

High-Dose Chemotherapy Plus Non-Cryopreserved Autologous Peripheral Blood Stem Cell Transplantation Rescue for Patients With Refractory or Relapsed Hodgkin Disease

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ABSTRACT

A simplified schedule of high-dose chemotherapy consisting of cyclophosphamide ($60 \text{ mg} \cdot \text{kg}^{-1} \cdot \text{d}^{-1}$ for 2 days), etoposide ($15 \text{ mg} \cdot \text{kg}^{-1} \cdot \text{d}^{-1}$ for 2 days), and carboplatin ($400 \text{ mg}/\text{m}^2$ per day for 2 days) plus autologous non-cryopreserved peripheral blood stem cells (PBSCs) was used for treatment of patients with relapsed ($n = 25$) and refractory ($n = 3$) Hodgkin disease. The use of such PBSCs mobilized by granulocyte colony-stimulating factor after high-dose myeloablative therapy resulted in a rapid, complete, and sustained hematopoietic recovery. The median time to achieve an absolute neutrophil count $>0.5 \times 10^9/\text{L}$ was 13 days (range, 7-18 days). The median time to a self-sustained platelet count $>20 \times 10^9/\text{L}$ was 15 days (range, 7-20 days). Twelve of the 28 patients (43%) were alive and without disease at a median follow-up of 16 months (range, 9-86 months) for all surviving patients. The estimated 2-year overall survival and disease-free survival for all patients were 45% and 42%, respectively. Thirteen patients died of relapse or progressive disease, 2 died of infection, and 1 was still surviving in relapse by the time of the analysis. The median time to relapse was 10 months (range, 3-28 months) from PBSC infusion. High-dose chemotherapy with short-duration chemotherapy and non-cryopreserved bone marrow is an effective and safe treatment modality for patients with relapsed or resistant Hodgkin lymphoma.

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KEY WORDS

High-dose chemotherapy • Resistant Hodgkin disease • Relapsed Hodgkin disease • Non-cryopreserved peripheral blood stem cells • Peripheral blood stem cell transplantation

INTRODUCTION

Approximately 60% to 70% of patients with advanced Hodgkin disease (HD) can be cured with combination chemotherapy [1,2]. However patients who relapse after attaining a complete remission (CR) with chemotherapy or who do not respond to induction chemotherapy and have primary refractory disease are rarely cured with conventional salvage chemotherapy or salvage radiotherapy [3,4]

High-dose therapy followed by autologous stem cell transplantation has been shown to produce long-term disease-free survival (DFS) in selected patients with advanced refractory HD [5,6]. However, the con-

ventional procedures for collection and freezing of peripheral blood stem cells (PBSCs) are time-consuming and expensive [7] and methods to simplify bone marrow transplantation procedures are needed mainly in developing countries

Hematopoietic stem cells appear to maintain viability for several days after collection if carefully stored. Preti et al [8], in a nonrandomized retrospective analysis, compared the engraftment kinetics of 54 patients who received cryopreserved marrow cells with those of 45 patients who received refrigerated cells. The refrigerated cells were stored for a median of 4 days (range, 3-9 days). The cryopreserved cells were stored a median of 69 days (range, 5-981 days) at

–80°C using a cryopreserving mixture of dimethylsulfoxide and hydroxyethyl starch. No significant difference in engraftment kinetics was found between the 2 groups [8].

Refrigerated storage was supplanted by cryopreservation because most transplantation centers develop multiday conditioning regimens. There is limited experience with autologous transplantation of non-cryopreserved hematopoietic stem cells. We present the results of 28 patients with Hodgkin lymphoma who were preconditioned with cyclophosphamide, etoposide, and carboplatin and received non-cryopreserved unmanipulated PBSC transplantation (PBSCT) for their advanced or refractory disease.

METHODS

Between 1996 and 2004, 28 patients with relapsed or refractory HD underwent high-dose therapy and autologous PBSCT at the Bone Marrow Transplant Unit of the Faculty of Medicine, Mansoura University (Mansoura, Egypt). The diagnosis and histologic subtype of HD were based on biopsy results.

Study Definitions and Evaluation

Patients were defined to have induction failure if they had received induction chemotherapy with or without salvage therapy and were never documented to be in a CR. A sensitive relapse was defined as $\geq 50\%$ reduction in the bidimensional measurements of the disease with the use of conventional salvage chemotherapy or radiotherapy. A resistant relapse was defined as $< 50\%$ reduction in the size of the tumor with the use of conventional salvage chemotherapy or radiotherapy. The duration of initial remission was calculated from time of documented CR to time of relapse.

Patients were clinically restaged at the time of entry into the study. Restaging procedures were repeated at day +30, day +90 after transplantation, or as clinically indicated. Follow-up tumor restaging was performed every 3 to 6 months for 2 years and then yearly unless recurrence was suspected. Patients who survived to at least day +30 and had no evidence of tumor by clinical and radiologic evaluation for ≥ 1 month were classified as having a CR. Patients who were received transplants in CR without evidence of disease were considered to be in continuous CR after transplantation. Partial remission (PR) was defined as a reduction $\geq 50\%$ of measurable disease for ≥ 1 month.

Patient Selection

Patients were eligible for PBSCT if they did not achieve CR after first-line chemotherapy (induction failure) or if they had relapsed after a standard che-

Table 1. Patient Characteristics

Item	Frequency (%)
Patients, n	28
Age, median (range)	30 (16-50)
Sex	
Male	19 (68)
Female	9 (32)
Stage at diagnosis	
II	5 (18)
III	15 (54)
IV	8 (28)
B symptoms at diagnosis	12 (43)
Extranodal disease at diagnosis	
Yes	9 (32)
No	19 (68)
Histology	
Lymphocyte predominance	2 (7)
Nodular sclerosis	5 (18)
Mixed cellularity	13 (46)
Lymphocyte depletion	8 (29)
Bone marrow involvement	
Yes	7 (25)
No	21 (75)

motherapy regimen. Patients whose first relapse occurred > 12 months after completion of induction chemotherapy were not candidates for transplantation unless they did not respond to standard salvage chemotherapy or had relapse at extranodal sites. Additional eligibility criteria included age < 60 years, Karnofsky Performance Status score ≥ 70 , left ventricular ejection fraction $> 50\%$, forced expiratory volume in 1 second $> 50\%$, and no major organ dysfunction unrelated to their underlying lymphoma.

The patient characteristics at time of diagnosis are listed in Table 1. The median age was 30 years (range, 16-50 years). There were 19 males and 9 females. Fifteen patients (54%) had stage III disease, 8 patients (28%) had stage IV disease, and 12 patients (43%) had "B" symptoms at time of diagnosis. Nine patients (32%) had extranodal involvement at diagnosis. The pathologic examination revealed lymphocytic predominance in 2 patients (7%), nodular sclerosis in 5 patients (18%), mixed cellularity in 13 patients (46%), and lymphocyte depletion in 8 patients (29%). Bone marrow involvement was detected in 7 patients (25%).

Details of prior therapy are presented in Table 2. All patients were treated with induction chemotherapy for a minimum of 3 cycles or until they had achieved a CR or minimal disease state. The median number of prior chemotherapy regimens was 4 (range, 3-8). Four patients (14%) had received prior radiotherapy at initial presentation or as salvage therapy for relapse. Conventional salvage chemotherapy was given to all relapsed patients (20 patients [71%] received dexamethasone, cytarabine, and cisplatin and 8 patients [29%] received etoposide, methylprednisone, cytarabine, and cisplatin). Most patients received 2-3 cycles

Table 2. Chemotherapy Regimen Before Transplantation

Regimen	Frequency (%)
No. of chemotherapy regimens, median (range)	4 (3-8)
Induction chemotherapy	
MOPP/ABV	17 (61)
ABVD	6 (21)
C-MOPP	5 (18)
Salvage chemotherapy	
DHAP	20 (71)
ESHAP	8 (29)
Radiation therapy	
Yes	4 (14)
No	24 (86)

MOPP/ABV indicates cyclophosphamide, vincristine, procarbazine, prednisone, doxorubicin, bleomycin, and vinblastine; ABVD, doxorubicin, bleomycin, vinblastine, and dacarbazine; C-MOPP, cyclophosphamide, vincristine, procarbazine, and prednisone; DHAP, dexamethazone, cytarabine, and cisplatin; and ESHAP, etoposide, methylprednisone, cytarabine, and cisplatin.

of salvage therapy to achieve maximal response before transplantation.

The patient characteristics at time of transplantation are presented in Table 3. Eight patients (28%) received transplants in second CR. Seventeen patients (61%) received transplants after second or subsequent relapse. The median duration of first CR to first relapse was 10 months (range, 5-16 months). The median time from diagnosis to transplantation was 13 months (range, 5-20 months). Twelve of relapsed patients (43%) had sensitive relapse. Three patients (11%) underwent transplantation after induction with or without salvage chemotherapy or chemoradiotherapy failure. The median Karnofsky Performance Status score was 90% (range, 80%-100%). Seven patients (25%) had extranodal diseases at time of transplantation. The median time from diagnosis to transplantation was 13 months (range, 5-20 months).

Mobilization of Progenitor Cells (Figure 1)

Patients received a single dose of cyclophosphamide (1.5 g/m²) followed after 5 days by 5 μg · kg⁻¹ · d⁻¹ recombinant human granulocyte colony-stimulating factor (G-CSF). Recombinant human G-CSF was continued until the end of leukapheresis.

Collection and Preservation of PBSCs (Figure 1)

After administration of cyclophosphamide, patients had complete blood cell counts measured daily and began leukapheresis when the total white blood counts recovered to 3 × 10⁹/L, usually at the ninth to the eleventh day from the start of mobilization. Leukapheresis was performed daily until the collection of >2 × 10⁸ mononuclear cells/kg and >3 × 10⁶ CD34⁺ cells/kg patient weight. The apheresis solution was

acid citrate dextrose and the final product was stored at 4°C for a maximum of 72 hours without special additives. Viability was assessed by trypan blue dye exclusion test 24, 48, and 72 hours from harvesting.

Conditioning Regimen (Figure 1)

The conditioning regimen for transplantation was started after the end of leukapheresis and given over 2 days. It contained cyclophosphamide (60 mg · kg⁻¹ · d⁻¹ for 2 days), etoposide (15 mg · kg⁻¹ · d⁻¹ for 2 days), and carboplatin (400 mg/m² per day for 2 days). Twenty-four hours after the end of chemotherapy, the non-cryopreserved stem cells were reinfused.

Statistical Analysis

Neutrophil recovery was defined as the first of 2 consecutive days of neutrophil counts >0.5 × 10⁹/L. Platelet recovery was defined as the first of 7 consecutive days with a platelet count >20 × 10⁹/L unsupported by transfusion. DFS was defined as the time from PBSC reinfusion to the time of disease progression or last follow-up. Overall survival (OS) was defined as the time from reinfusion of PBSCs to death or date of last follow-up. Survival distributions were assessed with Kaplan-Meier estimates [9].

RESULTS

Viability Assessment (Table 4)

Trypan blue dye exclusion showed >90% viability in all samples tested 24, 48, or 72 hours from harvesting.

Table 3. Patient Characteristics at Time of Transplantation

Item	Frequency (%)
Status at transplantation	
Second CR	8 (28)
Relapse	
Sensitive relapse	12 (43)
Resistant relapse	5 (18)
Refractory disease	3 (11)
Duration of first CR, median (range)	10 (5-16 months)
Time from diagnosis to transplant, median (range)	13 (5-20 months)
KPS score	
80	7 (25)
90	15 (54)
100	6 (21)
Extranodal disease	
Yes	7 (25)
No	21 (75)

CR indicates complete remission; KPS, Karnofsky Performance Status.

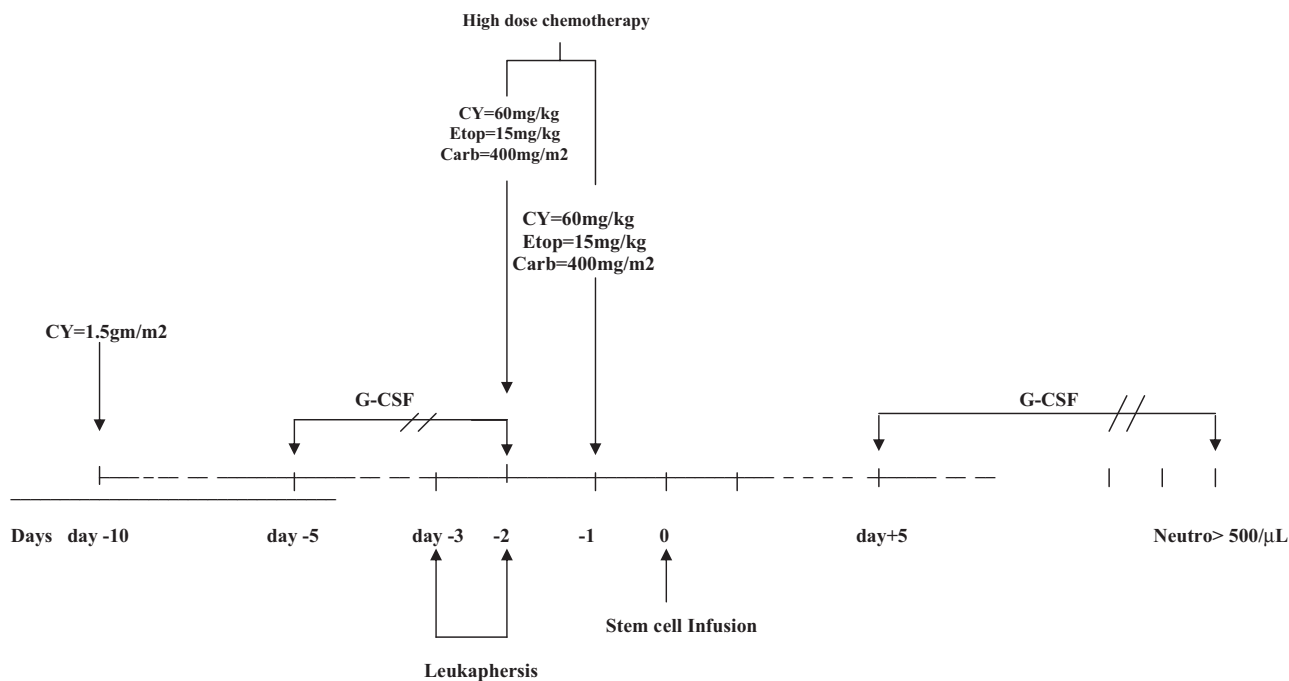


Figure 1. Schema of procedure. Carb indicates carboplatin; CY, cyclophosphamide; Etop, etoposide; G-CSF, granulocyte colony-stimulating factor; Neutro, neutrophil count.

Hematopoietic Recovery (Table 4)

Successful PBSC collection was achieved in all patients. The median number of CD34⁺ cells collected from patients was $6.4 \times 10^6/\text{kg}$ (range, 3.8-24.6 $\times 10^6/\text{kg}$). The use of such G-CSF mobilized PBSCs after high-dose myeloablative therapy resulted in a rapid, complete, and sustained hematopoietic recovery in all patients. The median time to achieve an absolute neutrophil count $>0.5 \times 10^9/\text{L}$ was 13 days (range, 7-18 days). The median time to a self-sustained platelet count $>20 \times 10^9/\text{L}$ was 15 days (range, 7-20 days).

Assessment of Response

All patients were assessable for response. No early deaths had occurred. At 30 days, 19 patients (68%), including the 8 patients who underwent transplantation in CR, were in CR. Five patients (19%) achieved PR. Two patients (7%) had stable disease and 2 patients (7%) did not respond. Without additional treatment, restaging at 3 months showed 22 patients (78%) in CR, 1 patient (4%) in PR, 1 patient (4%) with

stable disease, and 4 patients (14%) with disease progression.

OS and DFS

Twelve of the 28 patients (43%) were alive and disease free at a median follow-up of 16 months (range, 9-86 months) for all surviving patients. The estimated 2-year OS and DFS for all patients were 45% and 42%, respectively (Figure 2). Thirteen patients died of relapse or progressive disease, 2 patients died of infection, and 1 patient was still surviving in relapse by the time of analysis. No evidence of graft failure was present by the time of death of any patient. The median time to relapse was 10 months (range, 3-28 months) from PBSC infusion.

DISCUSSION

High-dose therapy followed by autologous stem cell transplantation has been shown to produce long-term DFS in selected patients with advanced refrac-

Table 4. Viability Data

	Viability %			CD34 Cells $\times 10^6/\text{kg}$	Time to Platelets Recovery (days)	Time to Neutrophiles Recovery (days)	No. of RBCs Transfusions	No. of Platelets Transfusions
	24 hrs	48 hrs	72 hrs					
Median	97.6	96	95	6.4	15	13	5	7
Range	92.4-99.4	91.6-98	91-96	3.8-24.6	7-20	7-18	4-7	5-9

RBC indicates red blood cell.

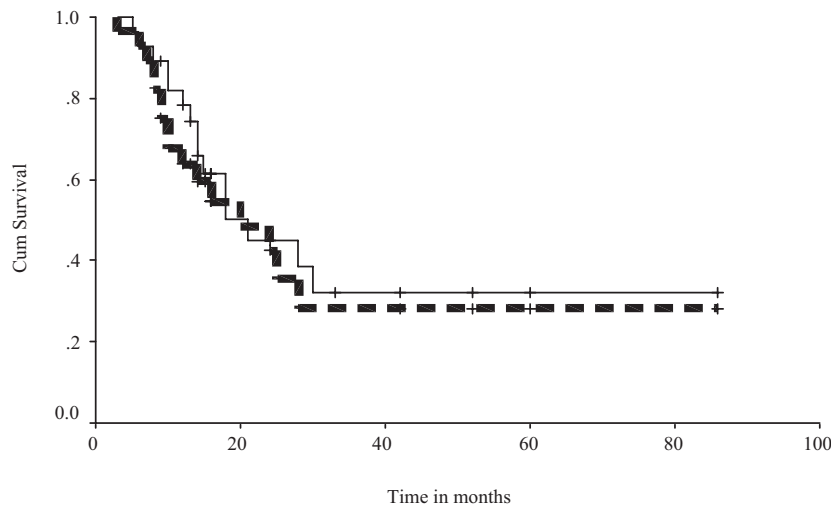


Figure 2: kaplan-Meier product estimate of the cumulative probability of DFS and OS

Figure 2. Kaplan-Meier product estimate of the cumulative probability of DFS (dashed line) and OS (solid line).

tory or relapsed HD. Approximately 40% to 72% long-term DFS has been reported from several series [10]. However, the conventional procedures for collection and freezing of PBSCs are time-consuming and expensive. Methods to simplify bone marrow transplantation procedures are needed mainly in developing countries.

Refrigerated storage for short-term preservation of bone marrow is an alternative to cryopreservation. In an *in vitro* analysis, the stem cell population was concentrated and the nucleated cell recovery, viability, and colony-forming potential were compared after refrigerated storage of whole bone marrow and buffy coat with cryopreserved bone marrow stored for the same interval. Although the nucleated cell recovery for cryopreserved marrow was significantly greater than that for refrigerated storage, the viability and colony-forming potential of the refrigerated storage were superior or equivalent, independent of prior processing [8]. Conversely, no clinically meaningful variation in post-transplantation course and engraftment kinetics between refrigerated storage and cryopreserved stem cells was found [8,11,12].

A simplified schedule of high-dose chemotherapy consisting of melphalan plus VP16 given over 12-18 hours in addition to autologous non-cryopreserved autologous bone marrow transplant was used for treatment of patients with relapsed and refractory disease and as first-line treatment for poor-prognosis HD. This high-dose chemotherapy with short-duration chemotherapy regimen and non-cryopreserved bone marrow was an effective and safe treatment modality for patients with relapsed and poor-prognosis HD [13]. Our study confirms that non-frozen stem

cells are useful to support patients with HD that is treated with myeloablative therapies.

One potential advantage to refrigerated storage is that it may provide an opportunity for extended exposure to growth factors and/or purging agents *in vitro* before transplantation [8]. Storage of PBSCs overnight at 4°C allows pooling of consecutive-day collections, resulting in decreased costs and processing time without compromising neutrophil and platelet engraftment after infusion of CD34⁺-selected progenitor cells [14].

The use of such non-cryopreserved stem cells may yield cost savings when performing autologous stem cell transplantation. In our institution, the cost of the entire transplantation was approximately 12% less when using non-cryopreserved cells than when using a comparable treatment regimen with cryopreserved cells. The difference is mostly attributed to the lower cost of storing and infusing the non-cryopreserved cells. Over a 10-year period, Ruiz-Arguelles et al [15,16] performed autotransplantations using non-cryopreserved and unmanipulated PBSCs mobilized from the bone marrow to the peripheral blood by means of filgrastim and using a single-day conditioning regimen with high-dose (200 mg/m²) melphalan. Their simplified method to perform autografting in patients and avoid cryopreservation of cells also resulted in a decrease in the cost of the autologous hematopoietic stem cell transplantation methods [15,16]. Therefore, it may represent a suitable approach in the management of patients requiring an autologous stem cell transplant in developing countries.

Most studies have used a high-dose combination

chemotherapy regimen. The most commonly used combinations are cyclophosphamide, carmustine, and etoposide [17,18] or bis-chloroethyl-nitrosourea, carmustine (BCNU), etoposide, cytarabine, and melphalan [19,20]. Excellent results were reported with the addition of cisplatin to a modified regimen of cyclophosphamide, carmustine, and etoposide [21,22]. Total body irradiation in combination with high-dose cyclophosphamide was used especially for those who had not received previous radiotherapy [23,24]. Hyperfractionated total lymphoid irradiation in combination with high-dose etoposide and cyclophosphamide has been shown to be efficacious in previously unirradiated patients with relapsing or chemotherapy-resistant HD [5]. At present, there is no evidence to suggest the superiority of 1 regimen over another.

In this study, we used the combination of cyclophosphamide, etoposide, and carboplatin followed by PBSCT.

Cyclophosphamide is a member of the oxazaphosphorine group of nitrogen mustard derivatives and is the most widely used alkylating agent in bone marrow transplantation, based in part on its broad range of antineoplastic activity and immunomodulatory properties [25]. The pharmacokinetics of unchanged cyclophosphamide in the plasma have shown considerable interpatient variability, with a terminal half-life of 3-9 hours [26]. Etoposide (VP16) is the podophylotoxin derivative most commonly used in standard clinical oncologic practice and as a component of many high-dose chemotherapy regimens for hematopoietic malignancies [27]. Caution must be exercised when high-dose etoposide is delivered immediately after high-dose cisplatin, because acute cisplatin exposure can reduce the systemic clearance of etoposide by approximately 25%, leading to a substantially greater than expected etoposide area under the curve and toxicity level [28]. In contrast, high-dose carboplatin exposure may not alter the disposition of high-dose etoposide [29]. The pharmacokinetics of etoposide have been studied over the entire range of intravenous regimens, from 0.1 to 3 g/m². Etoposide, which is highly protein bound, has a biexponential decay with a mean terminal half-life ($t_{1/2}$) of 4-8 hours after high-dose therapy [30]. Carboplatin is an analog of cisplatin in which the 2 chloride ligands are replaced by the carboxylate moiety. Carboplatin disappearance is caused, almost exclusively, by renal excretion [31]. The disappearance of carboplatin from plasma is characterized by a triexponential decay, with a $t_{1/2\alpha}$ of approximately 20 min, a $t_{1/2\beta}$ of about 1.5 hours, and a $t_{1/2\gamma}$ of approximately 7.5-20 hours. Essentially, all unbound platinum species have been cleared from the plasma within 24 hours after completion of a high-dose carboplatin infusion [32].

The combination was beneficial in the treatment of relapsed or refractory HD. DFS over 2 years can be

achieved in some patients. The results are at least equivalent to other published regimens including those based on total body irradiation and to other studies with nearly the same follow-up time [33]. The regimen appears to be a particularly attractive alternative for patients who have received dose-limiting radiotherapy and it should be evaluated further in prospective, randomized trials. Being short term, it allows the use of non-cryopreserved stem cell transplantation.

In conclusion, this simplified approach for performing autografting in patients, avoiding purging procedures, and cryopreservation of the cells is feasible and results in a substantial decrease of the cost of autologous hematopoietic stem cell transplantation methods. On the other hand, the results of the study are encouraging in so far that the DFS and OS are comparable to other protocols of high dose chemotherapy and PBSCT.

REFERENCES

1. Duggan DB, Petroni GR, Johnson JL, et al. Randomized comparison of ABVD and MOPP/ABV hybrid for the treatment of advanced Hodgkin's disease: report of an intergroup trial. *J Clin Oncol.* 2003;21:607-614.
2. Diehl V, Franklin J, Pfreundschuh M, et al, for the German Hodgkin's Lymphoma Study Group. Standard and increased-dose BEACOPP chemotherapy compared with COPP-ABVD for advanced Hodgkin's disease. *N Engl J Med.* 2003;348:2386-2395.
3. Martin A, Fernandez-Jimenez MC, Caballero MD, et al. Long-term follow-up in patients treated with Mini-BEAM as salvage therapy for relapsed or refractory Hodgkin's disease. *Br J Haematol.* 2001;113:161-171.
4. Radman I, Basic N, Labar B, et al. Long-term results of conventional-dose salvage chemotherapy in patients with refractory and relapsed Hodgkin's disease (Croatian experience). *Ann Oncol.* 2002;13:1650-1655.
5. Yahalom J, Gulati SC, Toia M, et al. Accelerated hyperfractionated total-lymphoid irradiation, high-dose chemotherapy, and autologous bone marrow transplantation for refractory and relapsing patients with Hodgkin's disease. *J Clin Oncol.* 1993; 11:1062-1070.
6. Josting A, Katay I, Rueffer U, et al. Favorable outcome of patients with relapsed or refractory Hodgkin's disease treated with high-dose chemotherapy and stem cell rescue at the time of maximal response to conventional salvage therapy (Dex-BEAM). *Ann Oncol.* 1998;9:289-295.
7. Hwang WL, Tsai CS, Jour JH, Kuo LH, Chang YM. Autologous peripheral blood stem cell transplantation using simplified procedures for collection and freezing. *Zhonghua Yi Xue Za Zhi (Taipei).* 1999;62:529-535.
8. Preti RA, Razis E, Ciavarella D, et al. Clinical and laboratory comparison study of refrigerated and cryopreserved bone marrow for transplantation. *Bone Marrow Transplant.* 1994;13:253-260.
9. Kaplan EL, Meier P. Non parametric estimation from incomplete observations. *J Am Stat Assoc.* 1958;53:157.

10. Lazarus HM, Loberiza FR Jr, Zhang MJ, et al. Autotransplants for Hodgkin's disease in first relapse or second remission: a report from the Autologous Blood and Marrow Transplant Registry (ABMTR). *Bone Marrow Transplant.* 2001;27:387-396.
11. Ahmed T, Wuest D, Ciavarella D, et al. Marrow storage techniques: a clinical comparison of refrigeration versus cryopreservation. *Acta Haematol.* 1991;85:173-178.
12. Sierra J, Conde E, Iriando A, et al. Frozen vs. nonfrozen bone marrow for autologous transplantation in lymphomas: a report from the Spanish GEL/TAMO Cooperative Group. *Ann Hematol.* 1993;67:111-114.
13. Seymour LK, Dansey RD, Bezwoda WR. Single high-dose etoposide and melphalan with non-cryopreserved autologous marrow rescue as primary therapy for relapsed, refractory and poor-prognosis Hodgkin's disease. *Br J Cancer.* 1994;70:526-530.
14. Lazarus HM, Pecora AL, Shea TC, et al. CD34+ selection of hematopoietic blood cell collections and autotransplantation in lymphoma: overnight storage of cells at 4 degrees C does not affect outcome. *Bone Marrow Transplant.* 2000;25:559-566.
15. Ruiz-Arguelles GJ, Lobato-Mendizabal E, Ruiz-Arguelles A, Perez-Romano B, Arizpe-Bravo D, Marin-Lopez A. Non-cryopreserved unmanipulated hematopoietic peripheral blood stem cell autotransplant program: long-term results. *Arch Med Res.* 1999;30:380-384.
16. Ruiz-Arguelles GJ, Gomez-Rangel D, Ruiz-Delgado GJ, Ruiz-Arguelles A, Perez-Romano B, Rivadeneyra L. Results of an autologous noncryopreserved, unmanipulated peripheral blood hematopoietic stem cell transplant program: a single-institution, 10-year experience. *Acta Haematol.* 2003;110:179-183.
17. Reece DE, Barnett MJ, Connors JM, et al. Intensive chemotherapy with cyclophosphamide, carmustine, and etoposide followed by autologous bone marrow transplantation for relapsed Hodgkin's disease. *J Clin Oncol.* 1991;10:1871-1879.
18. Stuart MJ, Chao NS, Horning SJ, et al. Efficacy and toxicity of a CCNU-containing high-dose chemotherapy regimen followed by autologous hematopoietic cell transplantation in relapsed or refractory Hodgkin's disease. *Biol Blood Marrow Transplant.* 2001;10:552-560.
19. Chopra R, McMillan AK, Linch DC, et al. The place of high-dose BEAM therapy and autologous bone marrow transplantation in poor-risk Hodgkin's disease. A single-center eight-year study of 155 patients. *Blood.* 1993;81:1137-1145.
20. Argiris A, Seropian S, Cooper DL. High-dose BEAM chemotherapy with autologous peripheral blood progenitor-cell transplantation for unselected patients with primary refractory or relapsed Hodgkin's disease. *Ann Oncol.* 2000;11:665-672.
21. Reece DE, Connors JM, Spinelli JJ, et al. Intensive therapy with cyclophosphamide, carmustine, etoposide \pm cisplatin, and autologous bone marrow transplantation for Hodgkin's disease in first relapse after combination chemotherapy. *Blood.* 1994;83:1193-1199.
22. Subira M, Sureda A, Martino R, et al. Autologous stem cell transplantation for high-risk Hodgkin's disease: improvement over time and impact of conditioning regimen. *Haematologica.* 2000;85:167-172.
23. Stiff PJ, Unger JM, Forman SJ, et al, for the Southwest Oncology Group. The value of augmented preparative regimens combined with an autologous bone marrow transplant for the management of relapsed or refractory Hodgkin disease: a Southwest Oncology Group phase II trial. *Biol Blood Marrow Transplant.* 2003;9:529-539.
24. Nademanee A, Molina A, Fung H, et al. High-dose chemo/radiotherapy and autologous bone marrow or stem cell transplantation for poor-risk advanced-stage Hodgkin's disease during first partial or complete remission. *Biol Blood Marrow Transplant.* 1999;5:292-298.
25. Chabner BA, Myers CE, Coleman CN, Johns DG. The clinical pharmacology of antineoplastic agents (second of two parts). *N Engl J Med.* 1975;292:1159-1168.
26. Huitema AD, Mathot RA, Tibben MM, Rodenhuis S, Beijnen JH. A mechanism-based pharmacokinetic model for the cytochrome P450 drug-drug interaction between cyclophosphamide and thioTEPA and the autoinduction of cyclophosphamide. *J Pharmacokinetic Pharmacodyn.* 2001;28:211-230.
27. Schmitz N, Gassmann W, Rister M, et al. Fractionated total body irradiation and high-dose VP 16-213 followed by allogeneic bone marrow transplantation in advanced leukemias. *Blood.* 1988;72:1567-1573.
28. Relling MV, McLeod HL, Bowman LC, Santana VM. Etoposide pharmacokinetics and pharmacodynamics after acute and chronic exposure to cisplatin. *Clin Pharmacol Ther.* 1994;56:503-511.
29. Kohl P, Koppler H, Schmidt L, et al. Pharmacokinetics of high-dose etoposide after short-term infusion. *Cancer Chemother Pharmacol.* 1992;29:316-320.
30. Newman EM, Doroshow JH, Forman SJ, Blume KG. Pharmacokinetics of high-dose etoposide. *Clin Pharmacol Ther.* 1988;43:561-564.
31. Murry DJ, Sandlund JT, Stricklin LM, Rodman JH. Pharmacokinetics and acute renal effects of continuously infused carboplatin. *Clin Pharmacol Ther.* 1993;54:374-380.
32. van Warmerdam LJ, Van der Wall E, ten Bokkel Huinink WW, Schornagel JH, Beijnen JH, Rodenhuis S. Pharmacokinetics and pharmacodynamics of carboplatin administered in a high-dose combination regimen with thioTEPA, cyclophosphamide and peripheral stem cell support (PSCS). *Proc Am Soc Clin Oncol.* 1995;14:A1503.
33. Lazarus HM, Crilley P, Ciobanu N, et al. High-dose carmustine, etoposide and cisplatin and autologous bone marrow transplantation for relapsed and refractory lymphoma. *J Clin Oncol.* 1992;10:1682-1689.