# Nucleotide sequence of a novel $\delta$-endotoxin gene cryIg of Bacillus thuringiensis ssp. galleriae 

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A gene crylg coding for entomocidal protein $\delta$-endotoxin of Bacillus thuringiensis ssp. galleriae str. 11-67 named CrylG has been cloned and sequenced (EMBL accession number X58120), The deduced amino acid sequence that contains 1156 amino acid residues shows only $28 \%$ of identical residues, when compared with other $\delta$-endotoxins of the Cryl family. The extent of identity is substantially higher for some regions of the sequence ('conserved blocks'), that presumably bear important structural or functional properties. This implies that CrylG $\delta$-endotoxin follows the same type of polypeptide chain folding as other Cryi proteins, whereas peculiarities of primary structure help to explain its unique specifcity.

CrylG: $\delta$-Endotoxin: Primary structure: Genomic library; Bacillus thuringiensis

## 1. INTRODUCTION

Entomocidal protein $\delta$-endotoxins are intensively studied in many laboratories. Their ability to selectively kill only certain species of insects creates a basis for their utilization in plant protection. Nevertheless the mode of the toxic action remains unknown.

Proteinaceous crystals of B. thuringiensis spp. galleriae str. 11-67 are composed of at least two $\delta$-endotoxins of $M_{\mathrm{r}} \sim 130 \mathrm{kDa}$ differing strongly in their immunological properties and entomocidal specificity, named 'positive' and 'negative' components according to the electrophoretic behavior of the respective true toxins [1]. Earlier we reported cloning of the gene coding for the 'positive' component, the endotoxin with unique specificity towards larvae of Galleria mellonella [2]. The partial amino acid sequence of this protein was determined by

Edman's method. Here we present the complete sequence of this gene named crylg, according to the principles of $\delta$-endotoxin classification proposed in [3].

## 2. METHODS

## 2.1. crylg gene cloning

A genomic library of Bacillus thuringiensis ssp. galleriae str. 11-67 was obtained on phasmid vector $\lambda$-pSLS. especially designed for procaryotic genomic library construction. using partial EcoRI digestion of total DNA [5]. Immunoscreening was performed with affinitypurified monospecific antiserum against pure CryIG protein. Restriction mapping of three immunopositive clones and preliminary localization of the toxin gene were reported earlier [1]. pOC10 phasmid. containing the full-size gene was used in further cloning procedures. The phage part of the pOC10 was removed by XbaI deletion, yielding pOK10 plasmid. Exact mapping and localization of the cryIg gene were performed using immunological tests (Fig. 1). KpnI-Xhal insert. carrying the full-size gene, the 521 base-pair $5^{\prime}$-flank and the 1.3 kb $3^{\prime}$-Hank was cloned into the pUCI 8 cloning vector. resulting in the


## CryIG

Fig. 1. Structural organization of the insert of pOK 10 plasmid, containing full-size CrylG gene.

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 ggiacetiticactgtattgiticacttccatcaggititcaaattgaanaactaantgitctacgatgtaagctattiatttatgacgaaggatacgigtaaaaatcgitattgagatiga






 tсtctctiatcacagcacctictctiactggaitaatttcaatagtatatgaccttataggtaaagtactaggaggtagtagtggacaatc catatcagattigictatatgtgacttat

 $L S I I D L R V S O S V L N D G I A D F N G S V L L Y R N Y L E A L D S W N K N$
 ctaattctgcttctcetgaagaactccgtacicgititagaatcgccgactcagaatitgatagaattitaacccgagggtctttaacgaatggiggctcgttagciagacaaaatgcce



 aatcaanactagtagagcttaitgaactatatactgattattgcgtacatiggtataatcgaggtitcaacgaactaagacaacgaggcactagigctacagctiggttagaatticata

 gatatcgtagagagatsacattgatggiattagataiagtagcatcatticaagtctigatattactaaitacccaatagaaacagattitcagitgagiagggicaittaiacagatc R Y R R E M T L MVL D.IVAS FS S L D I T N Y P I E T D F OL S RVI Y T D
 CAATTGGTYTTGTACATCGTAGTAGTCTTAGGGGAGANAGITGGTITAGCTTTGTTAATAGAGCTAATTTCTCAGATTTAGAAAATGCAATACCTAATCCTAGACCGTCTTGGtTtTTAA PIGFVHRSSLRGESWFSFVNRANFSDLENAIPNPDPSWF $\frac{L}{4}$
 ataatatgaitatatctactcgitcacttacaitcccggitaccciaagtactgatagagcgagggiatggtatggaagtcgagatcgaatitccccigctaattcacaatttattactg

 ELIS G O H T TA T O TILLG RNIFRVDSOACNLNDTTYGVNRAV
 ittaicatgatgcgagtgangiftctcaaagatccgigtacgaggggiataitcgaicaactgggatagataaccctagagttcaaantattaacactiatttacctggagaaaaticag FYH D A S E G S Q R S V Y E G Y I R T T G I D N P R V O NI N T Y L P G E N S
 ATATCCCAACTCCAGAAGACTATACTCATATATTAAGCCCAACAATAAATTTAACAGGAGGACTTAGACAAGTAGCATCTAATCGCCGTICATCTTTAGIAATGTATGGTTGGACACATA DIPTPEDYTHILSTTINLTGGLRQVASNRRSSLVMYGWTH








Fig. 2. The nucleotide sequence of a 4156 op tragment of Bacillus thuringiensis ssp. galleriae str, 11-67 DNA and the deduced amino acid sequence of CrylG $\delta$-endotoxin. The $N$-terminal amino acid sequence determined with the Edman method is doubly underlined. Inverted repeat in 3 '-flanking region is underlined. Also shown are the SD-sequence, Kmi and EcoRI sites.
pKP7 plasmid. which allowed expression of crylg gene in E coli using the Lac-promoter. Immunological and toxicity assays of the expression product confirmed its identity with CrylG toxin.

### 2.2. Sequence amelysis

Scquencing of pKP7 was carried out by the SEQUENASE version [4] of Sanger dideoxynuclectide method using a number of subclones in single- or double-strand form obtained in pUC or M13 vectors. The
sequence protocol included use of standard and custom synthetic primers. Sequence data were submitted to EMBL Data Library accession number X58120.

## 3. RESULTS AND DISCUSSION

The phasmid library on the $\lambda$-pSL5 vector has shown good expression properties, producing a quantity of protein, sufficient for immunoscreening in the phage form, although EcoRI site of $\lambda$-pSL5 is not situated in the region transcribed from the Lac-promoter. The strategy based on primary cloning of long inserts with consequent localization and subcloning of the full-size gene appeared to be productive.

Analysis of the DNA-derived amino-acid sequence shows its identity with the N -terminal fragment determined with Edman's method (Fig. 2). The sequence contains an open reading frame of 1156 codons with the AUG initiator codon and TAA terminator ochrecodon. The calculated $M_{\mathrm{r}}$ of the protein product is 129740 kDa ; experimental measurement of the $M_{\mathrm{r}}$ of CryIG has given essentially the same value [1].

Computer assay of 5 '-non-coding region of the CryIG gene failed to reveal sequences similar to known bacillar promoters, which might imply that expression of the crylg gene procuds via polycystronic mRNA. The Shine-Delgarno sequence GGAGGA was found 7 base pairs upstream from the initiator codon (Fig. 2). In the $3^{\prime}$-non-coding region 85 bp downstream from
terminator codon,a 17 -bp inverted repeat was found. The hairpin structure that might be presumed for this repeat cannot be considered a good terminator because the loop part of it is too long. i.e. 13 base pairs.

CryIG differs markedly from all other members [3] of the Cryl family. Its primary structure reveals only a marginal extent of identity when compared with other known members of the CryI family: it does not exceed $21 \%$ for the N -terminal half ( $1-670 \mathrm{bp}$ ) and $35 \%$ for the C-terminal half ( $671-1156 \mathrm{bp}$ ). The identical residues are not evenly distributed along the sequence forming pronounced stretches. In particular, all five conserved blocks of amino-acid residues, common for all CryI $\delta$-endotoxins, have also been found in the CryIG sequence. Characteristically, the identity extent for these blocks is substantially higher. i.e. $60-80 \%$.
A more detailed discussion of these data, especially of the alignment of sequences of Cryl proteins will appear elsewhere.

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