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In vitro studies of the antileishmanial activity of the newer 2-(substitutedphenoxy)-N-[(aryl)methylidene]acetohydrazide analogues



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ABSTRACT

A series of new 2-(substitutedphenoxy)-N-[(aryl)methylidene]acetohydrazide analogues (**8a**-n) were synthesized in search of potential therapeutics for leishmaniasis. All the compounds were characterized by infrared (IR), nuclear magnetic resonance (NMR) spectroscopy and mass spectrometry. The compounds were further evaluated for *in vitro* antileishmanial activity against promastigotes of *Leishmania donovani* as per the standard protocol reported elsewhere. 2-(2,4-Dichlorophenoxy)-N'-[[4-(morpholin-4-yl)phenyl]methylidene]acetohydrazide (**8k**) showed the most promising antileishmanial activity with IC₅₀ of 48.10 μ M, free from cytotoxicity (>153.08 μ M).

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1. Introduction

Leishmaniasis is a neglected infectious disease caused by protozoa of genus *Leishmania*. More than 20 species are found worldwide causing human leishmaniasis. The vector for the disease is phlebotomine (sand flies) and is manifested in three major clinical forms: cutaneous (CL), mucocutaneous and visceral leishmaniasis (VL). VL, the most severe form of leishmaniasis, is also known as kala-azar in India. The symptoms of VL include fever, hepatosplenomegaly and anaemia that may lead to death. The pathogen is endemic in 88 countries. Nearly 12 million people were infected and 2 million new cases occur every year. An estimated 350 million people are living at risk of contracting leishmaniasis. One of the major threats to control VL is its association with HIV infection (Trouiller and Olliaro, 1999; Alvar et al., 2012; WHO, 2012). Pentavalent antimonials remain the first-line, while polyene antifungal,

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amphotericin B, as a second-line treatment is currently used in many countries. AmBisome (liposomal amphotericin B) and miltefosine were also included in the treatment of VL (Reddy et al., 2007; Sindermann et al., 2004; Thakur et al., 1999). However, the use of these existing drugs is limited due to several complications, such as high cost, toxicity, parenteral administration, emergence and spread of drug resistance, and relapses in HIV-leishmania co-infected patients. Therefore, there is still a need for new efficacious and safe agents against leishmaniasis.

Hydrazides have emerged as biologically and pharmacological promising agents, with antileishmanial (Sagsehetti et al., 2014), antitubercular (Ramamurthy and Bhatt, 1989), anticonvulsant (Kaushik et al., 2010), antibacterial (Sridhar et al., 2002), antifungal (Mallikarjuna et al., 2009), anti-HIV (Vicini et al., 2009), antioxidant (Gurkok et al., 2009), and many more. Hence it was worth to synthesize these compounds.

2. Materials and methods

2.1. Chemistry

All the chemicals were supplied by Merck (Germany) and S. D. Fine Chemicals (India). Melting points were determined by open tube capillary method and were uncorrected. Purity of the compounds was checked by elemental analysis and the progress of reactions was monitored throughout by thin layer chromatography (TLC) plates (silica gel G) using mobile phase, hexane:ethylacetate (1:1), and the spots were identified by iodine vapours or UV light. IR spectra were recorded on a Schimadzu 8201 PC, FT-IR spectrometer (KBr pellets). ¹H NMR spectra were recorded on a Bruker AC 300 MHz spectrometer using TMS as internal standard in DMSO d_6 . Mass spectra were recorded on a Bruker Esquire LCMS using ESI and elemental analyses were performed on Perkin-Elmer 2400 Elemental Analyzer.

2.2. Synthesis of ethyl(substitutedphenoxy)acetate analogues (3a-d)

A mixture of equimolar amounts of the substituted phenol and ethyl chloroacetate was taken in a round bottom flask and suspended in 50-60 ml acetone, and anhydrous potassium carbonate (1-2 g) was added in the mixture. The mixture was refluxed for 24 h on the sand bath with vigorous stirring. The completion of the reaction was monitored by TLC using mobile phase hexane:ethylacetate (1:1). Initially, the colour of reaction mixture was colourless in case of phenol, while in other substituted phenols the colour was light yellow and the reaction proceeded until the reaction mixture became dark in colour. The reaction mixture, when cooled, was filtered under vacuum to remove solid potassium carbonate and the filtrate thus obtained was evaporated under vacuum. The residue thus obtained was dissolved into ethylacetate (10-15 ml) and washed with water twice. Ethylacetate layer was separated and dried over anhydrous sodium sulphate. The solvent was evaporated under vacuum and the products thus obtained were used for the next step.

2.3. Synthesis of 2-(substitutedphenoxy) acetohydrazide analogues (4a-d)

A solution of ethyl (substitutedphenoxy)acetate (0.01 mol) (**3a-d**) and hydrazine hydrate (0.02 mol) was taken in a round bottom flask and suspended in 50–60 ml ethanol. The mixture was refluxed for 5–6 h on a sand bath with vigorous stirring. The reaction was monitored throughout by TLC. The reaction was continued until the ethyl (substitutedphenoxy)acetate was consumed completely. The reaction mixture was poured in crushed ice filtered under vacuum and washed with water to remove solid 2-(substitutedphenoxy)acetohydrazide analogues (**4a-d**).

2.4. Synthesis of 4-(substitutedphenoxy) benzaldehyde analogues (7a-c)

A solution of substituted *p*-fluorobenzaldehyde (0.01 mol) and substituted phenol/morpholine (0.012 mol) was taken in a round bottom flask and suspended in 50–60 ml DMSO and anhydrous potassium carbonate. The mixture was refluxed for 14– 18 hrs on a sand bath with vigorous stirring. The reaction was monitored throughout by TLC. Initially, the reaction mixture was colourless in case of phenol, while in other phenols it was light yellow and the reaction became dark in colour as it proceeded and reached to completion. The reaction mixture was cooled, added water and ethylacetate in a separating funnel, and the ethylacetate layer was separated and evaporated under rotatory vacuum evaporator. The solid thus obtained was washed with ethanol, dried and used for the next step.

2.5. Synthesis of 2-(substitutedphenoxy)-N-[(aryl)methylidene]-acetohydrazide analogues (8a-n)

A solution of *p*-fluorobenzaldehyde (5)/substituted 4-(substitutedphenoxy)benzaldehyde analogues (7a-c) (1 mmol) and 2-(substitutedphenoxy)acetohydrazide analogues (4a-d) (1 mmol) was taken in a round bottom flask and suspended in 50–60 ml ethanol and TEA (1.5 mol). The mixture was refluxed for 10–14 h on sand bath with vigorous stirring. The reaction was monitored throughout by TLC. The reaction mixture was poured into the cold water and the product was extracted by ethylacetate using a separating funnel. The ethylacetate layer was then separated and evaporated under rotator vacuum evaporator. The solid thus obtained was washed and crystallized with ethanol.

2.5.1. 2-(2-Chlorophenoxy)-N'-{[4-(2,4-dichlorophenoxy) phenyl] methylidene}acetohydrazide (8a)

Yield 70%, mp. 148–150 °C, R_f 0.63 [hexane: ethylacetate (1:1)], IR (KBr) cm⁻¹: 3223.16 (NH), 1682.95 (C=O), 1573.97 (C=N), 1097.53 (-O-), 741.65 (C-Cl); ¹H NMR (DMSO d_6 , 300 MHz) δ ppm: 4.94 (s, 2H, CH₂), 6.72–7.74 (m, 11H, ArH), 7.99 (s, 1H, CH=N), 11.34 (s, 1H, CONH; D₂O exchangeable); ¹³C NMR (DMSO d_6 , 75 MHz) δ ppm: 154.83, 146.83, 143.31, 130.74, 129.94, 129.63, 129.40, 126.55, 124.97, 123.40, 121.88, 117.92, 114.20, 91.81; EI-MS (m/z) 449.75 (M⁺), 451.41 (M⁺+2), 453.81 (M⁺+4). Cal/Ana: [C (56.09) 56.05 H (3.36) 3.38 N (6.23) 6.24].

2.5.2. 2-(2-Chlorophenoxy)-N'-[(4-phenoxyphenyl) methylidene] acetohydrazide (**8b**)

Yield 69%, mp. 135–137 °C, $R_f 0.73$ [hexane: ethylacetate (1:1)], IR (KBr) cm⁻¹: 3186.51 (NH), 1680.05 (C=O), 1487.17 (C=N), 1164.08 (-O-), 743.58 (C-Cl); ¹H NMR (DMSO d_6 , 300 MHz) δ ppm: 5.09 (s, 2H, CH₂), 6.69–7.79 (m, 13H, ArH), 8.17 (s, 1H, CH=N), 11.04 (s, 1H, CONH₂; D₂O exchangeable); ¹³C NMR (DMSO d_6 , 75 MHz) δ ppm: 68.61, 115.72, 117.21, 117.90, 121.92, 122.56, 122.63, 126.91, 127.90, 128.51, 128.93, 129.92, 143.14, 154.71, 157.01, 159.33, 168.16; EI-MS (m/z) 381.41 (M⁺), 383.44 (M⁺+2). Cal/Ana: [C (66.23) 66.19 H (4.50) 4.53 N (7.36) 7.32].

2.5.3. 2-(2-Chlorophenoxy)-N'-{[4-(morpholin-4-yl)phenyl] methylidene} acetohydrazide (**8c**)

Yield 74%, mp. 128–130 °C, $R_f 0.76$ [hexane: ethylacetate (1:1)], IR (KBr) cm⁻¹: 3181.69 (NH), 1682.95 (C=O), 1493.92 (C=N), 1121.64 (-O-, morpholine), 747.44 (C-Cl); ¹H NMR (DMSO d_6 , 300 MHz) δ ppm: 2.40–2.45 (t, 4H, morpholine), 3.71–3.72 (t, 4H, morpholine), 4.84 (s, 2H, CH₂), 6.70–7.36 (m, 8H, ArH), 8.27 (s, 1H, CH=N), 11.41 (s, 1H, CONH; D₂O exchangeable); ¹³C NMR (DMSO d_6 , 75 MHz) δ ppm: 46.31, 66.11, 68.60, 114.41, 115.72, 122.51, 122.68, 123.33, 127.94, 129.92, 130.09, 143.01, 151.92, 154.71, 168.66; EI-MS (m/z) 373.44 (M⁺), 375.49 (M⁺+2). Cal/Ana: [C (61.04) 61.01 H (5.39) 5.43 N (11.24) 11.19].

2.5.4. 2-(2-Chlorophenoxy)-N'-[(4-fluorophenyl)methylidene] acetohydrazide (8d)

Yield 56%, mp. 122–124 °C, R_f 0.62 [hexane: ethylacetate (1:1)], IR (KBr) cm⁻¹: 3200.02 (NH), 1680.12 (C=O), 1457.16 (C=N), 1196.02 (-O-), 687.12 (C-F); ¹H NMR (DMSO d_6) δ ppm: 4.83 (s, 2H, CH₂), 6.81–7.46 (m, 8H, ArH), 7.99 (s, 1H, CH=N), 11.02 (s, 1H, CONH; D₂O exchangeable); ¹³C NMR (DMSO d_6 , 75 MHz) δ ppm: 68.51, 115.63, 115.71, 122.56, 122.61, 127.91, 129.41, 129.92, 130.82, 143.01, 154.72, 165.22, 168.01; EI-MS (*m*/z) 306.24 (M⁺), 308.37 (M⁺+2). Cal/Ana: [C (58.74) 58.69 H (3.94) 3.89 N (9.13) 9.09].

2.5.5. N'-{[4-(2,4-Dichlorophenoxy)phenyl]methylidene}-2-(2-methyl-phenoxy)acetohydrazide (**8e**)

Yield 84%, mp. 150–152 °C, R_f 0.54 [hexane: ethylacetate (1:1)], IR (KBr) cm⁻¹: 3220.27 (NH), 1678.13 (C=O), 1574.93 (C=N), 1099.46 (-O-), 714.65 (C--Cl); ¹H NMR (DMSO d_6 , 300 MHz) δ ppm: 2.23 (s, 3H, CH₃), 4.64 (s, 2H, CH₂), 6.82–7.79 (m, 11H, ArH), 7.98 (s, 1H, CH=N), 11.45 (s, 1H, CONH; D₂O exchangeable); ¹³C NMR (DMSO d_6 , 75 MHz) δ ppm: 14.41, 69.42, 114.21, 117.62, 120.33, 121.01, 124.51, 126.72, 126.81, 126.97, 127.21, 128.83, 128.96, 130.11, 131.01, 143.01, 150.11, 158.77, 159.31, 168.99; EI-MS (m/z) 429.45 (M⁺), 431.45 (M⁺+2) 433.44 (M⁺+4). Cal/Ana: [C (61.55) 61.51 H (4.23) 4.19 N (6.53) 6.49].

2.5.6. 2-(2-Methylphenoxy)-N'-[(4-phenoxyphenyl) methylidene] acetohydrazide (8f)

Yield 82%, mp. 145–147 °C, R_f 0.63 [hexane: ethylacetate (1:1)], IR (KBr) cm⁻¹: 3237.63 (NH), 1680.05 (C=O), 1497.78 (C=N), 1192.05 (-O--); ¹H NMR (DMSO d_6 , 300 MHz) δ ppm: 2.23 (s, 3H, CH₃), 5.12 (s, 2H, CH₂), 6.79–7.97 (m, 13H, ArH), 8.27 (s, 1H, CH=N), 11.44 (s, 1H, CONH; D₂O exchangeable); ¹³C NMR (DMSO d_6 , 75 MHz) δ ppm: 14.44, 69.92, 114.21, 117.51, 118.63, 121.01, 121.92, 124.51, 126.18, 126.99, 128.55, 129.91, 130.11, 143.12, 157.01, 158.71, 159.93, 168.66; EI-MS (*m*/z) 460.11 (M⁺), 461.09 (M⁺+1). Cal/Ana: [C (73.32) 73.29 H (5.59) 5.55 N (7.77) 7.72].

2.5.7. 2-(2-Methylphenoxy)-N'-{[4-(morpholin-4-yl)phenyl] methylidene} acetohydrazide (**8g**)

Yield 73%, mp. 136–138 °C, $R_f 0.65$ [hexane: ethylacetate (1:1)], IR (KBr) cm⁻¹: 3195.19 (NH), 1670.41 (C=O), 1553.71 (C=N), 1190.12 (-O-), 1123.57 (-O-, morpholine ring); ¹H NMR (DMSO d_6 , 300 MHz) δ ppm: 2.31 (s, 3H, CH₃), 2.96–2.98 (t, 4H, morpholine), 3.62–3.64 (t, 4H, morpholine), 4.82 (s, 2H, CH₂), 6.69– 7.62 (m, 8H, ArH), 7.92 (s, 1H, CH=N), 11.02 (s, 1H, CONH; D₂O exchangeable); ¹³C NMR (DMSO d_6 , 75 MHz) δ ppm: 14.45, 46.33, 66.42, 69.62, 114.21, 114.92, 121.01, 123.33, 124.55, 126.81, 130.11, 130.96, 143.09, 151.99, 158.72, 168.72; EI-MS (*m*/z) 353.11 (M⁺), 354.41 (M⁺+1). Cal/Ana: [C (67.97) 67.91 H (6.56) 6.49 N (11.89) 11.86].

2.5.8. N'-[(4-Fluorophenyl)methylidene]-2-(2-methylphenoxy) acetohydrazide (8h)

Yield 65%, mp. 164–166 °C, R_f 0.72 [hexane: ethylacetate (1:1)], IR (KBr) cm⁻¹: 3185.12 (NH), 1674.46 (C=O), 1453.71 (C=N), 1194.12 (-O-), 697 (C-F); ¹H NMR (DMSO d_6 , 300 MHz) δ ppm: 2.33 (s, 3H, CH₃), 4.92 (s, 2H, CH₂), 6.62–7.32 (m, 8H, ArH), 7.89 (s, 1H, CH=N), 10.97 (s, 1H, CONH; D₂O exchangeable); ¹³C NMR (DMSO d_6 , 75 MHz) δ ppm: 14.46, 69.42, 114.21, 115.56, 121.22, 125.09, 126.81, 129.92, 130.31, 131.11, 143.11, 158.71, 165.22, 168.99; EI-MS (m/z) 286.35 (M⁺), 287.49 (M⁺+1). Cal/Ana: [C (67.12) 67.07 H (5.28) 5.29 N (9.78) 9.76].

2.5.9. 2-(2,4-Dichlorophenoxy)-N'-{[4-(2,4-dichlorophenoxy) phenyl] methylidene}acetohydrazide (8i)

Yield 62%, mp. 138–140 °C, R_f 0.68 [hexane: ethylacetate (1:1)], IR (KBr) cm⁻¹: 3220.27 (NH), 1678.13 (C=O), 1474.63 (C=N), 1099.46 (-O-), 714.65 (C-Cl); ¹H NMR (DMSO d_6 , 300 MHz) δ ppm: 4.54 (s, 2H, CH₂), 6.72–7.71 (m, 10H, ArH), 7.97 (s, 1H, CH=N), 11.15 (s, 1H, CONH; D₂O exchangeable); ¹³C NMR (DMSO d_6) δ ppm: 66.18, 115.57, 117.76, 117.91, 123.40, 124.97, 126.55, 128.29, 129.41, 129.63, 129.94, 130.74, 143.68, 150.53, 153.29, 158.09, 168.63; EI-MS: (m/z) 481.51 (M⁺), 483.50 (M⁺+2), 485.49 (M⁺+4). Cal/Ana: [C (52.10) 52.05 H (2.91) 2.89 N (5.79) 5.77].

2.5.10. 2-(2,4-Dichlorophenoxy)-N'-[(4-phenoxyphenyl) methylidene] acetohydrazide (**8j**)

Yield 72%, mp. 144–146 °C, R_f 0.69 [hexane: ethylacetate (1:1)], IR (KBr) cm⁻¹: 3190.37 (NH), 1680.05 (C=O), 1586.50 (C=N), 1165.04 (-O-), 689.57 (C-Cl); ¹H NMR (DMSO d_6 , 300 MHz) δ ppm: 5.28 (s, 2H, CH₂), 7.02–7.73 (m, 12H, ArH), 7.99 (s, 1H, CH=N), 11.62 (s, 1H, CONH; D₂O exchangeable); ¹³C NMR (DMSO d_6 , 75 MHz) δ ppm: 68.66, 117.71, 118.11, 118.90, 121.01, 124.91, 126.91, 128.01, 128.91, 129.10, 129.94, 131.41, 143.33, 152.82, 157.01, 159.91, 168.62; EI-MS (m/z) 415.26 (M⁺), 417.43 (M⁺+2). Cal/Ana: [C (60.74) 60.71 H (3.88) 3.89 N (6.75) 6.73].

2.5.11. 2-(2,4-Dichlorophenoxy)-N'-{[4-(morpholin-4-yl) phenyl] methylidene}acetohydrazide (**8k**)

Yield 71%, mp. 130–132 °C, R_f 0.66 [hexane: ethylacetate (1:1)], IR (KBr) cm⁻¹: 3182.65 (NH), 1681.02 (C=O), 1528.65 (C=N), 1185.30 (-O, morpholine), 724.29 (C-Cl); ¹H NMR (DMSO d_6 , 300 MHz) δ ppm: 4.84 (s, 2H, CH₂), 2.40–2.45 (t, 4H, morpholine), 3.71–3.72 (t, 4H, morpholine), 6.70–7.36 (m, 8H, ArH), 8.27 (s, 1H, CH=N), 11.41 (s, 1H, CONH; D₂O exchangeable); ¹³C NMR (DMSO d_{6_7} , 75 MHz) δ ppm: 46.32, 66.01, 68.16, 114.41, 117.11, 123.01, 124.10, 128.11, 128.94, 130.19, 131.02, 143.12, 151.92, 152.81, 168.72; EI-MS (m/z) 408.54 (M⁺), 409.55 (M⁺+2), 411.49 (M⁺+4). Cal/Ana: [C (55.89) 55.85 H (4.69) 4.63 N (10.29) 10.25].

2.5.12. 2-(2,4-Dichlorophenoxy)-N'-[(4-fluorophenyl) methylidene] acetohydrazide (8l)

Yield 58%, mp. 172–174 °C, R_f 0.77 [hexane: ethylacetate (1:1)], IR (KBr) cm⁻¹: 3192.61 (NH), 1679.02 (C=O), 1483.21 (C=N), 1198.30 (-O—), 687.29 (C—F); ¹H NMR (DMSO d_6 , 300 MHz) δ ppm: 4.91 (s, 2H, CH₂), 6.72–7.31 (m, 7H, ArH), 7.87 (s, 1H, CH=N), 10.42 (s, 1H, CONH; D₂O exchangeable); ¹³C NMR (DMSO d_6 , 75 MHz) δ ppm: 68.12, 115.56, 117.11, 124.01, 128.11, 128.91, 129.44, 130.18, 131.41, 143.12, 152.81, 168.16; EI-MS (m/z) 340.11 (M^+), 341.23 (M^+ +1), 342.54 (M^+ +2). Cal/Ana: [C (52.81) 52.85 H (3.25) 3.23 N (8.21) 8.25].

2.5.13. 2-Phenoxy-N'-[(4-phenoxyphenyl)methylidene] acetohydrazide (8m)

Yield 60%, mp. 140–142 °C, R_f 0.78 [hexane: ethylacetate (1:1)], IR (KBr) cm⁻¹: 3083.31 (NH), 1680.05 (C=O), 1562.39 (C=N), 1192.05 (-O—); ¹H NMR (DMSO d_6 , 300 MHz) δ ppm: 5.04 (s, 2H, CH₂), 6.76–7.91 (m, 13H, ArH), 8.02 (s, 1H, CH=N), 11.14 (s, 1H, CONH; D₂O exchangeable); ¹³C NMR (DMSO d_6 , 75 MHz) δ ppm: 69.01, 114.31, 117.51, 117.61, 121.09, 121.91, 126.92, 128.50, 128.91, 129.79, 143.11, 157.11, 159.32, 168.19; EI-MS (m/z) 346.54 (M⁺), 347.49 (M⁺+1), 348.49 (M⁺+2). Cal/Ana: [C (72.82) 72.78 H (5.24) 5.23 N (8.09) 8.05].

2.5.14. N'-[(4-Fluorophenyl)methylidene]-2-

phenoxyacetohydrazide (8n)

Yield 62%, mp. 168–170 °C, $R_f 0.74$ [hexane: ethylacetate (1:1)], IR (KBr) cm⁻¹: 3231.63 (NH), 1679.05 (C=O), 1493.78 (C=N), 1191.05 (-O-), 688 (C-F); ¹H NMR (DMSO d_6 , 300 MHz) δ ppm: 4.98 (s, 2H, CH₂), 6.71–7.27 (m, 8H, ArH), 7.96 (s, 1H, CH=N), 11.12 (s, 1H, CONH; D₂O exchangeable); ¹³C NMR (DMSO d_6 , 75 MHz) δ ppm: 69.11, 114.31, 115.63, 121.11, 129.42, 129.89, 130.83, 143.12, 168.66; EI-MS (*m/z*) 272.11 (M⁺), 273.23 (M⁺+1). Cal/Ana: [C (66.17) 66.11 H (4.81) 4.83 N (10.29) 10.25].

2.6. In vitro antileishmanial activity

Promastigotes of Indian Leishmania donovani strain MHOM/IN/ 83/AG83 was obtained from the culture bank of Rajendra Memorial Research Institute of Medical Sciences (ICMR), Patna, India. The cryo-cells were revived and grown in RPMI 1640 medium (Sigma-Aldrich) supplemented with 10% foetal calf serum (FCS: Sigma-Aldrich) in BOD incubator at 25 °C.

Specimens were dissolved in dimethyl sulphoxide (DMSO) and further dilutions were made with RPMI 1640 medium (Navarro et al., 2010; Vitale et al., 1989). The concentrations of 500, 250, 125, 62.5, 31.25, 15.6, 7.8, 3.9, and 1.9 (µg/ml) were prepared from the stock solution of 10 mg/ml in DMSO solution by serial dilution methods. Promastigotes were adapted for growth at 25 °C in RPMI 1640 (Sigma-Aldrich) and supplemented with 20% heat inactivated foetal bovine serum (Miguel et al., 2007). Logarithmic phase cultures were used for experimental purposes, and the in vitro susceptibility assay was performed in sterilized 96-well culture micro titre plates (Nunc) (Rolón et al., 2006). A dose of 2.5×10^6 parasites was added to each well to a final volume of 200 ml, together with the drug concentration. Growth of promastigotes was monitored after 48 h by counting the number of motile promastigotes microscopically in a Neubauer chambers. The log dose and response were used to calculate the 50% inhibitory concentrations (IC₅₀) of compounds. IC₅₀ was calculated by linear regression analysis with 95% confidence limits. Tests were performed at least in triplicate on three different days in order to verify the results and amphotericin B was used as positive control.

2.7. Cytotoxicity

The active compounds were tested for cytotoxicity (IC_{50}) in VERO cells by serial double dilution technique. After 72 h exposure,



Fig. 1 - Protocol for the synthesis of acetohydrazide analogues (8a-n).

Table 1 – Antileishmanial activity and cytotoxicity evaluation of the newer acetohydrazide analogues (8a-n).				
S. no.	Compound	Structure	IC ₅₀ (μM)	IC ₅₀ (μM)
			L. donovaniª	Vero
1	8a		280.05 ± 34.71	ND
2	8b		59.29 ± 25.28	164.11
3	8c		242.06 ± 52.67	ND
4	8d		242.92 ± 37.88	ND
5	8e		183 ± 29.14	>145.59
6	8f		194.03 ± 73.58	>173.42
7	8g		171.44 ± 72.52	ND
8	8h		252.39 ± 32.34	ND
9	81		110.23 ± 32.74	ND
10	8j		144.60 ± 46.33	150.51
11	8k		48.10 ± 25.86	>153.08
12	81		270 ± 37.93	ND
13	8m		208.55 ± 27.60	ND
14	8n	or NNN F	466.89 ± 47.49	ND
15	Amphotericin B	-	0.69 ± 0.13	ND
^a Promastigotes of Leishmania donovani; ND: not determined.				

viability was assessed on the basis of cellular conversion of MTT into a formazan product using the Promega Cell Titer 96 Non-Radioactive Cell Proliferation method (Heifets et al., 1989).

3. Results and discussion

3.1. Chemistry

In the first step equimolar mixture of substituted phenol and ethyl chloroacetate was suspended in acetone anhydrous potassium carbonate and refluxed for 24 h to ethyl(substitutedphenoxy)acetate analogues (3a-d), which was further treated with hydrazine hydrate for 5-6 h in ethanol to obtain 2-(substitutedphenoxy)acetohydrazide analogues (4ad). On the other hand 4-(substitutedphenoxy)benzaldehyde analogues (7a-c) were synthesized from 4-fluorobenzaldehyde and substituted phenol and morpholine in DMSO and anhydrous potassium carbonate by refluxing for 14-18 h. We have used different reaction conditions for the synthesis of 4-(substitutedphenoxy)benzaldehyde analogues (7a-c) from the earlier reported method (Joshi et al., 2010; Tripathi and Kumar, 2013). In the final step the aromatic aldehydes (5 and 7a-c) and 2-(substitutedphenoxy)acetohydrazide analogues (4a-d) were refluxed for 10-14 h in ethanol and triethylamine (TEA) to obtain the final product 2-(substitutedphenoxy)-N-[(aryl)methylidene]acetohydrazide analogues (8a-n). The reaction was monitored throughout by TLC using chloroform-methanol (9:1) and hexane:ethylacetate (1:1) as mobile phase. The spots were located under iodine vapour/UV light. The reaction sequence is shown in Fig. 1. The synthesized compounds were characterized by spectral analysis and all the compounds were in full harmony with the proposed structures. In general the IR spectra afforded absorption at 3237–3181 cm⁻¹ due to N—H stretching, 1682–1670 cm⁻¹ band due to C=O, 1573–1453 cm⁻¹ band due to C=N and 1056-1198 cm⁻¹ due to ether stretching. In ¹H NMR the signals of the respective protons of the synthesized title compounds were verified on the basis of their chemical shifts and multiplicities in DMSO d_6 . The spectra showed a singlet at δ 2.23–2.33 ppm corresponding to CH₃ (Ar—CH₃); a multiplet at δ 2.40–3.72 ppm corresponding to morpholine protons; a singlet at δ 4.54–5.28 ppm to CH₂ (methylene bridge); a singlet at δ 6.69–7.97 ppm corresponding to aromatic protons; a singlet at δ 7.79–8.27 ppm corresponding to imine proton (CH=N); and a multiplet at δ 10.42–11.62 ppm (D₂O exchangeable) corresponding to amide (CONH).

3.2. Antileishmanial activity and cytotoxicity

Compounds were screened for *in vitro* antileishmanial activity against promastigotes of *Leishmania donovani* between 48.11 and 466.88 μ M. The most active compounds of the series was **8k** (IC₅₀ 48.11 ± 25.86 μ M) followed by **8b** (IC₅₀ 59.29 ± 25.28 μ M) while rest of the compounds showed moderate to low activity with IC₅₀ between 110.23 ± 32.74 and 466.88 ± 47.49 μ M (Table 1). The antileishmanial activity was carried out in μ g/ml and later converted in μ M. The SAR explored from the data showed that the terminal aryl group with 4-(4-phenyl)morpholine as in compound **8k** showed maximum

antileishmanial activity and the activity decreased with 4-phenoxybenzene, 4-fluorophenyl and 2,4-dichloro-1-(4phenoxy)benzene. The O-aryl group (acetohydrazide) with 2,4dichlorophenyl (as in compound **8k**) showed maximum activity followed by 2-chlorophenyl, 2-methylphenyl and unsubstituted phenyl respectively. Two compounds (**8b** and **8k**) showed promising activity against the promastigotes of *L. donovani* and SAR confirmed that the compound **8k** could be considered as lead for further optimization. Further 5 compounds were screened for cytotoxic studies. The results of cytotoxic studies showed that the active compounds (**8b** and **8k**) were found to be nontoxic with IC₅₀ of 164.11 and >153.08 µM respectively (Table 1).

4. Conclusion

All the compounds were synthesized in satisfactory yield. All the compounds were characterized by IR, NMR and mass spectral data. The antileishmanial activity was carried out for all the compounds against the promastigotes of *Leishmania donovani*. Standard protocol was followed for antileishmanial activity. Some of the compounds showed promising antileishmanial activity. The compound **8k** showed the maximum antileishmanial activity with could be considered as lead for antileishmanial activity. The cytotoxic studies also showed that the compound **8k** was found to be non-toxic. The detailed molecular modelling and chemical modifications studies are in progress in our laboratory.

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REFERENCES

- Alvar J, Velez ID, Bern C, Herrero M, Desjeux P, Cano J, et al. Leishmaniasis worldwide and global estimates of its incidence. PLoS ONE 2012;7:e35671.
- Gurkok G, Coban T, Suzen S. Melatonin analogue new indole hydrazide/hydrazone derivatives with antioxidant behavior: synthesis and structure-activity relationships. J Enzyme Inhib Med Chem 2009;24:506–15.
- Heifets LB, Flory MA, Lindholm-Levy P. Does pyrazinoic acid as an active moiety of pyrazinamide have specific activity against Mycobacterium tuberculosis? J Antimicrob Chemother 1989;33:1252–4.
- Joshi RS, Mandhane PG, Diwakar SD, Dabhade SK, Gill CH. Synthesis, analgesic and anti-inflammatory activities of some novel pyrazolines derivatives. Bioorg Med Chem Lett 2010;20:3721–5.

- Kaushik D, Khan SA, Chawla G, Kumar S. N'-[(5-chloro-3-methyl-1-phenyl-1H-pyrazol-4-yl)methylene] 2/4-substituted hydrazides: synthesis and anticonvulsant activity. Eur J Med Chem 2010;5:3943–9.
- Mallikarjuna BP, Sastry BS, Kumar GVS, Rajendraprasad Y, Chandrashekar SM, Sathisha K. Synthesis of new
 4-isopropylthiazole hydrazide analogs and some derived clubbed triazole, oxadiazole ring systems-a novel class of potential antibacterial, antifungal and antitubercular agents. Eur J Med Chem 2009;44:4739–46.
- Miguel DC, Yokoyama-Yasunaka JK, Andreoli WK, Mortara RA, Uliana SR. Tamoxifen is effective against Leishmania and induces a rapid alkalinization of parasitophorous vacuoles harbouring Leishmania (Leishmania) amazonensis amastigotes. J Antimicrob Chemother 2007;60:526–34.
- Navarro M, Gabbiani C, Messori L, Gambino D. Metal-based drugs for malaria, trypanosomiasis and leishmaniasis: recent achievements and perspectives. Drug Discov Today 2010;15:1070–8.
- Ramamurthy B, Bhatt MV. Synthesis and antitubercular activity of N-(2-naphthyl)glycine hydrazide analogues. J Med Chem 1989;32:2421–6.
- Reddy M, Gill SS, Kalkar SR, Wu W, Anderson PJ, Rochon PA. Oral drug therapy for multiple neglected tropical diseases: a systematic review. JAMA 2007;298:1911–24.
- Rolón M, Vega C, Escario JA, Gómez-Barrio A. Development of resazurin microtiter assay for drug sensibility testing of Trypanosoma cruzi epimastigotes. Parasitol Res 2006;99:103– 7.
- Sagsehetti JN, Shaik RI, Khan FA, Patil RH, Marathe SD, Gade WN, et al. Synthesis, antileishmanial activity and docking study of

N'-substitutedbenzylidene-2-(6,7-dihydrothieno[3,2-c]priding-5(4H)-yl)acetohydrazides. Bioorg Med Chem Lett 2014;24:1605– 10.

- Sindermann H, Croft SL, Engel KR, Bommer W, Eibl HJ, Unger C, et al. Miltefosine (Impavido): the first oral treatment against leishmaniasis. Med Microbiol Immunol 2004;193:173–80.
- Sridhar SK, Pandeya SN, Stables JP, Ramesh A. Anticonvulsant activity of hydrazones, Schiff and Mannich bases of isatin derivatives. Eur J Med Chem 2002;16:129–32.
- Thakur CP, Singh RK, Hassan SM, Kumar R, Narain S, Kumar A. Amphotericin B deoxycholate treatment of visceral leishmaniasis with newer modes of administration and precautions: a study of 938 cases. Trans R Soc Trop Med Hyg 1999;93:319–23.
- Tripathi L, Kumar P. Augmentation of GABAergic neurotransmission by novelN-(substituted)-2-[4-(substituted)benzylidene]hydrazinecarbothioamidesdA potential anticonvulsant approach. Eur J Med Chem 2013;64:477–87.
- Trouiller P, Olliaro P. Drug development output from 1975 to 1996 what proportion for tropical disease? Int J Infect Dis 1999;3:61–3.
- Vicini P, Incerti M, La Colla P, Loddo R. Anti-HIV evaluation of benzo[d]isothiazole hydrazones. Eur J Med Chem 2009;44:1801–7.
- Vitale RG, Mouton JW, Afeltra J, Meis JF, Verweij PE. Method for measuring postantifungal effect in Aspergillus species. Antimicrob Agents Chemother 1989;46:1960–5.
 WHO. World Health Organization,
 - <http://www.who.int/leishmaniasis/burden/en/>; 2012
 [accessed 17.10.14].