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## Review

## T-cell protein tyrosine phosphatase: A role in inflammation and autoimmunity

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## ABSTRACT

A cDNA of *PTPN2* encoding for T-cell protein tyrosine phosphatase (TC-PTP) was isolated and characterized as long as 20 years ago. However, findings suggesting a potentially exciting role of this enzyme in general autoimmunity have only recently been obtained. Genome-wide association scans of the human genome revealed the involvement of *PTPN2* in susceptibility to a several autoimmune disorders such as Crohn's disease, type 1 diabetes, and Graves' disease. Functional studies in immune cells revealed a key role of this enzyme in down-regulation of cytokine expression and inflammatory response, which provides an essential background to explaining the pathophysiological role of TC-PTP in autoimmunity. Thus, in addition to *PTPN22*, *PTPN2* is likely to represent a second member of the broad family of non-receptor PTPs contributing to general autoimmunity.

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## 1. Introduction

Many genes identified as causing or predisposing to autoimmunity encode proteins that are involved in lymphocyte, macrophage, or dendritic cell signal transduction. Among those, intracellular protein tyrosine phosphatases (PTPs) play a unique role as the key regulators of signal transduction. To date, around 110 PTP-encoding genes are found in the human genome [1]. All cells of the immune system exhibit high levels of tyrosine phosphorylation and express more PTP genes (up to 70) than any other tissue, with the possible exception of neurons [2]. Acute phenotypes in many PTP-knockout mice associated with deficient or hyperactive immune states and severe hematopoietic abnormalities suggest a crucial role of PTPs in maintaining immune balance [3].

In mice, deficient for T-cell protein tyrosine phosphatase (TC-PTP; EC 3.1.3.48), a product of the *ptpn2* gene, multiple defects in the lymphoid lineage were reported [4]. Analysis of T-cell populations derived from the spleen of TC-PTP<sup>-/-</sup> mice revealed a 2- to 3-fold lower proliferation rate compared with TC-PTP<sup>+/+</sup> lymphocytes [5]. The numbers of CD4<sup>+</sup>CD8<sup>+</sup> T cells and pre-B-cells were decreased. B lymphocytes exhibited impaired T-cell dependent responses thereby suggesting for alterations in immune regulation. In contrast, a dramatic increase in number of colony-forming macrophage units was observed [4]. TC-PTP<sup>-/-</sup> mice developed an inflammatory disorder characterized by chronic myocarditis, gas-

tritis, and nephritis due to the widespread tissue mononuclear cell infiltration. The mouse also exhibited a drastic increase in levels of nitric oxide and inflammatory cytokines such as interferon- $\gamma$  (IFN- $\gamma$ ), tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), and interleukin-2 (IL-2) that resulted in massive myocyte and hepatic necrosis [6]. These findings therefore suggest for marked anti-inflammatory properties of TC-PTP in the immune response.

Previously, a solid support was obtained for a role of two PTPs in autoimmunity. There are lymphoid tyrosine phosphatase (LYP), which is encoded by the *PTPN22* gene and implicated in a broad spectrum of autoimmune disorders, and CD45, whose genetic alterations are linked to multiple sclerosis and likely to be linked to autoimmune hepatitis [7]. Due to the recent advances in genome-wide association (GWA) studies, a new member could be added to the list of PTPs involved in autoimmunity. There is TC-PTP, a product of the human *PTPN2* gene. In this review, we summarize current knowledge suggesting the implementation of TC-PTP in inflammatory and autoimmune diseases.

## 2. TC-PTP expression results in two isoforms

The *PTPN2* gene maps to chromosome 18p11.3-p11.2 [8]. The 100-kb gene comprises 10 exons, with exons 1–7 encoding a conservative PTP catalytic domain [9]. The promoter of *PTPN2* is shown to contain multiple regulatory sites, including two putative responsive elements each for c-myc and PEA3, recognition sequences for the Sp1 and AP2 transcription factors and putative NF $\kappa$ B- and APF-binding sites [10]. The *PTPN2* promoter also has a suppressor element that inhibits expression of TC-PTP, while switching from G1 to S phase of the cell cycle. This therefore suggests that the

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transcription of *PTPN2* is regulated by cell-cycle progression reaching a maximal level at G1. TC-PTP is ubiquitously expressed, with the highest expression levels in hematopoietic tissues [11]. Expression of TC-PTP could be stimulated by various agents including mitogen concanavalin A [12] and anti-inflammatory cytokine IL-10 [13].

Alternative splicing results in two forms of TC-PTP that share a highly conservative PTP catalytic domain consisted of 272 amino acids (a.a.) but differ in the C termini [14,15]. The catalytic domain of TC-PTP shares a higher homology (74% amino acid identity) with the catalytic domain of PTP1B, the first protein tyrosine phosphatase ever being identified and characterized [16]. The less abundant 48-kDa isoform of the enzyme comprises 415 amino acid (a.a.) residues and is present in the endoplasmic reticulum (ER) and also in the nuclear membrane [17]. This isoform has a hydrophobic C-terminal tail that is necessary for binding two proteins, p23 and p25, targeting the enzyme to the endoplasmic reticulum [18]. Interestingly, the C-terminal part of TC-PTP negatively influences the enzyme activity. The removal of the hydrophobic C terminus of TC-PTP resulted in a 30-fold increase in activity [19]. In addition, limited proteolysis of TC-PTP released a highly active 33-kDa fragment, which again could be inhibited by the addition of the non-catalytic C-terminal segment of the 45-kDa TC-PTP [20]. The autoinhibition is a likely consequence of intramolecular interactions.

The major 45-kDa form of TC-PTP consists of 387 a.a. The C-terminal region of the 45-kDa isoform has a specific bipartite nuclear localization sequence that targets the enzyme to the nucleus [21]. This C-terminal region of TC45 is also involved in binding to DNA [15]. The nuclear form is able to shuttle between the nucleus and the cytoplasm in response to extracellular stimuli, thereby providing access for the nuclear enzyme to a larger variety of substrates, compared to the cytoplasmic TC-PTP isoform [22,23].

### 3. Regulation of TC-PTP activity

Activity of PTPs is regulated *in vivo* through oxidation and reduction reactions involving an invariant cysteine in an active site sequence motif [I/V]HCxxGxxR[S/T] [24]. In the catalytic domain of TC-PTP, the invariant cysteine is located at position 216 [9]. Oxidation of the Cys216 is likely to be attributable to the inactivation of TC45 by reactive oxygen species in response to insulin [25]. Since TC-PTP is involved in negative regulation of insulin-mediated signaling, the reversible oxidation of the Cys216 could be crucial in regulating TC-PTP activity in peripheral tissues such as the muscles and liver, which are major targets for the insulin action [26]. However, whether this mechanism plays a pivotal role in the regulation of TC-PTP activity in immune cells needs to be elucidated.

Phosphorylation/dephosphorylation is a common mechanism for regulating PTPs activity. Cyclin-dependent protein kinases (CDKs) preferably CDK1/cyclin-B1 were found to phosphorylate the Ser304 in TC45 not in TC48 [27]. The phosphorylation was cell cycle-dependent, increasing as cells progressed from G<sub>2</sub> into mitosis. However, the physiological relevance of this phosphorylation remains unclear since, the Ser304 resides outside the catalytic domain of TC-PTP, and modification of this amino acid residue does not have any apparent effect on the activity or intracellular localization of the enzyme [27]. The role of phosphorylation in the modulation of TC-PTP activity needs further clarification. In the enzyme, other steady-state phosphorylation sites may exist. Otherwise, in addition to the Ser340, other sites may be phosphorylated in response to specific stimuli, and then, together with Ser-304 phosphorylation, these may regulate TC45.

### 4. Role of TC-PTP in myeloid cells

A range of stimulatory signals and intracellular protein substrates of TC-PTP varies depending on the tissue where this

enzyme is expressed. For example, in the muscles and liver, TC-PTP functions as a negative regulator of insulin receptor-mediated signaling [28]. In hematopoietic tissues, TC-PTP regulates cellular proliferation *via* several pathways, including the dephosphorylation of growth factor receptors such as the epidermal growth factor [29] and the platelet-derived growth factor [30]. In myeloid cell lines, the enzyme downregulates colony-stimulating factor 1 (CSF-1)-mediated signaling [31]. CSF-1 binding to the CSF-1 receptor causes the autophosphorylation of the receptor molecule that leads to the recruitment of Src family kinases and other signaling molecules including Erk kinase, a key regulator of differentiation in a number of cell systems [32]. Erk activation then stimulates the proliferation of macrophage progenitor cells and their differentiation to mature macrophages [33]. TC-PTP dephosphorylates Tyr807 of the CSF-1 receptor, whose phosphorylation is shown to be crucial in the activation of Erk [34].

The treatment of human monocytes with CSF-1 was found to result in the tyrosine phosphorylation of p52<sup>Shc</sup> adaptor protein and its association with Grb2, which is required in GSF-1-mediated macrophage differentiation [35]. 45-kDa form of TC-PTP is able to specifically recognize and then dephosphorylate p52<sup>Shc</sup> phosphorylated on Tyr239 thereby preventing association of p52<sup>Shc</sup> with Grb2 [23]. Thus, TC-PTP could suppress CSF-1-dependent stimulation of the macrophage lineage through two mechanisms, i.e. *via* dephosphorylation of the CSF-1 receptor and the dephosphorylation of p52<sup>Shc</sup>. In addition, TC-PTP could also inhibit the development of macrophages through direct dephosphorylation and down-regulation of Src kinases [33].

### 5. Role of TC-PTP in lymphoid cells

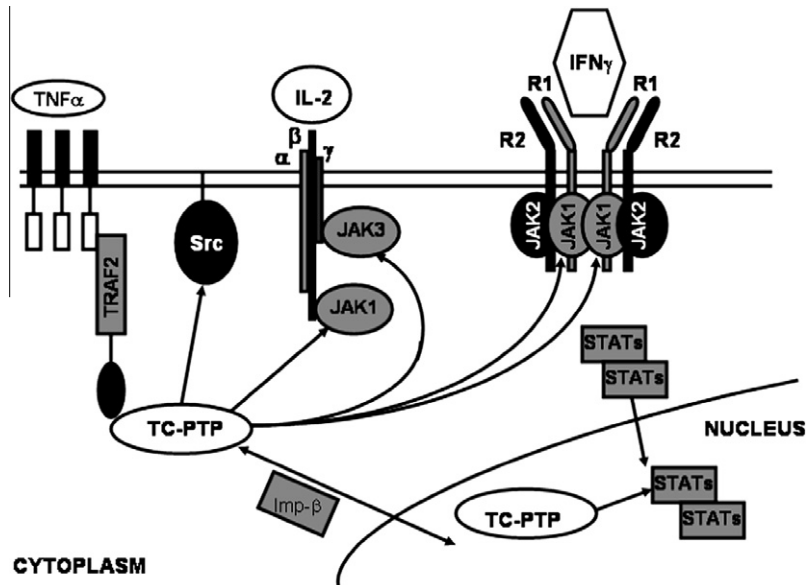
In lymphoid cells, TC-PTP possesses anti-inflammatory activity through the suppression of inflammatory response induced by a variety of proinflammatory cytokines. The enzyme downregulates IL-2 and IFN- $\gamma$ -mediated signaling (that leads to the activation of macrophages and induction of the inflammatory response) through dephosphorylation of JAK1 and JAK3 tyrosine kinases that participate in the activation of receptors for these cytokines (Fig. 1) [37]. In addition, both cytoplasmic and nuclear TC-PTP could suppress IFN- $\gamma$ -mediated gene expression, causing the dephosphorylation of the signal transducer and activator of transcription 1 (STAT-1) [38,39].

TC-PTP downregulates the expression of TNF- $\alpha$  and TNF- $\alpha$ -mediated signaling in immune cells. TC-PTP suppresses TNF- $\alpha$ -mediated activation of immune cells *via* binding to TRAF2, a component of the TNF- $\alpha$ -signaling pathway, which targets TC-PTP to Src kinase and then leads to the subsequent inhibition of Erk-dependent signaling (Fig. 1) [36].

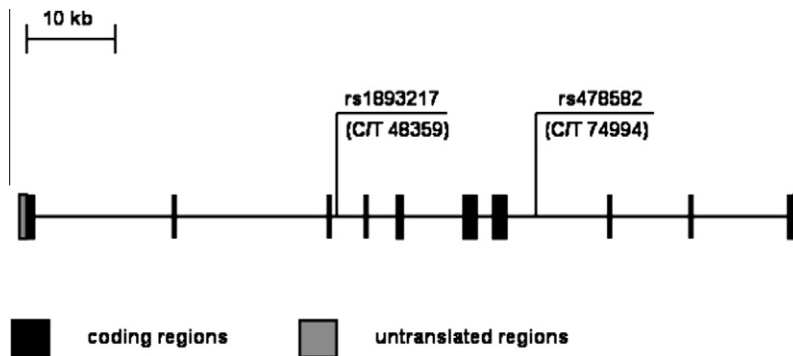
The nuclear 45-kDa TC-PTP isoform implies in the dephosphorylation of the downstream JAK effectors, the STATs that regulate transcription of many genes. In addition to STAT1, STAT3, STAT5a, and STAT5b are found to be substrates for TC-PTP [40,41]. Dephosphorylation of STAT5a/b by TC-PTP is associated with blocking prolactin signaling [40]. Dephosphorylation of STAT3 by TC-PTP suppresses the signaling pathway mediated by IL-6, a pleiotropic cytokine, which plays a role in hematopoiesis, immune system and inflammatory reactions [41].

### 6. Genetic analyses reveal a role of *PTPN2* as a putative susceptibility gene for general autoimmunity

In two recent GWA scans performed in Europeans, a highly significant evidence for association with type 1 diabetes (T1D) was obtained for the chromosome region 18p11 containing the *PTPN2* gene [42,43]. Two T1D-associated single nucleotide polymor-



**Fig. 1.** Substrates targeted by T-cell protein tyrosine phosphatase (TC-PTP). TC-PTP negatively regulates JAK/STAT signaling that leads to the induction of the inflammatory reaction. TC-PTP downregulates signaling mediated by proinflammatory cytokines (IFN- $\gamma$ , TNF- $\alpha$ , and IL-2) through the dephosphorylation of JAK1, JAK3 and Src kinases, which activate downstream signaling pathways induced by binding of the cytokines to their receptors. As a part of the complex with Importin- $\beta$  (Imp- $\beta$ ), TC-PTP is able to shuttle from the cytoplasm to the nucleus, where the enzyme can dephosphorylate STATs, the molecules involved in signal transduction and activation of the transcription of a number of genes.



**Fig. 2.** Structure of the protein tyrosine phosphatase, non-receptor 2 (*PTPN2*) gene encoding T-cell protein tyrosine phosphatase. The *PTPN2* gene encompasses nearly 100 kb on chromosome 18p11 and includes 10 exons. The location of two SNPs associated with type 1 diabetes is shown.

phisms (SNPs) of *PTPN2* are situated in intronic portions of the gene (e.g., rs478582 (C/T) and rs1893217 (C/T) in intron 3 and 7, respectively) (Fig. 2). Both SNPs are located within a 114-kb-long linkage disequilibrium block that includes the entire *PTPN2* gene [44]. The association between *PTPN2* and T1D was then confirmed by GWA analysis of the independent cohort of US Caucasians and the subsequent meta-analysis of the joint (American + European) population sample (Odds Ratio (OR) = 1.14;  $p = 8.73 \times 10^{-8}$  for marker rs2542151 (A/C) located near the *PTPN2* gene) [45].

Both markers within *PTPN2* (rs1893217 and rs478582) that showed association with T1D were found to be associated with Graves' disease (OR = 1.13 ( $p = 0.0251$ ) for rs1893217 and OR = 0.91 ( $p = 0.0239$ ) for rs478582) [42]. The marker rs2542151 (A/C) located near the *PTPN2* gene also showed a strong association with Crohn's disease (OR = 1.15;  $p = 3.16 \times 10^{-8}$ ) in a GWA-based search analysis of 1182 affected and 2024 control individuals from UK [46]. The association between *PTPN2* and Crohn's disease was then replicated by several large-scale population studies in Caucasians [47–50].

These findings suggest that the *PTPN2* gene could represent a general autoimmunity locus implicated in susceptibility to several autoimmune diseases. Although additional studies are required to replicate this association, as well as to analyze the functionality of the disease-associated markers, recent genetic analyses provide intriguing evidence for the presence of a genetic locus, with a likely involvement of *PTPN2*, that controls predisposition to autoimmunity on chromosome 18p11.

## 7. Conclusion

In summary, strong evidence in support of the implication of *PTPN2* in organ-specific autoimmunity is obtained for Crohn's disease and T1D. Findings about the role of this gene in susceptibility to Graves' disease require further confirmation. Disease-associated markers at *PTPN2* are located in non-coding regions, and their function is unclear. To date, the etiological variant(s) in *PTPN2* is

unknown, and therefore should be revealed through resequencing and other robust genetic approaches.

Generally, a role of the predisposing variants of *PTPN2* in autoimmunity might be attributed to less efficient suppression of the inflammatory response. The hypothesis can be supported by deleterious outcomes observed in the murine models deficient for *ptpn2* [4–6]. The major protective role of TC-PTP in autoimmunity is likely to be referred to the anti-inflammatory activity of this enzyme in the immune system and its inhibitory effects against macrophage hyperactivity. TC-PTP suppresses expression of major proinflammatory cytokines such as TNF- $\alpha$  and IL-6 whose overproduction accomplishes acute phase response in the pathogenesis of Crohn's disease, rheumatoid arthritis and other autoimmune inflammatory disorders [51]. Etiological variants of the *PTPN2* gene that are needed to be discovered and functionally characterized could alter the function of TC-PTP as an anti-inflammatory trigger and hence predispose to impaired immune response, a prerequisite of many autoimmune-related inflammatory diseases.

A role of the *PTPN22* gene encoding for lymphoid tyrosine phosphatase in a broad spectrum of autoimmune disorders was recently established [52]. Therefore, *PTPN2* is a likely candidate to extend the list of members of the large family of non-receptor PTPs involved in general autoimmunity.

## References

- [1] Alonso A, Sasin J, Bottini N, Friedberg I, Friedberg I, Osterman A, et al. Protein tyrosine phosphatases in the human genome. *Cell* 2004;117:699–711.
- [2] Mustelin T, Vang T, Bottini N. Protein tyrosine phosphatases and the immune response. *Nat Rev Immunol* 2005;5:43–57.
- [3] Hendriks WJAJ, Elson A, Harroch S, Stoker AW. Protein tyrosine phosphatases: functional inferences from mouse models and human diseases. *FEBS J* 2008;275:816–30.
- [4] You-Ten KE, Muise ES, Itie A, Wolfkamp SC, Henckaerts L, Schreiber S, et al. Impaired bone marrow microenvironment and immune function in T cell protein tyrosine phosphatase-deficient mice. *J Exp Med* 1997;186:683–93.
- [5] Dupuis M, De Jesus Ibarra-Sanchez M, Tremblay ML, Duplay P. Gr-1<sup>+</sup> myeloid cells lacking T cell protein tyrosine phosphatase inhibit lymphocyte proliferation by an IFN- $\gamma$ - and nitric oxide-dependent mechanism. *J Immunol* 2003;171:726–32.
- [6] Heinonen KM, Nestal FP, Newell EW, Charette G, Seemayer TA, Tremblay ML, et al. T-cell protein tyrosine phosphatase deletion results in progressive systemic inflammatory disease. *Blood* 2004;103:3457–64.
- [7] Vang T, Miletic AV, Arimura Y, Tautz L, Rickert RC, Mustelin T. Protein tyrosine phosphatases in autoimmunity. *Annu Rev Immunol* 2008;26:29–55.
- [8] Johnson CV, Cool DE, Glaccum M, Green N, Fischer EH, Bruskin A, et al. Isolation and mapping of human T-cell protein tyrosine phosphatase sequences: localization of genes and pseudogenes discriminated using fluorescence hybridization with genomic versus cDNA probes. *Genomics* 1993;16:619–29.
- [9] Cool DE, Tonks NK, Charbonneau H, Walsh KA, Fischer EH, Krebs EG. cDNA isolated from a human T-cell library encodes a member of the protein-tyrosine-phosphatase family. *Proc Natl Acad Sci USA* 1989;86:5257–61.
- [10] Wee C, Muise ES, Coquelle O, Ennis M, Wagner J, Lemieux N, et al. Promoter analysis of murine T-cell protein tyrosine phosphatase gene. *Gene* 1999;237:351–60.
- [11] Ibarra-Sanchez MJ, Wagner J, Ong MT, Lampron C, Tremblay ML. Murine embryonic fibroblasts lacking TC-PTP display delayed G1 phase trough detective NF- $\kappa$ B activation. *Oncogene* 2001;20:4728–39.
- [12] Rajendrakumar GV, Radha V, Swamp G. Stabilization of a protein-tyrosine phosphatase mRNA upon mitogenic stimulation of T lymphocytes. *Biochim Biophys Acta* 1993;1216:205–12.
- [13] Williams L, Bradley L, Smith A, Foxwell B. Signal transducer and activator of transcription 3 is the dominant mediator of anti-inflammatory effects of IL-10 in human macrophages. *J Immunol* 2004;172:567–76.
- [14] Lorenzen JA, Dadabay CY, Fischer EH. COOH-terminal sequence motifs target the T cell protein tyrosine phosphatase to the ER and nucleus. *J Cell Biol* 1995;131:631–43.
- [15] Kamatkar S, Radha V, Nambirajan S, Reddy RS, Swarup G. Two splice variants of a tyrosine phosphatase differ in substrate specificity, DNA binding and subcellular location. *J Biol Chem* 1996;271:26755–61.
- [16] Tonks NK, Diltz CD, Fischer EH. Purification of the major protein-tyrosine-phosphatases of human placenta. *J Biol Chem* 1988;263:6722–30.
- [17] Iversen LF, Moller KB, Pedersen AK, Petersen GH, Petersen AS, Andersen HS, et al. Structure determination of T cell protein tyrosine phosphatase. *J Biol Chem* 2002;277:19982–90.
- [18] Gupta V, Swarup G. Evidence for a role of transmembrane protein p25 in localization of protein tyrosine phosphatase TC48 to the ER. *J Cell Sci* 2006;119:1703–14.
- [19] Zander NF, Cool DE, Diltz CD, Rohrschneider LR, Krebs EG, Fischer EH, et al. Suppression of v-fms-induced transformation by overexpression of a truncated T-cell protein tyrosine phosphatase. *Oncogene* 1993;8:1175–82.
- [20] Hao LN, Tiganis T, Tonks NK, Charbonneau H. The noncatalytic C-terminal segment of the T cell protein tyrosine phosphatase regulates activity via an intramolecular mechanism. *J Biol Chem* 1997;272:22–9.
- [21] Bourdeau A, Dube N, Tremblay ML. Cytoplasmic protein tyrosine phosphatases, regulation and function: the roles of PTP1B and TC-PTP. *Curr Opin Cell Biol* 2005;17:203–9.
- [22] Tillmann U, Wagner J, Boerboom D, Westphal H, Tremblay ML. Nuclear localization and cell cycle regulation of a murine protein tyrosine phosphatase. *Mol Cell Biol* 1994;14:3030–40.
- [23] Tiganis T, Bennett AM, Ravichandran KS, Tonks NK. Epidermal growth factor receptor and the adaptor protein p52Shc are specific substrates of T-cell protein tyrosine phosphatase. *Mol Cell Biol* 1998;18:1622–34.
- [24] Fauman FB, Saper MA. Structure and function of the protein tyrosine phosphatases. *Trends Biochem Sci* 1996;21:413–7.
- [25] Meng TC, Buckley DA, Galic S, Tiganis T, Tonks NK. Regulation of insulin signaling through reversible oxidation of the protein-tyrosine phosphatases TC45 and PTP1B. *J Biol Chem* 2004;279:37716–25.
- [26] Galic S, Hauser C, Kahn BB, Haj FG, Neel BG, Tonks NK, et al. Coordinated regulation of insulin signaling by the protein tyrosine phosphatases PTP1B and TCPTP. *Mol Cell Biol* 2005;25:819–29.
- [27] Bukczynska P, Klingler-Hoffmann M, Mitchelhill KI, Haj FG, Neel BG, Tonks NK, et al. The T-cell protein tyrosine phosphatase is phosphorylated on Ser-304 by cyclin-dependent protein kinases in mitosis. *Biochem J* 2004;380:939–49.
- [28] Galic S, Klingler-Hoffmann M, Fodero-Tavoletti MT, Puryer MA, Meng TC, Tonks NK, et al. Regulation of insulin receptor signaling by the protein tyrosine phosphatase TCPTP. *Mol Cell Biol* 2003;23:2096–108.
- [29] Tiganis T, Flint AJ, Adam SA, Tonks NK. Association of the T-cell tyrosine phosphatase with nuclear import factor p97. *J Biol Chem* 1998;272:21548–57.
- [30] Persson C, Savenhed C, Bourdeau A, Tremblay ML, Markova B, Böhmer FD, et al. Site-selective regulation of platelet-derived growth factor beta receptor tyrosine phosphorylation by T-cell protein tyrosine phosphatase. *Mol Cell Biol* 2004;24:2190–201.
- [31] Simoncic PD, Bourdeau A, Lee-Loy A, Rohrschneider LR, Tremblay ML, Stanley ER, et al. T-cell protein tyrosine phosphatase (Tcptp) is a negative regulator of colony-stimulating factor 1 signaling and macrophage differentiation. *Mol Cell Biol* 2006;26:4149–60.
- [32] Stanley ER, Berg KL, Einstein DB, Lee PS, Pixley FJ, Wang Y, et al. Biology and action of colony-stimulating factor-1. *Mol Reprod Dev* 1997;46:4–10.
- [33] Gobert Gosse S, Bourgin C, Liu WQ, Garbay C, Mouchiroud G. M-CSF stimulated differentiation requires persistent MEK activity and MAPK phosphorylation independent of Grb2-Sos association and phosphatidylinositol 3-kinase activity. *Cell Signal* 2005;17:1352–62.
- [34] Csar XF, Wilson NJ, McMahon KA, Marks DC, Beecroft TL, Ward AC, et al. Proteomic analysis of macrophage differentiation p46/52(Shc) tyrosine phosphorylation is required for CSF-1-mediated macrophage differentiation. *J Biol Chem* 2001;276:26211–7.
- [35] Kharbanda S, Saleem A, Yuan Z, Emoto Y, Prasad KV, Kufe D. Stimulation of human monocytes with macrophage colony-stimulating factor induces a Grb2-mediated association of the focal adhesion kinase pp125FAK and dynamin. *Proc Natl Acad Sci USA* 1995;92:6132–6.
- [36] Van Vliet C, Bukczynska PE, Puryer MA, Sadek CM, Shields BJ, Tremblay ML, et al. Selective regulation of tumor necrosis factor-induced Erk signaling by Src family kinases and the T cell protein tyrosine phosphatase gene. *Nat Immunol* 2005;6:253–60.
- [37] Simoncic PD, Lee-Loy A, Barber DL, Tremblay ML, McGlade CJ. The T cell protein phosphatase is a negative regulator of Janus family kinases 1 and 3. *Curr Biol* 2002;12:446–53.
- [38] ten Hoeve J, de Jesus Ibarra-Sanchez M, Fu Y, Zhu W, Tremblay M, David M, et al. Identification of a nuclear Stat1 protein tyrosine phosphatase. *Mol Cell Biol* 2002;22:5662–8.
- [39] Zhu W, Mustelin T, David M. Arginine methylation of STAT1 regulates its dephosphorylation by T cell protein tyrosine phosphatase. *J Biol Chem* 2002;277:35787–90.
- [40] Aoki N, Matsuda T. A nuclear protein-tyrosine phosphatase TC-PTP is a potential negative regulator of the PRL-mediated signaling pathway: dephosphorylation and deactivation of signal transducer and activator of transcription 5a and 5b by TC-PTP in nucleus. *Mol Endocrinol* 2002;16:58–69.
- [41] Yamamoto T, Sekine Y, Kashima K, Kubota A, Sato N, Aoki N, et al. The nuclear isoform of protein-tyrosine phosphatase TC-PTP regulates interleukin-6-mediated signaling pathway through STAT3 dephosphorylation. *Biochem Biophys Res Commun* 2002;297:811–7.
- [42] Todd JA, Walker NM, Cooper JD, Smyth DJ, Downes K, Plagnol V, et al. Robust associations of four new chromosome regions from genome-wide analyses of type 1 diabetes. *Nat Genet* 2007;39:857–64.
- [43] Wellcome Trust Case Control Consortium. Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. *Nature* 2007;447:661–78.
- [44] The International HapMap Consortium. A haplotype map of the human genome. *Nature* 2005;437:1299–320.
- [45] Cooper JD, Smyth DJ, Smiles AM, Plagnol V, Walker NM, Allen JE, et al. Meta-analysis of genome-wide association study data identifies additional type 1 diabetes risk loci. *Nat Genet* 2008;40:1399–401.

- [46] Parkes M, Barrett JC, Prescott NJ, Tremelling M, Anderson CA, Fisher SA, et al. Sequence variants in the autophagy gene *IRGM* and multiple other replicating loci contribute to Crohn's disease susceptibility. *Nat Genet* 2007;39:830–2.
- [47] Franke A, Balschun T, Karlsen, Hedderich J, May S, Lu T, et al. Replication of signals from recent studies of Crohn's disease identifies previously unknown disease loci for ulcerative colitis. *Nat Genet* 2008;40:713–5.
- [48] Smyth DJ, Plagnol V, Walker NM, Cooper JD, Downes K, Yang JH, et al. Shared and distinct genetic variants in type 1 diabetes and celiac disease. *N Engl J Med* 2008;359:2767–77.
- [49] Barrett JC, Hansoul S, Nicolae DL, Cho JH, Duerr RH, Rioux JD, et al. Genome-wide association defines more than 30 distinct susceptibility loci for Crohn's disease. *Nat Genet* 2008;40:955–62.
- [50] Weersma PK, Stokkers PC, Cleynen I, Wolfkamp SC, Henckaerts L, Schreiber S, et al. Confirmation of multiple Crohn's disease susceptibility loci in a large Dutch–Belgian cohort. *Am J Gastroenterol* 2009;104:630–8.
- [51] Cronstein BN. Interleukin-6 – a key mediator of systemic and local symptoms in rheumatoid arthritis. *Bull NYU Hosp Jt Dis* 2007;65(Suppl. 1):S11–5.
- [52] Vang T, Miletic AV, Bottini N, Mustelin T. Protein tyrosine phosphatase PTPN22 in human autoimmunity. *Autoimmunity* 2007;40:453–61.