

Hypothesis

New uses for old drugs. Auranofin, a clinically established antiarthritic metallodrug, exhibits potent antimalarial effects *in vitro*: Mechanistic and pharmacological implications

Anna Rosa Sannella^a, Angela Casini^b, Chiara Gabbiani^b, Luigi Messori^{b,*}, Anna Rita Bilia^c, Francesco Franco Vincieri^c, Giancarlo Majori^a, Carlo Severini^a

^a Department of Infectious, Parasitic and Immunomediated Diseases, Vector-Borne Diseases and International Health Section, Istituto Superiore di Sanità, Viale Regina Elena 299, 00161 Rome, Italy

^b Department of Chemistry, University of Florence, Via della Lastruccia 3, 50019 Sesto Fiorentino, Florence, Italy

^c Department of Pharmaceutical Sciences, University of Florence, Via U. Schiff 6, 50019 Sesto Fiorentino, Florence, Italy

Received 28 January 2008; revised 8 February 2008; accepted 14 February 2008

Available online 21 February 2008

Edited by Barry Halliwell

Abstract The clinically established gold-based antiarthritic drug auranofin (AF) manifests a pronounced reactivity toward thiol and selenol groups of proteins. In particular, AF behaves as a potent inhibitor of mammalian thioredoxin reductases causing severe intracellular oxidative stress. Given the high sensitivity of *Plasmodium falciparum* to oxidative stress, we thought that auranofin might act as an effective antimalarial agent. Thus, we report here new experimental results showing that auranofin and a few related gold complexes strongly inhibit *P. falciparum* growth *in vitro*. The observed antiplasmodial effects probably arise from direct inhibition of *P. falciparum* thioredoxin reductase. The above findings and the safe toxicity profile of auranofin warrant rapid evaluation of AF for malaria treatment in animal models.

© 2008 Federation of European Biochemical Societies. Published by Elsevier B.V. All rights reserved.

Keywords: Gold drugs; Malaria; Thioredoxin reductase; *Plasmodium falciparum*

1. Introduction

Since the 1930s, a number of injectable gold(I) thiolates drugs, in particular myochrysin, sanochrysin, allochrysin, and solganol [1,2], have been widely employed in the clinics for the treatment of rheumatoid arthritis (the so called *Chrysotherapy*). Later on, in 1979, a new antiarthritic gold(I) phosphine compound, ((2,3,4,6-tetra-*O*-acetyl-1-(thio-kS)- β -D-glucopyranosato)(triethylphosphine) gold(I)), auranofin (AF), (Fig. 1) was introduced in the clinics, with the significant advantage of an oral administration [3]. Today, *chrysotherapy* still represents an effective option for the treatment of severe forms of rheumatoid arthritis though it is being progressively replaced by new and more sophisticated therapeutic approaches.

In spite of their long and established clinical use, the mechanism of action of antiarthritic gold drugs is still largely

controversial. However, recent studies point out that antiarthritic gold drugs directly interfere with the functioning of the immune system, at various levels [4,5]. In addition, a number of biochemical studies revealed that gold drugs behave as potent “thiol reactive species” and act as effective inhibitors of a number of proteases (mostly cysteine proteases) [6,7], involved in the progression of rheumatoid arthritis (RA). It also emerged that auranofin and some related gold drugs are strong inhibitors of mammalian thioredoxin reductases (TrxR), both in the cytosol and in the mitochondria [8–10]. Notably, thioredoxin reductase inhibition is associated with important modifications of the intracellular redox balance and, in particular, with induction of severe oxidative stress and relevant cytotoxic effects *in vitro* [11,12].

2. Auranofin: a possible antimalarial agent?

This latter feature of AF, i.e. its ability to act as a strong inhibitor of thioredoxin reductase (in the nM range), suggests that AF might display additional pharmacological uses beyond RA therapy. Indeed, thioredoxin reductases are emerging in the last years as an important class of likely drug targets for a variety of diseases [13]. Various research groups formerly analyzed the possible use of auranofin and related gold(I) complexes as cytotoxic and anticancer drugs. We explore here whether AF may manifest any significant antiplasmodial effect and thus be of potential interest for malaria treatment.

Malaria is today a major health emergency on the international scale and also a real challenge for modern pharmacology [14]. Owing to the frequent appearance of drug-resistant *Plasmodium* strains, the classical antimalarial drugs tend to become rapidly obsolete, generating a continuous need for new therapeutic agents, possibly endowed with innovative mechanisms of action. Remarkably, *Plasmodium falciparum* is known to be particularly sensitive and vulnerable to oxidative stress; in particular, *P. falciparum* thioredoxin reductase has been validated as a “druggable” target for the development of new antimalarials [15]. As *P. falciparum* thioredoxin reductase bears a number of functionally relevant cysteines (but no selenol groups) [16], we thought that *thiol-reactive* gold drugs

*Corresponding author. Fax: +39 0 55 457 3385.
E-mail address: luigi.messori@unifi.it (L. Messori).

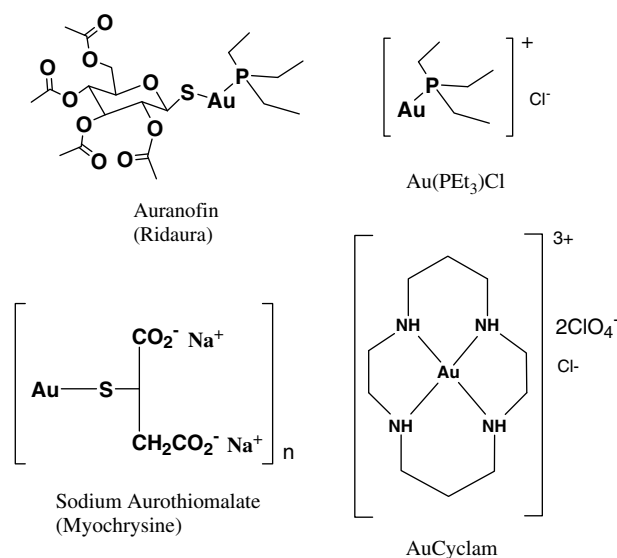


Fig. 1. Chemical structures of the selected gold complexes.

might display appreciable antiparasitic actions, through a direct and potent inhibition of this enzyme. These observations prompted us to measure the effects of a few representative gold drugs on *P. falciparum* growth *in vitro*. To our surprise, despite a certain number of metal complexes, and even gold compounds, had earlier been tested as antimalarial agents [17], we realized that classical antiarthritic gold drugs, to the best of our knowledge, had never been considered for this scope.

3. Direct evidence that auranofin and a few related gold compounds strongly inhibit *P. falciparum* growth

The antimalarial effects of auranofin were assayed *in vitro* through a validated experimental procedure based on the determination of *P. falciparum* lactate dehydrogenase (pLDH) activity (see Supporting Information Available for details). Measuring pLDH provides indeed a selective and easy screening method for the identification and quantitation of parasite growth in *in vitro* cultures [18]. For comparison purposes the effects of three additional and structurally diverse gold compounds, namely gold(I) triethylphosphine chloride (Au(PEt₃)Cl), sodium aurothiomalate and gold(III) cyclam (AuCyclam), reported in Fig. 1, were measured as well. Au(PEt₃)Cl is a derivative of auranofin where the thioglucose ligand is replaced by a chloride ion; aurothiomalate is the antiarthritic gold(I) drug *myochrysine*; at variance, AuCyclam is a gold(III) compound where the gold center is tightly coordinated to four nitrogens, within a roughly square planar arrangement [19]. Such a strong coordination environment reasonably results into a marked decrease of reactivity.

Results of *P. falciparum* growth assays, in the presence of the various gold compounds, are shown in Fig. 2, while Table 1 summarizes the concentrations inhibiting the cell viability by 50% (IC₅₀) determined for each species. It is evident that auranofin, even at very low concentrations, causes a strong and nearly complete inhibition of *P. falciparum* growth, with an IC₅₀ value of 142 nM; Au(PEt₃)Cl shows a similar trend but with a significantly higher IC₅₀ value of 2.1 μM. In contrast, aurothiomalate and AuCyclam are far less effective; their

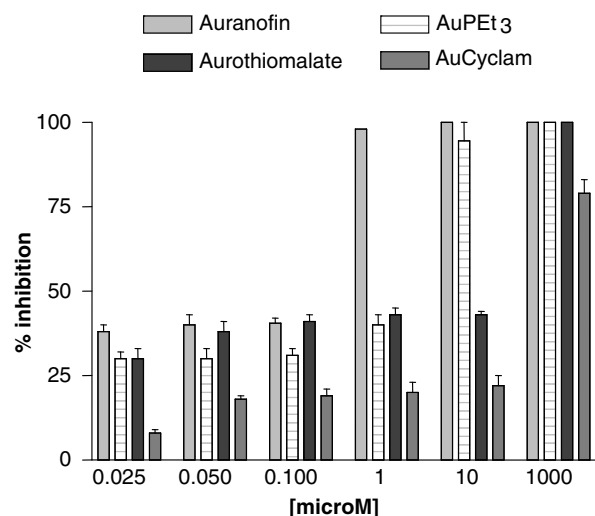


Fig. 2. Antiplasmodial activities of gold compounds. Percentages of inhibition of 3D7 *P. falciparum* strain growth against increasing concentrations of auranofin, Au(PEt₃)Cl, aurothiomalate and AuCyclam, respectively. The data are expressed assuming no inhibition for untreated controls.

Table 1
IC₅₀ values for auranofin, Au(PEt₃)Cl, aurothiomalate, AuCyclam, artemisinin (ART), ART/50 nM Auranofin and ART/100 nM auranofin against 3D7 *P. falciparum* strain

Name of compounds	3D7 <i>P. falciparum</i> strain IC ₅₀ (95% CI)
Auranofin	142 nM (132–152)
Au(PEt ₃)Cl	2.1 μM (2.0–2.2)
Aurothiomalate	168 μM (160–170)
AuCyclam	439 μM (428–440)
Artemisinin (ART)	16 nM (14.9–17.3)
ART/50 nM auranofin	12 nM* (8.9–12.3)
ART/100 nM auranofin	9 nM* (7–13)

**P* < 0.001.

IC₅₀ values being 168 μM and 439 μM, respectively (Table 1). It is of interest to observe that all gold compounds, starting from the lowest tested concentrations, cause a partial and rather constant inhibition of *P. falciparum* growth (~10–25%); this observation implies, most likely, the coexistence of two independent cytotoxic pathways, one of which is saturated very soon. This peculiar behavior clearly emerges from inspection of the recorded *P. falciparum* growth inhibition profiles; characteristic profiles obtained in the case of AF and Au(PEt₃)Cl are reported in Fig. 3.

In our opinion, the above reported results are of great interest since a very important antiparasitic activity is described for the first time for the antiarthritic gold(I) drug auranofin. This finding might imply that our working hypothesis is basically correct as the known and relevant inhibitory effects of AF toward thioredoxin reductase [9,10] are apparently reflected into a strong inhibition of *P. falciparum* growth.

Of course, further work is needed to confirm the validity of such hypothesis. In particular, specific studies on the inhibition of *P. falciparum* thioredoxin reductase by auranofin and related gold drugs would be warranted. In any case, if the relevant antiparasitic effects that we have observed here *in vitro*, will be confirmed by the animal studies that we are planning, the way would be open for the rapid clinical testing

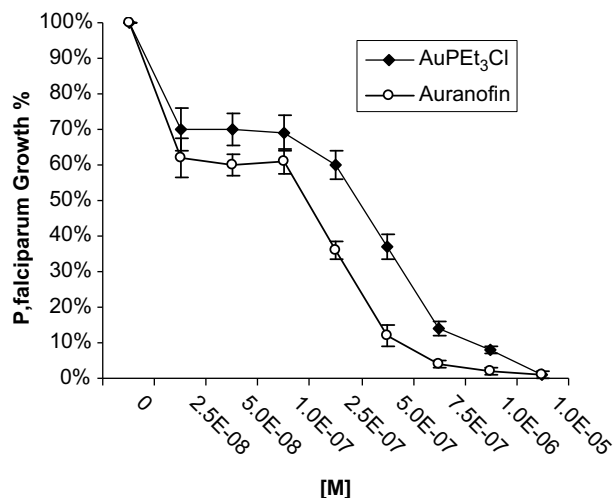


Fig. 3. Percentage of 3D7 *P. falciparum* growth as a function of the concentration (M) of auranofin or Au(PEt₃)Cl.

of auranofin in malaria treatment. Indeed, AF is already in the clinics since more than 25 years and presents a well known and quite safe toxicity profile: thus, its clinical use for a different therapeutic indication might be afforded very quickly and straightforwardly. Also, it is worthwhile reminding that AF is taken up very efficiently by erythrocytes [20], one of the major sites where a strong antiplasmodial action is required; this additional feature might render its *in vivo* antimalarial actions even more effective.

4. The combined effects of artemisinin and auranofin

Since we hypothesize that the specific antiproliferative effects of AF toward *P. falciparum* probably arise from induction of oxidative stress, we decided to measure its antiplasmodial effects in combination with artemisinin. Artemisinin (ART),

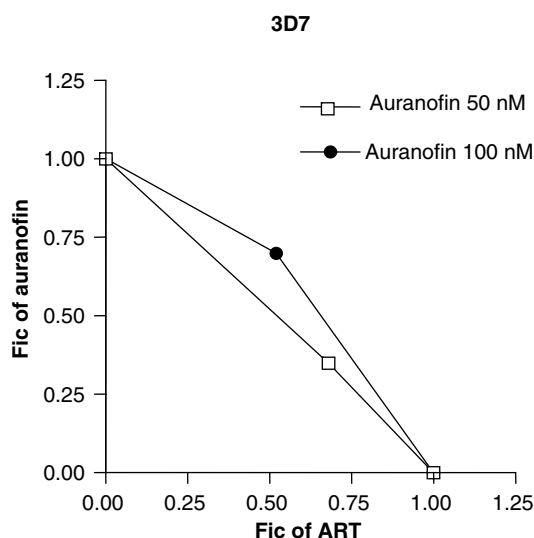


Fig. 4. Isobolograms showing the interaction between ART and auranofin when administered on 3D7 *P. falciparum* strain. ART was given at non-cytotoxic doses, ranging from 0.625 nM to 40 nM, in the presence of 0, 50 or 100 nM auranofin. Fic is the fractional inhibitory concentration.

today one of the most potent antimalarial agents available in the clinics (with an IC₅₀ value falling in the very low nanomolar range); is thought to work through induction of intracellular oxidative stress [21,22]. Thus, we wondered whether these two drugs, i.e. artemisinin and auranofin might manifest any significant synergism. Their possible interactions were evaluated through an established methodology relying on the so called “isobolographic” analysis [23,24]. Using AF, either 50 or 100 nM, in combination with artemisinin, the sum of FICs was 1.2 and 1.03, respectively, when administered on 3D7 *P. falciparum* strain. The results of isobolographic analysis are thus consistent with an *additive effect* of both auranofin concentrations, as shown in Fig. 4; any antagonism between these two agents may be ruled out.

5. Concluding remarks and perspectives

In conclusion, based on a simple mechanistic hypothesis and a few selective *P. falciparum* growth inhibition measurements *in vitro*, we have demonstrated for the first time that the antiarthritic drug AF displays very pronounced antiplasmodial effects, and thus merits further investigations as a potential antimalarial drug. Moreover, we have shown, through isobolographic analysis, that AF displays additive antimalarial effects when given in combination with artemisinin. Significant but less potent antiplasmodial effects have been measured for the other investigated gold compounds.

The observed antiplasmodial effects are probably mediated by severe oxidative stress originating from *P. falciparum* thio-reductase inhibition. Of course, further experimental work is absolutely warranted to support this latter statement; however the presence of a few functionally relevant cysteine residues at the active site of *P. falciparum* TrxR makes us confident that our working hypothesis is well grounded. If the effectiveness of antimalarial drug discovery strategies based on direct inhibition of thioredoxin reductase by metallic compounds will be eventually proved for AF, the way is open to the extensive testing of several other *thiol reactive* metallodrugs showing acceptable toxicity requirements.

Finally, we like to stress further that AF is in clinical use since more than 20 years. Looking for new uses of old (and thus *clinically established*) drugs represents today a very promising and effective strategy of modern drug discovery with obvious advantages. Indeed, much work concerning the safety profile can be avoided leading to a drastic reduction of times and costs. New therapeutic applications for some established drugs have been identified in the last years; a few relevant cases are described in the recent literature [25,26]. Auranofin might hopefully represent another successful example for this kind of strategy.

Acknowledgements: The authors gratefully acknowledge support from Ente Cassa di Risparmio di Firenze.

Appendix A. Supplementary material

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.febslet.2008.02.028.

References

- [1] Messori, L. and Marcon, G. (2004) Gold complexes in the treatment of rheumatoid arthritis. *Met. Ions Biol. Syst.* 41, 279–304.
- [2] Shaw III, C.F. (1999) Gold-based therapeutic agents. *Chem. Rev.* 99, 2589–2600.
- [3] Kean, W.F., Hart, L. and Buchanan, W.W. (1997) Auranofin. *Br. J. Rheum.* 36, 560–572.
- [4] Havarinasab, S., Johansson, U., Pollard, K.M. and Hultman, P. (2007) Gold causes genetically determined autoimmune and immunostimulatory responses in mice. *Clin. Exp. Immunol.* 150, 179–188.
- [5] Eisler, R. (2003) Chrysotherapy: a synoptic review. *Inflam. Res.* 52, 487–501.
- [6] Chircorian, A. and Barrios, A.M. (2004) Inhibition of lysosomal cysteine proteases by chrysotherapeutic compounds: a possible mechanism for the antiarthritic activity of Au(I). *Bioorg. Med. Chem. Lett.* 14, 5113–5116.
- [7] Weidauer, E., Yasuda, Y., Biswal, B.K., Cherny, M., James, M.N. and Bromme, D. (2007) Effects of disease modifying anti-rheumatic drugs (DMARDs) on the activities of rheumatoid arthritis-associated cathepsins K and S. *Biol. Chem.* 388, 331–336.
- [8] Gromer, S., Arscott, L.D., Williams Jr., C.H., Schirmer, R.H. and Becker, K. (1998) Human placenta thioredoxin reductase. Isolation of the selenoenzyme, steady state kinetics, and inhibition by therapeutic gold compounds. *J. Biol. Chem.* 273, 20096–20101.
- [9] Rigobello, M.P., Scutari, G., Boscolo, R. and Bindoli, A. (2002) Induction of mitochondrial permeability transition by auranofin, a gold(I)–phosphine derivative. *Br. J. Pharmacol.* 136, 1162–1168.
- [10] Rigobello, M.P., Messori, L., Marcon, G., Cinelli, M.A., Bragadin, M., Folda, A., Scutari, G. and Bindoli, A. (2004) Gold complexes inhibit mitochondrial thioredoxin reductase: consequences on mitochondrial functions. *J. Inorg. Biochem.* 98, 1634–1641.
- [11] Yoo, M.H., Xu, X.M., Carlson, B.A., Gladyshev, V.N. and Hatfield, D.L. (2006) Thioredoxin reductase 1 deficiency reverses tumor phenotype and tumorigenicity of lung carcinoma cells. *J. Biol. Chem.* 281, 13005–13008.
- [12] Lu, J., Chew, E.H., Holmgren, S. and Targeting, A. (2007) TrxR is a basis for cancer therapy by arsenic trioxide. *Proc. Natl. Acad. Sci. USA* 104, 12288–12293. (This paper strongly suggests that the anticancer activity of arsenic trioxide is by means of a Trx system-mediated apoptosis. This new article reinforces the view that thioredoxin reductase represents today an important cancer target).
- [13] Becker, K., Gromer, S., Schirmer, R.H. and Müller, S. (2000) Thioredoxin reductase as a pathophysiological factor and drug target. *Eur. J. Biochem.* 267, 6118–6125.
- [14] See for example the website <<http://www.cdc.gov/malaria/>>.
- [15] Jaeger, T. and Flohe, L. (2006) The thiol-based redox networks of pathogens: unexploited targets in the search for new drugs. *Biofactors* 27, 109–120.
- [16] Nickel, C., Rahlfs, S., Deponte, M., Koncarevic, S. and Becker, K. (2006) Thioredoxin networks in the malarial parasite *Plasmodium falciparum*. *Antioxid. Redox Signal.* 8, 1227–1239.
- [17] Sánchez-Delgado, R.A. and Anzellotti, A. (2004) Metal complexes as chemotherapeutic agents against tropical diseases: trypanosomiasis, malaria and leishmaniasis. *Mini Rev. Med. Chem.* 4, 23–30.
- [18] Makler, T., Piper, R.C. and Milhous, W.K. (1998) Lactate dehydrogenase and the diagnosis of malaria. *Parasitol. Today* 14, 376–377.
- [19] Messori, L., Abbate, F., Marcon, G., Orioli, P., Fontani, M., Mini, E., Mazzei, T., Carotti, S., O'Connell, T. and Zanello, P. (2000) Gold(III) complexes as potential antitumor agents: solution chemistry and cytotoxic properties of some selected gold(III) compounds. *J. Med. Chem.* 43, 3541–3548.
- [20] Herrlinger, J.D., Alsen, C., Beress, R., Hecker, U. and Weikert, W. (1982) Distribution of gold in serum erythrocytes and white blood cells after *in vitro* incubation and during chrysotherapy with different gold compounds. *J. Rheumatol. Suppl.* 8, 81–89.
- [21] O'Neill, P.M. and Posner, G.H. (2004) A medicinal chemistry perspective on artemisinin and related endoperoxides. *J. Med. Chem.* 47, 2945–2964.
- [22] Krungkrai, S.R. and Yuthavong, Y. (1987) The antimalarial action on *Plasmodium falciparum* of qinghaosu and artesunate in combination with agents which modulate oxidant stress. *Trans. R. Soc. Trop. Med. Hyg.* 81, 710–714.
- [23] Berenbaum, M.C. (1978) A method for testing for synergy with any number of agents. *J. Infect. Dis.* 137, 122–130.
- [24] Bell, A. (2005) Antimalarial drug synergism and antagonism. Mechanistic and clinical significance. *FEMS Microbiol. Lett.* 253, 171–184.
- [25] Chong, C.R. and Sullivan Jr., D.J. (2007) New uses for old drugs. *Nature* 448 (7154), 645–646.
- [26] During the preparation of this manuscript, a new article has appeared detailing the effectiveness of auranofin in the treatment of another important parasitic disease, schistosomiasis Kuntz, A.N., Davioud-Charvet, E., Sayed, A.A., Califf, L.L., Dessolin, J., Arnér, E.S. and Williams, D.L. (2007) Thioredoxin glutathione reductase from *Schistosoma mansoni*: an essential parasite enzyme and a key drug target. *PLoS Med.* 4, e206. (This additional finding gives further support to the idea that an established drug such as auranofin, in view of its peculiar reactivity, may be conveniently used for therapeutic indications other than the original one).