Effect of inhibition of prostaglandin E2 production on pancreatic infection in experimental acute pancreatitis

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Abstract
Objective. Acute pancreatitis is one of the important causes of systemic inflammatory response syndrome (SIRS). SIRS results in gut barrier dysfunction that allows bacterial translocation and pancreatic infection to occur. Indomethacin has been used to reduce inflammatory process and bacterial translocation in experimental models. The purpose of this study was to determine the effect of inhibition of prostaglandin E2 (PGE2) production on pancreatic infection.

Materials and methods. An experimental model of severe acute pancreatitis (AP) was utilized. The animals were divided into three groups: sham (surgical procedure without AP induction); pancreatitis (AP induction); and indomethacin (AP induction plus administration of 3 mg/kg of indomethacin). Serum levels of interleukin (IL)-6 and IL-10, PGE2, and tumor necrosis factor (TNF)-α were measured 2 h after the induction of AP. We analyzed the occurrence of pancreatic infection with bacterial cultures performed 24 h after the induction of AP. The occurrence of pancreatic infection (considered positive when the CFU/g was >10⁵), pancreatic histologic analysis, and mortality rate were studied.

Results. In spite of the reduction of IL-6, IL-10, and PGE2 levels in the indomethacin group, TNF-α level, bacterial translocation, and pancreatic infection were not influenced by administration of indomethacin. The inhibition of PGE2 production did not reduce pancreatic infection, histologic score, or mortality rate. Conclusion. The inhibition of PGE2 production was not able to reduce the occurrence of pancreatic infection and does not have any beneficial effect in this experimental model. Further investigations will be necessary to discover a specific inhibitor that would make it possible to develop an anti-inflammatory therapy.

Key Words: experimental acute pancreatitis, pancreatic infection, indomethacin, bacterial translocation, prostaglandin E2
escape of bacteria from the gut [7]. Additionally, it has been speculated that release of excessive amounts of inflammatory mediators leads to failure of the intestinal barrier [8]. Therefore, the occurrence of bacterial translocation in AP can be considered an indirect marker of systemic inflammatory response level.

Pancreatic infection is the most serious complication of severe AP, with a mortality rate up to 80%. The pathogenesis of pancreatic infection in AP remains poorly understood. Increased gut bacterial translocation in AP has been demonstrated in a number of previous studies, where bacterial translocation has been correlated with pancreatic infection [9,10]. Our group previously demonstrated the occurrence of bacterial translocation in experimental AP [11,12].

PEG2 and prostaglandin-derived mediators play an important role in mediating the systemic inflammatory response. Cyclooxygenases (COX) are the key enzymes of prostaglandin synthesis. Inhibition of PEG2 production could reduce the systemic inflammatory response and therefore have some benefits in AP [13,14].

Indomethacin is an unselective COX inhibitor capable of reducing PEG2 production. In experimental models of shock and burn injury, administration of PEG2 synthesis blocker prevented the occurrence of bacterial translocation. It has been reported that high levels of PEG2 are responsible for suppression of splenic T-cell proliferation during sepsis resulting in bacterial translocation [15]. However, the role of reduction of PEG2 production in cytokine production and pancreatic infection has not been evaluated in experimental AP.

To elucidate the role of PEG2 in severe AP we sought to analyze the effects of inhibition of PEG2 production on the local and systemic response in experimental severe AP.

**Materials and methods**

**Animals**

One hundred twenty male Wistar rats, weighing 225–250 g, were housed in individual cages. The temperature was kept at 20–22°C, a 12 h light-dark cycle was maintained, and all rats were fed with a standard rat chow and water ad libitum during the entire protocol; any fast period was observed. The experimental protocol was approved by the University of São Paulo Ethics Commission. All animals received care in accordance with ‘Guide for the Care and Use of Laboratory Animals.’

**Experimental design**

Animals were divided into three experimental groups: group I (sham group) – 40 rats submitted to surgical procedure without induction of AP plus intraperito-

eal administration of 0.5 ml of saline; group II (pancreatitis group) – 40 rats submitted to induction of AP by sodium taurocholate injection into the pancreatic duct plus intraperitoneal administration of 0.5 ml of saline; and group III (indomethacin group) – 40 rats submitted to induction of AP by sodium taurocholate injection into the pancreatic duct plus intraperitoneal administration of 3 mg/kg of indomethacin.

**Induction of acute pancreatitis**

The rats were operated under aseptic conditions, using ketamine anesthesia 0.2 ml/100 g (Ketalar®). The pancreas was exteriorized through a midline abdominal incision, the proximal bile duct was clamped at the liver hilum level, and the distal bile duct was cannulated using a 19G polyethylene catheter through the duodenal wall. AP was induced by intraductal retrograde injection of 0.5 ml of 2.5% sodium taurocholate (Sigma-Chemical Company™, St Louis, MO, USA), with pressure control. Injection flow was 0.2 ml/min. In the sham group, sodium taurocholate injection was omitted, but the surgical procedure was identical to the other groups, including bile duct cannulation.

**Administration of indomethacin and saline**

Intraperitoneal administration of 3 mg/kg of indomethacin (Sigma-Chemical Company™) was performed after the induction of AP and the same dose was repeated each 12 h.

Intraperitoneal administration of 0.5 ml of saline was performed after the induction of AP and was repeated every 12 h in groups I and II.

The administration of indomethacin and saline was carried out in aseptic conditions.

**Levels of cytokines**

Blood samples were collected 2 h after the induction of AP by needle aspiration. Plasma levels of IL-6, IL-10, PGE2, and TNF-α were determined by ELISA using kits provided by the manufacturing company (Biosource International Cytoscreen™) and expressed in pg/ml.

After induction of AP a systemic inflammatory response is initiated and inflammatory mediators can be measured and predict the severity of inflammation. Inflammatory mediators have a very short half-life and the high levels are observed in the first hours of systemic inflammatory response. In a previous publication the authors showed that 2 h is an adequate time to measure the cytokine levels in this experimental model [16].
Cultures and measurement of pancreatic infection

At 24 h after induction of AP, 20 rats from each group were submitted to a midline laparotomy incision under anesthesia and tissue samples were harvested from the pancreas, mesenteric lymph nodes, and liver. All samples were weighed and homogenized and aerobic cultures were made using blood agar plates and MacConkey agar plates and expressed in colony forming units (CFU) per gram. Blood and free peritoneal fluid were sampled using sterile syringes and cultured using an automated culture system (BACTEC PEDS Plus); all positive culture were inoculated on blood agar plates and MacConkey agar plates for bacterial identification. The presence of pancreatic infection was accepted when CFU/g was $>10^5$.

Histological analysis

Fragments of pancreas were harvested and fixed in 10% formaldehyde solution, embedded in paraffin, sectioned, and stained with hematoxylin and eosin (H&E) for light microscopy. The inflammatory changes in the pancreas were analyzed and classified by a previous described score [17].

Mortality study

Twenty animals from each group were kept alive for 7 days to determine the mortality rate. These animals were maintained in individual cages, received regular rat chow, and received saline solution or indomethacin according to the experimental protocol.

Statistical analysis

The data were analyzed using Graphpad Prism® version 2.1. Different groups were compared using Fisher’s exact test, ANOVA, Tukey, Mann-Whitney, Kruskal-Wallis, and Dunn for the appropriate categorical data. Statistical analyses of the mortality rate were performed using Kaplan–Meier curve and log-rank test. Standard error was used and differences were considered statistically significant when $p$ values were $<0.05$.

Results

All experiments were successfully concluded.

Cytokine levels

The levels of IL-6, IL-10, PGE2, and TNF-α were undetectable in the sham group. A significantly higher level of cytokines was observed in the pancreatitis group. Significant decreases in levels of IL-6, IL-10, and PGE$_2$ were observed after indomethacin administration when compared with the pancreatitis group ($p <0.05$). The administration of indomethacin was not able to reduce the level of TNF-α (Table I).

Cultures

Pancreas. Positive cultures were not observed in the sham group. A significantly increased rate of positive bacterial cultures was obtained from pancreas cultures in the pancreatitis and indomethacin groups. There were no significant differences in the incidence of positive cultures between these groups (Table II). A high incidence of Escherichia coli was observed in pancreas cultures in the pancreatitis and indomethacin groups.

Mesenteric lymph nodes. The pancreatitis and indomethacin groups had a significantly increased rate of positive bacterial cultures when compared with the sham group. Positive culture was not observed in the sham group. There were no significant differences in positive cultures and bacterial populations between the pancreatitis and indomethacin groups (Table II).

Liver. A higher incidence of positive cultures was observed in the pancreatitis and indomethacin groups, without significant differences in both groups. Positive culture was not observed in the sham group (Table II).

Blood and peritoneal cavity. A significantly increased rate of positive bacterial cultures was obtained from blood and peritoneal cultures in the pancreatitis and indomethacin groups in contrast with the sham group. There was no significant difference between the pancreatitis and indomethacin groups (Table II).

Pancreatic infection

Infection of pancreatic tissue was not observed in the sham group. A high incidence of pancreatic infection was observed in the pancreatitis group (60%) and was also observed after indomethacin administration (70%) (Figure 1).

<table>
<thead>
<tr>
<th>Group</th>
<th>IL-6</th>
<th>IL-10</th>
<th>PGE$_2$</th>
<th>TNF-α</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sham</td>
<td>0</td>
<td>0</td>
<td>151±115</td>
<td>0</td>
</tr>
<tr>
<td>Pancreatitis</td>
<td>184±19</td>
<td>201±34</td>
<td>6728±1360</td>
<td>175±14</td>
</tr>
<tr>
<td>Indomethacin</td>
<td>74±2*</td>
<td>47±13*</td>
<td>306±62*</td>
<td>155±20</td>
</tr>
</tbody>
</table>

Mean value of plasma levels of IL-6, IL-10, PGE$_2$, and TNF-α expressed in pg/ml. $p <0.05$, indomethacin group vs pancreatitis group, ANOVA.
Histologic analysis

Pancreatic edema, inflammatory infiltration, pancreatic necrosis, pancreatic hemorrhage, and extrapancreatic fat necrosis were not reduced after indomethacin administration ($p > 0.05$) (Figure 2).

Mortality study

Mortality was not observed in the sham group. Seven rats died after induction of AP in the pancreatitis group (35% mortality rate) and a significant reduction of mortality rate was not observed in the indomethacin group (30% mortality rate) (Figure 3).

Discussion

PGE2 has an inflammatory property and its role in AP is unclear. Several investigations have shown that high levels of PGE2 could be responsible for triggering factors of inflammation and suggested that PGE2 inhibition could have beneficial effects in AP [14]. Furthermore, experimental studies in burn models described the effects of PGE2 in bacterial translocation, proposing a suppression of intestinal T-cell proliferation allowing bacterial translocation caused by high levels of PGE2 [15,18].

All major inflammatory mediators have been studied in recent years and researchers are still looking for the elucidation of the key mediators in severe AP associated with the discovery of specific inhibitors that would make it possible to develop a clinically effective anti-inflammatory therapy [19,20]. Trying to answer this question and clarify the role of PGE2 in AP, we used indomethacin as a PGE2 synthesis blocker and correlated with other inflammatory mediators, such as IL-6, IL-10, and TNF-$\alpha$.

In the present study, the administration of indomethacin successfully inhibited PGE2 production and also decreased levels of IL-6 and IL-10. However, it was not able to reduce TNF-$\alpha$ levels. After administration of indomethacin the levels of TNF-$\alpha$ were kept high in the indomethacin group, demonstrating that indomethacin was incapable of interfering with TNF-$\alpha$ production. Reduction of PGE2 to very low levels in this experimental protocol did not reduce the severity of AP, the incidence of bacterial translocation and pancreatic infection, or the mortality rate.

These data suggest that AP is a very complex inflammatory process involving multiple mediators and that PGE2 does not play a fundamental role in this disease. With respect to the decreased serum levels of IL-6 and IL-10, these data also suggest that isolated reduction of IL-6 and IL-10 was not able to influence the systemic inflammatory response in AP and indicate that TNF-$\alpha$ might play a very important role in the inflammatory process of severe AP and might be a better marker of severity than other inflammatory mediators [21–24].

In septic burn models, high levels of PGE2 had been correlated with macrophage dysfunction and the administration of a COX inhibitor improved

### Table II. Occurrence of bacterial translocation 24 h after taurocholate-induced pancreatitis.

<table>
<thead>
<tr>
<th>Group</th>
<th>Pancreas</th>
<th>Mesenteric lymph nodes</th>
<th>Liver</th>
<th>Blood</th>
<th>Peritoneal cavity</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sham</td>
<td>0/10*</td>
<td>0/10*</td>
<td>0/10*</td>
<td>0/10*</td>
<td>0/10*</td>
</tr>
<tr>
<td>Pancreatitis</td>
<td>8/10</td>
<td>8/10</td>
<td>4/10</td>
<td>7/10</td>
<td>8/10</td>
</tr>
<tr>
<td>Indomethacin</td>
<td>7/10</td>
<td>6/10</td>
<td>5/10</td>
<td>8/10</td>
<td>8/10</td>
</tr>
</tbody>
</table>

The occurrence of bacterial translocation was considered positive in all positive cultures. *$p < 0.05$, sham group vs pancreatitis and indomethacin groups, Fischer test.*

Figure 1. Percentage of pancreatic infections in rats with taurocholate-induced pancreatitis 24 h after induction of pancreatitis. *$p < 0.05$, sham group vs pancreatitis and indomethacin groups, Fischer test.*

Figure 2. Histologic grading of lesions in histological analysis of pancreatic tissue of rats with taurocholate-induced acute pancreatitis. *$p < 0.05$, sham group vs pancreatitis and indomethacin groups, Kruskal-Wallis test.*
immunity and reduced bacterial translocation [15]. Our study did not show any reduction in bacterial translocation and pancreatic infection after indomethacin administration and marked PGE2 inhibition, suggesting that bacterial translocation in AP may have a different and more complex mechanism than those observed in septic-burn models.

The pancreatic infection was identified as the main determinant of morbidity and mortality in severe AP [25,26]. In necrotic pancreatic tissue, bacteria find ideal conditions to replicate and form a secondary infectious focus, which could fuel SIRS and increase the mortality rate [27]. The present study indicates that pancreatic infection is related to bacterial translocation when associated with AP.

Bacterial translocation is related to mucosal damage caused by hemodynamic and microvascular dysfunction, intravascular thrombosis, vasodilatation, hypotension, and endothelial damage triggered by TNF-α [28]. Our present data show high and similar levels of TNF-α in both groups that underwent induction of AP. Based on these data, it is possible to speculate that the intensity of the inflammatory process is mainly related to TNF-α level and this can explain the high occurrence of pancreatic infection and mortality rate in this experimental model [20,22].

It has been reported that PGE2 increases vaso-permeability and sequestration of inflammatory cells [29,30]. However, in this model the reduction of PGE2 to very low levels was not able to change the local inflammatory process based on pancreatic histologic analysis. The multiplicity of factors involved in the pathophysiology of AP could explain this finding. Furthermore, the mortality rate in this experimental rat model did not have any beneficial advantage in rats treated with indomethacin. Indeed, it is possible to hypothesize that PGE2 does not play a central role in the physiopathology of AP and PGE2 probably is not an important target for anti-inflammatory therapy in severe AP.

In conclusion, the inhibition of PGE2 production, even when associated with reduction of IL-6 and IL-10 levels, did not have any beneficial effect in this experimental model of AP. Further investigations will be necessary to discover specific inhibitors that would make it possible to develop clinically effective anti-inflammatory therapy in severe AP.

References


