



Are Genetically Robust Regulatory Networks Dynamically Different from Random Ones?

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Abstract

We study a genetic regulatory network model developed to demonstrate that genetic robustness can evolve through stabilizing selection for optimal phenotypes. We report preliminary results on whether such selection could result in a reorganization of the state space of the system. For the chosen parameters, the evolution moves the system slightly toward the more ordered part of the phase diagram. We also find that strong memory effects cause the Derrida annealed approximation to give erroneous predictions about the model's phase diagram. © 2010 Published by Elsevier Ltd.

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1. Introduction

Gene networks are extremely robust against genetic perturbations [1, 2]. For example, systematic gene knock-out studies on yeast showed that almost 40% of genes on chromosome V have no detectable effects on indicators like cell division rate [3]. Similar studies on other organisms agree with these results [1, 2]. It is also known that phenotypically, most species do not vary much, although they experience a wide range of environmental and genetic perturbations. This striking resilience makes one wonder about the origins, evolutionary consequences, and mechanistic causes of genetic robustness.

It has been proposed that genetic robustness evolved through stabilizing selection for a phenotypic optimum. Wagner showed that this in fact could be true by modeling a developmental process within an evolutionary scenario, in which the genetic interaction sequence represents organismal development, and the equilibrium configuration of the gene network represents the phenotype [4]. His results show that the genetic robustness of a population of model genetic regulatory networks can increase through stabilizing selection for a particular equilibrium configuration (phenotype) of each network.

In this paper we investigate the effects of the biological evolution of genetic robustness on the dynamics of gene regulatory networks in general. In particular, we want to answer the question whether the evolution process moves the system to a different point in the phase diagram. Below, we present some preliminary results.

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2. Model

We use a model by Wagner [4], which has also been used by other researchers with minor modifications. Each individual is represented by a regulatory gene network consisting of N genes. The expression level of each gene, s_i , has only two values, +1 or -1, expressed or not, respectively. The expression states change in time according to regulatory interactions between the genes. The time evolution of the system configuration represents an (organismal) developmental pathway. The discrete-time dynamics are given by a set of nonlinear difference equations representing a random threshold network (RTN),

$$s_i(t+1) = \begin{cases} \operatorname{sgn}\left(\sum_{j=1}^N w_{ij}s_j(t)\right), & \sum_{j=1}^N w_{ij}s_j(t) \neq 0 \\ s_i(t), & \sum_{j=1}^N w_{ij}s_j(t) = 0 \end{cases}, \quad (1)$$

where sgn is the sign function and w_{ij} is the strength of the influence of gene j on gene i . Nonzero elements of the $N \times N$ matrix \mathbf{W} are independent random numbers drawn from a standard normal distribution. (The diagonal elements of \mathbf{W} are allowed to be nonzero, corresponding to self-regulation.) The (mean) number of nonzero elements in \mathbf{W} is controlled by the connectivity density, c , which is the probability that a w_{ij} is nonzero.

The dynamics given by Eq. (1) can have a wide variety of features. For a specified initial configuration $\mathbf{s}(0)$, the system reaches either a fixed-point attractor or a limit cycle after a transient period. The lengths of transients, number of attractors, distribution of attractor lengths, etc. can differ from system to system, depending on whether the dynamics are ordered, chaotic, or critical. The fitness of an individual is defined by whether it can reach a developmental equilibrium, a certain fixed gene-expression pattern, \mathbf{s}^* , in a “reasonable” transient time. Further details of the model are explained in the next section.

3. Monte Carlo Simulations

3.1. Generation and Robustness Assessment of Random Networks

We studied populations of $\mathcal{N} = 400$ random networks (founding individuals) with $N = 10$. Each network was assigned a matrix \mathbf{W} and an initial configuration $\mathbf{s}(0)$. \mathbf{W} was generated as follows. Each w_{ij} was independently chosen to be nonzero with probability c . If so, it was assigned a random number drawn from a standard gaussian distribution, $N(\mu = 0, \sigma = 1)$. Then, each “gene” of the initial configuration, $s_i(0)$, was assigned either -1 or +1 at random, each with probability 1/2.

After \mathbf{W} and $\mathbf{s}(0)$ were created, the dynamics were started and the network’s stability was evaluated. If the system reached a fixed point, \mathbf{s}^* , in $3N$ timesteps, then it was considered stable and kept. Otherwise it was considered unstable, both \mathbf{W} and $\mathbf{s}(0)$ were discarded, and the process was started over and repeated until a stable network was generated. For each stable network, its fixed point, \mathbf{s}^* , was regarded as the “optimal” gene-expression state (phenotype) of the system. This is the only modification we made to Wagner’s model: he generated networks with preassigned $\mathbf{s}(0)$ and \mathbf{s}^* , whereas we accept any \mathbf{s}^* as long as it can be reached within $3N$ timesteps from $\mathbf{s}(0)$.

After generating $\mathcal{N} = 400$ individual stable networks, we analyzed their state-space structures and evaluated their robustness as discussed in subsection 3.3.

3.2. Evolution

In order to generate a breed of more robust networks, a mutation-selection process was simulated for all of the $\mathcal{N} = 400$ random, stable networks as follows. First, a clan of $\mathcal{N}' = 500$ identical copies of each network was generated. For each clan, a four-step process was performed for $T = 400$ generations:

1. Recombination: Each pair of the N rows of consecutive matrices in the clan were swapped with probability 1/2. Since the networks were already shuffled in step 4 (see below), there was no need to pick random pairs.
2. Mutation: Each nonzero w_{ij} was replaced with probability $1/(cN^2)$ by a new random number drawn from the same standard gaussian distribution. Thus, on average, one matrix element was changed per matrix per Monte Carlo step.

3. Fitness evaluation: Each network was run starting from the original initial condition, $\mathbf{s}(0)$. If the network reached a fixed point, \mathbf{s}^\dagger , within $3N$ timesteps, then its fitness, $f(\mathbf{s}^\dagger, \mathbf{s}^*) = \exp(-H^2(\mathbf{s}^\dagger, \mathbf{s}^*)/\sigma_s)$, was calculated. Here $H(\mathbf{s}^\dagger, \mathbf{s}^*)$, denotes the normalized Hamming distance between \mathbf{s}^\dagger and \mathbf{s}^* , and σ_s denotes the strength of selection, \mathbf{s}^* is the optimal gene-expression state, which is the final gene-expression state of the original network that “founded” the clan. We used $\sigma_s = 0.1$. If the network could not reach a fixed point, then it was assigned the minimum nonzero fitness value, $\exp(-1/\sigma_s)$.
4. Selection/Asexual Reproduction: The fitness of each network was normalized to the fitness value of the most fit network in the clan. Then, a network was chosen at random and duplicated into the descendant clan with probability equal to its normalized fitness. This process was repeated until the size of the descendant clan reached N' . Then the old clan was discarded, and the descendant clan was kept as the next generation. Note that this process allows multiple copies (offspring) of the same network to appear in the descendant clan, while some networks may not make it to the next generation due to genetic drift.

At the end of the $T = 400$ generation selection, any unstable networks were removed from the evolved clan.

3.3. Assessment of Robustness

The mutational robustness of a network was assessed slightly differently for random and evolved networks. For a random network, first, one nonzero w_{ij} was picked at random and replaced by a new random number with the same standard gaussian distribution. Then, the dynamics were started, and it was checked if the system reached the same equilibrium state, \mathbf{s}^* , within $3N$ timesteps. This process was repeated $5000c$ times using the original matrix (i.e., each mutated matrix was discarded after its stability was evaluated). The robustness of the original network before evolution was defined as the fraction of singly-mutated networks that reached \mathbf{s}^* .

For the evolved networks, clan averages were used. For each of $N^{\text{opt}} \leq 400$ networks in a clan, robustness was assessed as described above with one difference: the number of perturbations was reduced to $5000c/N^{\text{opt}}$ per network to keep the total number of perturbations used to estimate robustness of networks before and after evolution approximately equal. The mean robustness of the those N^{opt} networks was taken as the robustness of the founder network after evolution. Therefore, the robustness of a network after evolution is the mean robustness of its descendant clan of stable networks.

4. Results

As Wagner pointed out, the stabilizing selection described above increases the robustness of the model population against mutations [4]. However, it is not very clear what kind of a reorganization in the state space occurs during the evolution. Also, it is not known whether this robustness against mutations leads to robustness against environmental perturbations. In this paper, we focus on the effects of evolution in terms of moving the system to another point in the phase diagram. In other words, we investigate whether the system becomes more chaotic or more ordered after evolution.

A standard method for studying damage spreading in systems such as the one considered here is the Derrida annealed approximation [5, 6], in which one calculates changes with time of the overlap of two distinct states, $\mathbf{s}(t)$ and $\tilde{\mathbf{s}}(t)$,

$$x(t) = \frac{1}{2N} \sum_{i=1}^N |s_i(t) + \tilde{s}_i(t)|. \quad (2)$$

The change of the overlap over one time step for $N \gg \langle k \rangle = Nc$ is given by

$$x(t+1) = n(0)x(t) + n(1)x(t) + \sum_{k=2}^{\infty} n(k) \left[(x(t))^k + \sum_{l=1}^{k-1} \Pi_k(l) \mathcal{P}(k, l) \right], \quad (3)$$

where the Poisson distribution $n(k) = \kappa^k \exp(-\kappa)/k!$, is the probability of finding a gene, i , with k inputs, the binomial distribution $\Pi_k(l) = \binom{k}{l} (1-x(t))^l (x(t))^{k-l}$ is the probability of finding $k-l$ of these inputs in the overlapping parts of $\mathbf{s}(t)$ or $\tilde{\mathbf{s}}(t)$, and $\mathcal{P}(k, l) = 1 - \frac{2}{\pi} \arctan\left(\sqrt{l/(k-l)}\right)$ (for $k > l$) is the probability of the sum of $k-l$ matrix elements

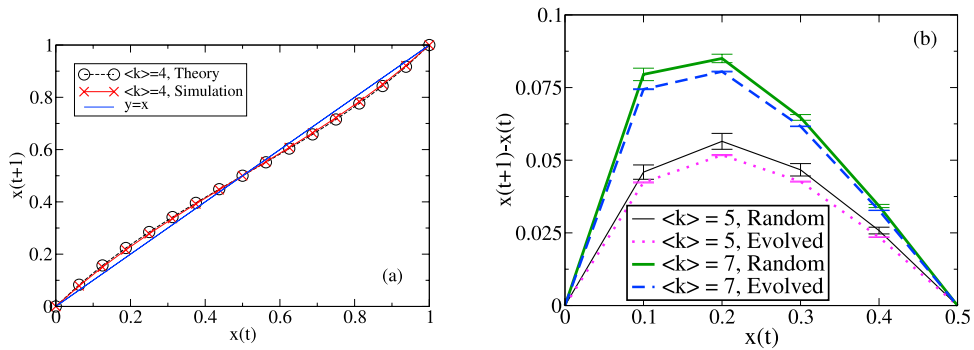


Figure 1: (a) $x(t+1)$ shown vs. $x(t)$ for $N = 16$ and $\langle k \rangle = 4$. The theory, Eq. (3), is in good agreement with the simulations. The deviations are due to the small size of the simulated system as the theoretical calculation assumes $N \gg \langle k \rangle$. (b) Damage-spreading rate, $x(t+1) - x(t)$ vs. $x(t)$, for random and evolved networks with $N = 10$ and $\langle k \rangle = 5$ and 7 , showing the difference between the “random” and “evolved” curves. Only the first half of the curves are shown since $x(t+1)$ vs. $x(t)$ is point-symmetric about $(1/2, 1/2)$. The results were averaged over 10 random networks and all of their evolved descendants (~ 300 evolved networks per random network). The evolved curves for each $\langle k \rangle$ lie very close to their “random” counterparts. However, they are outside twice the error bar range of each other at most data points.

being larger than the sum of l matrix elements, which are independent and $N(0, 1)$ distributed. Here, $\kappa = \langle k \rangle$, the mean number of inputs per node.

For most RTNs that have been studied so far [5, 6], Eq. (3) can be iterated as a map to give the full time evolution of the overlap. Changes in the fixed-point structure of this map with changing $\langle k \rangle$ would then signify phase transitions of the system. As seen in Fig. 1a, for $\langle k \rangle = 4$, such a map would have a stable fixed point at $x = 1/2$. One can also show that $\lim_{x(t) \rightarrow 0^+} dx(t+1)/dx(t) > 1$ for all $\langle k \rangle > 0$ (this implies $\lim_{x(t) \rightarrow 1^-} dx(t+1)/dx(t) > 1$ and $\lim_{x(t) \rightarrow 1/2} dx(t+1)/dx(t) < 1$), and so it would seem that the system has no phase transition and always stays chaotic for nonzero $\langle k \rangle$. However, simulations of damage spreading for longer times indicate that the system studied here has strong memory effects due to the update rule for spins with no inputs, given by the last line in Eq. (1), which retard the damage spreading [7]. In fact, like other RTNs the system undergoes a phase transition near $\langle k \rangle \approx 2$ from a chaotic phase at larger $\langle k \rangle$ to an ordered phase at smaller $\langle k \rangle$. The strong, retarding memory effects mean that Eq. (3) cannot be iterated as a map, and the naïve prediction based on the Derrida annealed approximation is erroneous.

Despite its irrelevance for the long-time damage spreading, the damage-spreading rate shown in Fig. 1b properly describes the short-time dynamical character of the system. However, as Eq. (3) assumes that the interaction constants, w_{ij} , are statistically independent, it may not apply to evolved networks as we do not know whether the selection process creates correlations between the matrix elements. Nevertheless, we can still compute $x(t+1)$ as a function of $x(t)$ numerically to see if there is a change in the degree of chaoticity (or order) of the dynamics. As seen in Fig. 1b, the damage-spreading rates for evolved networks are slightly (but statistically significantly) lower than for their random predecessors, which are thus slightly more chaotic.

To summarize, we have presented preliminary results on some general properties of a popular RTN model of a gene regulatory network and on how the biological evolution of genetic robustness affects its dynamics [8]. We have also shown that the update rule for spins without inputs leads to strong memory effects that invalidate naïve iteration of the Derrida annealed approximation as a map. The evolutionary process that improves the genetic robustness of such networks has only a very small effect on their dynamical properties: after evolution, the system moves slightly toward the more ordered part of the phase diagram.

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