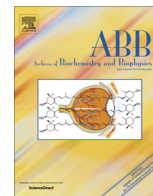




Contents lists available at ScienceDirect

## Archives of Biochemistry and Biophysics

journal homepage: [www.elsevier.com/locate/yabbi](http://www.elsevier.com/locate/yabbi)

# A randomized placebo-controlled study on the effects of lutein and zeaxanthin on visual processing speed in young healthy subjects

Emily R. Bovier<sup>a,b</sup>, Billy R. Hammond<sup>b,\*</sup><sup>a</sup> Department of Psychology, SUNY Oswego, Oswego, NY 13126, USA<sup>b</sup> Brain and Behavioral Sciences, The University of Georgia, Athens, GA 30602-3013, USA

## ARTICLE INFO

## Article history:

Received 11 September 2014

and in revised form 14 November 2014

Available online 4 December 2014

## Keywords:

Lutein

Zeaxanthin

CFF

Visual processing speed

## ABSTRACT

Speed of processing is a particularly important characteristic of the visual system. Often a behavioral reaction to a visual stimulus must be faster than the conscious perception of that stimulus, as is the case with many sports (e.g., baseball). Visual psychophysics provides a relatively simple and precise means of measuring visual processing speed called the temporal contrast sensitivity function (tCSF). Past study has shown that macular pigment (a collection of xanthophylls, lutein (L), meso-zeaxanthin (MZ) and zeaxanthin (Z), found in the retina) optical density (MPOD) is positively correlated with the tCSF. In this study, we found similar correlations when testing 102 young healthy subjects. As a follow-up, we randomized 69 subjects to receive a placebo ( $n = 15$ ) or one of two L and Z supplements ( $n = 54$ ). MPOD and tCSF were measured psychophysically at baseline and 4 months. Neither MPOD nor tCSF changed for the placebo condition, but both improved significantly as a result of supplementation. These results show that an intervention with L and Z can increase processing speed even in young healthy subjects.

© 2014 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/3.0/>).

## Introduction

One likely manifestation of biological thrift is that a single, typically widely available, molecule can often play many and diverse roles throughout nature. Lutein (L)<sup>1</sup> and zeaxanthin (Z), for instance, play a critical role in plant photosynthesis [4] and the embryonic development of chicks (giving the yellow to egg yolk; [18]). L and Z are antiatherogenic [6] but also help prevent photo-oxidative degradation of the skin [24]. L and Z are potent lipid-based antioxidants and anti-inflammatories [31] but also serve as optical filters within the macula of the eye [13]. They are ornamental [10] and yet found within human brain information processing areas such as the hippocampus [30]. Their diversity throughout nature is reflected by an equally impressive diversity within our biology.

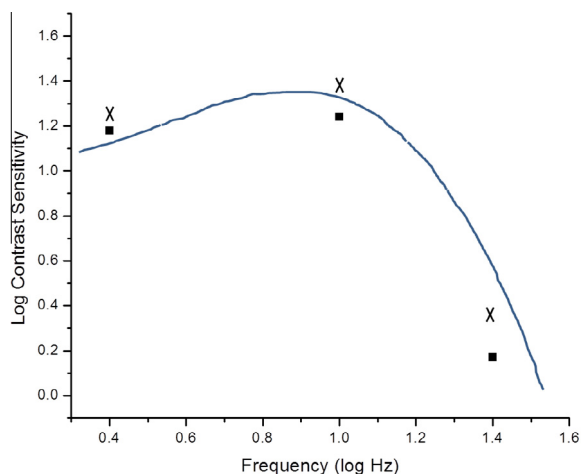
The behavioral effects associated with L and Z seem no less encompassing. Significant relations have been reported between

macular pigment optical density (MPOD; L and Z and meso-Z measured in the retina) and a large number of visual measures including glare disability and discomfort, photostress recovery, and chromatic contrast [13]. Measures of L and Z within the retina appear to be strongly linked to measures of L and Z in brain tissue [29] and MPOD has also been linked to measures that are mediated by brain such as cognition [15,9,23], auditory thresholds [33], balance time, reaction time [22], and temporal vision [11,21,2].

Taken together, L and Z seem important to biology, in general, and humans are no exception. In many cases, the basis for their functional effects has been well characterized. For example, in the eye, many effects are due simply to selective filtering. How (and really if) they influence brain function, however, is less clear. One possibility is simply protection from the accumulated effects of oxidative and inflammatory stress. Data linking reduced MPOD to dementia [19] and cognitive impairment [23] is consistent with that possibility. Another possibility, more relevant to younger individuals and palliative approaches, is a direct improvement by some type of local interaction with neural cells (the so-called neural efficiency hypothesis; [11,36,21]). Such interactions (as opposed to simply enhanced protection) would imply that supplementation over a relatively short time period (yet long enough to increase

\* Corresponding author.

E-mail address: [bhammond@uga.edu](mailto:bhammond@uga.edu) (B.R. Hammond).<sup>1</sup> Abbreviations used: tCSF, temporal contrast sensitivity function; L, lutein; MZ, meso-zeaxanthin; Z, zeaxanthin; MPOD, macular pigment optical density; LED, light-emitting diode.



**Fig. 1.** The average baseline tCSF values for subjects in the placebo (Xs) and treatment conditions (squares). The shape of the template curve depicted by the solid line was derived from Wooten et al. [35]. The similarity in shape suggests our temporal measures were a valid estimate of the general temporal contrast sensitivity function.

MPOD) in young healthy subjects would yield behavioral improvements in tasks that are generally mediated by the central nervous system.<sup>2</sup> To test this idea, we measured MPOD and temporal vision (specifically, the temporal contrast sensitivity function, tCSF) in a group of young healthy subjects.

We chose the tCSF because the stimuli can be designed to obviate individual differences mediated by optical effects (e.g., influence of light absorption by retinal L and Z is eliminated by using wavelengths not absorbed by MP) and because the retina is known to follow temporally varying stimuli much faster than brain, hence, high-frequency thresholds are determined by the rate limiter which, in this case, appears to be visual cortex [32]. An example of the tCSF, with the specific points we assessed, is shown in Fig. 1. Renzi and Hammond [21] originally found that MPOD was correlated with tCSF. In the first phase of our study, we correlated tCSF with MPOD in 102 subjects. In the second phase, we utilized a placebo-controlled design and randomized subjects to receive either placebo ( $n = 15$ ) or a xanthophyll-containing supplement ( $n = 54$ ).

## Method

### Subjects

Young adults (ages 18–32 years) were recruited the University of Georgia and Athens, GA community for a four-month double-blind supplementation trial. At the time of enrollment, subjects were randomly assigned (simple randomizing without replacement) to one of three treatment groups. The treatment groups were either 20 mg Z/day ( $N = 29$ ; EyePromise Zeaxanthin, ZeaVision, LLC; Chesterfield, MO) or a “multi” condition 26 mg Z + 8 mg L + 190 mg mixed n-s fatty acids/day ( $N = 25$ ; EyePromise vizual EDGE, ZeaVision, LLC; Chesterfield, MO). A total of 15 subjects received a placebo. Supplements were provided to subjects in an unmarked bottle, and they were instructed to follow dosage instructions listed under the cap for each day when taken with a meal.

All methods and procedures were approved by the University of Georgia’s Institutional Review Board and adhered to the principles in the Declaration of Helsinki. Subjects provided written consent.

<sup>2</sup> Although a purely protective effect cannot be ruled out. Even young subjects can often have quite high levels of inflammatory and oxidative stress [5] and functional improvements would likely result from their amelioration.

**Table 1**

Baseline correlations between macular pigment and measures of temporal vision and temporal contrast sensitivity ( $N = 102$ ).

	Macular pigment (30' eccentricity)	
	r-Value	p-Value (one tailed)
<i>Foveal temporal contrast sensitivity</i>		
1.4 log hertz	0.29	<0.005
1.0 log hertz	0.27	<0.005
0.4 log hertz	0.26	<0.005
<i>Parafoveal temporal contrast sensitivity</i>		
1.4 log hertz	0.21	<0.025
1.0 log hertz	0.26	<0.005
0.4 log hertz	0.26	<0.005

MPOD and temporal visual function were measured on two separate occasions during a single week in order to determine a stable baseline value (we then used the average for the baseline correlations shown in Table 1). At the second visit, subjects received the masked pill bottles and were instructed to take the contents with a meal and to refrain from making substantial changes to their diet during the intervention. Compliance was assessed by questioning the subjects twice during and once at the conclusion of the intervention.

### Assessment of macular pigment

Macular pigment optical density (MPOD) was determined at 30-min retinal eccentricity using customized heterochromatic flicker photometry with a table-top device described by [34]. In brief, a 460 nm light-emitting diode (LED) is presented in square-wave alternation with a 570 nm LED creating the perception of flicker, which is presented at an individually-customized rate. The difference in energy of the 460 nm LED required to eliminate flicker in the fovea (where macular pigment accumulates) compared to the parafovea (an area of the retina without macular pigment) was used to derive MPOD.

### Assessment of temporal contrast sensitivity

Temporal contrast sensitivity was assessed by the customized, LED-driven tabletop device described by Wooten et al. [35]. The test stimulus consisted of a 1-degree 660 nm target at the center of a 5.5-degree 660 nm surround, separated by a 4 arc minute gap. A fixation point at the center of the target was used for foveal measurements. Unlike MP density, which was only measured in the central 1-degree, we also assessed parafoveal temporal sensitivity. To obtain these measures, subjects fixated a small red point placed at 7-degree in the nasal visual field. Subjects viewed the stimuli through a 3 mm artificial pupil. Measurements of temporal contrast sensitivity occurred at 0.4, 1.0, and 1.4 log frequency (i.e., the LEDs were presented in sine-wave at 2.51, 10, and 25 Hz, respectively). Temporal contrast sensitivity values were derived from temporal contrast thresholds, or the depth of modulation at which the target first appeared to flicker. Depth of modulation refers to the amplitude modulation of the sine wave, or the difference between the maximum and minimum luminance of the wave. For each frequency setting, the target was initially set at 0% depth of modulation (and therefore perceptually fused) and increased until the subject reported flicker detection, for a total of five ascending trials for each frequency setting.

### Statistical analyses

Results were analyzed with SPSS 17.0. The baseline relations were assessed using a Pearson product moment correlational

**Table 2**  
Macular pigment at baseline and after 4 months of supplementation for each group.

Group	N	Baseline	Final	Change	t-Value	p-Value
Placebo	15	0.37 ± 0.15	0.35 ± 0.16	−0.02	1.28	0.22
Zeaxanthin	29	0.40 ± 0.15	0.49 ± 0.16	+0.09	4.44 <sup>2</sup>	<0.01
Multi	25	0.33 ± 0.15	0.42 ± 0.16	+0.09	4.67 <sup>2</sup>	<0.01

analysis with one-tailed criteria. Changes in temporal contrast sensitivity after supplementation were assessed with paired samples *t*-tests with significance also set at *p* < 0.05. Bonferroni corrections for multiple *t*-tests were made based on condition and treatment group.

## Results

As shown in Table 1, and similar to past studies, we found strong statistical relationships between MPOD and tCSF across the three frequencies that we tested. These strong relations were found both in the fovea, where MP is dense, and the parafovea. This initial relation motivated the second phase of the study which was designed to assess whether increasing MPOD could actually result in a change in temporal processing. Sixty-nine subjects were randomly assigned to receive either placebo or a pure zeaxanthin supplement or xanthophyll-containing multi for 4 months. As shown in Table 2, MPOD did not change in the placebo group but did

increase significantly in both supplement conditions. In general, both conditions increased MPOD by nearly 0.10 log units.

The increase in MPOD translated to a concomitant increase in temporal processing speed for the supplemented subjects. These data are shown in aggregated form to increase statistical power (see Table 3). As shown in the table, foveal tCSF values did not change for the placebos but improved significantly in the treatment group. A similar finding was found for the parafoveal assessments: no statistically significant change for the placebos but significant increases for the treatment group (see Table 3). Significant changes were maintained when analyses were performed for each treatment condition considered separately (see Table 4).

## Discussion

In this study we found that MPOD was positively correlated with temporal processing speed even when young healthy subjects were targeted. This latter point is significant since young healthy subjects are typically considered to be at peak efficiency and might be expected to be most resistant to change due to ceiling effects. Nonetheless, both MPOD and tCSF increased by about 0.10 log units. This magnitude of change translates to about a 20% increase in MPOD and a similar average increase (~20%) in temporal processing speed. The intervention resulted in a larger change than one might predict based on the relatively moderate (but

**Table 3**  
Changes in measures of foveal and parafoveal temporal contrast sensitivity for subjects either in an active treatment group (*N* = 54) or the placebo group (*N* = 15).

Temporal frequency	Group	Baseline	Final	Change	t-Value	p-Value <sup>a</sup>
Foveal 1.4 log hertz	Treatment	0.17 ± 0.15	0.25 ± 0.14	+0.08	5.53	<0.003
	Placebo	0.32 ± 0.18	0.32 ± 0.19	0.00	0.34	2.22
Foveal 1.0 log hertz	Treatment	1.24 ± 0.13	1.37 ± 0.16	+0.13	6.44	<0.003
	Placebo	1.34 ± 0.14	1.37 ± 0.10	+0.03	0.80	1.32
Foveal 0.4 log hertz	Treatment	1.18 ± 0.15	1.27 ± 0.13	+0.09	4.71	<0.003
	Placebo	1.25 ± 0.16	1.27 ± 0.10	+0.02	0.37	2.16
Parafoveal 1.4 log hertz	Treatment	0.39 ± 0.16	0.44 ± 0.15	+0.05	2.89	0.02
	Placebo	0.40 ± 0.23	0.45 ± 0.21	+0.05	1.45	0.51
Parafoveal 1.0 log hertz	Treatment	1.10 ± 0.13	1.19 ± 0.13	+0.09	6.48	<0.003
	Placebo	1.14 ± 0.15	1.20 ± 0.11	+0.06	1.88	0.24
Parafoveal 0.4 log hertz	Treatment	0.99 ± 0.14	1.09 ± 0.11	+0.10	5.41	<0.003
	Placebo	1.06 ± 0.16	1.11 ± 0.11	+0.05	1.35	0.60

<sup>a</sup> *p*-Values reflect Bonferroni correction for multiple comparisons.

**Table 4**  
Changes in measures of foveal and parafoveal temporal contrast sensitivity for subjects in different active treatment groups.

Temporal frequency	Group <sup>a</sup>	Baseline	Final	Change	t-Value	p-Value <sup>b</sup>
Foveal 1.4 log hertz	Zeaxanthin	0.18 ± 0.19	0.25 ± 0.17	+0.07	3.51	0.006
	Multi	0.17 ± 0.10	0.24 ± 0.10	+0.07	4.61	<0.003
Foveal 1.0 log hertz	Zeaxanthin	1.25 ± 0.17	1.42 ± 0.17	+0.17	6.27	<0.003
	Multi	1.24 ± 0.07	1.30 ± 0.11	+0.06	3.09	0.02
Foveal 0.4 log hertz	Zeaxanthin	1.19 ± 0.19	1.29 ± 0.15	+0.10	3.12	0.01
	Multi	1.17 ± 0.07	1.24 ± 0.09	+0.07	4.52	<0.003
Parafoveal 1.4 log hertz	Zeaxanthin	0.40 ± 0.19	0.43 ± 0.15	+0.03	0.96	1.05
	Multi	0.38 ± 0.12	0.46 ± 0.15	+0.08	3.35	0.009
Parafoveal 1.0 log hertz	Zeaxanthin	1.08 ± 0.16	1.19 ± 0.15	+0.11	5.24	<0.003
	Multi	1.12 ± 0.06	1.19 ± 0.10	+0.07	3.90	0.003
Parafoveal 0.4 log hertz	Zeaxanthin	0.99 ± 0.15	1.09 ± 0.13	+0.10	4.78	<0.003
	Multi	0.99 ± 0.15	1.09 ± 0.09	+0.10	3.17	0.01

<sup>a</sup> Zeaxanthin Group, *N* = 28; Multi Group, *N* = 25.

<sup>b</sup> *p*-Values reflect Bonferroni correction for multiple comparisons.

consistent) cross-sectional relations which tend to only explain about 10% of the variance.

These empirical data are consistent with the idea that increasing central levels of xanthophyll carotenoids can have a generally positive effect upon brain function. We define this as salubrious based on the observation that aging [26] and degenerative disease (e.g., Alzheimer's; [7]; minimal hepatic encephalopathy; [25]; Multiple Sclerosis; [8]) tend to disproportionately affect (i.e., slow) temporal processing. Faster visual processing tends to be positively correlated with, for example, improved sports performance [12] and driver safety [20], reading speed [14], and executive cognitive function [1].

At this stage, there is not enough information to productively speculate on precisely how L and Z might influence processing speed. A few observations, however, are worth noting. The first is that we found significant improvements in the parafovea (see Tables 3 and 4), where MP density is minimal, further suggesting that the locus of the effect is, at least in part, the brain itself. Further, given the time course that is typically seen (e.g., on how long it takes to increase MPOD) it is unlikely that L and Z act as nervous system stimulants like caffeine (stimulants tend to increase visual processing speeds quickly and relatively transiently; [28]). Although it is possible that the pigments are acting to create structural change (e.g., enhancing gap junction communication) within or across neurons or glia, and this would lead to a more lasting improvement in processing speed, it is not clear whether the amounts within brain tissue are sufficient for this purpose (L and Z in brain is expressed in picomolar amounts as opposed to retina which is nanomolar; [29]). Direct effects on DNA could amplify the effects of dietary L and Z on brain and there is some evidence that L and Z may have such capabilities [27,3,17]. In any event, we do know that L and Z are in brain and their presence appears to be more the result of active as opposed to passive mechanisms (since amounts are in excess of what one would predict based on dietary intake; [16]). These observations, combined with the empirical results, implies that L and Z can directly alter brain function, likely throughout the lifespan.

More generally, these data fit in with a widening body of literature that has linked diet to central nervous system function even in young subjects. It can be generally remarked that improving diet is not simply to prevent acquired or deficiency disease, but rather to optimize function throughout life.

### Acknowledgment

This study was funded by ZeaVision, LLC.

### References

- [1] K. Ball, J.D. Edwards, L.A. Ross, J. Gerontol. B Psychol. Sci. Soc. Sci. 62 (1) (2007) 19–31 (special issue).
- [2] E.R. Bovier, L.M. Renzi, B.R. Hammond, PLoS ONE 9 (9) (2014) e108178.
- [3] Q. Bian, S. Gao, J. Zhou, J. Qin, A. Taylor, E.J. Johnson, G. Tang, J.R. Sparrow, D. Gierhart, F. Shang, Free Radical Biol. Med. 53 (6) (2012) 1298–1307.
- [4] C.I. Cazzonelli, B.J. Pogson, Trends Plant Sci. 15 (5) (2010) 266–274.
- [5] K.J. Chuang, C.C. Chan, T.C. Su, C.T. Lee, C.S. Tang, Am. J. Respir. Crit. Care Med. 176 (4) (2007) 370–376.
- [6] M.M. Ciccone, F. Cortese, M. Gesualdo, S. Carbonara, A. Zito, G. Ricci, G. Riccioni, Mediators Inflamm. 2013 (2013) 1–11.
- [7] S. Curran, J. Wattis, Hum. Psychopharmacol. Clin. Exp. 15 (2) (2000) 103–112.
- [8] M.L. Daley, R.L. Swank, C.M. Ellison, Arch. Neurol. 36 (5) (1979) 292–295.
- [9] J. Feeney, C. Finucane, G.M. Savva, H. Cronin, S. Beatty, J.M. Nolan, R.A. Kenny, Neurobiol. Aging 34 (11) (2013) 2449–2456.
- [10] E. García-de Blas, R. Mateo, J. Viñuela, L. Pérez-Rodríguez, C. Alonso-Alvarez, Physiol. Biochem. Zool. 86 (5) (2013) 483–498.
- [11] B.R. Hammond, B.R. Wooten, Ophthalmic Physiol. Opt. 25 (4) (2005) 315–319.
- [12] B.R. Hammond, L.M. Fletcher, Am. J. Clin. Nutr. 96 (5) (2012) 1207S–1213S.
- [13] B.R. Hammond, L.M. Fletcher, J.G. Elliott, Invest. Ophthalmol. Vis. Sci. 54 (1) (2013) 476–481.
- [14] M.D. Jackson, J.L. McClelland, J. Exp. Psychol. Gen. 108 (2) (1979) 151.
- [15] E.J. Johnson, K. McDonald, S.M. Caldarella, H.Y. Chung, A.M. Troen, D.M. Snodderly, Nutr. Neurosci. 11 (2) (2008) 75–83.
- [16] E.J. Johnson, Nutr. Rev. 72 (9) (2014) 605–612.
- [17] M. Kuchan, F. Wang, Y. Geng, B. Feng, C. Lai, Lutein stimulates the differentiation of human stem cells to neural progenitor Cells in vitro. Presented at Advances and Controversies in Clinical Nutrition, Washington, DC, 2013. Abstract No.23.
- [18] A. Lucas, J. Morales, A. Velando, J. Exp. Biol. 217 (8) (2014) 1253–1262.
- [19] J.M. Nolan, E. Loskutova, A.N. Howard, R. Moran, R. Mulcahy, J. Stack, S. Beatty, J. Alzheimers Dis. 42 (4) (2014) 1191–1202.
- [20] C. Owsley, K. Ball, G. McGwin Jr, M.E. Sloane, D.L. Roenker, M.F. White, E.T. Overley, JAMA 279 (14) (1998) 1083–1088.
- [21] L.M. Renzi, B.R. Hammond, Ophthalmic Physiol. Opt. 30 (4) (2010) 351–357.
- [22] L.M. Renzi, E.R. Bovier, B.R. Hammond, Nutr. Neurosci. 16 (6) (2013) 262–268.
- [23] L.M. Renzi, M.J. Dengler, A. Puente, L.S. Miller, B.R. Hammond Jr, Neurobiol. Aging 35 (7) (2014) 1695–1699.
- [24] R.L. Roberts, Am. J. Lifestyle Med. 7 (3) (2013) 182–185.
- [25] M. Romero-Gómez, J. Córdoba, R. Jover, J.A. del Olmo, M. Ramírez, R. Rey, V. Felipe, Hepatology 45 (4) (2007) 879–885.
- [26] T.A. Salthouse, Psychol. Rev. 103 (3) (1996) 403.
- [27] J.M. Serpeloni, I.M.D.S. Cólus, F.S.D. Oliveira, A.F. Aissa, A.Z. Mercadante, M.L.P. Bianchi, L.M.G. Antunes, Food Chem. Toxicol. (2014).
- [28] J.M. Smith, H. Misiak, Psychopharmacology 47 (2) (1976) 175–182.
- [29] R. Vishwanathan, M. Neuringer, D.M. Snodderly, W. Schalch, E.J. Johnson, Nutr. Neurosci. 16 (1) (2013) 21–29.
- [30] R. Vishwanathan, M.J. Kuchan, S. Sen, E.J. Johnson, J. Pediatr. Gastroenterol. Nutr. (2014).
- [31] M.X. Wang, J.H. Jiao, Z.Y. Li, R.R. Liu, Q. Shi, L. Ma, Atherosclerosis 227 (2) (2013) 380–385.
- [32] E.F. Wells, G.M. Bernstein, B.W. Scott, P.J. Bennett, J.R. Mendelson, Exp. Brain Res. 139 (1) (2001) 106–110.
- [33] J.C. Wong, H.S. Kaplan, B.R. Hammond, Nutr. Neurosci. (2014).
- [34] B.R. Wooten, B.R. Hammond, R.I. Land, D.M. Snodderly, Invest. Ophth. Vis. Sci. 40 (1999) 2481–2489.
- [35] B.R. Wooten, L.M. Renzi, R. Moore, B.R. Hammond, Biomed. Opt. Express 1 (1) (2010) 47–58.
- [36] J.P. Zimmer, B.R. Hammond Jr., Clin. Ophthalmol. 1 (1) (2007) 25.