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Therapeutic potential and pharmacological activities of *Atractylodes lancea* (Thunb.) DC.

Nut Koonrungsesomboon¹, Kesara Na-Bangchang², Juntra Karbwang^{1*}

¹Department of Clinical Product Development, Institute of Tropical Medicine, Nagasaki University, Japan

²Graduate Program in Bioclinical Sciences, Chulabhorn International College of Medicine, Thammasat University, Thailand

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ABSTRACT

The rhizome of *Atractylodes lancea* (*A. lancea*) (Thunb.) DC. (AL) is extensively used in Chinese, Thai, and Japanese traditional medicines as crude extracts/decoctions or a component in various herbal formulations. Various pharmacological activities of AL and its major constituents have been demonstrated *in vitro*, *ex vivo*, and in animal models. Results from the toxicity studies in animal models suggest safety profile of AL and its active constituents. Despite extensive use with positive impression in many diseases, there has not been a clinical study that can conclusively support its efficacy and safety profile in human. This review comprehensively summarizes current information on the pharmacological activities of AL and their active constituents including anticancer, anti-inflammatory, antimicrobial and antipyretic activities, as well as activities on central nervous, cardiovascular, and gastrointestinal systems.

1. Introduction

According to the World Health Organization (WHO) report in 2011[1], traditional medicine addresses up to two-third of the world's population's primary health care needs. One major component of traditional medicine is the use of herbal medicine. A common issue of herbal medicine is the limitation of information on their pharmacological activities and their constituents. Traditionally, the use of herbal medicine was based on empirical treatment and then passed on from generation to generation. In the past 20 years, there were more studies on pharmacological activities and the constituents of many herbal medicines, but the information is often published in local journals and is not extensively disseminated. The limited access to these information prevented many herbal medicines from being developed to their full potential.

The rhizome of *Atractylodes lancea* (*A. lancea*) (Thunb.)

DC. (AL) has been used widely in many countries for various indications. This compound is called "Cangzhu" in China, "Khod-Kha-Mao" in Thailand, and "So-jutsu" in Japan. In Chinese traditional medicine, this rhizome is used extensively for the treatment of several diseases such as rheumatic diseases, digestive disorders, night blindness, and influenza. These traditional uses are explained by the compound's ability to eliminate dampness, strengthen the spleen, expel wind-cold from the superficial parts of the body, and clear away the common cold[2]. In Thai traditional medicine, the dried rhizome of AL has been used to treat fever and the common cold[3]. Moreover, it has also been used as a component in Thai traditional medicine in order to relieve gastrointestinal symptoms including dyspepsia, flatulence, nausea, and noninfectious diarrhea. In Japan, the rhizome of AL is a component in several Kampo medicines, *eg.*, Juzen-taiho-to[4] and Saireito[5,6].

History of extensive use of this herb in mankind has facilitated the development of this herb to its full therapeutic potential. This has brought about this review article, whose purpose is to aid the readers in gaining a better understanding of the potential and toxicity of this medicinal plant and to contribute to appropriate decision-making in further development of AL. This review article

*Corresponding author: Juntra Karbwang, MD, PhD, Department of Clinical Product Development, Institute of Tropical Medicine, Nagasaki University, Japan.

Tel: +81-95-819-7558

Fax: +81-95-819-7846

E-mail: karbwangj@nagasaki-u.ac.jp

will focus on the pharmacological activities of the crude extract of AL rhizome including its major constituents: β -eudesmol, hinesol, atractylone and atractylodin[7–9].

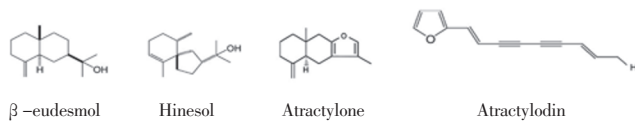


Figure 1. The chemical structures of major components of *A. lancea* (Thunb.) DC.

2. Pharmacological activities of *A. lancea* (Thunb.) DC.

2.1. Anticancer activities

Several conventional anticancer drugs being used in patients with cancers are derived from plants. These include vinblastine, vincristine, etoposide, teniposide, paclitaxel, vinorelbine, docetaxel, topotecan, and irinotecan, all of which have been approved by the US Food and Drug Administration[10]. Moreover, there are several herbal medicines of which their promising anticancer activities were demonstrated in laboratory experiments and clinical trials[11]. Recently, it appears that the rhizome of AL is a promising candidate herbal plant for further development as anticancer drugs, particularly as an alternative treatment in patients with cholangiocarcinoma (CCA), the cancer of bile duct.

The anticancer activities of AL particularly anti-CCA have been demonstrated in several studies both *in vitro* and *in vivo*. Of a total of 28 plants and 5 herbal formulations used in Thai traditional medicine investigated for their cytotoxic activities, the crude ethanolic extract of AL rhizome was shown to exhibit the most potent and selective activity against CCA cell line (CL-6) with IC_{50} (concentration which inhibits cell growth by 50%) of (24.09 ± 3.40) (mean \pm SD) μ g/mL and SI (selectivity index) of 8.6[12]. Results of the *in vitro* screening of tumoricidal properties of international medicinal herbs conducted in the United States also confirmed the anticancer activity of AL in murine neuroblastoma cells originally derived from a spontaneous malignant tumor with moderate to strong activity with LC_{50} (50% lethal concentration, the concentration which causes 50% cell death) of 0.704 mg/mL[13]. These two studies have caught researchers' attentions to further investigate the anticancer property of AL. Based on calcein-AM and Hoechst 33342 assays, the cytotoxic activity of the ethanolic extract of AL against CL-6 was found to be more potent and more selective than the standard anticancer 5-fluorouracil (5-FU)[14]. Additionally, AL also exhibited significant inhibitory effects on clonogenic survival, tube formation, and invasion of CL-6 cells through a basement membrane model

in a dose-dependent manner. However, this compound did not significantly exhibit antioxidative activity determined by the radical-scavenging activity of 2,2-diphenyl-1-picrylhydrazyl radical (DPPH). With regards to antitumor property of AL in animal models, the ethanolic extract at the concentrations of 1 000, 3 000, and 5 000 mg/kg body weight significantly inhibited tumor growth in CCA-xenografted nude mice[15]. The tumor size of AL-treated group was reduced to about 10% of that in the control group on day 40 after treatment (mean \pm SD: tumor volumes: (550 ± 13) and $(20\ 661 \pm 126)$ mm³ for AL-treated and control group, respectively). At the highest dose of 5 000 mg/kg body weight, AL significantly inhibited lung metastasis by about 95%, while in the control group lung metastasis accounted for about 90% of total lung mass. All dose levels provided about 2-fold prolongation of the survival time of mice compared with the control group (mean \pm SD: 83.30 ± 0.88 and 40.00 ± 0.57 d in AL-treated and control group, respectively).

Lines of evidence have suggested that either anti-angiogenic or apoptotic-related activity or both, might at least in part contribute to cytotoxic activity of AL. Tsuneki *et al*[16] investigated the anti-angiogenic activity of β -eudesmol, the main constituent of AL, both *in vitro* and *in vivo*. The proliferation of various endothelial cells including porcine brain microvascular endothelial cells (PBMEC) derived from cerebral microvessel, human dermal microvascular endothelial cells (HDMEC) derived from peripheral microvessels, and human umbilical vein endothelial cells (HUVEC) derived from peripheral veins, were markedly inhibited by β -eudesmol at concentrations ranging from 50 to 100 μ M. Moreover, β -eudesmol also showed a broad spectrum of anti-angiogenic effects not only on blockade of the phosphorylation of extracellular signal-related kinase (ERK) 1/2 induced by basic fibroblast growth factor (bFGF) or vascular endothelial growth factor (VEGF), but also on prevention of endothelial tube formation and inhibition of cell migration stimulated by bFGF. In animal model, β -eudesmol significantly inhibited angiogenesis of subcutaneously implanted Matrigel plugs in mice and adjuvant-induced granuloma in mice[16]. These results were consistent with the observations by Ma *et al*[17], showing an inhibitory effect of β -eudesmol (50–100 μ M) in HUVEC induced by VEGF and bFGF. Apart from HUVEC, Hela (human cervical cells), the proliferation of SGC-7901 (human gastric cancer cells), and BEL-7402 (human liver cancer cells) were also inhibited by β -eudesmol (10–100 μ M) in a time- and dose-dependent manner. Furthermore, β -eudesmol (2.5–5 mg/kg) significantly inhibited tumor growth in mice implanted with H22 and S180 tumor cells and also obviously inhibited vascular index (calculated by carmine content in the tumor tissues divided by tumor tissue weight)[17]. Recently, Zhao *et al*[18] demonstrated that AL extract inhibited the growth of human gastric cancer cells in a dose- and time-dependent manner, and proposed that the cytotoxic mechanism of AL was related to apoptosis

and cell cycle arrest through mitochondria-dependent and death receptor-dependent apoptotic pathways. Further investigation should be focused on the mechanism of action of anticancer property of AL in CCA, identification of its active constituents, as well as confirmation of its clinical efficacy and safety in CCA patients.

2.2. Pharmacological activities on nervous system

Although neither serious adverse effect on central nervous system (CNS) nor any morbidity has been reported in human so far, the use of AL in human should be with caution in patients with nervous problems due to its various effects on nervous system. The pharmacological activity of the rhizome extract of AL on central nervous system has been demonstrated in various animal models with regards to its effects on general behavior and spontaneous movement, anti-electroshock convulsion, and potentiation of hypnotic action of hexobarbital sodium^[19]. AL extract at the highest dose of 5 000 mg/kg body weight significantly interfered with muscle relaxation in mice similar to that produced by the reference drug diazepam (4 mg/kg body weight)^[15]. The acetone extract of AL rhizome also showed an anti-anoxic effect in potassium cyanide (KCN)-induced anoxia in mice^[20]. Nine out of ten (90%) mice treated with the AL extract at the dose of 1 500 mg/kg body weight survived, while none in the control group survived (0/10: 0%). The anti-anoxic action of AL rhizome extract was shown to be due mainly to its active constituent β -eudesmol. Six out of ten mice (60%) treated with β -eudesmol at the dose of 300 mg/kg body weight survived, whereas none in control group survived (0/10: 0%).

The effect on post-synaptic neuromuscular junction (NMJ) of β -eudesmol was shown to be primarily through the blockage of nicotinic acetylcholine receptors (nAChR) via accelerated desensitization^[21–23]. The potentiating effect of β -eudesmol on NMJ was greater in diabetic than in normal muscles^[24,25]. β -eudesmol has been proposed as a promising compound for potentiating neuronal function. It was shown to induce neurite outgrowth from rat pheochromocytoma cells (PC-12) via mitogen-activated protein kinase (MAPK) activation^[26].

2.3. Pharmacological activities on cardiovascular system

AL extract at the dose levels of 1 000, 3 000, and 5 000 mg/kg body weight significantly reduced the heart rate of rats, but only the highest dose (5 000 mg/kg body weight) significantly decreased both systolic and diastolic blood pressure^[15]. However, the mechanism of the anti-hypertensive effect of AL is still unknown. The anti-platelet activity of AL has been demonstrated in collagen-induced platelet aggregation model^[27]. Since it did not inhibit adrenaline/ADP- or adrenaline/5-HT-induced platelet aggregation, its mechanism of action has been thought to be via suppression

of collagen-induced signal pathway, the upstream of the release of thromboxane A₂ (TXA₂) from platelets. Altogether, results suggest that care should be taken when using AL extract or its active constituents in patients with platelet disorders or coagulopathy.

2.4. Pharmacological activities on gastrointestinal system

The pharmacological effects of AL and its constituents on gastrointestinal system support their clinical use for alleviation of digestive symptoms in traditional medicine. AL extract has been shown to delay gastric emptying and stimulate small intestinal motility. The mechanisms of its action on these activities could be through either the inhibition of both dopamine D₂ and 5-HT₃ receptors^[28], or activation of vagal tone and inhibition of corticotropin-releasing factor (CRF)^[29]. The main activity was shown to be due to the atractylodin component^[30].

AL extract at the dose levels of 1 000, 3 000, and 5 000 mg/kg body weight produced an anti-ulcer effect at similar potency as the reference drug omeprazole given at a dose of 20 mg/kg body weight^[15]. Results from a previous study in pylorus-ligated rats suggest that the mechanism of action of AL extract on anti-ulcer activity might be mediated through inhibition of gastric secretion and reduction of effects on histamine-induced ulceration and stress-induced ulceration^[31]. β -eudesmol is thought to be an active compound which exerts inhibitory effect on gastric secretion stimulated by histamine. The compound could prevent gastric ulceration as effectively as cimetidine at the same dose level (10 mg/kg body weight)^[32]. Apart from β -eudesmol, the anti-ulcer activity of AL was also shown with hinesol, another main constituent in AL extract at the dose of 100 mg/kg body weight. Further investigation should be performed to elucidate the mechanisms of action of AL and its constituents on gastrointestinal system.

2.5. Other pharmacological activities

The anti-inflammatory activity of AL might be due to the contribution of several of its active constituents through various mechanisms. The lipophilic extract from AL rhizome exhibited potent inhibitory effect against 5-lipoxygenase (5-LOX) and cyclooxygenase-1 (COX-1) with IC₅₀ of 2.9 and 30.5 μ g/mL, respectively^[33]. Isolated compound that exhibited potent inhibitory activities against both enzymes was shown to be atractylochromene (IC₅₀ for 5-LOX and COX-1 = 0.6 and 3.3 μ M, respectively). Despite relatively low potency on COX-1 (IC₅₀ = 64.3 μ M), quinone, another isolated compound, showed a selective inhibitory activity against 5-LOX (IC₅₀ = 0.2 μ M). Atractylone also exhibited inhibitory effects against 5-LOX but with potency about 100-fold lower than quinone (IC₅₀ = 25.1 μ M). The study conducted by Seo *et al.*^[34] demonstrated that the anti-inflammatory effect of β -eudesmol was via regulation of

interleukin-6 (IL-6) production and expression through regulation of the p38 MAPK and nuclear factor (NF)- κ B. In addition, it also suppressed receptor-interacting protein 2 (RIP2)/caspase-1 activation induced by phorbol 12-myristate 13-acetate calcium ionophore A23187 (PMACI).

The antimicrobial activity of AL against various microorganisms has been demonstrated in various studies including *Staphylococcus aureus*[35], *Escherichia coli*[35,36], *Saccharomyces cerevisiae*, and *Candida albicans*[36]. Moreover, the growth of some *fungi* species, such as *Rhodotorula glutinis* and *Saprolegnia*, was also inhibited by the volatile oil extract of AL[37]. The activity on *Rhizopus* and *Absidia* was however, relatively weak.

Although AL extract did not produce any significant central or peripheral analgesic effects, it was shown to produce an antipyretic effect at a dose of 5 000 mg/kg body weight in the rat model[15]. This antipyretic activity supports its use for relieve fever and cold as indicated in Thai traditional medicine.

3. Safety profiles of *A. lancea* (Thunb.) DC.

AL rhizome showed safety profiles in various animal models. Following administration of AL extract at the high dose level of 5 000 mg/kg body weight in rats and mice, no significant toxicity except stomach irritation and general CNS depressant signs (reduced alertness and locomotion and diminished response to touch and balance) was observed[15]. Results from the acute and subacute toxicity tests both in rats and mice indicated safety profiles of AL in a broad range of dose levels (1 000–5 000 mg/kg body weight).

Several clinical studies of AL have been conducted in patients with different diseases/symptoms using AL in the forms of various formulations[4,38–41]; however, there has been no clinical study conducted using AL extract or its major constituents alone. This thus signifies the needs for further investigations in clinical trials to prove their clinical efficacy and safety profiles in humans. Despite the lack of clinical studies to directly support its safety in human,

Table 1

The pharmacological activities of *A. lancea* (Thunb.) DC. and its compounds

Pharmacological activity	Model	Active ingredient	Mechanism of action	Reference
Anti-tumour activities				
Cytotoxic activity	<i>In vitro</i>	50% Ethanol extract 50 μ g/mL		[12]
	<i>In vitro</i>	50% Ethanol extract 50 μ g/mL		[14]
	<i>In vitro</i>	Petroleum ether fraction, ethyl acetate fraction, <i>n</i> -butanol fraction, and water fraction of AL 0.0625–1 mg/mL – Induction of cell apoptosis via [18] the mitochondrial pathway		[13]
	<i>In vitro</i>	100% Ethanol extract 5 mg/mL	Prenylated dihydrobenzofuran derivative	[44]
Anticancer activity	Mice	50% Ethanol extract 1 000–5 000 mg/kg		[15]
Anti-angiogenic activity	<i>In vitro</i>	β -eudesmol 50 and 100 μ M	β -eudesmol	– Inhibition of the endothelial cell proliferation [16] – Suppression of DNA synthesis – Inhibition of endothelial cell migration – Inhibition of tube formation by endothelial cells – Blockage of bFGF- and VEGF-induced ERK1/2 activation (only at the concentration of 100 μ M) – Inhibition of phosphorylation of CREB induced by VEGF in the growth factor signaling pathway

	Mice	β -eudesmol 0.90 μ mol/kg	β -eudesmol	[16]
	<i>In vitro</i>	β -eudesmol 50 and 100 μ M	β -eudesmol	[17]
				– Inhibition of the growth factor signaling pathway by depressing activation of ERK–MAPK
				– Suppression of CREB activation in growth factor signaling pathway
	Mice	β -eudesmol 2.5–5 mg/kg	β -eudesmol	[17]
	<i>In vitro</i>	50% Ethanol extract 25–100 μ g/mL		[14]
Anti-clonogenic activity	<i>In vitro</i>	50% Ethanol extract 12.5–50 μ g/mL		[14]
Inhibitory activity on cell invasion				
	<i>In vitro</i>	50% Ethanol extract 12.5–150 μ g/mL		[14]
Pharmacological activities on nervous system				
NMJ blocking activity	<i>Ex vivo</i>	β -eudesmol 200 μ M	β -eudesmol	[21]
				– Blockade of nicotinic ACh receptors by accelerating the desensitization of the nicotinic ACh receptor
	<i>Ex vivo</i>	β -eudesmol 20 μ M	β -eudesmol	[23]
				– Blockade of closed state of nicotinic ACh receptors by accelerating the desensitization of the nicotinic ACh receptor
	<i>Ex vivo</i>	β -eudesmol 20 μ M	β -eudesmol	[22]
				– Depression of the regenerative release of ACh during repetitive stimulation
	<i>Ex vivo</i>	β -eudesmol 80 μ M	β -eudesmol	[25]
CNS activity on neuronal differentiation				
	<i>In vitro</i>	β -eudesmol 100 and 150 μ M	β -eudesmol	[26]
				– Induction of neurite outgrowth mediated by MAPK activation
Anti-anoxic activity				
	Mice	β -eudesmol 300 mg/kg	β -eudesmol	[20]
Motor coordination impairment				
	Mice	50% Ethanol extract 5 000 mg/kg		[15]
CNS depressant activity	Mice	Benzene extract 200–1 000 mg/kg		[19]
Pharmacological activities on cardiovascular system				
Anti-hypertensive activity	Rats	50% Ethanol extract 5 000 mg/kg		[15]
Anti-platelet activity	<i>In vitro</i>	Crude extract 30–1 000 μ g/mL		[27]
				– Inhibition of collagen-induced signal pathway, which is upstream of the release of TXA ₂ from platelets
Pharmacological activities on gastrointestinal system				
Anti-ulcer activity	Rats	50% Ethanol extract 1,000–5 000 mg/kg		[15]
	Rats	Benzene extract 500 mg/kg		[19]
	Rats	50% Methanol extract 200 mg/kg		[31]
				– Inhibition of gastric secretion by histamine H ₂ -receptor blocking

	Rats	β -eudesmol 50 mg/kg	β -eudesmol	– Inhibition of gastric secretion [32] by histamine H ₂ -receptor blocking
	Rats	Hinesol 100 mg/kg	Hinesol	– Inhibit gastric secretion by [32] unknown mechanism
Improvement of the Rats delayed gastric emptying		Ethanol extract 30–120 mg/kg		– Inhibition of the CRF release [29] – Activation of vagal pathway – Involvement in the release of gastrointestinal hormones such as motilin, gastrin and somatostatin
	Rats	Water extract 250 mg/kg and Atractylodin and its Atractylodin and its derivatives derivatives 0.1–0.3 mg/kg		[30]
Intestinal motility stimulation	Mice	Water extract 500–1 000 mg/kg and β -eudesmol 50–100 mg/kg	β -eudesmol	– Inhibition of the dopamine D ₂ [28] receptor and the 5-HT ₃ receptor
Other pharmacological activities				
Anti-inflammatory activity	Rats	50% Ethanol extract 5 000 mg/kg		[15]
	<i>In vitro</i>	β -eudesmol 2, 20 μ M	β -eudesmol	– Regulation of IL-6 through [34] regulation of the p38 MAPK and NF- κ B – Suppression of RIP2 expression and caspase-1 activation
	<i>In vitro</i>		Atractylchromene, Quinone, Atractylon	Inhibition against 5-LOX and [33] COX-1
Antipyretic activity	Mice	Atractylenolide I 300 mg/kg	Atractylenolide I	[45]
Antimicrobial activity	Rats	50% Ethanol extract 5 000 mg/kg		[15]
– against <i>E. coli</i> , <i>S. cerevisiae</i> , and <i>C. albicans</i>	<i>In vitro</i>	95% Ethanol extract 200 mg/mL		[36]
– against <i>E. coli</i> , <i>S. aureus</i>	<i>In vitro</i>		Atractylodin derivatives	[35]
– against <i>Rhodotorulaglutinis</i> and <i>Saprolegnia</i>	<i>In vitro</i>			[37]

AL, *Atractylodes lancea*; bFGF, basic fibroblast growth factor; VEGF, vascular endothelial growth factor; ERK, extracellular signal-regulated kinase; CREB, cyclic adenosine monophosphate (cAMP) response element binding protein; NMJ, neuromuscular junction; ACh, acetylcholine; TXA₂, thromboxane A₂; CRF, Corticotropin-releasing factor; IL, interleukin; MAPK, mitogen-activated protein kinase; NK- κ B, nuclear factor- κ B; RIP2, receptor-interacting protein 2; LOX, lipoxygenase; COX, cyclooxygenase.

available information has indicated no serious adverse event when they were administered in humans. Ayurved Siriraj herbal recipe Chantaleela which consists of 60.6 mg AL in each tablet (250 mg/tablet) was administered to healthy male and female volunteers at the dose of 545.4 mg of AL/d for 1 d (divided into 3 doses, administered every 8 h). No adverse event was observed in any subject for 10 d follow-up[42]. Moreover, observational study conducted in China showed a safety profile of “Fufang Cangzhu Tang”, a Chinese herbal formula which contains 15 g *Atractylodes* rhizome decocted into 300 mL of liquor and separately administered orally

twice a day for 8 weeks in 32 senile patients with obesity or overweight complicated with impaired glucose tolerance[43].

4. Conclusion

AL rhizome has been shown to exhibit various pharmacological activities including anticancer activities, activities on nervous and gastrointestinal systems, as well as anti-hypertensive, anti-platelet, anti-ulcer, anti-inflammatory, antimicrobial, and antipyretic activities.

Despite extensive use with positive impression, there has not been a clinical study that can conclusively support its efficacy and safety profile. Further investigations should focus on the application of AL in patients with different diseases/symptoms. In addition, more investigation is required to identify the specific mechanisms of certain pharmacological activities, including anticancer activities of AL, and its active constituents.

Conflict of interest statement

The authors declare that there are no conflicts of interest.

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References

- [1] Gao PF, Watanabe K. Introduction of the World Health Organization project of the International Classification of Traditional Medicine. *Zhong Xi Yi Jie He Xue Bao* 2011; **9**(11): 1161–1164.
- [2] Qian S, Wang L, Duan J, Feng H. The research progress in chemical constituents and biological activities of *Atractylodes lancea* DC. 2006; **25**: 8–11.
- [3] Chayamarit K. *Thai medicinal plants*. Bangkok: Department of Forestry; 1995.
- [4] Saiki I. A Kampo medicine “Juzen-taiho-to”–prevention of malignant progression and metastasis of tumor cells and the mechanism of action. *Biol Pharm Bull* 2000; **23**(6): 677–688.
- [5] Kishida Y, Miki H, Nishii T, Inoue T, Nishida S, Yoshikawa H, et al. Therapeutic effects of Saireito (TJ-114), a traditional Japanese herbal medicine, on postoperative edema and inflammation after total hip arthroplasty. *Phytomedicine* 2007; **14**(9): 581–586.
- [6] Kitamoto M, Kato K, Sugimoto A, Kitamura H, Uemura K, Takeda T, et al. Sairei-to ameliorates rat peritoneal fibrosis partly through suppression of oxidative stress. *Nephron Exp Nephrol* 2011; **117**(3): e71–e81.
- [7] Ji L, Ao P, Pan JG, Yang JY, Yang J, Hu SL. GC-MS analysis of essential oils from rhizomes of *Atractylodes lancea* (Thunb.) DC. and *A. chinensis* (DC.) Koidz. *Zhongguo Zhong Yao Za Zhi* 2001; **26**(3): 182–185.
- [8] Zhou J, Fang L, Wang X, Zhang J, Guo L-p, Huang L-q. Comparison of the volatile compounds of crude and processed *Atractylodes rhizome* analyzed by GC-MS. *Afr J Pharm Pharmacol* 2012; **6**: 2155–2160.
- [9] Ouyang Z, Zhang L, Zhao M, Wang P, Wei Y, Fang J. Identification and quantification of sesquiterpenes and polyacetylenes in *Atractylodes lancea* from various geographical origins using GC-MS analysis. *Rev Bras Farmacogn/Braz J Pharmacogn* 2012; **22**: 957–963.
- [10] Dholwani KK, Saluja AK, Gupta AR, Shah DR. A review on plant-derived natural products and their analogs with anti-tumor activity. *Indian J Pharmacol* 2008; **40**(2): 49–58.
- [11] Ruan WJ, Lai MD, Zhou JG. Anticancer effects of Chinese herbal medicine, science or myth? *J Zhejiang Univ Sci B* 2006; **7**(12): 1006–1014.
- [12] Mahavorasirikul W, Viyanant V, Chaijaroenkul W, Itharat A, Na-Bangchang K. Cytotoxic activity of Thai medicinal plants against human cholangiocarcinoma, laryngeal and hepatocarcinoma cells *in vitro*. *BMC Complement Altern Med* 2010; **10**: 55.
- [13] Mazzio EA, Soliman KF. *In vitro* screening of tumoricidal properties of international medicinal herbs: part II. *Phytother Res* 2010; **24**(12): 1813–1824.
- [14] Plengsuriyakarn T, Viyanant V, Eursitthichai V, Itharat A, Na-Bangchang K. *In vitro* investigations on the potential roles of Thai medicinal plants in treatment of cholangiocarcinoma. *Int Res J Pharm Pharmacol* 2012; **2**: 52–63.
- [15] Plengsuriyakarn T, Viyanant V, Eursitthichai V, Picha P, Kupradinun P, Itharat A, et al. Anticancer activities against cholangiocarcinoma, toxicity and pharmacological activities of Thai medicinal plants in animal models. *BMC Complement Altern Med* 2012; **12**: 23.
- [16] Tsuneki H, Ma EL, Kobayashi S, Sekizaki N, Maekawa K, Sasaoka T, et al. Antiangiogenic activity of beta-eudesmol *in vitro* and *in vivo*. *Eur J Pharmacol* 2005; **512**(2–3): 105–115.
- [17] Ma EL, Li YC, Tsuneki H, Xiao JF, Xia MY, Wang MW, et al. Beta-eudesmol suppresses tumour growth through inhibition of tumour neovascularisation and tumour cell proliferation. *J Asian Nat Prod Res* 2008; **10**(1–2): 159–167.
- [18] Zhao M, Wang Q, Ouyang Z, Han B, Wang W, Wei Y, et al. Selective fraction of *Atractylodes lancea* (Thunb.) DC. and its growth inhibitory effect on human gastric cancer cells. *Cytotechnology* 2013.
- [19] Yamahara J, Sawada T, Tani T, Nishino T, Kitagawa I. Biologically active principles of crude drugs. Pharmacological evaluation of the crude drug “Zhu” (author’s transl). *Yakugaku Zasshi* 1977; **97**(8): 873–879.
- [20] Yamahara J, Matsuda H, Naitoh Y, Fujimura H, Tamai Y. Antianoxic action and active constituents of *Atractylodes lancea* rhizoma. *Chem Pharm Bull (Tokyo)* 1990; **38**(7): 2033–2034.

- [21]Kimura M, Nojima H, Muroi M, Kimura I. Mechanism of the blocking action of beta-eudesmol on the nicotinic acetylcholine receptor channel in mouse skeletal muscles. *Neuropharmacology* 1991; **30**(8): 835–841.
- [22]Nojima H, Kimura I, Kimura M. Blocking action of succinylcholine with beta-eudesmol on acetylcholine-activated channel activity at endplates of single muscle cells of adult mice. *Brain Res* 1992; **575**(2): 337–340.
- [23]Chiou LC, Chang CC. Antagonism by beta-eudesmol of neostigmine-induced neuromuscular failure in mouse diaphragms. *Eur J Pharmacol* 1992; **216**(2): 199–206.
- [24]Muroi M, Tanaka K, Kimura I, Kimura M. beta-eudesmol (a main component of *Atractylodes lancea*)-induced potentiation of depolarizing neuromuscular blockade in diaphragm muscles of normal and diabetic mice. *Jpn J Pharmacol* 1989; **50**(1): 69–71.
- [25]Kimura M, Diwan PV, Yanagi S, Kon-no Y, Nojima H, Kimura I. Potentiating effects of beta-eudesmol-related cyclohexylidene derivatives on succinylcholine-induced neuromuscular block in isolated phrenic nerve-diaphragm muscles of normal and alloxan-diabetic mice. *Biol Pharm Bull* 1995; **18**(3): 407–410.
- [26]Obara Y, Aoki T, Kusano M, Ohizumi Y. Beta-eudesmol induces neurite outgrowth in rat pheochromocytoma cells accompanied by an activation of mitogen-activated protein kinase. *J Pharmacol Exp Ther* 2002; **301**(3): 803–811.
- [27]Nasu Y, Iwashita M, Saito M, Fushiya S, Nakahata N. Inhibitory effects of *Atractylodes lanceae* rhizoma and Poria on collagen- or thromboxane A₂-induced aggregation in rabbit platelets. *Biol Pharm Bull* 2009; **32**(5): 856–860.
- [28]Kimura Y, Sumiyoshi M. Effects of an *Atractylodes lancea* rhizome extract and a volatile component β-eudesmol on gastrointestinal motility in mice. *J Ethnopharmacol* 2012; **141**(1): 530–536.
- [29]Zhang H, Han T, Sun LN, Huang BK, Chen YF, Zheng HC, et al. Regulative effects of essential oil from *Atractylodes lancea* on delayed gastric emptying in stress-induced rats. *Phytomedicine* 2008; **15**(8): 602–611.
- [30]Nakai Y, Kido T, Hashimoto K, Kase Y, Sakakibara I, Higuchi M, et al. Effect of the rhizomes of *Atractylodes lancea* and its constituents on the delay of gastric emptying. *J Ethnopharmacol* 2003; **84**(1): 51–55.
- [31]Kubo M, Nogami M, Nishimura M, Moriura T, Arichi S. Origins, processing, and qualities of crude drugs (1). Preventive effects of a Chinese crude drug, Zhu, on experimental stomach ulcer and its pharmacological evaluation. I. *Yakugaku Zasshi* 1983; **103**(4): 442–448.
- [32]Nogami M, Moriura T, Kubo M, Tani T. Studies on the origin, processing and quality of crude drugs. II. Pharmacological evaluation of the Chinese crude drug “zhu” in experimental stomach ulcer. (2). Inhibitory effect of extract of *Atractylodes lancea* on gastric secretion. *Chem Pharm Bull (Tokyo)* 1986; **34**(9): 3854–3860.
- [33]Resch M, Steigel A, Chen ZL, Bauer R. 5-Lipoxygenase and cyclooxygenase-1 inhibitory active compounds from *Atractylodes lancea*. *J Nat Prod* 1998; **61**(3): 347–350.
- [34]Seo MJ, Kim SJ, Kang TH, Rim HK, Jeong HJ, Um JY, et al. The regulatory mechanism of β-eudesmol is through the suppression of caspase-1 activation in mast cell-mediated inflammatory response. *Immunopharmacol Immunotoxicol* 2011; **33**(1): 178–185.
- [35]Chen Y, Wu Y, Wang H, Gao K. A new 9-nor-atractylodin from *Atractylodes lancea* and the antibacterial activity of the atractylodin derivatives. *Fitoterapia* 2012; **83**(1): 199–203.
- [36]Wat CK, Johns T, Towers GH. Phototoxic and antibiotic activities of plants of the Asteraceae used in folk medicine. *J Ethnopharmacol* 1980; **2**(3): 279–290.
- [37]Wang Y, Dai CC, Chen Y. Antimicrobial activity of volatile oil from *Atractylodes lancea* against three species of endophytic fungi and seven species of exogenous fungi. *Ying Yong Sheng Tai Xue Bao* 2009; **20**(11): 2778–2784.
- [38]Kamiyama H, Takano S, Ishikawa E, Tsuboi K, Matsumura A. Anti-angiogenic and immunomodulatory effect of the herbal medicine “Juzen-taiho-to” on malignant glioma. *Biol Pharm Bull* 2005; **28**(11): 2111–2116.
- [39]Liu Y, Jia Z, Dong L, Wang R, Qiu G. A randomized pilot study of atractylenolide I on gastric cancer cachexia patients. *Evid Based Complement Alternat Med* 2008; **5**(3): 337–344.
- [40]Wang CC, Lin SY, Cheng HC, Hou WC. Pro-oxidant and cytotoxic activities of atractylenolide I in human promyeloleukemic HL-60 cells. *Food Chem Toxicol* 2006; **44**(8): 1308–1315.
- [41]Wang GT. Treatment of operated late gastric carcinoma with prescription of strengthening the patient’s resistance and dispelling the invading evil in combination with chemotherapy: follow-up study of 158 patients and experimental study in animals. *Zhong Xi Yi Jie He Za Zhi* 1990; **10**(12): 712–716.
- [42]Itthipanichpong R, Lupreechaset A, Chotewuttakorn S, Akarasereenont P, Onkoksoong T, Palo T, et al. Effect of Ayurved Siriraj herbal recipe Chantaleela on platelet aggregation. *J Med Assoc Thai* 2010; **93**(1): 115–122.
- [43]Shi J, Hu Y, Wang Q. Fufang cangzhu tang for treatment of senile obesity or overweight complicated with impaired glucose tolerance – a clinical observation in 32 cases. *J Tradit Chin Med* 2006; **26**(1): 33–35.
- [44]Duan JA, Wang L, Qian S, Su S, Tang Y. A new cytotoxic prenylated dihydrobenzofuran derivative and other chemical constituents from the rhizomes of *Atractylodes lancea* DC. *Arch Pharm Res* 2008; **31**(8): 965–969.
- [45]Endo K, Taguchi T, Taguchi F, Hikino H, Yamahara J, Fujimura H. Antiinflammatory principles of *Atractylodes* rhizomes. *Chem Pharm Bull (Tokyo)* 1979; **27**(12): 2954–2958.