenization, the solution was incubated at 4°C for 10 min, then centrifuged for 10 min at 16,000 g. The resulting supernatant was then removed for application of the assay.

Total nitrate/nitrite concentration was measured with a NO colorimetric assay kit according to the instructions of the manufacturer (Abcam).



same vessel (n = 20, p = 0.95). **(B)** In post-surgery patients, the presence of PVAT had a significant anticontractile effect on the small artery contractility (n = 15, *p < 0.01). KPSS = high-potassium physiological saline; Log [NE] (M) = log of the molar concentration of norepinephrine.

Immunohistochemistry. MACROPHAGE MARKERS AND TNF-a. Immediately after dissection, tissue from each gluteal biopsy consisting of skin and subcutaneous fat was placed in 4% paraformaldehyde in phosphate buffered saline for 24 h and subsequently processed to paraffin wax blocks. Consecutive 5µm sections were de-waxed, rehydrated, and immunostained for 2 macrophage markers-CD68 [KP1] (Dako, Glostrup, Denmark; dilution 1:100), and CD68 [PGM1] (Biocare Medical, Concord, California; dilution 1:100)-and the proinflammatory cytokine TNF-a (Abcam; dilution 1: 100). Antigen retrieval was performed, followed by blocking of endogenous peroxidase and nonspecific protein binding with Dako blocking solutions. Tissue sections were incubated with primary antibodies for 18 h, followed by anti-rabbit/antimouse EnVision-HRP (Dako) and finally by Vector SG chromogen kit (Vector Laboratories, Burlingame, California), used to disclose the presence of the macrophages and TNF-a. Color images were captured with a Go-3 QImaging camera (QImaging Corporation, Vancouver, Canada) mounted on



(A) Blocking peptide for polycional antibody to adiponectin receptor 1 increases vessel contractility to norepinephrine (n = 7, p <0.0001). (B) Incubation with superoxide dismutase (SOD) and catalase rescues the PVAT anticontractile effect in samples taken from pre-surgery patients (n = 7, p < 0.001). (C) Post-surgery, inhibition of nitric oxide synthase by incubation with N5-[imino(methylamino) methyl]-L-ornithine, citrate (L-NMMA) leads to increased contractility of vessel segments with intact PVAT (n = 4, *p < 0.001). Abbreviations as in Figure 1.

a Leitz Diaplan microscope (Leica, Wetzlar, Germany). Cells stained with macrophage markers present in adipose tissue were counted, and the results were expressed as cells/mm². For assessment of TNF- α , quantitative analysis of immunostaining was obtained, converting color images to greyscale and with a macro subroutine in an ImagePro version 6.2 image analysis program (MediaCybernetics United Kingdom, Marlow, United Kingdom). The extent of staining was expressed as a percentage of the entire area photographed. Adipocyte size was quantified on microphotographs obtained on Zeiss Axio Imager M2 microscope equipped with Axio-Cam camera and AxioVision Rel. 4.8 program by free-hand tracing the margins of 100 consecutive cells (on average)/case to avoid selection bias (total: 3,000 cells).

Immunohistochemistry for AdipoR1. Five µm deparaffinized formalin-fixed tissue sections were microwaved in citrate buffer pH 6.0, treated with 0.3% hydrogen peroxide, blocked with Protein Block (Dako), and incubated with 1 of the anti-AdipoR1 antibodies, goat polyclonal to AdipoR1 (Abcam), or a rabbit monoclonal to AdipoR1 (Epitomics, Burlingame, California), followed by anti-rabbit/anti-mouse EnVision-HRP (Dako) and Vector SG chromogen kit (Vector Laboratories). The results of immunostaining were compared, and the monoclonal rabbit antibody was selected for further use. Slides were examined and images captured on a Zeiss Axio Imager M2 microscope. **Statistical analysis.** The statistical presentation includes paired and unpaired tests. In the case of ordinal tests, medians and quartiles are used, and for parametric tests mean \pm SD is used. Cumulative concentration-response curves were constructed with data obtained by wire myography and analyzed with a 2-way analysis of variance and a Bonferroni post hoc test for each dose. A p value of <0.05 was considered statistically significant. Analyses were performed with the GraphPad Prism 5 software (GraphPad, La Jolla, California).

Results

Study design and participants. Six months after surgery and weight loss, the patients had a significantly lower waist circumference (n = 15; p < 0.0001), BMI (p < 0.0001), and systolic blood pressure (p = 0.0025) (Table 1).

Post-operatively, there was a significant reduction in insulin (p = 0.0042), fasting glucose (p = 0.0312), and hemoglobin A1c (p < 0.0071) and an improvement in pancreatic beta-cell function (homeostatic model assessment of beta-cell function; p < 0.01) and insulin resistance (homeostasis model assessment of insulin resistance; p = 0.0023). After surgery serum levels of adiponectin increased significantly (p = 0.0197), and leptin decreased (p = 0.0014), but resistin levels did not change significantly (p = 0.0796).



Inflammatory infiltrates composed mostly of macrophages were present in wreath-like arrangements (**A**), in groups (**B**), or were scattered. Staining for macrophage marker (CD68 (KP1) (**C**) allowed for performing cell counts. There was a significant reduction in the number of CD-68 staining macrophages post-surgery (n = 14, 7.3 \pm 1.1 vs. 4.1 \pm 0.7, *p < 0.01) (**D**). Scale bar = 20 μ m.

Weight-loss surgery restores PVAT anticontractile function. In severely obese patients before surgery and weight-loss, PVAT did not significantly alter the norepinephrine-induced contractility of the small arteries in comparison with skeletonized segments of the same vessels (n = 15, p = 0.96) (Fig. 1A).

In samples taken from patients 6 months after surgery and weight loss, PVAT had a significant anticontractile effect on vessels when compared with vessels without PVAT (n = 15, p < 0.01) (Fig. 1B). The vessels with intact PVAT after surgery had a very similar response to cumulative doses of norepinephrine as the PVAT-intact vessels harvested from healthy lean volunteers (p = 0.52, n = 7) (Fig. 1B). The maximal degree of constriction is lower in PVAT-intact vessels in healthy and post-surgery samples as compared with skeletonized vessels, but the degree of constriction of skeletonized vessels to norepinephrine is similar in segments from pre- and post-surgery (Online Fig. 1).

Anticontractile property of PVAT after weight loss is abolished by adiponectin blockade. Samples taken from patients 6 months after surgery were incubated with blocking peptide for AdipoR1. This had no effect on segments without PVAT, but in vessels with PVAT there was a significant increase in vessel contractility (p < 0.0001, n = 7) (Fig. 2A). Free radical scavengers can rescue PVAT anticontractile function in obesity. In patients with severe obesity, the presence of perivascular adipose tissue had no effect on vessel contractility in comparison with vessel segments devoid of PVAT. Incubation of the vessels with the free radical scavengers superoxide dismutase and catalase resulted in a shift in the curve to resemble that of vessels with intact PVAT taken from healthy individuals (p < 0.001, n = 7) (Fig. 2B).

Increased NO bioavailability after surgery. There was a significant increase in NO in PVAT of severely obese patients 6 months after surgery (1.016 vs. 1.196 nmol/ μ l, p = 0.029, n = 5).

In post-surgery samples, de-endothelialized vessel segments were incubated with N5-[imino(methylamino) methyl]-L-ornithine, citrate. The incubation had no effect on vessels without PVAT, but in vessels with PVAT, there was a significant increase in contractility to cumulative doses of norepinephrine (p < 0.001, n = 4) (Fig. 2C).

Reduced PVAT inflammation, adipocyte size, and AdipoR1 after bariatric surgery. The perivascular adipose tissue was stained, to quantify the inflammation, for both CD68 staining macrophages (Fig. 3) and the cytokine TNF- α (Figs. 4A to 4C).

There was a significant reduction in the CD68 [KP1] staining macrophage numbers after surgery (7.3 \pm 1.1 vs. 4.1 \pm 0.7, p = 0.0067, n =14) (Figs. 4C and 4D), but the change in CD68 [PGM1] staining was not statistically changed. There was a significant reduction in the percentage of adipose tissue area staining for TNF- α (1.41 \pm 0.34 vs. 0.68 \pm 0.09, p < 0.05) (Figs. 4A to 4C) after surgery.



Immunostaining for tumor necrosis factor (TNF)- α showed that adipocytes themselves were positive for this cytokine, to a higher extent pre-surgery (**A**) than post-surgery (**B**). Note positive staining of microvessels in the pre-operative biopsy. There is a significant reduction in the percentage of adipose tissue area that stains for TNF- α pre-surgery versus post-surgery (1.41 ± 0.34 vs. 0.68 ± 0.09, *p < 0.05) (**C**). The average adipocyte area was 7,672 ± 369.6 μ m² pre-surgery and 3,955 ± 207.5 μ m² (*p < 0.0001) post-surgery (**D**). Scale bar = 20 μ m.

surgery and 3,955 \pm 207.5 μ m² (p < 0.0001) post-surgery (Fig. 4D). The change in average adipocyte area correlated with the change in BMI after surgery (R² = 0.356, p = 0.0243, n = 14) (Online Fig. 2).

There was a significant reduction in circulating markers of inflammation including hsCRP (p < 0.001), interleukin-6 (p = 0.013), monocyte chemoattractant protein-1 (283.1 vs. 258.3 pg/ml; p = 0.013), and the adhesion molecule E-selectin (12.7 vs. 7.47 ng/ml; p < 0.001). However, there was no significant change in circulating TNF- α levels (p = 0.519) (Table 1).

Discussion

The present study investigated the effect of bariatric surgery on the vasodilatory properties of PVAT. We designed the study after our earlier observation that, in severely obese patients, although there is an accumulation of adipose tissue, there is also a paradoxical inhibition of the beneficial PVAT vasodilation (7). There are now 3 main findings presented here. First, bariatric surgery reverses the obesity-induced damage to PVAT anticontractile function. Second, the functional recovery of the PVAT is independent of the endothelium. Third, bariatric surgery restores the function of PVAT by reducing adipose inflammation and increasing local adiponectin and NO bioavailability (Figure 5). The observations advance our understanding of the mechanisms by which obesity disrupts the vasodilating function of adipose tissue and how this pathology might be reversed by clinical intervention.

The cardiovascular complications of obesity are undoubtedly among the most pressing issues facing clinicians today (19). To date, the most common recommended intervention has been the encouragement of dietary

modification. Effective and sustained dieting does lead, without doubt, to significant weight loss, but unfortunately the weight loss is not sustained (20). Furthermore, the impact of dietary modification on cardiovascular outcomes is unclear (13,21). For patients with severe obesity, the most effective method of achieving significant and sustained weight reduction is by weight-reducing surgery (14). In addition, the weight loss after surgery also significantly correlates with improvements to blood pressure (22,23), left ventricular mass (24), exercise capacity (25), and glucose metabolism (26). Changes to weight in patients are associated with profound modulation of the cytokine and inflammatory profiles of adipose tissue. In this regard, it is now established that, in obese patients, adipose tissue undergoes dual processes of hypoxia and inflammation, leading to a reduction in the secretion of adipocytokines such as adiponectin. The hypoxia is thought to be due to adipocyte hypertrophy (27) twinned with reductions to capillary density and angiogenic capacity (28). This results in up-regulation of hypoxia inducible factor-1 and inflammatory differentiation of macrophages, which subsequently secrete TNF- α (29-31). Differentiated macrophages in obesity also release superoxide anions, which further contribute to local vascular dysfunction by diminishing availability of NO (32).

The functional damage we observe in PVAT and its recovery after bariatric surgery are entirely consistent with this hypoxia/inflammation in fat hypothesis. Thus, in our previous studies we observed adipocyte hypertrophy and increased TNF staining in adipose tissue associated with loss of PVAT vasodilatory function. We now show that, after bariatric surgery, there is restoration of PVAT vasodilatory capacity to a degree that is similar to that observed in healthy non-obese participants. It is interesting to note that this

Table 1 Patient De	Jennographic Data and Biomarker Prome				
	Healthy	Pre-Surgery	Post-Surgery	p Value	
Age, yrs	$\textbf{51.0} \pm \textbf{11.5}$	$\textbf{53.2} \pm \textbf{6.73}$	$\textbf{53.2} \pm \textbf{6.73}$		
Waist, cm	$\textbf{92.0} \pm \textbf{11.6}$	$\textbf{140.90} \pm \textbf{14.85}$	$\textbf{115.5} \pm \textbf{13.3}$	<0.0001	
BMI, kg/m ²	$\textbf{25.40} \pm \textbf{2.68}$	$\textbf{51.58} \pm \textbf{7.47}$	$\textbf{37.960} \pm \textbf{7.175}$	<0.0001	
Systolic BP, mm Hg	$\textbf{135.0} \pm \textbf{17.2}$	$\textbf{146.1} \pm \textbf{28.8}$	$\textbf{132.70} \pm \textbf{22.15}$	0.0025	
hsCRP, mg/l	$\textbf{1.33} \pm \textbf{1.09}$	$\textbf{6.321} \pm \textbf{2.444}$	$\textbf{2.747} \pm \textbf{2.282}$	0.0027	
IL-6, pg/ml	$\textbf{2.070} \pm \textbf{0.811}$	$\textbf{2.830} \pm \textbf{1.755}$	$\textbf{0.644} \pm \textbf{0.774}$	0.0052	
TNF-α, pg∕ml	$\textbf{4.83} \pm \textbf{1.75}$	$\textbf{35.14} \pm \textbf{40.30}$	$\textbf{27.14} \pm \textbf{37.30}$	0.12	
Adiponectin, mg/l	$\textbf{2.190} \pm \textbf{0.633}$	$\textbf{1.845} \pm \textbf{0.699}$	$\textbf{2.410} \pm \textbf{0.828}$	0.011	
Leptin, ng/ml	$\textbf{9.67} \pm \textbf{7.76}$	$\textbf{59.9} \pm \textbf{17.7}$	$\textbf{24.91} \pm \textbf{10.42}$	0.0014	
Resistin, ng/ml	$\textbf{9.26} \pm \textbf{3.05}$	$\textbf{15.780} \pm \textbf{6.823}$	$\textbf{13.550} \pm \textbf{4.605}$	0.0796	
E-selectin, ng/ml	$\textbf{131.0} \pm \textbf{5.06}$	$\textbf{12.530} \pm \textbf{5.581}$	$\textbf{8.753} \pm \textbf{3.374}$	0.0033	
Insulin, mU/I	$\textbf{7.18} \pm \textbf{4.11}$	$\textbf{27.83} \pm \textbf{16.17}$	$\textbf{14.83} \pm \textbf{10.12}$	0.004	
Glucose, mmol/I	$\textbf{5.080} \pm \textbf{0.377}$	$\textbf{6.825} \pm \textbf{3.585}$	$\textbf{5.858} \pm \textbf{2.355}$	0.0312	
HbA1C, %	$\textbf{5.740} \pm \textbf{0.689}$	$\textbf{7.0} \pm \textbf{2.119}$	$\textbf{6.330} \pm \textbf{1.603}$	0.0071	
НОМА-В, %	$\textbf{90.2} \pm \textbf{26.9}$	$\textbf{243.0} \pm \textbf{175.7}$	$\textbf{169.9} \pm \textbf{143.7}$	0.01	
HOMA-IR, %	$\textbf{0.933} \pm \textbf{0.557}$	$\textbf{7.838} \pm \textbf{4.213}$	$\textbf{3.860} \pm \textbf{2.683}$	0.0023	

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Values are mean \pm SD. The p values compare the matched baseline and post-surgery values and were determined with paired t tests. BP = blood pressure; BMI = body mass index; HbA1C = hemoglobin A1c; H0MA-B = homeostatic model assessment of beta-cell function; H0MA-IR = homeostasis model assessment of insulin resistance; hsCRP = high-sensitivity C-reactive protein; IL = interleukin; TNF = tumor necrosis factor.



functional restoration of the PVAT occurs even though the patients are still severely obese. The improvement in PVAT function after surgery is associated with smaller adipocytes (in the context of a dramatic weight loss), a reduction in PVAT TNF, and increased local adipose tissue NO bioavailability. Our findings are consistent with other studies that have shown significant reductions in macrophage numbers as well as monocyte chemoattractant protein-1 and hypoxia inducible factor-1 α in the stromal vascular fraction of white adipose tissue after bariatric surgery (33). Also, importantly, we show that the functional vasodilatory improvement in the subcutaneous fat is due to restitution of PVAT-derived adiponectin and NO bioavailability.

The functional improvement of PVAT after bariatric surgery was independent of the vascular endothelium, because all arteries in this study were denuded of endothelium. Also

notably, there were no differences in constriction of arteries devoid of fat to norepinephrine before and after surgery. Taken together, these observations indicate that the improvement in adipose-vascular coupling after surgery was due to improvements in the vasodilating capacity of the PVAT rather than changes to isolated arterial contractility or endothelial function. Study limitations. There are a number of limitations to this study that need to be considered in parallel with the findings. We did not monitor the level of exercise performed by the participants at baseline or after surgery, with respect to our patient selection. At 6 months after surgery, the patients had lost a significant amount of weight and were presumably more mobile. As such, we acknowledge that the improvement in PVAT function might be as a consequence of increased levels of mobility. Second, it would have been preferential to recruit participants who were losing weight as part of a caloriecontrolled diet to compare changes to PVAT function due to this intervention. This was not possible within the limitations of our faculty, but we hope to perform this study in the future. Similarly, a drawback of this study is that we were not able to perform a 6-month follow-up biopsy on severely obese patients who received no intervention. However, we assume that there would have been no change in PVAT function over this time. We have studied subcutaneous small arteries under the assumption that all small arteries and the surrounding PVAT share similar properties. A separate consideration is that we have detected changes only to subcutaneous gluteal PVAT after surgery. We did not study the function of PVAT from other anatomical sites (i.e., mesenteric or skeletal). As such, we acknowledge that changes to subcutaneous vascular function might not be reflected in other vascular territories, but previous studies have shown similar remodeling profiles of human cerebral and mesenteric arteries to those seen in arteries taken from gluteal subcutaneous biopsies. Finally, it should be noted that norepinephrine was the only contractile agent used. We acknowledge that it would have been preferable to study a variety of vasoconstrictors. However, it should be noted that studies from other groups have shown that PVAT antagonizes contraction to U46619 (34), angiotensin II (35,36), phenylephrine (35,37), and serotonin (35,36).

Conclusions

Despite these limitations, however, we believe that we present convincing evidence that PVAT-induced vasodilation of small arteries can be fully restored after bariatric surgery. Furthermore, this restoration of function is due to a reduction in inflammation within the adipose tissue, which leads to increased adiponectin secretion. The data offer an insight into 1 mechanism by which bariatric surgery improves vascular function. We anticipate that further work will harness this observation to improve therapeutic options for the severely obese patient.

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REFERENCES

- 1. WHO. Obesity and Overweight (Factsheet no 311). Available at: http://www.who.int/mediacentre/factsheets/fs311/en/index.html. Accessed August 11, 2009.
- WHO. Population-Based Prevention Strategies for Childhood Obesity. Report of the World Health Organization Forum and Technical Meeting 2009. Available at: http://www.who.int/dietphysicalactivity/ childhood/child-obesity-eng.pdf. Accessed June 2013.
- Poirier P, Giles TD, Bray GA, et al. Obesity and cardiovascular disease: pathophysiology, evaluation, and effect of weight loss: an update of the 1997 American Heart Association Scientific Statement on Obesity and Heart Disease from the Obesity Committee of the Council on Nutrition, Physical Activity, and Metabolism. Circulation 2006;113:898–918.
- Jonk AM, Houben AJ, de Jongh RT, Serne EH, Schaper NC, Stehouwer CD. Microvascular dysfunction in obesity: a potential mechanism in the pathogenesis of obesity-associated insulin resistance and hypertension. Physiology (Bethesda) 2007;22:252–60.
- Kerr SM, Livingstone MB, McCrorie TA, Wallace JM. Endothelial dysfunction associated with obesity and the effect of weight loss interventions. Proc Nutr Soc 2011;70:418–25.
- 6. Williams IL, Wheatcroft SB, Shah AM, Kearney MT. Obesity, atherosclerosis and the vascular endothelium: mechanisms of reduced nitric oxide bioavailability in obese humans. Int J Obes Relat Metab Disord 2002;26:754–64.
- Greenstein AS, Khavandi K, Withers SB, et al. Local inflammation and hypoxia abolish the protective anticontractile properties of perivascular fat in obese patients. Circulation 2009;119:1661–70.
- Fang L, Zhao J, Chen Y, et al. Hydrogen sulfide derived from periadventitial adipose tissue is a vasodilator. J Hypertens 2009;27:2174–85.
- Lee YC, Chang HH, Chiang CL, et al. Role of perivascular adipose tissue-derived methyl palmitate in vascular tone regulation and pathogenesis of hypertension. Circulation 2011;124:1160–71.
- Withers BS, Agabiti-Rosei C, Livingstone DM, et al. Macrophage activation is responsible for loss of anticontractile function in inflamed perivascular fat. Arterioscler Thromb Vasc Biol 2011;31:908–13.
- Poirier P, Cornier MA, Mazzone T, et al. Bariatric surgery and cardiovascular risk factors: a scientific statement from the American Heart Association. Circulation 2012;123:1683–701.
- Butner KL, Nickols-Richardson SM, Clark SF, Ramp WK, Herbert WG. A review of weight loss following Roux-en-Y gastric bypass vs restrictive bariatric surgery: impact on adiponectin and insulin. Obes Surg 20:559–68.
- Sjostrom L, Peltonen M, Jacobson P, et al. Bariatric surgery and longterm cardiovascular events. JAMA 2012;307:56–65.
- Heneghan HM, Meron-Eldar S, Brethauer SA, Schauer PR, Young JB. Effect of bariatric surgery on cardiovascular risk profile. Am J Cardiol 2011;108:1499–507.
- Mulvany MJ, Halpern W. Contractile properties of small arterial resistance vessels in spontaneously hypertensive and normotensive rats. Circ Res 1977;41:19–26.
- Lu C, Zhao AX, Gao YJ, Lee RM. Modulation of vein function by perivascular adipose tissue. Eur J Pharmacol 2011;657:111–6.
- Dubrovska G, Verlohren S, Luft FC, Gollasch M. Mechanisms of ADRF release from rat aortic adventitial adipose tissue. Am J Physiol Heart Circ Physiol 2004;286:H1107–13.
- Verlohren S, Dubrovska G, Tsang SY, et al. Visceral periadventitial adipose tissue regulates arterial tone of mesenteric arteries. Hypertension 2004;44:271–6.

- Whitlock G, Lewington S, Sherliker P, et al. Body-mass index and cause-specific mortality in 900 000 adults: collaborative analyses of 57 prospective studies. Lancet 2009;373:1083–96.
- Mann T, Tomiyama AJ, Westling E, Lew AM, Samuels B, Chatman J. Medicare's search for effective obesity treatments: diets are not the answer. Am Psychol 2007;62:220–33.
- Hofso D, Nordstrand N, Johnson LK, et al. Obesity-related cardiovascular risk factors after weight loss: a clinical trial comparing gastric bypass surgery and intensive lifestyle intervention. Eur J Endocrinol 2010;163:735–45.
- 22. Frezza EE, Wei C, Wachtel MS. Is surgery the next answer to treat obesity-related hypertension? J Clin Hypertens (Greenwich) 2009;11: 284–8.
- Aghamohammadzadeh R, Heagerty AM. Obesity-related hypertension: epidemiology, pathophysiology, treatments, and the contribution of perivascular adipose tissue. Ann Med 2012;44 Suppl 1:S74–84.
- 24. Cavarretta E, Casella G, Cali B, et al. Cardiac remodeling in obese patients after laparoscopic sleeve gastrectomy. World J Surg 2013;37: 565–72.
- Wilms B, Ernst B, Thurnheer M, Weisser B, Schultes B. Differential changes in exercise performance after massive weight loss induced by bariatric surgery. Obes Surg 2013;23:365–71.
- Ballantyne ĞH, Wasielewski A, Saunders JK. The surgical treatment of type II diabetes mellitus: changes in HOMA Insulin resistance in the first year following laparoscopic Roux-en-Y gastric bypass (LRYGB) and laparoscopic adjustable gastric banding (LAGB). Obes Surg 2009; 19:1297–303.
- Trayhurn P, Wang B, Wood IS. Hypoxia in adipose tissue: a basis for the dysregulation of tissue function in obesity? Br J Nutr 2008;100: 227–35.
- Pasarica M, Sereda OR, Redman LM, et al. Reduced adipose tissue oxygenation in human obesity: evidence for rarefaction, macrophage chemotaxis, and inflammation without an angiogenic response. Diabetes 2009;58:718–25.
- Halberg N, Khan T, Trujillo ME, et al. Hypoxia-inducible factor 1alpha induces fibrosis and insulin resistance in white adipose tissue. Mol Cell Biol 2009;29:4467–83.
- Stuart Wood I, de Heredia FP, Wang B, Trayhurn P. Cellular hypoxia and adipose tissue dysfunction in obesity. Proc Nutr Soc 2009;68: 370–7.
- Yokota T, Oritani K, Takahashi I, et al. Adiponectin, a new member of the family of soluble defense collagens, negatively regulates the growth of myelomonocytic progenitors and the functions of macrophages. Blood 2000;96:1723–32.
- 32. Aghamohammadzadeh R, Withers S, Lynch F, Greenstein A, Malik R, Heagerty A. Perivascular adipose tissue from human systemic and coronary vessels: the emergence of a new pharmacotherapeutic target. Br J Pharmacol 2012;165:670–82.
- Cancello R, Henegar C, Viguerie N, et al. Reduction of macrophage infiltration and chemoattractant gene expression changes in white adipose tissue of morbidly obese subjects after surgery-induced weight loss. Diabetes 2005;54:2277–86.
- Gao YJ, Zeng ZH, Teoh K, et al. Perivascular adipose tissue modulates vascular function in the human internal thoracic artery. J Thorac Cardiovasc Surg 2005;130:1130–6.
- Löhn M, Dubrovska G, Lauterbach B, Luft FC, Gollasch M, Sharma AM. Periadventitial fat releases a vascular relaxing factor. FASEB J 2002;16:1057–63.
- Malinowski M, Deja MA, Golba KS, Roleder T, Biernat J, Wos S. Perivascular tissue of internal thoracic artery releases potent nitric oxide and prostacyclin-independent anticontractile factor. Eur J Cardiothorac Surg 2008;33:225–31.
- Gao YJ, Holloway AC, Zeng ZH, et al. Prenatal exposure to nicotine causes postnatal obesity and altered perivascular adipose tissue function. Obes Res 2005;13:687–92.

Key Words: bariatric • cardiovascular • obesity.

> APPENDIX

For supplementary figures, please see the online version of this article.