

REVIEW

Discovery and development of natural heat shock protein 90 inhibitors in cancer treatment

Institute of Materia Medica, Chinese Academy of Medical Sciences Chinese Pharmaceutical Association

Acta Pharmaceutica Sinica B

www.elsevier.com/locate/apsb www.sciencedirect.com

Yong Li^{a,b}, Dongmei Zhang^{a,b}, Jun Xu^{a,b}, Junmin Shi^{a,b}, Lei Jiang^c, Nan Yao^{a,b}, Wencai Ye^{a,b,*}

^aCollege of Pharmacy, Jinan University, Guangzhou 510632, China ^bGuangdong Province Key Laboratory of Pharmacodynamic Constituents of Traditional Chinese Medicine and New Drugs Research, Jinan University, Guangzhou 510632, China ^cSchool of Biomedical Science, The Chinese University of Hong Kong, Hong Kong, China

Received 13 February 2012; revised 24 March 2012; accepted 30 March 2012

KEY WORDS

Heat shock protein 90 (Hsp90); Tumor; Hsp90 inhibitors; Geldanamycin **Abstract** Heat shock protein 90 (Hsp90) is a highly conserved molecular chaperone that plays a vital role in the signal transduction of cancers. Hsp90 inhibitors are able to inhibit Hsp90 or the complex of Hsp90 and co-chaperones resulting in the degradation of Hsp90-dependent client proteins through the ubiquitination-proteasome pathway, thereby leading to the growth inhibition of tumor cells. This review will briefly discuss the molecular structure and biological function of Hsp90, and focus on a summary of recent progress in the development and testing of natural Hsp90 inhibitors and their different means by which they interact with Hsp90.

© 2012 Institute of Materia Medica, Chinese Academy of Medical Sciences and Chinese Pharmaceutical Association. Production and hosting by Elsevier B.V. All rights reserved.

*Corresponding author. Tel.: +86 20 85220936; fax: +86 20 85221559. E-mail address: chywc@yahoo.com.cn (Wencai Ye).

2211-3835 © 2012 Institute of Materia Medica, Chinese Academy of Medical Sciences and Chinese Pharmaceutical Association. Production and hosting by Elsevier B.V. All rights reserved.

Peer review under responsibility of Institute of Materia Medica, Chinese Academy of Medical Sciences and Chinese Pharmaceutical Association. http://dx.doi.org/10.1016/j.apsb.2012.03.009



Production and hosting by Elsevier

1. Introduction

Heat shock proteins (Hsps) are a family of highly conserved stress proteins which can be induced by environmental stress such as heat, hypoxia, DNA damage or UV radiation to regulate cell metabolism and protect prokaryotic and eukaryotic cells from harmful exogenous stimulation. They are also known as molecular chaperones due to their ability to maintain proper protein folding, and facilitate the assembly of protein polymers and the formation of mature proteins.

The highly conserved molecular chaperone Hsp90 is one of the most abundant of proteins in almost all eukaryotic cells, accounting for 1–2% of total cellular proteins¹. Hsp90 has two major cytoplasmic isoforms Hsp90 α (inducible form) and Hsp90 β (constitutive form), as well as a Raf-associated protein Hsp90N which is associated with cellular transformation². Other Hsp90 analogues include glucose regulated protein 94 (Grp94) in the endoplasmic reticulum and tumor necrosis factor receptor-associated protein 1 (TRAP1) in the mitochondrial matrix^{1,2}.

Hsp90, in general, exists as a homodimer $(\alpha/\alpha \text{ or } \beta/\beta)$ in cells. The structure of Hsp90 is composed of three domains: the highly conserved NH₂-terminal nucleotide binding domain (25 kDa, Hsp90N), the middle domain (35 kDa, Hsp90M) and the C-terminal domain (10 kDa, Hsp90C). Hsp90N is an important binding site for ATP/ADP, which is associated with forming the complex of Hsp90 and co-chaperones *via* conformational transition³. Hsp90M is the binding site for nuclear localization signal and client proteins, and is involved in the specific recognition of client proteins and regulating molecular chaperones for appropriate substrate activation including ATP hydrolysis⁴. The C-terminal of Hsp90 is the site of dimerization. This region contains a pentapeptide domain (MEEVD) which is also the binding site of co-chaperones of Hsp90 such as Hop and Sti 1⁵.

In eukaryotic plasma, Hsp90 is considered to mediate the folding, stabilization, activation and assembly of its client proteins, including steroid receptors, protein kinases and transcription factors, through the formation of complexes with other co-chaperones such as Hsp70, Hop and p23⁶. A client protein is bound to Hsp70/Hsp40 to form an early complex, and then Hsp90 along with the co-chaperone Hop binds to this early complex to form an intermediate complex. When ADP is replaced by ATP, Hsp90 undergoes a conformational change which releases Hsp70/Hsp40 and Hop, thus allowing the ATP-dependent association of other co-chaperones, including p23 and the immunophilins to form a mature complex. When the ATP-binding site of Hsp90 is occupied by competitive inhibitors, the Hsp90 complex is disrupted and the client proteins can be degraded through the ubiquitination-proteasome pathway^{6,7}.

Cancer is a disease associated with genetic instability which allows cancer cells to acquire six distinguishing characteristics, including self-sufficiency in growth signaling, resistance to apoptosis, insensitivity to growth inhibitory signaling, sustained angiogenesis, tissue invasion and metastasis, and limitless proliferative potential. Many signaling proteins related to these hallmarks of cancer are client proteins of Hsp90, such as Akt, Her-2, p53, CDK4, Raf-1, IKK, HIF-1 α , Src (Table 1). In addition, the rapid growth of cancer cells requires the participation of large quantities of cytosolic heat shock proteins. Thus, it is useful to inhibit cancer growth and metastasis by suppressing the formation of complexes of Hsp90 and its client proteins, which

 Table 1
 Relationship between Hsp90 client proteins and hallmarks of cancer.

Hallmark of cancer	Hsp90 client protein
Evasion of apoptosis	Akt,Rip,P53,Survivin,
	Apaf-1,Bcl-2,IGF-IR
Sustained angiogenesis	VEGFR, HIF1,Akt,
	Fit-3,FAK,Src
Limitless replicative potential	n-TERT, telomerase
Tissue invasion and metastasis	c-MET,MMP2
Self-sufficiency in growth signals	EGFR/Her-2,Raf,
	Bcr-Abl,ErbB-2,
	Src,Akt,MEK
Insensitivity to anti-growth	Plk-1,Cdk4,Cdk6,
signals	Myt-1,cyclin D

not only reduces the tolerance of cancer cells to chemotherapeutics, but also synergistically enhances the efficacy of multiple anticancer drugs. Hsp90 has been considered as a potent molecular target in the development of new anticancer agents. Since the first natural anticancer drug vinblastine was applied in clinic, many chemotherapeutic agents from natural products have been discovered and developed, including paclitaxel, camptothecin, epothilone, bleomycin and cantharidin, *etc.* Current Hsp90 inhibitors are derived from natural products, such as geldanamycin, radicicol, novobicon, epigallocatechin-3-gallate, celastrol and hypericin (Fig. 1). This review will briefly describe the discovery and development of natural product Hsp90 inhibitors.

2. Inhibitors of Hsp90 N-terminal ATP binding domain

2.1. Geldanamycin (GA) and derivatives

Geldanamycin(1) (Fig. 1), originally isolated from the culture filtrates of Streptomyces hygroscopicus, inhibits the growth of protozoa, bacteria and fungus⁸. GA was initially found to arrest cancer cell proliferation by inhibiting Src tyrosine kinase, but it was unable to directly inhibit the activity of purified kinase⁹. Further studies revealed that the anti-proliferative activity of GA resulted from the binding of GA to Hsp90. GA inhibits client protein binding to Hsp90 through competitive coupling with ATP/ADP binding "pocket" of the Hsp90 N-terminal domain, leading to the degradation of the client proteins via the ubiquitination-proteasome pathway (Fig. 2). The selectivity of GA for Hsp90 is due to the special ATP binding "pocket" of Hsp90 as compared to other nucleotidebinding proteins. Structure-activity analysis has revealed that the 2,3-double bond of GA was indispensible for stable conformation of Hsp90; 11-hydroxy and 17-methoxy moieties contribute to formation of hydrogen bonds and increase the binding affinity of GA or its derivatives for Hsp90, thereby leading to an enhancement of anticancer activities¹⁰. These results suggest that the effectiveness of Hsp90 inhibitors is closely related to their binding ability with Hsp90.

The structure of geldanamycin has been modified to reduce its hepatotoxicity and increase its water-solubility. The site of focus is the 17 position of the benzoquinone group to generate more potent derivatives, including 17-allylamino-17-demethoxy geldanamycin (17-AAG, 2) (Fig. 1) and 17-N,N-dimethylaminoethylamino-17-demethoxy geldanamycin

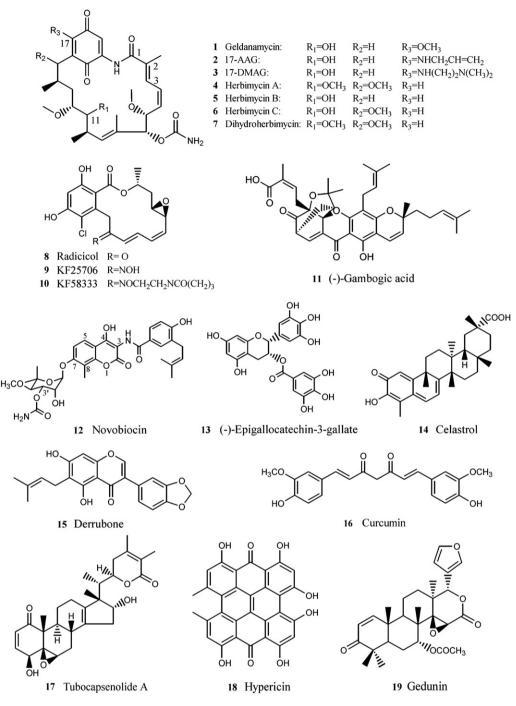


Figure 1 Chemical structures of Hsp90 inhibitors.

(17-DMAG, 3) (Fig. 1) which are in phase II/III or I clinical trials respectively (Table 2).

17-AAG (17-allyamino-17-demethoxygeldanamycin), in which methoxymoiety of GA is replaced by an ally amino group, is the first Hsp90 inhibitor used in a clinical study with hepatotoxicity is apparently less than that of geldanamycin. 17-AAG can be metabolized by cytochrome P450 CYP3A4 to 17-amino-demethoxygeldanamycin (17-AG) *in vivo* which maintains Hsp90 inhibitory activity and with higher stability than 17-AAG¹¹. 17-AAG binds to purified full-length dimeric Hsp90 in the low micromolar range, whereas it inhibits tumor cell growth in the low nanomolar range. This difference could be explained by the

finding that benzoquinone ansamycins largely accumulated in tumor cells with high micromolar concentration¹². Further studies revealed that 17-AAG had a higher affinity for Hsp90 derived from the breast carcinoma cell line BT474 ($IC_{50}=6 \text{ nM}$) than Hsp90 from primary human renal epithelial cells ($IC_{50}=600 \text{ nM}$) and 400 nM) and normal fibroblasts ($IC_{50}=400 \text{ nM}$). This indicated that Hsp90 in tumor cells existed in the form of active multi-chaperone complexes with Hsp70, Hsp40, Hop or p23, whereas Hsp90 in normal cells was in an uncomplexed inactive form¹³. In contrast to this suggestion, a recent study proposed that the affinity between GA and Hsp90 alone or the Hsp90 complex was similar, and that GA and its derivatives inhibited

241

Hsp90 through slow but tight binding interaction with high affinity and low dissociation¹⁴. An alternative explanation was also provided by Maroney et al.¹⁵, as they revealed that the benzoquinone ring of ansamycins could be reduced to dihydroquinone by NAD(P)H: Quinone oxidoreductase 1. The reduced product had approximately 40-fold greater affinity for Hsp90 in vitro than its oxidized form and exhibited enhanced anticancer activity, which contributed to the accumulation of ansamycin dihydroquinones in cancer cells. Additionally, quantum chemical calculations and mutation analysis demonstrated that geldanamycin could undergo keto-enoltautomerization catalyzed by Hsp90 which enhanced its binding to Hsp90¹⁶. It was also found that Hsp90 did not catalyze the trans-cis isomerisation of GA and confirmed that the binding capability of the dihydroquinone form of GA and its analogues to Hsp90 was more potent than the affinity of their oxidized quinone forms¹⁷.

17-AGG (oxidized form) entered clinical trials in 1999, and its reduced form entered clinical trials in 2005. Further clinical research and application of 17-AAG was limited due to its

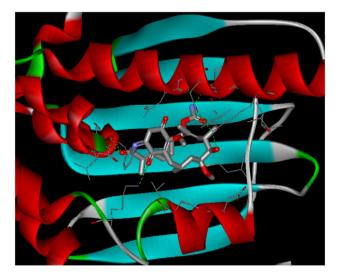


Figure 2 Binding mode of GA to Hsp90.

poor solubility. 17-DMAG, a new generation of Hsp90 inhibitor developed by Kosan Biosciences, has better water solubility, tissue dispersibility and bio-availability¹⁸. In comparison with 17-AAG, 17-DMAG possesses more potent anticancer activity against many cell lines *in vitro* and *in vivo*, and some scholars have suggested that this difference might be attributed to the greater affinity of Hsp90 for the benzoquinone ansamycins or their metabolites¹⁹. Unfortunately, 17-DMAG was also revealed to have gastrointestinal, liver, bone marrow, renal and gallbladder toxicity in dogs²⁰. Thus, Kosan Biosciences stopped the clinical development of 17-DMAG in the beginning of 2008. However, the further clinical tests are still being carried out by other research groups because of its favorable water solubility and various antitumor activities against different tumor types.

2.2. Herbimycin and its analogues

Herbimycin is also a member of benzoquinone ansamycin antibiotics isolated from the culture filtrates of *Streptomyces hygroscopicus*. Herbimycin A (4) (Fig. 1) was primarily isolated from the fermentation broth of *Streptomyces hygroscopicus* (No. AM-3672)²¹, and later herbimycin B (5), C (6) (Fig. 1) and dihydroherbimycin were obtained. Herbimycin was a selective inhibitor of a tyrosine kinase, and had an inhibitory effect on cancer angiogenesis and proliferation^{22,23}. Yang et al.²⁴ have compared the anti-proliferative activities of herbimycin analogues on Vero and B16-F10 cells as follows: herbimycin C>herbimycin A>dihydroherbimycin A (7) (Fig. 1). Further studies suggested that the underlying anti-tumor mechanism was also competitive binding to the N-terminal domain of Hsp90 to arrest the ATPase cycle²⁵. To date there is no clinical report on herbimycin and its analogues.

2.3. Radicicol (RA) and its analogues

Radicicol (8) (Fig. 1) belongs to the macrocyclic lactone antibiotics originally extracted from the filtrate of fungus *Monosporiumbonorden*. It can inhibit cancer cell proliferation and

Table 2H	sp90 inhibitors	in clinical	trials.
----------	-----------------	-------------	---------

	Phase	Indication	NCT ID
17-AAG	II/III	Multiple myeloma	NCT00514371, NCT00546780
	II	Pancreatic cancer	NCT00577889
	II	Breast cancer	NCT00096109, NCT00118092
	II	Melanoma	NCT00087386 NCT00088374
	II	Kidney tumor	
17-DMAG	Ι	Solid tumor	NCT00088868, NCT00089362
	Ι	Leukemia lymphoma	NCT01126502 ^a
Gambogic acid	IIa	Cancers	2004L00333(SFDA)
EGCG	Ι	Small cell lung cancer	NCT01317953 ^a
	II	Breast cancer	NCT00917735 ^a
	II	Prostate cancer	NCT00676780, NCT00596011ª
Curcumin	II	Pancreatic cancer	NCT00192842
	II	Breast cancer	NCT01042938
	I/II	Osteosarcoma	NCT00689195 ^a

^aThis study is currently recruiting participants.

angiogenesis *in vitro* and *in vivo* through depletion of $p60^{v-src}$ protein kinase and disruption of K-ras-activated aberrant signaling pathways^{26–28}. Similar to benzoquinone ansamycins, it can also competitively bind to the N-terminal domain of Hsp90 to disrupt Hsp90 complex formation²⁹. Results from isothermal titration calorimetry suggested that the affinity of radicicol to Hsp90N was about 50-fold greater than that of geldenamycins¹⁰. Radicicol showed more potent antitumor activity than geldenamycins *in vitro*, but weaker activity *in vivo*, which could be explained by the fact that the presence of epoxy and α , β , γ , δ -unsaturated carbonyl groups reduced the stability of radicicol³⁰.

Structure-activity relationship analysis indicated that the benzene hydroxyl and macrocyclic ring were required for biological activity of radicicol³¹. A series of stable radicicoloxime derivatives have been synthesized by means of reducing the nucleophilicity of unsaturated carbonyl groups. Soga et al.^{32,33} have indicated that KF25706 (9) (Fig. 1) could selectively deplete the client proteins of Hsp90 via inhibition of v-Src and K-ras-activated signaling pathways and KF58333 (10) (Fig. 1) could suppress VEGF expression and tumor angiogenesis through an HIF-1α-dependent pathway. These results indicate that the oxime moiety plays a significant role in sustaining the stability and enhancing the biological activity of radicicoloxime analogues. In contrast to geldanamycin and its derivatives, radicicol has no hepatotoxicity, which implies that hepatotoxicity of benzoquinone ansamycins is not a common characteristic of all Hsp90 inhibitors³⁴. Thus, radicicol and its analogues are promising drug candidates for further development as nonbenzoquinone ansamycin Hsp90 inhibitors. Unfortunately, there is no clinical report of these compounds to date.

2.4. Gambogic acid

Gambogic acid (11) (Fig. 1) is the main component of gamboge used for treating tumors or infections. It has been reported that gambogic acid was an inhibitor of Hsp90 through binding to the N-terminal domain of Hsp90 in a non-competitive manner similar to that of geldanamycin³⁵. A phase IIa clinical trial to explore tolerability and safety of gambogic acid in cancer patients *via* different intravenous administration methods was completed in 2008 in China (Table 2).

3. Inhibitors of Hsp90 C-terminal ATP-binding domain

3.1. Novobiocin and analogues

Novobiocin (12) (Fig. 1), a coumarin antibiotic isolated from several *Streptomyces* strains, exhibited anti-bacterial activity through binding with topoisomerase II to inhibit ATP hydrolysis³⁶. Given that Hsp90 is also an ATPase-dependent protein, novobiocin might be also an inhibitor of Hsp90. Novobiocin interacted with the Hsp90C ATP-binding domain to disrupt Hsp90 dimerization and induce conformational changes of the Hsp90 complex, finally leading to the degradation of client proteins *via* the ubiquitination-proteasome pathway. However, novobiocin at a concentration of 700 μ M induced depletion of Hsp90 clients in SKBr3 cells, which was much weaker than Hsp90N inhibitors including benzoquinone ansamycins and radicicol analogues³⁷. Further

reasonable modifications should be carried out to enhance the bioactivity of novobiocin.

The structure of novobiocin is composed of three domains, including benzamide moiety, a coumarin moiety and noviose moiety. Blagg et al.³⁸ have synthesized a series of analogues and elucidated their structure-activity relationships: 3-secondary amine and 7-noviose moieties were indispensable for their bioactivities; 8-methoxy was useful to increase the bioactivity but 5-substitution may decrease the bioactivity; and the coumarinlactone moiety was not critical for Hsp90 inhibition. 4-Hydroxyl and 3'-carbamate of noviose moieties could impair the inhibitory activity towards Hsp90 but were essential for inhibition of DNA topoisomerase³⁹. In contrast to Blagg's reports, Renoir et al.⁴⁰ found that antitumor activities of these derivatives were significantly enhanced through substituting a tosyl moiety at C-4 or C-7 position of coumarin rings and highlighted that C-4 and/or C-7 positions of coumarin may be required for degradation of Hsp90 client proteins. Currently, novibiocon has mainly been used to treat bacterial infection in the clinic.

3.2. (-)-Epigallocatechin-3-gallate (EGCG)

(-)-Epigallocatechin-3-gallate (EGCG) (13) (Fig. 1) is a major polyphenolcatechin of green tea and possesses excellent antitumor activity, although its mechanism has not yet been clarified. Palermo et al.41 first reported EGCG could inhibit aryl hydrocarbon receptor binding of Hsp90. Furthermore, Li et al.⁴² proposed that EGCG may disrupt the association of Hsp90 with co-chaperone Hsc70 and p23 via direct binding to the Hsp90 C-terminal domain, and degrade Hsp90-related client proteins including Akt, Cdk4, Raf-1, Her-2 and pERK. Additionally, EGCG was also proven to be able to inhibit the growth and development of tumors in vitro and in vivo by suppressing the expression of Hsp70 and Hsp90⁴³. Gasiewicz et al.⁴⁴ suggested that EGCG directly inhibited the dimerization and modulated the conformation of Hsp90 by binding at or adjacent to the C-terminal ATP-binding site, just not as N-terminal Hsp90 inhibitors with destabilization of the Hsp90 complex. These reports indicate that EGCG could treat various tumors and enhance the anti-cancer activity of chemotherapeutics. Recently, a phase I clinical trial is testing the safety of EGCG for extensive-stage small lung cancer in those who achieved the objective tumor response after firstline therapy (Table 2).

4. Inhibitors of Hsp90 and interaction with co-chaperones

Co-chaperones play a significant role in modulating the formation and maturation of Hsp90 and its client proteins. The cell division cycle protein 37 (CDC37) is an important cochaperone, which helps the efficient recruitment of client proteins (mainly protein kinases) to the chaperone machinery and facilitates the formation of mature client proteins. Therefore, inhibition of CDC37/Hsp90 complex may efficiently decrease the expression of client proteins in cancer cells.

Celastrol (14) (Fig. 1), also called tripterine, was isolated from the root extracts of Thunder God Vine which is used to treat inflammatory and autoimmune diseases (e.g., rheumatoid arthritis, sclerosis and systemic lupus erythematosus) in traditional Chinese medicine. It was recently reported to

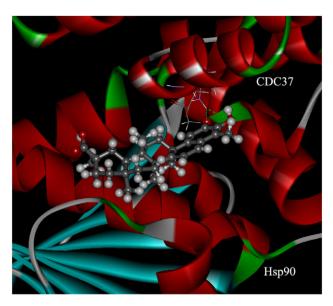


Figure 3 Binding mode of celastrol to Hsp90/CDC37.

possess antitumor activity⁴⁵. Zhang et al.⁴⁶ have found that celastrol showed an anti-pancreatic cancer effect in vitro and in vivo through disruption of the interaction of CDC37 with Hsp90 without affecting ATP binding to Hsp90, thereby inducing the degradation of Hsp90 client proteins (Fig. 3). According to results from ¹H, ¹⁵N-HSQC NMR, the thiol group of CDC37 N-terminal may react with the quinonemethide of celastrol through a Michanel addition at the C6 position, resulting in a change of CDC37 conformation and disruption of the Hsp90 complex formation. In addition, there is a large hydrophobic core at the interface of CDC37/Hsp90 complex and celastrol could disrupt the hydrophobic core at C293 and block the formation of Hsp90/CDC37 complex. Thus, the N-terminal of CDC37 and the middle domain may be the main interaction sites of celastol⁴⁷. Extracts of Thunder God Vineis used to treat rheumatoid arthritis, Crohn's disease and immunological rejection of kidney transplantation in IIb clinical trials, whereas the anti-tumor effect of celastrol has been studied only in preclinical tests.

Derrubone (15) (Fig. 1) is a natural prenylated isoflavone originally isolated from *Derris robusta*. In high-throughput screening, Blagg et al.⁴⁸ discovered that it induced potent cell growth inhibition and Her2 degradation in human breast cells (MCF-7 and SkBr3) without describing its specific mechanism. Furthermore, they synthesized chimeric analogues of novibiocin and derrubone sharing several key structural features, and evaluated their bioactivities. They also found that the iso-flavone motif of derrubone and the coumarin moiety of vovibiocin exerted different modes of action on binding to Hsp90, however, the C-3 position of the bicyclic ring of both was required for inhibiting Hsp90⁴⁹. Derrubone, as a lead Hsp90 inhibitor, is expected to have a promising development.

Curcumin (16) was isolated from Zingiberaceae family plants, including *Curcuma aromatica*, *Curcuma longa* and *Curcuma phaeocaulis*. It possesses anti-oxidant, anti-inflammatory and anti-tumor activities. Curcumin can also induce apoptosis in cancer cells by different signaling pathways and markedly enhance the sensitization of cells to other anticancer drugs⁵⁰. Jung et al.⁵¹ found that curcumin induced degradation of ErbB2 (a client of Hsp90), suggesting that Hsp90 or an

Hsp90 complex may be a target of curcumin. Lee et al.52 indicated that curcumin treatment resulted in dissociation of the Hsp90/p23 complex from human telomerase reverse transcriptase (hTERT), down-regulation of hTERT expression and inhibition of nuclear localization of telomerase to modulate the growth and development of cancer. Giommarelli et al.⁵³ observed that curcumin could directly inhibit Hsp90 and facilitate dissociation of client proteins (EGFR, Akt, surviving and Raf-1) from Hsp90 complexes. But given that curcumin had a low affinity for Hsp90 and led to the downregulation of Hsp70, it was indicated that the mechanism behind its Hsp90 inhibitory effect may be not identical to that of 17-AAG. In addition, it could also enhance the sensitivity of tumor cells to HDAC inhibitors and synergistically induce cellular growth inhibition and apoptosis. These results suggest that curcumin may be a promising agent to treat cancers in clinic. Currently, several clinical trials have been reported in the treatment of colorectal cancer, rectal cancer and pancreatic cancer (Table 2). Unfortunately, the application of curcumin in the clinic was limited owing to its poor bioavailability. It is anticipated that structural modifications will markedly improve its bioavailability and antitumor activity.

5. Other Hsp90 inhibitors

Tubocapsenolide A (17) (Fig. 1), a member of withanolide compounds, was isolated from *Tubocapsicum anomalum*. It could selectively induce thiol oxidation of Hsp90 and Hsp70 to destabilize and degrade Hsp90 client proteins, leading to cell cycle arrest and apoptosis in breast cancer MDA-MB-231 cells⁵⁴.

Hypericin (18) (Fig. 1) was isolated from the European plant *Hypericum perforatum*, which is used in anti-virus and anti-tumor treatments. Blank et al.⁵⁵ proposed that hypericin enhanced ubiquitinylation of Hsp90 to degrade Hsp90 client proteins. Phase I/II or II clinical trials have been conducted to test the effectiveness, safety, and tolerance of hypericin for patients with recurrent malignant gliomas or cutaneous T-cell lymphaomas, as well as psoriasis.

Gedunin (19) (Fig. 1) was derived from the Indian neem tree (*Azadirachta indica*). Hieronymus et al.⁵⁶ reported that gedunin modulated Hsp90 to inhibit proliferation of cancer cells, but it did not competitively bind to the Hsp90 ATP-binding pocket as seen with geldanamycin. Blagg et al.⁵⁷ synthesized 19 gedunin analogues in an effort to elucidate the structure–activity relationships of the gedunin–Hsp90 interaction. Unfortunately, anti-proliferative activities of these derivatives were much lower than that of gedunin in breast cancer cell lines MCF-7 and SkBr3. Further study is underway to obtain more gedunin analogues with potent chemotherapeutic properties.

6. Conclusions

A significant hallmark of cancer cells in the process of growth and development is infinite proliferation and immortalization, which requires associated chaperones for regulating protein synthesis. Studies also find Hsp90 and co-chaperones are abnormally expressed in cancer cells, thus it is expected that the identification and development of Hsp90 inhibitors will have a promising future. However, whether Hsp90 inhibitors also influence Hsp90 function in normal cells while they inhibit Hsp90 and Hsp90 complexes of tumor cells remains to be determined. Some studies have shown that 17-DMAG is widely distributed in different tissues, but it is unclear why it is metabolized much more slowly in cancer cells than normal cells⁵⁸. The affinity of ansamycins for Hsp90 in cancer cells is apparently 20–50 times stronger than it is in normal cells. This may be explained by the idea that Hsp90 in cancer cells is in the form of multi-chaperone complexes that modulate client protein maturation, while Hsp90 in normal cells is in an uncoupled inactive form. Another explanation is that the ATPase activity of Hsp90 in tumor cells is markedly higher than that in normal cells¹³. Nevertheless, the molecular mechanisms of interaction of Hsp90 inhibitors with Hsp90 warrants further study.

Most chemotherapeutics have serious side effects, such as geldanamycin and its derivatives demonstrating hepatotoxicity, poor solubility, nausea and vomiting in clinical use. In China, traditional Chinese medicines in cancer treatment offer unique advantages and are widely accepted and used in the clinic. The multiple effects of Hsp90 inhibitors on the treatment of tumor are consistent with the characteristics of traditional Chinese medicines in which multi-level, multi-target approaches are valued. Therefore, natural Hsp90 inhibitors can be useful in the enhancement of antitumor activity and decreased toxicity and resistance to drugs when combined with other chemotherapeutic agents. Hsp90 inhibitors have been isolated from Chinese herbs and yielded multiple bioactive constituents, including curcumin, hypericin, celastrol, etc. Although the biological activities of these compounds are less than those of geldanamycin and its analogues, their toxicities are also lower. Furthermore, these compounds not only enrich the chemical structure library of Hsp90 inhibitors, but also provide lead compounds for further synthesis of more selective and less toxic Hsp90 inhibitors. It is anticipated that the focus on discovery and development of Hsp90 inhibitors from traditional Chinese medicines through chemical separation and computer-aided drug design combined with high-throughput screening technology will vield further success in the development of Hsp90 inhibitors for use in cancer.

Acknowledgments

This work was supported by National Science Foundation of China (90913020 and 30901847), Science and Technology Program of China (2012ZX09103-101-053) and Science and Technology Star of Zhujiang of Guangzhou City (Dongmei Zhang).

References

- Csermely P, Schnaider T, Soti C, Prohaszka Z, Nardai G. The 90-kDa molecular chaperone family: structure, function, and clinical applications. A comprehensive review. *Pharmacol Ther* 1998;9:129–68.
- Sreedhar AS, Kalmar E, Csermely P, Shen YF. Hsp90 isoforms: functions, expression and clinical importance. *FEBS Lett* 2004;562: 11–5.
- Young JC, Moarefi I, Hartl FU. Hsp90: a specialized but essential protein-folding tool. J Cell Biol 2001;154:267–73.
- Meyer P, Prodromou C, Hu B, Vauqhan C, Roe SM, Panaretou B, et al. Structural and functional analysis of the middle segment of Hsp90: implications for ATP hydrolysis and client protein and cochaperone interactions. *Mol Cell* 2003;11:647–58.

- Minami Y, Kimura Y, Kawasaki H, Suzuki K, Yahara I. The carboxy-terminal region of mammalian Hsp90 is required for its dimerization and function *in vivo*. *Mol Cell Biol* 1994;14:1459–64.
- Whitesell L, Lindquist SL. Hsp90 and the chaperoning of cancer. Nat Rev Cancer 2005;5:761–72.
- Hahn JS. The Hsp90 chaperone machinery: from structure to drug development. BMB Rep 2009;42:623–30.
- 8. Deboer C, Meulman PA, Wnuk RJ, Peterson DH. Geldanamycin, a new antibiotic. *J Antibiot* 1970;**23**:442–7.
- Whitesell L, Shifrin SD, Schwab G, Neckers LM. Benzoquinonoid ansamycins possess selective tumoricidal activity unrelated to src kinase inhibition. *Cancer Res* 1992;52:1721–8.
- Roe SM, Prodromou C, O'Brien R, Ladbury JE, Piper PW, Pearl LH. Structural basis for inhibition of the Hsp90 molecular chaperone by the antitumor antibiotics radicicol and geldanamycin. J Med Chem 1999;42:260–6.
- Sausville EA, Tomaszewski JE, Ivy P. Clinical development of 17-allylamino, 17-demethoxygeldanamycin. *Curr Cancer Drug Targets* 2003;3:377–83.
- Guo WC, Reigan P, Siegel D, Zirrolli J, Gustafson D, Ross D. The bioreduction of a series of benzoquinone ansamycins by NAD(P)H: quinone oxidoreductase 1 to more potent heat shock protein 90 inhibitors, the hydroquinone ansamycins. *Mol Pharmacol* 2006;**70**:1194–203.
- Kamal A, Thao L, Sensintaffar J, Zhang L, Boehm MF, Fritz LC, et al. A high-affinity conformation of Hsp90 confers tumour selectivity on Hsp90 inhibitors. *Nature* 2003;425:407–10.
- Gooljarsingh LT, Fernandes C, Yan K, Zhang H, Grooms M, Johanson K, et al. A biochemical rationale for the anticancer effects of Hsp90 inhibitors: slow, tight binding inhibition by geldanamycin and its analogues. *Proc Natl Acad Sci USA* 2006;103:7625–30.
- Maroney AC, Marugan JJ, Mezzasalma TM, Barnakov AN, Garrabrant TA, Weaner LE, et al. Dihydroquinone ansamycins: toward resolving the conflict between low *in vitro* affinity and high cellular potency of geldanamycin derivatives. *Biochemistry* 2006;45:5678–85.
- Lee YS, Marcu MG, Neckers L. Quantum chemical calculations and mutational analysis suggest heat shock protein 90 catalyzes *trans-cis* isomerization of geldanamycin. *Chem Biol* 2004;11:991–8.
- Onuoha SC, Mukund SR, Coulstock ET, Senqerova B, Shaw J, McLauqhlin SH, et al. Mechanistic studies on Hsp90 inhibition by ansamycin derivatives. J Mol Biol 2007;372:287–97.
- Biamonte MA, van de Water R, Arndt JW, Scannevin RH, Perret D, Lee WC. Heat shock protein 90: inhibitors in clinical trials. *J Med Chem* 2010;53:3–17.
- Smith V, Sausville EA, Camalier RF, Fiebig HH, Burqer AM. Comparison of 17-dimethylaminoethylamino-17-demethoxy-geldanamycin (17DMAG) and 17-allylamino-17-demethoxygeldanamycin (17AAG) *in vitro*: effects on Hsp90 and client proteins in melanoma models. *Cancer Chemother Pharmacol* 2005;56: 126–37.
- Glaze ER, Lambert AL, Smith AC, Page JG, Johson WD, McCormick DL, et al. Preclinical toxicity of a geldanamycin analog, 17-(dimethylaminoethylamino)-17-demethoxygeldanamycin (17-DMAG), in rats and dogs: potential clinical relevance. *Cancer Chemother Pharmacol* 2005;56:637–47.
- Omura S, Iwai Y, Takahashi Y, Sadakane N, Oiwa H, Haseqawa Y, et al. Herbimycin, a new antibiotic produced by a strain of *Streptomyces. J Antibiot* 1979;**32**:255–61.
- 22. Honma Y, Okabekado J, Hozumi M, Uehara Y, Mizuno S. Induction of erythroid-differentiation of k562 human-leukemic cells by herbimycin A, an inhibitor of tyrosine kinase-activity. *Cancer Res* 1989;49:331–4.
- McCollum GW, Rajaratnam VS, Bullard LE, Yang R, Penn JS. Herbimycin A inhibits angiogenic activity in endothelial cells and reduces neovascularization in a rat model of retinopathy of prematurity. *Exp Eye Res* 2004;**78**:987–95.
- 24. Yang HL, Huang WY, Yang YP. Purification, structure elucidation and antitumor activity of minor antibiotics M3 and M4 in the

fermentation broth of *Streptomyces hygrosopicus* NND-52. *Chin J Antibiot* 2006;**31**:274–7.

- Sakagami M, Morrison P, Welch WJ. Benzoquinoid ansamycins (herbimycin A and geldanamycin) interfere with the maturation of growth factor receptor tyrosine kinases. *Cell Stress Chaperones* 1999;4:19–28.
- Oikawa T, Onozawa C, Kuranuki S, Iqarashi Y, Sato M, Ashino H, et al. Dipalmitoylation of radicicol results in improved efficacy against tumor growth and angiogenesis *in vivo. Cancer Sci* 2007;98:219–25.
- Kwon HJ, Yoshida M, Fukui Y, Horinouchi S, Beppu T. Potent and specific-inhibition of p60v-src protein-kinase both *in vivo* and *in vitro* by radicicol. *Cancer Res* 1992;**52**:6926–30.
- Soga S, Kozawa T, Narumi H, Akinaqa S, Irie K, Matsumoto K, et al. Radicicol leads to selective depletion of Raf kinase and disrupts K-Ras-activated aberrant signaling pathway. *J Biol Chem* 1998;273:822–8.
- Schulte TW, Akinaga S, Soga S, Sullivan W, Stensqard B, Toft D, et al. Antibiotic radicicol binds to the N-terminal domain of Hsp90 and shares important biologic activities with geldanamycin. *Cell Stress Chaperones* 1998;3:100–8.
- Agatsuma T, Ogawa H, Akasaka K, Asai A, Yamashita Y, Mizukami T, et al. Halohydrin and oxime derivatives of radicicol: synthesis and antitumor activities. *Bioorg Med Chem* 2002;10:3445–54.
- Turbyville TJ, Wijeratne EMK, Liu MX, Burns AM, Seliqa CJ, Luevano LA, et al. Search for Hsp90 inhibitors with potential anticancer activity: isolation and SAR studies of radicicol and monocillin I from two plant-associated fungi of the Sonoran Desert. J Nat Prod 2006;69:178–84.
- 32. Soga S, Neckers LM, Schulte TW, Shiotsu Y, Akasaka K, Narumi H, et al. KF25706, a novel oxime derivative of radicicol, exhibits *in vivo* antitumor activity *via* selective depletion of Hsp90 binding signaling molecules. *Cancer Res* 1999;**59**:2931–8.
- 33. Kurebayashi J, Otsuki T, Kurosumi M, Soqa S, Akinaqa S, Sonoo H. A radicicol derivative, KF58333, inhibits expression of hypoxia-inducible factor-1 alpha and vascular endothelial growth factor, angiogenesis and growth of human breast cancer xenografts. *Jpn J Cancer Res* 2001;**92**:1342–51.
- 34. Yamamoto K, Garbaccio RM, Stachel SJ, Solit DB, Chiosis G, Rosen N, et al. Total synthesis as a resource in the discovery of potentially valuable antitumor agents: cycloproparadicicol. *Angew Chem (Int Ed Engl)* 2003;42:1280–4.
- Davenport J, Manjarrez JR, Peterson L, Krumm B, Blagg BS, Matts RL. Gamboic acid, a natural product inhibitor of Hsp90. *J Nat Prod* 2011;74:1085–92.
- Gellert M, Odea MH, Itoh T, Tomizawa J. Novobiocin and coumermycin inhibit DNA supercoiling catalyzed by DNA gyrase. *Proc Natl Acad Sci USA* 1976;73:4474–8.
- Marcu MG, Schulte TW, Neckers L. Novobiocin and related coumarins and depletion of heat shock protein 90-dependent signaling proteins. J Natl Cancer Inst 2000;92:242–8.
- Donnelly AC, Mays JR, Burlison JA, Nelson JT, Vielhauer G, Holzbeierluin J, et al. The design, synthesis, and evaluation of coumarin ring derivatives of the Novobiocin scaffold that exhibit antiproliferative activity. J Org Chem 2008;73:8901–20.
- Burlison JA, Neckers L, Smith AB, Maxwell A, Blagg BS. Novobiocin: redesigning a DNA gyrase inhibitor for selective inhibition of Hsp90. J Am Chem Soc 2006;128:15529–36.
- 40. Le Bras G, Radanyi C, Peyrat JF, Brion JD, Alami M, Marsaud V, et al. New novobiocin analogues as antiproliferative agents in breast cancer cells and potential inhibitors of heat shock protein 90. J Med Chem 2007;50:6189–200.
- 41. Palermo CM, Westlake CA, Gasiewicz TA. Epigallocatechin gallate inhibits aryl hydrocarbon receptor gene transcription

through an indirect mechanism involving binding to a 90 kDa heat shock protein. *Biochemistry* 2005;**44**:5041–52.

- 42. Li Y, Zhang T, Jiang Y, Lee HF, Schwartz SJ, Sun D. (-)-Epigallocatechin-3-gallate inhibits Hsp90 function by impairing Hsp90 association with cochaperones in pancreatic cancer cell line Mia Paca-2. *Mol Pharm* 2009;6:1152–9.
- 43. Tran P, Kim SA, Choi HS, Yoon JH, Ahn SG. Epigallocatechin-3gallate suppresses the expression of Hsp70 and Hsp90 and exhibits anti-tumor activity *in vitro* and *in vivo*. BMC Cancer 2010;10:276.
- Yin ZY, Henry EC, Gasiewicz TA. (-)-Epigallocatechin-3-gallate is a novel Hsp90 inhibitor. *Biochemistry* 2009;48:336–45.
- Salminen A, Lehtonen M, Paimela T, Kaarniranta K. Celastrol: molecular targets of Thunder God Vine. *Biochem Biophys Res Commun* 2010;**394**:439–42.
- Zhang T, Hamza A, Cao X, Wang B, Yu S, Zhan CG, et al. A novel Hsp90 inhibitor to disrupt Hsp90/Cdc37 complex against pancreatic cancer cells. *Mol Cancer Ther* 2008;7:162–70.
- Sreeramulu S, Gande SL, Gobel M, Schwalbe H. Molecular mechanism of inhibition of the human protein complex Hsp90-Cdc37, a kinome chaperone-cochaperone, by Triterpene Celastrol. *Angew Chem (Int Ed Engl)* 2009;48:5853–5.
- Hadden MK, Galam L, Gestwicki JE, Matts RL, Blagg BS. Derrubone, an inhibitor of the Hsp90 protein folding machinery. *J Nat Prod* 2007;**70**:2014–8.
- Mays JR, Hill SA, Moyers JT, Blagg BS. The synthesis and evaluation of flavone and isoflavone chimeras of novobiocin and derrubone. *Bioorg Med Chem* 2010;18:249–66.
- Hatcher H, Planalp R, Cho J, Torti FM, Torti SV. Curcumin: from ancient medicine to current clinical trials. *Cell Mol Life Sci* 2008;65:1631–52.
- Jung YJ, Xu WP, Kim H, Neckers L. Curcumin-induced degradation of ErbB2: a role for the E3 ubiquitin ligase CHIP and the Michael reaction acceptor activity of curcumin. *Biochim Biophys Acta* 2007;**1773**:383–90.
- Lee JH, Chung IK. Curcumin inhibits nuclear localization of telomerase by dissociating the Hsp90 co-chaperone p23 from hTERT. *Cancer Lett* 2010;290:76–86.
- 53. Giommarelli C, Zuco V, Favini E, Pisano C, Dal Piaz F, De Tommasi N, et al. The enhancement of antiproliferative and proapoptotic activity of HDAC inhibitors by curcumin is mediated by Hsp90 inhibition. *Cell Mol Life Sci* 2010;67:995–1004.
- 54. Chen WY, Chang FR, Huang ZY, Chen JH, Wu YC, Wu CC, Tubocapsenolide A. a novel withanolide, inhibits proliferation and induces apoptosis in MDA-MB-231 cells by thiol oxidation of heat shock proteins. *J Biol Chem* 2008;**283**:17184–93.
- 55. Blank M, Mandel M, Keisari Y, Meruelo D, Lavie G. Enhanced ubiquitinylation of heat shock protein 90 as a potential mechanism for mitotic cell death in cancer cells induced with hypericin. *Cancer Res* 2003;63:8241–7.
- Hieronymus H, Lamb J, Ross KN, Peng XP, Clement C, Rodina A, et al. Gene expression signature-based chemical genomic prediction identifies a novel class of Hsp90 pathway modulators. *Cancer Cell* 2006;10:321–30.
- Brandt GEL, Schmidt MD, Prisinzano TE, Blagg BS. Gedunin, a novel Hsp90 inhibitor: semisynthesis of derivatives and preliminary structure-activity relationships. J Med Chem 2008;51:6495–502.
- Eiseman JL, Lan J, Lagattuta TF, Hamburger DR, Joseph E, Covey JM, et al. Pharmacokinetics and pharmacodynamics of 17demethoxy 17- (2-dimethylamino)ethyl amino geldanamycin (17DMAG, NSC 707545) in C.B-17 SCID mice bearing MDA-MB-231 human breast cancer xenografts. *Cancer Chemother Pharmacol* 2005;55:21–32.