Crystallization and preliminary X-ray analysis of restriction endonuclease FokI bound to DNA

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Abstract FokI is a type IIs restriction endonuclease which recognizes an asymmetric DNA sequence and cleaves DNA a short distance away from the sequence. The enzyme is bipartite in nature with its DNA recognition and cleavage functions located on distinct domains. We report here cocrystals of the complete FokI enzyme (579 amino acids) bound to a 20-bp DNA fragment containing its recognition sequence. The complex is amongst the largest protein-DNA complexes to be crystallized, and required macroseeding techniques for optimal crystal growth. The cocrystals diffract to at least 2.8 Å in resolution and belong to space group P2\(_1\) with unit cell dimensions of \(a = 67.9\ \text{Å}, b = 119.8\ \text{Å}, c = 69.1\ \text{Å}, \beta = 96.6^\circ\). Using specific amino acid analysis we show that asymmetric unit contains a single FokI molecule bound to the 20-bp DNA fragment. This paper reports the first cocrystals of a type IIs restriction endonuclease.

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Key words: FokI; Crystallization; Restriction endonuclease, type IIs; X-ray diffraction analysis

1. Introduction

FokI from Flavobacterium okeanokites is a member of type IIs subclass of restriction enzymes. Unlike the more common type II enzymes such as BamHI and EcoRI which recognize palindromic DNA sequences and cleave within those sequences, FokI recognizes an asymmetric DNA sequence 5'-GGATG-T3' and cleaves 9 and 13 base pairs away from the recognition sequence [1] (Fig. 1). Furthermore, FokI exists as a monomer in solution while BamHI and EcoRI exist as dimers [2]. Biochemical studies show that FokI has a bipartite structure, containing two functionally independent domains, one which serves as the DNA binding domain (41 kDa), and one which serves as the non-specific cleavage domain (25 kDa) [3]. Due to its bipartite nature, FokI has been used to create artificial enzymes by attaching its cleavage domain to the DNA binding domains of transcription factors [4,5]. FokI can also be used to cleave single stranded DNA at predetermined sites by using suitably tailored oligonucleotide adapters [6]. To understand how the bipartite enzyme recognizes and cleaves DNA, we have cocrystallized the intact enzyme with a 20-bp DNA fragment containing its recognition sequence.

2. Materials and methods

The FokI endonuclease gene [7,8] was expressed under the control of P\(_{lac}\) promoter in E. coli with a copy of the FokI methylase gene residing on the chromosome. The cells were grown at 37°C to mid-log phase and then induced with 0.3 mM isopropyl \(\beta\)-thiogalactoside (IPTG). 3 h after induction the cells were harvested by centrifugation at 4°C, suspended in 4 vols. of buffer A (10 mM potassium phosphate (pH 6.5), 1 mM EDTA, 1 mM EGTA, and 1 mM DTT, 0.2 M NaCl). The cells were disrupted by sonication, and the cellular debris was removed by centrifugation. The resulting supernatant was applied to a phosphocellulose (P-11, Whatman) column and eluted with a 0.2-1.0 M NaCl gradient. The fractions which contained active FokI endonuclease were identified by incubating a sample with DNA followed by analysis with agarose gel electrophoresis. The active fractions were pooled and dialyzed against buffer B (10 mM potassium phosphate (pH 7.3), 0.1 M NaCl, 1 mM EDTA, 1 mM EGTA, 1 mM DTT, and 10% glycerol). The dialysate was then applied to a CM Sepharose column and eluted with a 0.1-0.8 M NaCl gradient. In this and the following chromatographic steps, the FokI endonuclease was the major protein species and was followed as a ~66 kDa band by SDS PAGE. The pool of FokI endonuclease protein was diluted 7-fold with buffer C (10 mM sodium potassium phosphate (pH 6.8), 1 mM EDTA, 1 mM EGTA, 1 mM DTT, 10% glycerol), and then applied to a heparin Sepharose column and eluted with a 0.0-0.8 M KCl gradient developed in buffer C. The eluted endonuclease was diluted with 3 vols. of buffer C and then extensively dialyzed against buffer D (10 mM Tris-HCl (pH 7.8), 50 mM KCl, 1 mM EDTA, 1 mM EGTA, 1 mM DTT, 10% glycerol). The dialysate was then applied to a DEAE Sepharose column and eluted with a 50 mM to 0.8 M KCl gradient. The resulting FokI endonuclease had a specific activity of ~30,000 units per mg of protein (one unit is defined as the amount of enzyme required to completely digest 1 µg of lambda DNA in a 50 µl reaction mixture of 50 mM potassium acetate, 20 mM Tris-acetate (pH 7.9), 10 mM magnesium acetate, 1 mM DTT). A single liter of bacterial cells yields about 3 mg of purified protein (Fig. 2). For crystallization the enzyme is concentrated to 10 mg/ml by ultrafiltration, aliquoted and stored at -70°C.

DNA was synthesized on an Applied Biosystems 391 PCRMate automated synthesizer using phosphoramidite chemistry. We retained the 5' trityl groups to facilitate later HPLC purification. Derivatized oligonucleotides were synthesized by substituting iodouracil phosphoramidites (Peninsula Laboratories) for the thymidine phosphoramidites. Oligonucleotides were cleaved and deprotected with NH\(_2\)OH as per manufacturer's protocols. The DNA was purified by HPLC with a Rainin PureDNA column which facilitates removal of failed sequences and then detritylation with trifluoroacetic acid [9]. In some instances, oligonucleotides were purified with denaturing PAGE, extracted, desalted and further purified by HPLC. After HPLC, samples were desalted, resuspended in Bis-Tris buffer (25 mM pH 6.8) with 0.1 M KCl and annealed together by heating to 85°C for 5 min, followed by slow cooling.

3. Results and discussion

For cocrystallization trials we purified two 20-bp DNA oligomers, one with blunt ends and the other with a single AT base pair overhang (Table 1). Both oligomers encom-
passed the FokI recognition sequence GGATG at one end and the potential cleavage sites 9 and 13 base pairs away at the other end. The best cocrystals were obtained with the overhanging 20-mer, using macroseeding techniques [10]. The procedure for growing these cocrystals is as follows: Freshly thawed FokI is mixed with the DNA at a molar ratio of 1:1 and allowed to equilibrate on ice for 20 min. The mixture is set up as 2 μl hanging drops containing 0.15 mM protein, 0.75 mM DNA, 1.1 M ammonium sulfate, 0.5 M MES (pH 6.0), 0.2 M KCl, 10 mM potassium phosphate, 0.5 mM EDTA, 0.5 mM EGTA, and 5% glycerol against reservoir solutions containing 2.2 M ammonium sulfate and 1.0 M MES (pH 6.0). Small cocrystals, measuring ~0.03 mm × 0.2 mm × 0.15 mm, are observed within 5 days at 20°C. The small cocrystals are macroseeded by first washing them in a solution containing 1.4 M ammonium sulfate and 0.1 M MES (pH 6.0), and then transferring a single crystal in 1 μl of wash solution to 1 μl of fresh protein/DNA mixture. The cocrystals grow optimally with reservoir solutions containing 1.75 M ammonium sulfate and 1.0 M MES (pH 6.0). The size, quality, and reproducibility of the cocrystals are much improved with sitting drop geometry as opposed to hanging drops [11]. After 2–3 days at 20°C, the seeded cocrystals grow to 1.0 mm × 0.2 mm × 0.15 mm in dimensions and resemble parallelepipeds in shape (Fig. 3). The cocrystals are highly birefringent and stable in their sitting drops for many months. The presence of protein and DNA in the cocrystals was confirmed by PAGE after thoroughly washing crystals in their reservoir solution.

Cocrystals were analyzed by precession photography on a Rigaku rotating anode X-ray source operating at 100 mA and 50 kV. Cocrystals obtained with the blunt-ended oligomer diffracted weakly to a resolution of ~4 Å and were not further characterized. Cocrystals obtained with the overhanging oligomer exhibited much stronger diffraction. These cocrystals belong to space group P21 with unit cell dimensions of a = 67.9 Å, b = 119.8 Å, c = 69.1 Å, and β = 96.6°. Very strong reflections at 3.4 Å along the a axis, characteristic of DNA, suggest that the DNA axis is parallel to the a axis. In accordance, the length of the axis is approximately the length of a 20-bp B-form DNA oligomer.

Assuming a model in which a single FokI molecule is bound to the 20-bp DNA duplex, \( V_{\text{m}} \) [12] calculations left an ambiguity as to the number of protein/DNA complexes contained in the crystallographic asymmetric unit. One protein/DNA complex per asymmetric unit (asu) gives a \( V_{\text{m}} \) of 3.5 Å³/Da, while two complexes give a \( V_{\text{m}} \) of 1.75 Å³/Da. Both of these values are within the range found in macromolecular crystals [13]. Therefore, to determine \( Z_{\text{sym}} \) (the number of molecules per asu) we used volume-specific amino acid analysis [14]. Two large cocrystals were carefully measured to determine their volume and then dissolved and subjected to amino acid analysis. The results from this analysis gave \( Z_{\text{sym}} \) values of 0.91 and 1.07, strongly suggesting that the asymmetric unit contains a single protein/DNA complex.

For data collection, we developed a procedure to flash freeze the cocrystals at 110 K by first soaking them for 8 h in a stabilization solution (2.3 M ammonium sulfate, 1.0 M MES (pH 6.0)) containing 20% glycerol. The fragility of the crystals required transient addition of 0.125% glutaraldehyde [15]. The unit cell dimensions were found to change during freezing to \( a = 65.59, b = 119.34, c = 71.52 \) Å, and \( \beta = 101.4° \). We have measured native data at the Cornell High Energy Synchrotron Source (Beamline A1) from a single flash frozen crystal, utilizing the charge coupled device (CCD) detector. The data extends to 2.8 Å in resolution and merges with an overall \( R_{\text{merge}} \) of 7.5%. We have also measured data from heavy atom derivatives including an iodinated derivative in which contains iodouracil residues in place of thymine residues along the DNA duplex. The structure of FokI/DNA complex prom-

### Table 1

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<th>DNA oligomers used in cocrystallization studies with FokI</th>
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*Underlines indicate the FokI recognition sequence.*
ises to provide a framework for the design of chimeric enzymes with novel specificities.

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**References**


