

## The Marek's Disease Virus (MDV) Unique Short Region: Alphaherpesvirus-Homologous, Fowlpox Virus-Homologous, and MDV-Specific Genes

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Despite its previous classification as a gammaherpesvirus, primarily due to its lymphotropism, Marek's disease virus (MDV), an oncogenic avian herpesvirus, is phylogenetically more related to the "neurotropic" alphaherpesviruses, characterized by its prototype, herpes simplex virus (HSV) (Buckmaster *et al.*, 1988, *J. Gen. Virol.* 69, 2033–2042). In this report we present the DNA sequence of an 11,286-bp DNA segment encompassing the entire 11,160-bp-long  $U_s$  region of the oncogenic avian herpesvirus, Marek's disease virus. Eleven open reading frames (ORFs) likely to code for proteins were identified; of these, 7 represent homologs exclusive to alphaherpesvirus S component genes. These include MDV counterparts of HSV US1 (ICP22), US2, US3 (a serine-threonine protein kinase), US6, US7, and US8 (HSV glycoproteins gD, gI, and gE, respectively), and US10. Three additional ORFs were identified with no apparent relation to any sequences currently present in the SwissProt or GenBank/EMBL databases, while a fourth was found to exhibit significant homology to an uncharacterized fowlpox virus (FPV) ORF. Having precisely identified the  $IR_s-U_s$  and  $U_s-TR_s$  junctions, we have corrected and clarified their previously reported locations. By characterizing genes encoding three new alphaherpesvirus-related homologs (US1, US8, and US10), completing the sequence for a fourth (US7), and identifying 2 new MDV-specific ORFs (SORF1 and SORF3) and a fowlpox homolog (SORF2), our sequence analysis of the "virulent" GA strain of MDV (vMDV) extends upon that of a 5255-bp segment located in the  $U_s$  region of the "very virulent" RB1B strain of MDV (vMDV) (Ross *et al.*, 1991, *J. Gen. Virol.* 72, 939–947; 949–954). These two sequences were found to exhibit 99% identity at both nucleotide and predicted amino acid levels. Combined with the fact that MDV  $U_s$  sequences failed to show statistically significant CpG deficiencies, our analysis is consistent with MDV bearing a closer phylogenetic relation to alphaherpesviruses than to gammaherpesviruses. Because alphaherpesvirus-specific  $U_s$  region genes are primarily nonessential for virus replication, they are thought to be important biological property determinants. Thus, our sequence provides a foundation for further MDV studies aimed at resolving the apparent discrepancy between MDV's genetic and biologic properties. © 1995 Academic Press, Inc.

### INTRODUCTION

Marek's disease virus (MDV) is a highly pathogenic herpesvirus of chickens, which can cause: (i) T-cell lymphomas as early as 3 weeks postinfection; (ii) peripheral neural lesions, characterized by lymphoproliferative infiltration and demyelination, occasionally leading to paralysis and/or blindness; (iii) various phenomena of acquired immunodeficiency; and (iv) atherosclerosis in normocholesterolemic chickens, bearing a remarkable resemblance to the human disease, in both character and distribution of arterial lesions (reviewed in Calnek and Witter, 1991). Marek's disease (MD) most commonly refers to the lymphoproliferative conditions above and is noteworthy for being the first naturally occurring lymphomatous neoplasm to be effectively controlled by vaccination (Churchill *et al.*, 1969).

Because of similar biological properties, especially its lymphotropism, MDV and its antigenically related, apathogenic vaccine virus, herpesvirus of turkeys (HVT), have until recently been classified as gammaherpesviruses (Roizman *et al.*, 1981; Roizman, 1992). In contrast to gammaherpesviruses, MDV and HVT have genome structures more closely resembling those of alphaherpesviruses (Cebrian *et al.*, 1982; Fukuchi *et al.*, 1985; Igarashi *et al.*, 1987). Consistent with their structural relatedness to alphaherpesviruses, recent data indicate that MDV and HVT are phylogenetically more related to alphaherpesviruses than gammaherpesviruses (Buckmaster *et al.*, 1988). This raises interesting questions regarding the seeming incongruence between MDV's genetic and biologic properties. To understand the nature of these differences and to identify new glycoproteins potentially important in virus–host cell interactions and mechanisms of protective immunity against MD, we have become particularly interested in the MDV  $U_s$  region. This stems from the observation that alphaherpesvirus  $U_s$  regions are known to contain a cluster of glycoprotein genes and appear to specify determinants for pathogenesis and viral dissemination, rather than those essential for virus production (Roizman, 1990a). These determi-

Sequence data from this article have been deposited with the EMBL/Gen Bank Data Libraries under Accession No. L22174.

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nants are encoded by a cluster of "nonessential" or *supplementary essential* (Roizman, 1990a) genes which are likely to account for many of the unique *in vivo* properties characteristic for a given alphaherpesvirus. The natural host-MDV system affords a unique opportunity to examine the *in vivo* role and function of this putative class of supplementary essential genes.

The alphaherpesvirus U<sub>S</sub> region is flanked by a pair of inverted repeat sequences (inverted and terminal repeat short, IR<sub>S</sub> and TR<sub>S</sub>, respectively, or simply, repeat short, R<sub>S</sub>). Together, these components make up the S region. Alphaherpesvirus U<sub>S</sub> and other S region genes originate from an area specific to members of this group, arguably their most divergent coding region. Second they specify a cluster of glycoprotein genes important for virus-cell interactions and mechanisms of protective immunity. A similar cluster in MDV would be particularly significant given the paucity of details regarding protective immunity against naturally occurring MD tumors.

The DNA sequence of a 5255-bp segment from U<sub>S</sub> region of the "very virulent" RB1B strain of MDV (vMDV) was recently reported (Ross *et al.*, 1991). This region was found to contain open reading frames (ORFs) homologous to proteins encoded by HSV US2, US3 (protein kinase), US6 (glycoprotein D), part of US7 (glycoprotein I), and an additional MDV-specific ORF. In this report, we extend these results and present a sequence analysis of the entire 11.2-kbp MDV U<sub>S</sub> region ("virulent" GA strain, vMDV).

## MATERIALS AND METHODS

### Recombinant plasmids, M13 subcloning, and DNA sequencing

Pathogenic MDV GA strain subclones included *EcoRI*-O, -I, and -V cloned into pBR328 (Gibbs *et al.*, 1984) (pE328-O, pE328-I, and pE328-V, Fig. 1B); *BamHI*-A and *BamHI*-P1, cloned into pACYC184 and pBR322, respectively (Fukuchi *et al.*, 1985) (pBACYC-A and pB322-P1 (Fig. 1B), kindly provided by Dr. Meihan Nonoyama of the Tampa Bay Research Institute, St. Petersburg, FL); and GA-02, a phage clone containing a partially digested MDV *Sau3A* insert cloned into the *SaI* site of EMBL3, kindly provided by Dr. Paul J. A. Sondermeijer, Intervet International, Boxmeer, The Netherlands. The latter clone contains most of *BamHI*-A, all of *BamHI*-P1, and additional 3' flanking sequences, including some of those present in pE328-V. This phage clone was used to generate the pUC18 subclone, pSP18-A (Fig. 1B). This clone contains a 2.5-kb *SaI* insert with approximately 20 bp of EMBL-3's multiple cloning site at its 3' end. Together, the above clones (Fig. 1B) were used to generate M13mp18 and -19 subclones for use as templates for nucleotide sequencing.

DNA sequencing of both strands was performed by the dideoxy chain-termination method (Sanger *et al.*,

1977) using single-stranded M13 templates. Reaction products were synthesized and labeled using a 17-mer M13 primer, a modified T7 DNA polymerase (Sequenase), [<sup>35</sup>S]thio-dATP (NEN), and appropriate deoxy- and dideoxynucleotides according to instructions by the manufacturer (Sequenase sequencing kit; United States Biochemical Corp., Cleveland, OH) and electrophoresed through 7% polyacrylamide/50% urea/Tris-Borate-EDTA gels. Remaining sequence gaps were determined by substituting M13 primers with synthetic 17-mer oligonucleotides (under similar reaction conditions, 0.5 pmol/reaction).

### Analysis of sequence data

Sequences were assembled and analyzed with an IBM Personal System 2/Model 50 microcomputer utilizing Genepro (Version 4.10; Riverside Scientific Enterprises, Seattle, WA) sequence analysis software packages or programs obtained from the University of Wisconsin Genetics Computer Group (UWGCG, Versions 6.2 and 7.0; Devereaux *et al.*, 1984) and run through a VAX 8650 mini-computer. Homology searches of the SwissProt (Release 18.0), GenBank (Release 71.0), and EMBL (Release 30.0) databases were performed using the UWGCG programs FASTA and TFASTA (Pearson and Lipman, 1988).

## RESULTS

### Defining the MDV U<sub>S</sub> region and the location of the unique-repeat region junctions

Figure 1 contains a map of the area that was sequenced. This segment is bounded by a pair of *PvuII* sites and spans the 3' half of *BamHI*-A, extending an additional 1.5 kbp to the right of the 3' end of *BamHI*-P<sub>1</sub> (Figs. 1B and 2). This 11,286-bp segment spans the entire U<sub>S</sub> region, which is 11,160 bp in length and flanked at the 5' and 3' ends by a 63-bp stretch of IR<sub>S</sub> and TR<sub>S</sub> DNA, respectively, each inversely complementary to the other (Figs. 1B and 2). Based on Southern blot analysis, the IR<sub>S</sub>-U<sub>S</sub> junction was previously localized to a 1.4-kb *BglI* fragment (Fukuchi *et al.*, 1985) located in the second of five *EcoRI* subfragments of *BamHI*-A (for *BamHI*-A/*EcoRI* map, see Wen *et al.*, 1988). Our sequence analysis demonstrates that the IR<sub>S</sub>-TR<sub>S</sub> junction is instead located in the middle of the third *EcoRI* subfragment, approximately 2-3 kbp downstream from the position reported above. This is further supported by Southern blot analysis of genomic DNA from the same strain (GA) as above (data not shown). Consequently, the U<sub>S</sub>-TR<sub>S</sub> junction was localized 263 bp downstream of the US8 termination codon (following position 11,223, Fig. 2).

### Nucleotide sequence and identification of open reading frames

The overall guanine plus cytosine ratio of the region sequenced was found to be 41%, somewhat below the

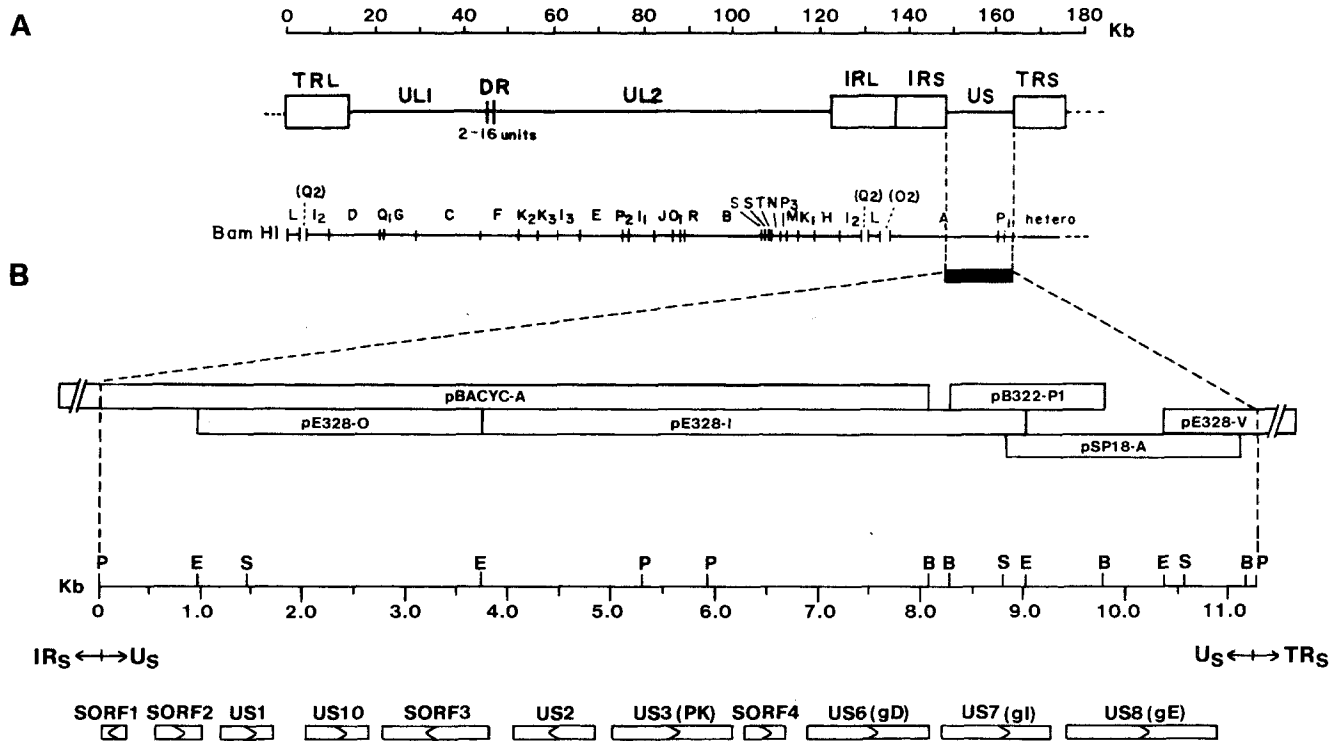


Fig. 1. Map location of area sequenced and organization of MDV U<sub>S</sub> ORFs. (A) MDV genome structure and *Bam*HI restriction map outlining area sequenced. Restriction enzymes: B, *Bam*HI; E, *Eco*RI; P, *Pst*I; and S, *Sal*I. Upper boxes define plasmid clones with *Bam*HI, *Eco*RI, or *Sal*I-bound inserts that were used to generate M13mp18 and -19 templates for sequencing. (B) Organization of MDV U<sub>S</sub> ORFs. Lower boxes (with arrows) represent location of MDV ORFs. Arrows define direction of transcription/translation. Names of ORFs are displayed above boxes. Basis for nomenclature is outlined under Results.

reported genomic value of 46% (Calnek and Witter, 1991). Observed frequencies of CpG dinucleotides in the whole sequence, or in the coding regions only, did not differ significantly from those expected from their mononucleotide compositions (data not shown). This result agrees with those obtained from alphaherpesviruses, while sharply contrasting with the CpG deficiencies associated with all gammaherpesviruses thus far studied (Efsthathiou *et al.*, 1990; Honess *et al.*, 1989).

The region sequenced contains at least 11 ORFs likely to code for proteins (Fig. 1B; basis for names is defined below). This prediction was primarily based on homology and positional organization comparisons to other alphaherpesvirus genes, as well as the observation that alphaherpesviruses generally tend to contain relatively tightly packed, unspliced coding regions (Davison and Scott, 1986; McGeoch *et al.*, 1985, 1987, 1988; Telford *et al.*,

1992). Methods for detecting protein coding regions based on the use of MDV-derived codon frequency tables (using these and previously published MDV sequences; Binns and Ross, 1989; Chen *et al.*, 1992; Jones *et al.*, 1992; Ross *et al.*, 1989; Scott *et al.*, 1989) or analysis of compositional bias (using the UWGCG programs CODONPREFERENCE and TESTCODE) were inconclusive. However, as pointed out previously (Ross *et al.*, 1991), MDV-encoded ORFs do exhibit a detectable bias for A-T residues in the wobble position. Furthermore, using the UWGCG program FRAMES, together with the MDV-derived codon frequency table above, the 11 identified ORFs clearly show a significantly low pattern of rare codon usage not observed following computer-based translation of the remaining reading frames (data not shown).

The predicted amino acid sequences of the above

Fig. 2. Nucleotide and predicted amino acid sequences. The nucleotide sequence is given as the rightward 5' to 3' strand only (numbered 1 to 11,286). IR<sub>S</sub> and TR<sub>S</sub> sequences are located at the 5' and 3' ends, respectively, and are depicted using lowercase symbols; U<sub>S</sub> sequences are in uppercase. Rightward- and leftward-directed predicted amino acid sequences are shown above and below the corresponding nucleotide sequences, respectively, in single-letter code. The name of each ORF is given to the left of the first line of its respective sequence. Amino acid sequences are numbered from the N terminus, beginning with the first in-frame methionine codon and ending with the amino acid at the C-terminus, which precedes the termination codon. Dotted lines identify potential polyadenylation signals. Putative signal peptide and transmembrane domain regions of MDV US6 (gD), -US7 (gI), and -US8 (gE) are overlined at the amino and carboxy ends, respectively. Signal peptide overlining continues through to the last amino acid to the left of the predicted cleavage site (von Heijne, 1985). Potential N-glycosylation sites (N-X-S/T) are indicated by dashed lines.

		<----IRs	
	1	cagctgctgattttccccgtgcatctcataccgccatttttgggtagaggtatatttttattAGCCAAATCGTATTCCTGGAAGTTTGACAAAACCTGT	100
		89 * G F R I G P L K V F G T	78
	101	CAAACCTGACACGGTCCAAGCGAAACTCGAAAAAAAAGGGGGGGGGGAGAATATTCTGTAGGACCGGCAGAAGTTCTCAAGGCAGAGGAAAGATACACA	200
	78	L S V R D L R F E F F F P P P P S Y E T P G A S S R L A S S L Y V	45
	201	TTATTTTTTGTAGATTTAGGCAAGTTTTGCAGAACCTGCAGGGAATGTATACACCATCAAATCTACTCGACTTATTGCTTGAGTCCAATTTAACAGAAA	300
	44	N N K T L N L C T K C F R C P I Y V G D F R S S K N S S D L K V S	12
SORF1	301	TAAAATATATTGATGTTGCCACATATGCATCCTCGCATATGGGGTGGGACACAGGACGATTATATCCCCAGACATGAACCTCAAACCTGCCATTTTGT	400
	11	I L I Y Q H Q S M H M	1
	401	CCCATCATTGGAGAGACAAAATCGCATACATCCTACTTATCGCACACATTGGATGTCGGTCTTTATTTCAGGCCATATCAGCTTTCACGGGGCAAATTCG	500
SORF2	1	M Q R Q T G H M E D K K R T G L E S Q G T E N A F S D	27
	501	TATTCATAGATCCGTCATCGATGCAGCGCCAAACCGGACATATGGAAGCAAAAAGAGAACCGGTTTGGAAATCGCAGGGGACCGAAGTGCATTTTTCAGA	600
	28	G R D G K D G L L H E G I N E P I L I P S T I A D L E G I R E L V	60
	601	TGGCAGAGATGGCAAAGATGGATTGTTACATGAAGGAATTAATGAGCCATTTTATTCCGCTACCATCGCAGATCTCGAGGGGATTCGTGAATTGGTC	700
	61	R K F R G R L L P F E K C P D F C L R I G G L E A S F H K G Q E E L	94
	701	CGAAAATTCGCTGCTACTGCCCTTTGAAAAGTGTCCGATTTTGTCTGAGAATTTGGGGTTGGAGGCCAGCTTCATAAAGGCAGGAGGAGC	800
	95	L E Y C E A L Y L P Q P V K M E I V G I V D D V P C L A T G M Q L	127
	801	TGTTAGAGTATTGTGAAGCACTTATTACCACAACCTGTAAGATGAAATAGTAGGCATTGTAGACGATGTCCATGTCTGGCAACGGGGATGCAATT	900
	128	L I L V A E G G E V Y A Y E E D T L H K L A T S F S E F L E I G V	160
	901	ACTCATCTTGTGCCAGGGGAGAGGTATATGCCTATGAAGAAGTACTGCAATAGTCCAGGAGTTTTCCGAATCCTTGAATTTGGAGTG	1000
	161	K S L G R E V Y H C G E Y I E Q V V H * 179	
	1001	AAATCTTTAGGGAGGAGGTTTACCATTGTGGAGAATATAGAGCAAGTAGTACATTAGGGCTGGGTAAAGACCAAGTAATTTTTCACCGGATATCA	1100
	1101	CGTGATGTAATTTAGCAATTTATTGTTCTAGCAGAAGATAAAGCTGTAAGTATATAATACAGGCCAAAGTCTCCAATTTACACTTGAGCAGAAAAAC	1200
US1	1	M S R D R D R A R P D T R I S S S S D N E S D D E D	25
	1201	CTGCTTTCCGCTCCATCGGAGGCAACATGAGTCGTGATCGAGATCGAGCCAGACCCGATACACGATTATCATCGTCAGATAATGAGAGCGACGACGAAGA	1300
	26	Y Q L P H S H P E Y G S D S S D Q D F E L N N V G K F C P L P W K	58
	1301	TTATCACTGCCAATTCACATCCGGAATATGGCAGTGACTCGTCCGATCAAGACTTGAACATTAATAATGTGGGCAAAATTTGTCCTCCTACCATGGAAA	1400
	59	P D V A R L C A D T N K L F R C F I R C R L N S G P F H D A L R R A	92
	1401	CCCAGTGCCTCGGTTATGTGGGATACAAACAACTATTTTCGATGTTTATTTCGATGTCGACTAAATAGCGGTCGTTCCACGATGCTCTTCGGAGAG	1500
	93	L F D I H M I G R M G Y R L K Q A E W E T I M N L T P R Q S L H L	125
	1501	CACTATTCGATATTCATATGATTGGTGAATGGGATATCGACTAAAACAAGCCGAAATGGGAAACTATCATGAATTTGACCCACGCCAAGTCTACATCT	1600
	126	R R T L R D A D S R S A H P I S D I Y A S D S I F H P I A A S S G	158
	1601	GCGCAGGACTCTGAGGATGCTGATAGTCGAAGCGCCATCCTATATCCGATATATGCTCCGATAGCATTTTTACCCAATCGCTGCTCCTCGGGA	1700
	159	T I S S D C D V K G M N D L S V D S K L H * 179	
	1701	ACTATTTCTCAGACTCGGATGTAAGAAGTGAACGATTTGTCGGTAGACAGTAATTCGATTAACATCCAGACTTGAAGAGAAGCTCTTATTATAT	1800
	1801	AATTTAAATTTGTTAGACATAGAGCCGACATTTCTTGTATCTAATGAGATAAAAATAAGATTTGGATTTATTGTCATGATCTGTTGCAACAAAACG	1900
	1901	CTGACCCCCCATCCATGAAGGGCGTGTCAAATAACGTTGCTTTTTGTTGATATGAAGATATTAAATGTGGCGTGAGCCTAATGAGAGGAGAA	2000
US10	1	M A M W S L R R K S S R S V Q	15
	2001	CGTGTTTGAATACTGGAGACGAGCCCGTGTAAAGATTAACAATATTTGGAGAGGTATGGCCATGTGGTCTCTACGGCGCAAATCTAGCAGGAGTGTGCAA	2100
	16	L R V D S P K E Q S Y D I L S A G G E H V A L L P K S V R S L A R T	49
	2101	CTCCGGTAGATTCTCCAAGAAGACAGAGTATGATATACTTTCTCCGGCGGGGAACATGTTGCGCTATTGCTCAAAATCTGATCGCAGCTAGCCAGGA	2200
	50	I L T A A T I S Q A A M K A G K P P S S R L W G E I F D R M T V T	82
	2201	CCATATTAACCGCGCTACGATCTCCAGGCTGCTATGAAAGCTGGAACACCACCTCGTCTCGTTTGTGGGGTGAATATTCGACAGAATGACTGTCAC	2300
	83	L N E Y D I S A S P F H P T D P T R K I V G R A L R C I E R A P L	115
	2301	GCTTAACGAATATGATATTTCTGCTTCGCCATTCACCCGACAGACCCGACGAGAAAAATTTAGGCGGGCTTTACGGTGTATTGAACGTGCTCCTCTT	2400
	116	T H E E M D T R F T I M M Y W C C L G H A G Y C T V S R L Y E K N V	149
	2401	ACACACGAAGAAATGGACACTCGGTTTACTATCATGATGATTGGTGTGCTTGGACATGCTGGATACTGTACTGTTTCGGCTTATATGAGAAGAATG	2500
	150	R L M D I V G S A T G C G I S P L P E I E S Y W K P L C R A V A T	182
	2501	TCCGCTTATGGACATAGTAGTTCCGCAACGGGCTGTGGAATAAGTCCACTCCCGAATAGAGTCTTATTGGAACCTTTATGTCGTGCGGCTGCTAC	2600
	183	K G N A A I G D D A E L A H Y L T N L R E S P T G D G E S Y L * 213	
	2601	TAAGGGAAATGCAGCAATCGGTGATGCTGAATTTGGCACATTATCTGACAAATCTTCGGAAATCGCCAACAGGAGACGGGAATCCTACTTATAACTA	2700
	2701	ATCGACAATTAATAGGATTTTAGGAAAACTGCTACTAACGTTGTTAAATAATAAATTTTATTTCATAAAGGCATTACAGTGTGTCATGATT	2800
	2801	GTATGATTAATATGGGTATGCATGAGGATTAATCGATTGAAACTTTGCTAAATGCTGTAGGATTTACTATTATTAGTCTGGATCGAGGCGGACG	2900
		351 * I P Y A H P N S R N F S Q R F T Q L I K S N M L R S R P P R	322
	2901	TAAATGGAGATTGCGGCAAATGTAGGGGTGCTGGTACATAAGACCTCCAACATCCATCGACTCATCGGCCTCGCTCCAATGGATATGTTGATGTACCT	3000
	321	L H L N R C I Y P H Q Y M L G V D M R S M P R R G F P Y T S T G	289
	3001	TGTAAGATTATGACATTAGAAGATCGATGGTGAATAGTGGGATCTATCCATGCTATTCTCAATATGCATGATATGCAATGTTCCCGGTTAGGTTTGA	3100
	288	Q L T I V N S S R H H I T P D I D M S N E I N C S I C H E R N P K	256
	3101	TAAGATCATGTATGTTCTATAATAACAACCTCCTCTCAGAAGAAATCAATTTTATTATGTCACCTGCTTGGATATTCAGTTTCTGTCAATCGATTCCG	3200
	255	I L D H I T R Y Y L E E E S S D N I K H G S D K S I G T E T L R N A	222

	3201	TTGCATTTCGGTGCAGCATGCTTGTATGGCATTTCCTATGCTATCATCCGGCAGGCTAAGGGTGTCTATACTCGCACACAGGTAGAGCAAGAACCAGC	3300
	221	Q M Q T C C T K I A N G I S D D P L G L P T R Y E C V P L A L V V	189
	3301	GCATATCGAGCTACCTCTATTGCCCGCTAAGGACATTTCTTGCAGACTGATTGTGCATGAACATATTTTCGTGATTGTGTGCATATAACCCCTTGTGA	3400
	188	A Y R A V E I A G S L V N R A S Q I T M F M N R T N H R D Y G K N	156
	3401	TTCATGGAAGCATTGTGGTCCAGTTTTCAGATGAAATGAAAACAATCGGGGAAAAATGGTCCCACCTGTTTCATCTTCAATGCATCTCTCACATC	3500
	155	I G I S L M T T W N E L H F S F L A P L F P G V Q K M K L A D R V D	122
	3501	CCAAGTCTATAGAATATTTCCACTGACCAGTTTCGGTAAGATCAGTTTCTGTAATTTGTGATAGTTTCAATCGAAAACATTTTGTCCATCATGGCA	3600
	121	W T R Y F I R W Q G T E T L D T E T F N T I T E I S F M K D M M A	89
	3601	AAAAATCTATAGGCAGACCAGATAACCAATTTGACACCACATATCCTTGTGTATATCAAACGATGTAATAGATCCCTCGTTAGTAGATATGGTACATAAAA	3700
	88	F F R Y A S W I V M Q C W M D K H I D F S T I S G E N T S I T C L	56
	3701	GGCCTAATCTCTCGGGCTCCATACATTGAACGATTCTTCTGTGAATTCATCAACAACACATGCCAAAAATTTACATTAGTAATCTTTCTCGGTGG	3800
	55	L G L R E R A E M C Q V I G E T F E D V V V H W F N V N T I K R P P	22
SORF3	3801	CTTACCAAATCGTCTCTTGGTATATCCATATCATCGAACATTGTAGCATTGACTCTGCTCATCGTTGTCTTTCAAATGCGCTCGATTGTTGAATCTCTC	3900
	21	K G F R G R P I D M D D F M T A N V R S M	1
	3901	CTGATGTTAGAAGTATATGGAAGATAGCCTGGATACATAAGTATAGTATAGAGGGTGTGTTATTGCACTAATATACAAATTATACGTGACACTATAGCGAC	4000
	4001	GGTTGAGCGATGCACCTAATCGTAATGTGTATACGCCCATCATGTAATTATATCTAATTGGTAGCAAGTAGGCTCGAATAACAGCTAATGACTAC	4100
		270 * H S	269
	4101	CGGCTCTACATTTTTCTGTATTCGTGACTTTCTGTGCGAGTGAACGAACCGGAATGCAATCGCATCTCTATCTTCTTTCTGCAACATTTTCCACA	4200
	268	G A R C K K Q I R S K G T A T Y R V P I A I A D R D E K K C C K G C	235
	4201	ACAGAATAATCTCGCGGGTACTACTCATTGAGGTGGTTCGATTCCGGAGGTTTTAGAGGATTSGGTGGGGACCCGAGGATTTGTATACATACC	4300
	234	C F L R G P T S S M Q P P E I E P P K L P N P P S G L I K Y V C V	202
	4301	ATACACTGTGCAAAAATCGCTCTATCTTCTGGGGTGTGCAACTTCGGTCCCATTGATAGTGTCAAGAGAGTTGAATATTGTGGGAAATGGCCACG	4400
	201	M D S D C F H A R D E P T D A F K P P E W T S T L L T Q I N D P I A W	169
	4401	GCATACCGGACCGGTCAGCAGACTTTGATTGCAAGTAACCTTTTGGCAAAGGAATACATTCGAGCGCAATGGCACAATATCTGCCGCCCAACTAT	4500
	168	P M G S W T G S V K I A L L R K P L P I C E L A I A C I D A A G V I	135
	4501	CCACAAGCTATGTGGAGCATTACCAGAAACTTCAAGTTCACATCAAATATCCAGATAGAACATCCTGCCATTCTGTGGAACATCCTGCAACATCTTCA	4600
	134	W L S H P A N G S V E S E L M L Y G S L V D Q W E T S C G A V D E	102
	4601	AATAGCCGCACTATAACGAATCCCTAGTTCGGCCAATCCGGTACCACGAACCTCCAGTTCATCTGGTGGCTTTGTCCTTACTATCGGTCGATGTTGCC	4700
	101	F L R V I F S D R T G A L G T G R V G T G D P P K T R V I P R H Q	69
	4701	GAGGAAGAATTAACATGGGTTTGGCAAAACGGAATAGGCTGCGAGCTCTGGCGATTATGGGACACCCACATCATCCTGTATTTGTTCCATACATTGCTT	4800
	68	R P L I L M P K A F R F L D A A R A I I P V G V D D Q I Q E M C Q K	35
	4801	TATAAGGAATCCATAAAGTAGATGCAGCATCTAGACTTCTCTGGCAATCGATCGCAATCATCTAGAAGTGTGACTATAGTTATCGGACACCCC	4900
	34	I L F I W L T S A A D R S R G P L R D C E D L L T V I T I M S V G	2
US2	4901	ATCTTACCTCCACCAATAATCTTTTTATTGTTAATAACTGGCCGGTCTGATCTCCAATCTTATACTCTGGTAGAATATGAACAGGGTTAAAACTA	5000
US3	1	M	
(PK)	1	M S S T P E A E T M E C G I S S S K V H D S K T N T T Y G	29
	5001	GGTAATAGACTGGATGTCTTCGACTCCGGAGGCGAAGACGATGGAATGTGGCATTTCCTCGTCAAGTACACGACTCTAAAATAACTACTACCTACGGA	5100
	30	I I H N S I N G T D T T L F D T F P D S T D N A E V T G D V D D V K	63
	5101	ATTATACATAACAGCATCAATGGTACGGATACGAGCTGTTTGTACTTTCCGACAGTACCGATAACCGGGAAGTACGGGGATGTGGACGATGTGA	5200
	64	T E S S P E S Q S E D L S P F G N D G N E S P E T V T D I D A V S	96
	5201	AGACTGAGAGCTCTCCGAGTCCCAATCTGAAGATTGTCACTTTTGGGAACGATGGAATGAATCCCCGAAACGGTACGGACATTGATGCAAGTTTC	5300
	97	A V R M Q Y N I V S S L P P G S E G Y I Y V C T K R G D N T K R K	129
	5301	AGCTGTGCAATGCAGTATAACATGTTTCATCGTTACCGCCCGGATCTGAAGGGTATATCTATGTTGTACAAAGCGTGGGATAATACCAAGAGAAA	5400
	130	V I V K A V T G G K T L G S E I D I L K K M S H R S I I R L V H A Y	163
	5401	GTCATTGTGAAAGCTGTGACTGGTGGCAAAACCTTGGGAGTAAATGATATATAAAAAATGCTCACCGCTCCATAATTAGATTAGTTACATGCTT	5500
	164	R W K S T V C M V M P K Y K C D L F T Y I D I M G P L P L N Q I I	196
	5501	ATAGATGGAATCGACAGTTTGTATGGTAATGCCTAAATACAAATGCGACTTGTTCAGTACATAGATATCATGGACCATTGCCACTAAATCAAATAAT	5600
	197	T I E R G L L G A L A Y I H E K G I I H R D V K T E N I F L D K P	229
	5601	TACGATAGAACGGGGTTGCTTGGAGCATTGGCATATATCCACGAAAAGGTATAATACATCGTGTGTAATAACTGAAAATATATTTTGGATAAACCT	5700
	230	E N V V L G D F G A A C K L D E H T D K P K C Y G W S G T L E T N S	263
	5701	GAAAATGATGATTTGGGGACTTTGGGGCAGCATGTAATTAGATGAACATACAGATAAACCCAAATGTTATGGATGGAGTGGAACTCTGGAACCAATT	5800
	264	P E L L A L D P Y C T K T D I W S A G L V L F E M S V K N I T F F	296
	5801	CGCCTGAACCTGCTTGCATCTGATCCATACTGTACAAAACCTGATATAGGAGTGCAGGATTAGTTCTGTTGAGATGTCAGTAAAAAATAACCTTTTT	5900
	297	G K Q V N G S G S Q L R S I I R C L Q V H P L E F P Q N N S T N L	329
	5901	TGGCAACAAGATAACGGCTCAGGTTCTCAGCTGAGATCCATAATTAGATGGCTGCAAGTCCATCCGTTGGAATTTCCACAGAACAATTTCAACAACTTA	6000
	330	C K H F K Q Y A I Q L R H P Y A I P Q I I R K S G M T M D L E Y A I	363
	6001	TGCAAACTTCAAGCAGTACGGATTACGATACCATATGCAATCCCTCAGATTATACGAAAGAGTGGTATGACGATGGATCTGGAATATGCTA	6100
	364	A K M L T F D Q E F R P S A Q D I L M L P L F T K E P A D A L Y T	396
	6101	TTGCAAAAATGCTCACATTCGATCAGGAGTTTAGACCATCTGCCCAAGATATTTAATGTTGCCTCTTTTACTAAAGAACCCGCTGACGCAATTATACAC	6200
	397	I T A A H M * 402	
	6201	GATAACTGCCGCTCATATGTAACACCCGTCAAAATAACTTCAATGATTCATTTTATAATATATACTACCGGTTACCTGCAATAATGACAACATTCGAA	6300

Fig. 2—Continued

SORF4	1		M A P S G P T P Y S H R P Q I K H Y G T F S D	23
	6301	GTCTTTGAAGATTCGCAGACCTTTTTTGGCAATGGCACCTTCGGGACCTACGCCATATCCACAGACCGCAAATAAGCATTATGGAACATTTTCGGAT	.....	6400
	24	C M R Y T L N D E S K V D D R C S D I H N S L A Q S N V T S S M S V		57
	6401	TGCATGAGATATACTCTAAACGATGAGAGTAAGGTAGATGATAGATGTTTCAGACATACATAACTCCTTAGCACAATCCAATGTTACTTCAAGCATGTCTG		6500
	58	M N D S E E C P L I N G P S M Q A E D P K S V F Y K V R K P D R S		90
	6501	TAATGAACGATTCGGAAGAATGTCCATTAATAAATGGACCTTCGATGCAGGCAGAGGACCCCTAAAAGTGTTCCTTATAAAGTTCGTAAGCCTGACCGAAG		6600
	91	R D F S W Q N L N S H G N S G L R R E K Y I R S S K R R W K N P E		123
	6601	TCGTGATTTTTCATGGCAAATCTGAACTCCCATGGCAATAGTGGTCTACGTCGTGAAAAATATATACGTTCCCTCTAAGAGGCGATGGAAGAATCCCGAG		6700
	124	I F K V S L K C E S I G A G N G I K I S F S F F * 147		
	6701	ATATTTAAGGTATCTTTGAAATGTGAATCAATGGCGCTGGTAACGGAATAAAAAATTCATTCTCATTTTCTAACATTATAATATATCAGATCGTTTCT		6800
	6801	TATATACTTATTTTCATCGTCGGGATATGACTAACGTATACTAAGTTACAAGAAACAACCTGCTTAACGTCGAACATAACGGAATAAAAAATATATATAGC		6900
US6 (gd)	1		M N R Y R Y E S I F F R Y I S S T R M I	20
	6901	GTCTCTATAACTGTTATATGGCACCTTTAGAGCTTCGGTATGAATAGATACAGATATGAAAGTATTTTTTTAGATATATCTCATCCACGAGAATGA		7000
	21	L I I C L L L G T G D M S A M G L K K K D N S P I I P T L H P K G N		53
	7001	TTCTTATAACTGTGTTACTTTGGGAACTGGGACATGTCGCAATGGGACTTAAGAAAGACAATTCCTCGATCATTCCACATCCCGAAAGGTAA		7100
	54	E N L R A T L N E Y K I P S P L F D T L D N S Y E T K H V I Y T D		86
	7101	TGAAAACCTCCGGCTACTCTCAATGAATACAAAATCCCGTCTCCACTGTTGATACACTTGACAATTCATATGAGACAAAACACGTAATATATACGGAT		7200
	87	N <sup>-</sup> C <sup>-</sup> S F A V L N P F G D P K Y T L L S L L L M G R R K Y D A L V A W		120
	7201	AATGTAGTTTTGCTGTTTTGAATCCATTTGGCGATCCGAAATATACGCTTCTCAGTTTACTGTTGATGGGACGACGCAAAATATGATGCTCTAGTAGCAT		7300
	121	F V L G R A C G R P I Y L R E Y A N <sup>-</sup> C <sup>-</sup> S T N E P F G T C K L K S L		153
	7301	GGTTTGTCTTGGGACAGCATGTGGGAGACCAATTTATTTACGTGAATATGCCAATGCTCTACTAATGAACATTTGGAACCTTGTAATTAAGTCCCT		7400
	154	G W W D R R Y A M T S Y I D R D E A L K L I A A P S R E L S G L Y		186
	7401	AGGATGGGGATAGAAGATATGCAATGACGAGTTATCGATCGAGATGAAATGAAATGATTTCGAGCACCCAGTCGAGCTAAGGTGATATAT		7500
	187	T R L I I I N G E P I S S D I L L T V K G T C S F S R R G I K D N K		220
	7501	ACGCGTTAATAATTTAATGGAGAACCATTTCGAGTGACATTTACTGACTGTTAAAGGAACATGTAGTTTTTCGAGACGGGGATAAAGGATAACA		7600
	221	L C K P F S F F V N <sup>-</sup> G <sup>-</sup> T T R L L D M V R T G T P R A H E E N V K Q		253
	7601	AACATGCAAACCGTTCAGTTTTTTGTCATGTTACAACACGGCTGTTAGACATGGTGCGAACAGGAACCCCGAGAGCCCATGAAGAAAATGTGAAGCA		7700
	254	W L E R N G G K H L P I V V E T S M Q Q V S N L P R S F R D S Y L		286
	7701	GTGGCTTGAACGAAATGGTGGTAAACATCTACCAATCGTCGTAACATCTATGCAACAAGTCTCAAATTTGCCGAGAAGTTTAGAGATTTCATATTTA		7800
	287	K S P D D D K Y N D V K M T S A T T N N <sup>-</sup> I <sup>-</sup> T T S V D G Y T G L T N R		320
	7801	AAATCACCTGACGAGATAAATAATGACGTCAAAATGACATCGGCCACTACTAATAACATTACCACCTCCGTGGATGGTTACTGGACTCACTAATC		7900
	321	P E D F E K A P Y I T K R P I A I S V E E A S S Q S P K I S T E A K K		353
	7901	GGCCCGAGGACTTTGAGAAAGCACCATACATAACTAAACGACCPATCTCTGTCGAGGAGGCAATCCAGTCAATCACCAAAAATACSTAGAAA		8000
	354	S R T Q T I I S L V V L C V M F C F T V I G S G I W I L R K H R K		386
	8001	ATCCCGAACGCAAATAAATAATTCCTAGTGTCTATGCGTCATGTTTGTTCATTGTAATCGGGTCTGGTATATGGATCCTTCGCAAAACCCGCAA		8100
	387	T V M Y D R R R P S R R A Y S R L * 403		
	8101	ACGGTGTATGATAGACGCTCGTCCATCAAGACGGGCATATTCGCCCTATAACACGTGTTGGTATGGGCGTGCCTATAGTCATAAGAAGTTGAC		8200
US7 (gl)	1		M Y V L Q L L F W I R L F R	14
	8201	TACATTGATCAATGACATTATAGCTTCTTTGGTCAGATAGACGGCGTGTGTGATTCGGATGATGTAACAATATATTTTGGATCCGCCCTCTTTC		8300
	15	G I W S I V Y T G T S V T L S T D Q S A L V A F R G L D K M V N V		47
	8301	GAGGACTCTGGTCTATAGTTTACTGGAACATCTGTTACGTTATCAACGGACCAATCTGCTCTTGTTCGGTTCGCGGATAGATAAAATGGTGAATGT		8400
	48	R G Q L L F L G D Q T R T S S Y T G T T E I L K W D E E Y K C Y S		80
	8401	ACGCGGCAACTTTTATTCCTGGGCGACCGACTCGGACTTCTATACAGGAACGAGCAATCTTGAATGGGATGAAGAATATAAATGCTATTCC		8500
	81	V L H A T S Y M D C P A I D A T V F R G C R D A V V Y A Q P H G R V		114
	8501	GTTCTACATGCGACATCATATATGGATTGCTCTGCTATAGACGCCACGGTATTACAGAGCTGTAGAGACGCTGGTATATGCTCAACCTCATGGTAGAG		8600
	115	Q P F P E K G T L L R I V E P R V S D T G S Y Y I R V S L A <sup>-</sup> G R N		147
	8601	TACAACCTTTCCGAAAGGGGAACATTTGTGAGAATGTGCAACCCAGAGTATCAGATACAGGACGCTATTACATACGTGTATCTCTCGCTGGAAGAAA		8700
	148	M <sup>-</sup> S <sup>-</sup> D I F R M V V I I R S S K S W A C N <sup>-</sup> H <sup>-</sup> S A S S F Q A H K C I R		180
	8701	TATGAGCGATATTTAGAATGGTGTATTATAAGGAGTAGCAAATCTGGGCTGTAATCACTCTGCTAGTTCAATTCAGGCCATAAATGTATTCCG		8800
	181	Y V D R M A F E N Y L I G H V G N L L D S D S E L H A I Y N <sup>-</sup> I <sup>-</sup> T P Q		214
	8801	TATGTCGACCGTATGGCCCTTGAATAATCTGATTTGACATGTAGGCAATTTGTCGAGCAGTGACTCGGAATTCATGCAATTTATAATATTACTCCCC		8900
	215	S I S T D I N I V T T P F Y D N S G T I Y S P T V F N L F N N <sup>-</sup> N <sup>-</sup> S		247
	8901	AATCCATTTCCACAGATATAAATATTGTAACGACTCCATTTTACGATAATTCGGGAACAATTTATTCACCTACGGTTTTTAATTTGTTAATAACAATTC		9000
	248	H V D A M N <sup>-</sup> S <sup>-</sup> T G M W N T V L K Y T L P R L I Y F S T M I V L C T		280
	9001	CCATGTCGATGCAATGAAATCGACTGGTATGGAATACCGTTTTAAAATATACCCCTCCAAGGCTTATTTACTTTTCTACGATGATGTAATGATA		9100
	281	I A L A I Y L V C E R C R S P H R R I Y I G E P R S D E A P L I T S		314
	9101	ATAGCATTGGCAATTTATTTGGTCTGTGAAAGGTGGCGCTCTCCCATCGTAGGATATACATCGGTGAACCAAGATCTGATGAGGCCCACTCATCACTT		9200
	315	A V N E S F Q Y D Y N V K E T P S D V I E K E L M E K L K K K V E		347
	9201	CTGCAGTTAACGAATCATTTCAATATGATTATAATGTAAGGAAACTCCTTCAGATGTTATGAAAAGGAGTTGATGGAAGAACTGAAGAAGAAAGTCGA		9300

Fig. 2—Continued

348	L L E R E E C V * 355		
9301	ATTGTTGGAAGAGAAGAATGTGTATAGTTTGAGAACTATTATAGGTAGGTGGTACCTGTTAGCTTAGTATAAGGGGAGGAGCCGTTTCTTGTITTTAA	9400	
US8			
(gE)			
1			
9401	AGACACGAACACAAGGCCGTAAGTTTATATGTGAATTTTGTGCATGCTGCGAGTCAGCGTCATAATGTGTGTTTCCAAATCTGATAATAGTGACGA	9500	M C V F Q I L I I V T T
13	I K V A G T A N I N H I D V P A G H S A T T T I P R Y P P V V D G	45	
9501	CGATCAAAGTAGCTGGAACGCCAACATAAATCATATAGACGTTCTGCAGGACATTCTGTACAACGACGATCCCGGATATCCACCAGTTGTCGATGG	9600	
46	T L Y T E T W T W I P N H C N E T A T G Y V C L E S A H C F T D L	78	
9601	GACCTTTACACCGAGCGTGGACATGGATTCCCAATCACTGCAACGAAACGGCAACAGGCTATGTATGCTGGAAAGTGCTCACTGTTTACCAGATTG	9700	
79	I L G V S C M R Y A D E I V L R T D K F I V D A G S I K Q I E S L S	112	
9701	ATATTAGGAGTATCCTGCATGAGGTATGCGGATGAAATCGTCTTACGAAGTATAAATTTATTGTCGATGCGGGATCCATTAACAAATAGAATCGCTAA	9800	
113	L N G V P N I F L S T K A S N K L E I L N A S L Q N A G I Y I R Y	145	
9801	GTCTGAATGGAGTTCGAATATATTCTACTACGAAAGCAAGTAAACAAGTTGGAGATACTAAATGCTAGCCTACAAATGCGGGTATCTACATTCCGTA	9900	
146	S R N G T R T A K L D V V V V G V L G Q A R D R L P Q M S S P M I	178	
9901	TTCTAGAAATGGACGAGGACTGCAAAAGCTGGATGTTGTTGTGGTGGCGTTTGGGTCAAGCAAGGGATCGCCTACCCCAATGTCAGTCCTATGATC	10000	
179	S S H A D I K L S L K N F K A L V Y H V G D T I N V S T A V I L G P	212	
10001	TCATCCACGCCGATATCAAGTTGTCATTAATAAACTTTAAAGCATTAGTATATCACGTTGGAGATACTATCAATGCTCGACGGCGGTTACTAGGAC	10100	
213	S P E I F T L E F R V L F L R Y N P T C K F V T I Y E P C I F H P	245	
10101	CTTCTCGGAGATTCACATTTAGGTTGTTGTTCTCCGTTATAATCCAACGTCGAAGTTCGTCACGATTATGAACCTTGATATTTACCC	10200	
246	K E P E C I T T A E Q S V C H F A S N I D I L Q I A A A R S E N C	278	
10201	CAAAGAACCAGAGTGTATTACTACTGCAGAACAATCGGTATGTCATTTCGCATCCAACATTGACATTCTGCAGATAGCCGCCGACGTTCTGAAAATTGT	10300	
279	S T G Y R R C I Y D T A I D E S V Q A R L T F I E P G I P S F K M K	312	
10301	AGCACAGGGTATCGTAGATGATTTATGACACGGCTATCGATGAATCTGTGACGGCCAGATTAACATTCATAGAACCAGGAATTCCTTCTTTAAATGA	10400	
313	D V Q V D D A G L Y V V V A L Y N G R P S A W T Y I Y L S T V E T	345	
10401	AAGATGTCAGGTAGACGATGCTGGATTGATGTGGTTGTGGCTTATACAATGGACGTCGAAGTGCATGGACTTACATTTATTGTCAACGGTGGAAAC	10500	
346	Y L N V Y E N Y H K P G F G Y K S F L L Q N S S I V D E N E A S D W	378	
10501	ATATCTTAATGTATAAAAACACCACAAGCCGGGATTTGGGTATAAATCTTCTACAGAACAGTAGTATCGTCGACGAAATGAGGCTAGCGATTGG	10600	
379	S S S S I K R R N G T I I Y D I L L T S L S I G A I I I V I V G G	412	
10601	TCCAGTCCGTCATTAACCGGAGAAATAATGGTACTATCATTTATGATATTTACTCACATCGCTATCAATTGGGGCGATTATTATCGTCATAGTAGGG	10700	
413	V C I A I L I R R R R R R R R T R G L F D E Y P K Y M T L P G N D L	445	
10701	GTGTTGTATTGCCATATTAATTAGGCGTAGGAGACGACGTCGCACGAGGGGTATTTCGATGAATATCCCAATATATGACGCTACCAGGAAACGATCT	10800	
446	G G M N V P Y D N T C S G N Q V E Y Y Q E K S A K M K R M G S G Y	478	
10801	GGGGGCATGAATGTACCGTATGATAATACATGCTCTGGTAACCAAGTTGAATATTATCAAGAAAAGTCGGCTAAAATGAAAAGAATGGGTTCGGGTTAT	10900	
479	T A W L K N D M P K I R K R L D L Y H * 497		
10901	ACCGCTTGCTAAAAATGATATGCCGAAAATAGGAAAACGCTTAGATTTATACCACTGATATGTACATATTTAACTTAATGGGATATAGTATATGGAC	11000	
11001	GTCTATATGACGAGAGTAAATAAACTGACAATGCAAAATGAAGCTGATCTATATTGTGCTTTATATTGGGACAAAACCTCGCACAAAGCTCATTCAACACA	11100	
11101	TCCACTCTGGACAGCTTCATGTAAAAATAAAGTAAATCAATGATAATGGGAGAGAAGATGTGAGCAAGGATCCATGGTGTCTGCTTTTTATAGA	11200	
11201	TACTACCGCAATGCTACATATAAataaaaaataacctctacccaaaaatggcggtatgagatgcacggggaaaatacgcagctg 11286		
	TRs---->		

Fig. 2—Continued

ORFs (beginning from the first ATG codon) are shown relative to the nucleotide sequence in Fig. 2. Since their gene products are yet to be characterized, in order to simplify identification, 7 have been named (Fig. 1B, Table 1) based on homology (see below) to HSV-1-encoded U<sub>S</sub> ORFs (McGeoch *et al.*, 1985). When appropriate, the letters MDV will preface the homolog's name to indicate the ORF's origin. The four nonalphaherpesvirus-related ORFs have been arbitrarily named SORFs 1, -2, -3 and -4 (unique Short region Open Reading Frame). Due to the A-T-rich nature of MDV, there are numerous TATA-like sequences for transcriptional initiation, more than are likely to have functional relevance. Therefore, these sites have not been highlighted in Fig. 2. All of the ORFs contain potential AUG (ATG) codons in a favorable context for translational initiation (purine in -3 position and/or guanine in +4 position; Kozak, 1989). Such a context is lacking for the first methionine codons of SORF2 and US3 (PK), but is observed among secondary methionine codons which correspond to amino acid positions 8 and

10, respectively (Fig. 2). Potential polyadenylation signals, according to HSV-1 usage patterns (McGeoch, 1991), are identified in Fig. 2; ORF locations, lengths (AA), and predicted molecular masses of the putative translational products are outlined in Table 1.

#### Database- and computer-assisted homology comparisons

Using the computer program FASTA or TFASTA (Pearson and Lipman, 1988), each of the 11 predicted amino acid sequences was screened against the SwissProt protein database or GenBank/EMBL nucleic acid databases, respectively, in addition to recently published pseudorabies virus (PRV) (van Zijl *et al.*, 1990) and equine herpesvirus-1 (EHV-1) (Colle *et al.*, 1992; Elton *et al.*, 1991; Holden *et al.*, 1992a,b; Telford *et al.*, 1992) S segment gene sequences not present in these databases. Optimized FASTA/TFASTA scores greater than 100 were initially considered as potential candidates possessing a

TABLE 1  
Summary of MDV U<sub>s</sub> ORF Data

Name	ORF start	ORF stop Codons	M <sub>r</sub> <sup>a</sup>	FASTA scores with				Properties/references	
				HSV-1	VZ	PRV	EHV-1		
SORF1	331	62	89	10.1	—	—	—	—	
SORF2	521	1060	179	20.1	—	—	—	Homologous to FPV ORF4 <sup>c</sup>	
US1	1227	1766	179	20.4	101	160	218	209	Regulatory protein <sup>d</sup>
US10	2056	2697	213	23.6	134	147	—	260	Virion protein <sup>e</sup>
SORF3	3863	2805	351	40.6	—	—	—	—	—
US2	4902	4090	270	29.7	335	—	168	355	—
US3	5014	6222	402	44.7	611	616	563	551	Protein kinase <sup>f</sup>
SORF4	6332	6775	147	16.8	—	—	—	—	—
US6	6943	8154	403	42.6 <sup>b</sup>	211	—	279	246	Membrane glycoprotein D (gD) <sup>g</sup>
US7	8261	9328	355	38.3 <sup>b</sup>	145	228	188	242	Membrane glycoprotein I (gI) <sup>g</sup>
US8	9467	10960	497	53.7 <sup>b</sup>	192	376	217	402	Membrane glycoprotein E (gE) <sup>h</sup>

<sup>a</sup> Predicted, in absence of posttranslational modifications.

<sup>b</sup> Based on sequences that follow the predicted signal peptide cleavage site.

<sup>c</sup> FASTA = 237; Tomley *et al.* (1988).

<sup>d</sup> Holden *et al.* (1992a); Jackers *et al.* (1992); Sears *et al.* (1985).

<sup>e</sup> Holden *et al.* (1992b); McGeoch *et al.* (1988).

<sup>f</sup> Leader and Purves (1988); Purves *et al.* (1991); Zhang and Leader (1990).

<sup>g</sup> Campadelli-Fiume *et al.* (1990); Johnson and Spear (1989); Long *et al.* (1992); Peeters *et al.* (1993).

<sup>h</sup> Dubin *et al.* (1991); Johnson *et al.* (1988); Whealy *et al.* (1993); Zsak *et al.* (1992).

significant degree of amino acid similarity. The results of this analysis are in Table 1; the scores obtained are comparable to those of previously established S region homologies. Further evidence for homology was derived from dot matrix analyses and % similarity/identity analysis (using GAP; data not shown). Homologies between MDVs and their alphaherpesvirus S region counterparts ranged between 40 and 60% similarity and between 20 and 40% identity (data not shown). Apart from MDV US3, 6 ORFs (MDV US1, -10, -2, -6, -7 and -8) were found to be exclusively homologous to alphaherpesvirus S segment genes; in contrast, SORF1, -3, and -4 failed to show statistically significant homology with any sequences in either of the two databases. On the other hand, using SORF2 as a probe for FASTA analysis, a FASTA score of 237 was obtained, indicating homology to an uncharacterized fowlpox virus (FPV) ORF (e.g., FPV ORF4; Tomley *et al.*, 1988). Upon alignment, these sequences were found to exhibit 67% similarity and 42% identity over the 100 AA aligned (Fig. 3). Like other US3 homologs, MDV's counterpart exhibits homology to the serine-threonine protein kinase superfamily (Hanks *et al.*, 1988), as evidenced by a relatively large number of FASTA scores between 150 and 250. Nevertheless, these scores were three- to four-fold lower than those obtained between US3 homologs of HSV, VZV, and PRV (Table 1). The US3 gene family of herpesvirus protein kinases appear to define a distinct subfamily within the serine-threonine protein kinase superfamily; it is thought that related cellular counterparts exist and await future characterization (Hanks *et al.*, 1988). Homologies to HSV-2 US2, -3, -6, and -7 are not

presented in this report, inasmuch as their ORFs exhibit greater than 70% identity to their HSV-1 counterparts (McGeoch *et al.*, 1987) and result in homologies with MDV that basically resemble those with HSV-1. MDV US6 exhibits demonstrable homology to HSV-2 US4 (FASTA = 100) and its PRV counterpart, gX (FASTA = 90). This is consistent with earlier findings suggesting duplication and divergence of S component glycoprotein genes from common precursors (McGeoch, 1990).

#### Analysis of MDV glycoproteins, gD, gI, and gE

In comparing the gB homologs of seven different herpesviruses included in the alpha-, beta-, and gammaherpesvirus subfamilies, there is complete conservation of 10 cysteine residues (Ross *et al.*, 1989). Alphaherpesvirus S component glycoproteins have also been found to contain similar patterns of conserved cysteine residues (McGeoch, 1990). HSV-1 US6 (gD) contains seven cysteine residues; six appear critical for correct folding, antigenic structure, and extent of oligosaccharide processing (Wilcox *et al.*, 1988; Long *et al.*, 1992). Not only are these same six cysteines conserved among gD homologs of HSV-2 (McGeoch *et al.*, 1987), PRV (Petrovskis *et al.*, 1986a), EHV-1 (Audonnet *et al.*, 1990; Flowers *et al.*, 1991; Telford *et al.*, 1992), BHV-1 (Tikoo *et al.*, 1990), and simian herpes B virus (SHBV; Bennett *et al.*, 1992), but they are conserved by the MDV gD homolog as well (data not shown). Similar cysteine conservation patterns apply to alphaherpesvirus US7 (gI) and US8 (gE) homologs (McGeoch, 1990) and their MDV counterparts (data not shown).



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MDV SORF2  83 LEASFHKGQEEL...LEYCEALYLPQPVKMEIVGIVDDVPCLATGMQLLI 129
           ::  ::  :|||  |:| | |||||  :|:|:|::|  :  |:|
FPV ORF4   1 MDRNINLPEEELKYIKECCEVLYLPQPTRMDIIGVMNDS.ISWNENLII 49

MDV SORF2  130 LVAEGGEVYAYEEDTLHKLATSFSEFLEIGVKSLGREVYHCGEYIEQVVH 179
           |: |:| :| |:|: | |:| : || |||: || ||||| | | :
FPV ORF4   50 LMSEDGKIYVYDDEALYKVADTMEEFSEIGLINLGNVEVYHCREDIKPLPE 99

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Fig. 3. Homology between MDV SORF2 and FPV ORF4. GAP (UWCGC) analysis aligning area conserved between SORF2 and fowlpox virus ORF4 (Tomley *et al.*, 1988). Amino acid numbers (with respect to predicted 5' ATG) of aligned sequences are listed at the beginning and end of each line. Bars and double dots identify identical and similar amino acid matches, respectively. The area aligned was 67% similar, 42% identical; a FASTA score of 237 was obtained.

Careful inspection of the N-terminal regions of the MDV gD, gI, and gE homologs has revealed that all contain the three basic building blocks of signal peptide sequences: a basic, positively charged N-terminal region (n-region), a central hydrophobic region (h-region), and a more polar terminal region (c-region) that seems to define the cleavage site (von Heijne, 1985). Figure 2 shows the likely position of these sites (von Heijne, 1986). Also included are the locations of other characteristic features of membrane glycoproteins, namely, the presence of potential N-glycosylation sites (i.e., N-X-S/T) and putative hydrophobic transmembrane and charged cytoplasmic domains near the C-terminal end. Like other gI-homologs, MDV's counterpart contains a relatively long cytoplasmic domain. However, in contrast to the other gD homologs, MDV gD's signal peptide contains a longer n-region (18 residues) that is unusually highly charged (+4; Fig. 2) considering an overall mean value of +1.7 among eukaryotes, which generally does not vary with length (von Heijne, 1986). Although a methionine codon exists directly before the hydrophobic h-region at position 6997 in Fig. 2 (as in PRV's gD homolog; Petrovskis *et al.*, 1986a), the scanning model for translation (Kozak, 1989) favors usage of the more 5'-proximal initiation codon (at position 6943, Fig. 2).

#### Comparison of MDV sequences to those previously published

Comparison of sequences of the "virulent" GA strain of MDV (Fig. 2) with those derived from a 5.5-kbp region of the "very virulent" RB1B strain of MDV (Ross *et al.*, 1991) has revealed over 99% identity at both the nucleic acid and the predicted amino acid levels. One difference results in an extension of 5 additional amino acids at the 5' end of the GA US6/gD (M-N-R-Y-R) relative to its RB1B sequence counterpart (ORF5; Ross *et al.*, 1991). In addition to the 5-AA extension, the next four positions in the GA strain (Y-E-S-I) would differ from the corresponding RB1B positions (M-K-V-F). Differences in these signal sequences could account for differences in gD processing between these two strains. The predicted amino acid sequence of the GA US2 (Fig. 2) is identical to that

published in a recent report (Cantello *et al.*, 1991), except for the presence of an alanine in place of an arginine at position 143. This minor difference is due to the inversion of a guanine and a cytosine relative to each other in the two GA sequences.

The RB1B counterpart of SORF4 (ORF4 in their report) was recently proposed to be a probable homolog of HSV-1 gG (Ross and Binns, 1991). It is tempting to propose such a homology, given their similar locations relative to other  $U_s$  region genes. We have further tested this proposed homology by similarly aligning these two sequences with GAP, following repeated shuffles of either of the two sequences while maintaining length and composition (using the /RANDOMIZATIONS command line option for 100 randomizations). This analysis was performed twice (each time with one of the two sequences shuffled). In doing so, we failed to find a significant difference in homology score ratios between the actual versus randomized alignments ( $1.12 \pm 0.07$ ). In some cases, the homologies of the randomized alignments actually exceeded the proposed MDV ORF4/HSV-1 gG alignment. Therefore we do not consider the proposed homology to be statistically significant. In fact, when using the type of stringency in the above example, equally significant homologies are encountered with almost any given protein database search (data not shown). While we cannot absolutely rule out that the two sequences are evolutionarily related, any functional homology would appear absent, since the supposed MDV gG homolog lacks hydrophobic domains representing signal peptide and transmembrane domain regions. Thus, it would appear that, at the very least, selection pressure for the maintenance of a common glycoprotein function appears to have been lost in this case.

## DISCUSSION

### New findings

In this report, we have characterized the sequences of 3 new MDV  $U_s$  region ORFs homologous to HSV US1 (ICP22), US8 (gE), and US10; 2 new MDV-specific  $U_s$  ORFs (SORF1 and -3); a fowlpox virus homolog (SORF2);

and a complete HSV US7-homologous sequence. This extends upon the sequence analysis of a 5255-bp segment located in the U<sub>S</sub> region of the RB1B strain (Ross *et al.*, 1991; Ross and Binns, 1991). Between two oncogenic serotype 1 strains, the "very virulent" RB1B- and the "virulent" GA strain, only minor sequence differences were found over their common 5.3-kbp region. With completion of the entire 11,160-bp U<sub>S</sub> sequence (GA strain), we have precisely determined the IR<sub>S</sub>-U<sub>S</sub> and U<sub>S</sub>-TR<sub>S</sub> junctions (Fig. 2); these were somewhat of a surprise, since previous workers (Fukuchi *et al.*, 1985) using the same MDV strain as ours (GA), mapped the IR<sub>S</sub>-TR<sub>S</sub> junction to a different fragment located 2–3 kb upstream of the correct location.

Alphaherpesvirus S regions are characterized by a set of homologs which are specific to members of this taxonomic subfamily (Davison and Taylor, 1987; McGeoch, 1990). The identification of seven alphaherpesvirus S region homologs in this study is consistent with MDV bearing a closer relation to alphaherpesviruses than gamma-herpesviruses (Buckmaster *et al.*, 1988). Failure to identify CpG dinucleotide deficiencies among MDV U<sub>S</sub> region sequences is further consistent with this proposal (Efstathiou *et al.*, 1990; Honess *et al.*, 1989).

#### Potential importance of alphaherpesvirus S component differences in determining biological divergence and pathogenesis

Since MDV has been traditionally regarded as a gamma-herpesvirus, much of the previous work interpreting MDV's properties has proceeded by analogy with the association between EBV and B-cells (Wen *et al.*, 1988, for example). Because of the closer genetic relationship between MDV and other alphaherpesviruses, we agree with others (Lawrence *et al.*, 1990) that the lymphotropic properties of MDV and HVT are unlikely to be determined by molecules homologous to those of EBV.

Upon further examination, more parallels exist between the "lymphotropic" MDV and the "neurotropic" alphaherpesviruses than previously appreciated. Lymphotropism (and epitheliotropism) is probably common to all herpesviruses and is largely responsible for the widespread dissemination of HSV and VZV in cases involving neonatal and immunocompromised patients, often resulting in death (Nahmias and Roizman, 1973; Grose, 1982). These infections are characterized by a biphasic viremia similar to that observed in MDV-infected chickens; in the absence of maternal antibodies, young chickens can often die from an early mortality syndrome lacking any tumor involvement (Jakowski *et al.*, 1970; Witter *et al.*, 1980). A biphasic viremia has also been established for VZV infections involving immunocompetent patients as well (Grose, 1981; Ozaki *et al.*, 1986). With respect to T-cell tropism, MDV and HSV are similar; replication of each is restricted to activated, Ia-bearing T-cells

(Braun *et al.*, 1984; Calnek, 1986). Like MDV, equine herpesvirus-1, an alphaherpesvirus, can also establish latent infections in T-lymphocytes (Welch *et al.*, 1992). This lends support to an earlier proposal characterizing EHV-1 as a T-lymphotropic herpesvirus (Scott *et al.*, 1989). In addition to latent T-lymphocyte infections, MDV also appears to establish latent infections in both Schwann and satellite cells (Pepose *et al.*, 1981) like VZV (Croen *et al.*, 1988). Such complexities suggest that a biologically based classification system is overly simplistic, potentially misleading, and guided by biases that are dictated by the manner in which these viruses are studied.

To account for the different biological expressions that exist, a renewed focus on molecular differences between MDV and other alphaherpesviruses may be in order. In this regard, the MDV U<sub>S</sub> region (and adjoining repeats) may be particularly important. Fifty-three of the 55 unique long (U<sub>L</sub>) region genes of HSV-1 possess an equivalent in VZV (McGeoch *et al.*, 1988); a considerable number of these are related to beta- and gamma-herpesvirus genes as well (29 of 67 EBV genes are counterparts to VZV U<sub>L</sub> genes; Davison and Taylor, 1987). In contrast, alphaherpesvirus S components are specific for members of this taxonomic subfamily and appear to represent their most divergent coding region (Davison and Wilkie, 1983; Davison and McGeoch, 1986; Telford *et al.*, 1992). In comparing MDV with other alphaherpesviruses, significant divergence also extends to the U<sub>L</sub>-flanking repeat regions (Buckmaster *et al.*, 1988) which are known to be expressed in tumor cells (Jones *et al.*, 1992; Schat *et al.*, 1989; Sugaya *et al.*, 1991). A comparison of the genetic organization of selected alphaherpesvirus S segment genes is presented in Fig. 4. It is based on previously published reports on EHV-1 Ab4p field isolate strain (Telford *et al.*, 1992), HSV-1 (McGeoch *et al.*, 1985), VZV (Davison and Scott, 1986), and PRV (Petrovskis *et al.*, 1986a,b; Petrovskis and Post, 1987; van Zijl *et al.*, 1990; Zhang and Leader, 1990); other alphaherpesvirus S segment genes corresponding to BHV-1 (Tikoo *et al.*, 1990); EHV-1 Ky-A-cell culture strains (Audonnet *et al.*, 1990; Breeden *et al.*, 1992; Colle *et al.*, 1992; Elton *et al.*, 1991; Flowers *et al.*, 1991; Flowers and O'Callaghan, 1992; Holden *et al.*, 1992a,b), EHV-4 (Cullinane *et al.*, 1988; Nagesha *et al.*, 1993), HSV-2 (McGeoch *et al.*, 1987), SHBV (Bennett *et al.*, 1992; Killeen *et al.*, 1992), and simian varicella virus (SVV; Fletcher and Gray, 1993) have been described elsewhere. Despite obvious similarities, there are marked differences in (i) gene content, organization, and localization; (ii) sequence conservation; and (iii) positioning of IR<sub>S</sub>-U<sub>S</sub> and U<sub>S</sub>-TR<sub>S</sub> junctions. Nevertheless, these overall gene layouts are consistent with a model to account for the divergence of alphaherpesviruses from a common ancestor by a number of homologous and semi-homologous recombination events which result in expansion or contraction of the inverted repeat regions and a concomitant loss or gain of U<sub>S</sub> gene(s) (Davison and

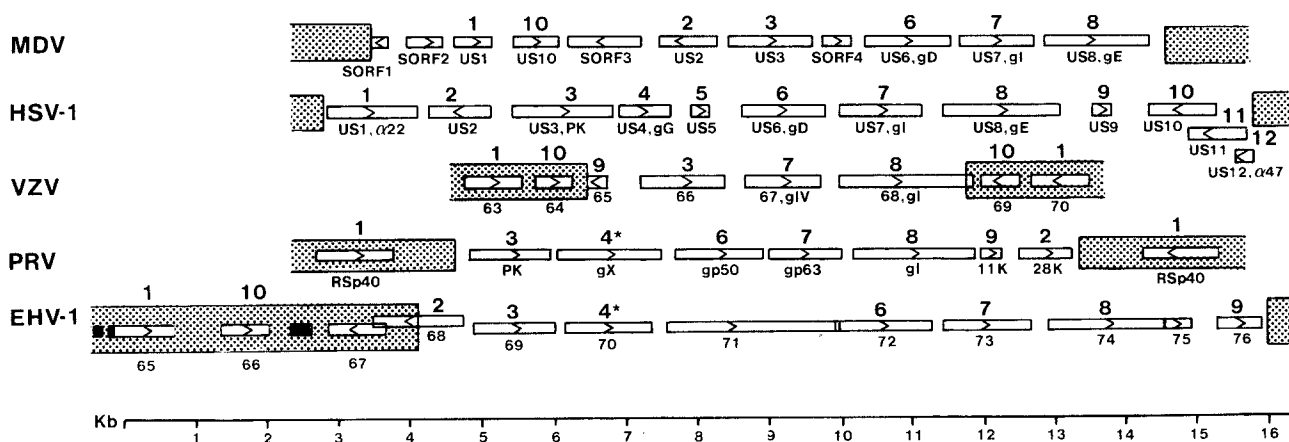


FIG. 4. Comparison of MDV and alphaherpesvirus S region genes. Based on published S region ORFs (Davison and Scott, 1986; McGeoch *et al.*, 1985; Petrovskis *et al.*, 1986a,b; Petrovskis and Post, 1987; Telford *et al.*, 1992; van Zijl *et al.*, 1990; Zhang and Leader, 1990). Numbers above boxes refer to homologs based on relation to HSV-1  $U_S$  ORF nomenclature (McGeoch *et al.*, 1985). Polypeptide designations common to each system are listed below each of those boxes where applicable. Larger, stippled boxes refer to identified  $IR_S$ ,  $TR_S$ , and/or  $R_S$  regions. Bolder areas identify repeat sequences present in the EHV-1 Ab4p field isolate strain. Asterisks refer to homologs which show relatedness to HSV-1 US4, rather than HSV-1 US4.

McGeoch, 1986). In the case of VZV, homologs of six HSV-1  $U_S$  region genes are missing (US2, US4, US5, US6, US11, US12). Unlike all other alphaherpesviruses thus far analyzed (Fig. 4; Fletcher and Gray, 1993; Killeen *et al.*, 1992), MDV appears to lack a US9 homolog. The HSV-1 US9 gene is known to encode a differentially phosphorylated 12- to 20-kDa tegument protein which becomes associated with nucleocapsids at or soon after their formation in the nuclei of infected cells (Frame *et al.*, 1986). A recent study has suggested that PRV's US9 homolog has a function associated with envelopment at the nuclear membrane (Pol *et al.*, 1991). Lacking such a homolog might contribute to MDV's characteristic inability to become stably enveloped in tissues other than the feather follicle epithelium.

#### Presence of MDV-specific and fowlpox virus-homologous genes

MDV contains at least 3 ORFs unrelated to any others presently described (SORFs 1, -3, and -4; Fig. 4). Other S component ORFs have been identified that are specific to a given alphaherpesvirus and/or its common-host relative. Such genes have been identified in HSV-1/HSV-2 (US11 and US12; Davison and McGeoch, 1986; McGeoch *et al.*, 1985) and in EHV-1/EHV-4 (ORF67/ $IR_6$ , ORF 71/ $EUS_4$ , and ORF 75; Telford *et al.*, 1992; Colle *et al.*, 1992; Breeden *et al.*, 1992; Nagesha *et al.*, 1993). Further sequence analysis of other alphaherpesvirus S regions will be necessary to determine whether such genes are truly unique to these herpesviruses and whether they confer a species-specific growth advantage.

SORF3, located in the *EcoRI*-O subfragment (Fig. 1B), specifies a 351-AA MDV-specific ORF. Considering its location, preliminary transcriptional mapping of the other

genes mapping in *EcoRI*-O (e.g., MDV US1 and -10; P. Brunovskis, unpublished observations) and previously reported data (Schat *et al.*, 1989), it appears possible that SORF3 may code for the 1.1-kb  $A_3$  transcript, one of four immediate-early transcripts consistently identified in all MDV tumor cell lines tested (Schat *et al.*, 1989).

A major surprise from this work was finding a FPV-related ORF. We are not aware of any other examples of such conservation across virus family lines, except a few cases that include cellular counterparts as well. MDV's FPV homolog, SORF2, was found to be 67% similar and 42% identical (over 100 AA) to FPV ORF4 (Tomley *et al.*, 1988). With a FASTA score of 237 and the alignment in Fig. 3, the level of conservation is more striking than that generally characterizing alphaherpesvirus S region homologies (Fig. 4). Interestingly, compared with FPV ORF4, SORF2 contains an amino-terminal extension of 82 AA; conversely, ORF4 carries a carboxy-terminal extension of 41 AA. The block of conserved sequences may encode one or more functional domains that have independently evolved following host cell acquired gene transfer. On the other hand, it is intriguing to consider the possibility of virus-virus gene transfer. Individual cells have been found to be simultaneously infected by MDV and FPV (Tripathy *et al.*, 1975). Given the different modes of replication for MDV and FPV (e.g., nuclear vs. cytoplasmic) such a possibility could point to a possibly novel form of gene transfer.

#### MDV $U_S$ region genes as potential determinants for pathogenesis and tissue tropism

Recent studies have shown that 11 of 12 open reading frames contained in the HSV-1  $U_S$  region are dispensable for growth *in vitro* (Longnecker *et al.*, 1987; Roizman and

Sears, 1990). These, and other "dispensable" genes appear to specify functions for optimal survival, maintenance, and dissemination among the host (and its population at large), rather than the presence of functions necessary for replication (Longnecker *et al.*, 1987; Roizman and Sears, 1990). The significant divergence of alphaherpesvirus S components may reflect this region's capacity for determining distinct tissue and host cell growth potentialities. Previous results have suggested that the product of HSV US1 (ICP22) encodes a determinant for tissue tropism, since its function appears to be dispensable for growth in some cell lines, but not others (Sears *et al.*, 1985). Considering the extensive genetic divergence among a cluster of different glycoprotein homologs, each potentially subject to glycosylation, phosphorylation, palmitoylation, myristylation, and/or sulfation (Grose, 1990; Spear, 1984), a potentially large window exists for the creation of multiply distinct virus-cell interactions which can affect host range, tissue tropism, invasiveness, and cell-cell spread. Previous results have demonstrated that "nonessential" alphaherpesvirus glycoproteins encode functions associated with virulence (Lomniczi *et al.*, 1984; Meignier *et al.*, 1988; Mettenleiter *et al.*, 1988; Roizman and Sears, 1990). This may reflect their ability to promote the infection and spread of virus *in vivo* (Lomniczi *et al.*, 1984; Longnecker *et al.*, 1987; Mettenleiter *et al.*, 1988; Pol *et al.*, 1991; Card *et al.*, 1992). Consistent with this proposal is the observation that a specific deletion of PRV gI (homolog of HSV gE) and/or PRV gp63 (homolog of HSV gI) was found to reduce the spread of infection in both rat (Card *et al.*, 1992; Whealy *et al.*, 1993) and pig (Kimman *et al.*, 1992) central nervous systems. This defect could reflect the inability of PRV gI mutants to promote cell-cell spread (Zsak *et al.*, 1992).

If MDV U<sub>s</sub> region genes specify virulence determinants, these could indirectly affect oncogenic potential by affecting any number of critical events which precede tumor induction. Previous studies have shown that oncogenic potential appears to be directly correlated with cell-associated viremia levels and the capacity to cause immunosuppression (Calnek and Witter, 1991). The sequence of events leading to transformation include (i) an initial lytic growth phase in B-cells, which is thought to cause activation and expansion of T-cells; (ii) a latent growth phase involving infected T-cells; (iii) a second wave of lytic infection, coincident with permanent immunosuppression; and (iv) oncogenic transformation (Calnek, 1986). Attenuated MDV strains (derived from oncogenic serotype 1 strains), as well as nononcogenic MDV and HVT strains (serotypes 2 and 3, respectively), are deficient in inducing the early cytolytic infection of B-cells in chickens, suggesting that their cell tropisms differ from those of oncogenic strains (Schat *et al.*, 1985; Shek *et al.*, 1982). This is reflected in evidence that attenuation of MDV leads to a marked reduction in infectivity and/or replication in lymphocytes (Schat *et al.*, 1985).

## In conclusion

The current herpesvirus classification system has been described as "simple, fortuitously appropriate and defective" (Roizman, 1990b). It has been further suggested that "the delineation and evolutionary relatedness of genes responsible for biological properties may be a more significant criterion for both evolutionary relatedness and classification than the arrangement and evolution of genes conserved throughout the family Herpesviridae" (Roizman, 1990b, 1992). While such a view is currently open to debate, inasmuch as alphaherpesvirus-specific U<sub>s</sub> regions specify a cluster of "dispensable" functions thought to be important biological property determinants, our sequence provides a foundation for further studies to resolve the apparent discrepancy between MDV's genetic and biologic properties.

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*Note added in proof.* After submission of this paper, we learned of an article describing the nucleotide sequence of an 8.9-kb region of the MDV GA strain comprising 72% (8020 bp) of the MDV U<sub>s</sub> region, along with 905 bp of flanking IR<sub>s</sub> sequences (Sakaguchi *et al.*, *Virus Genes* 6(4) 365-378). Their published IR<sub>s</sub>-U<sub>s</sub> and U<sub>s</sub>-TR<sub>s</sub> junction sites were identical to ours. Moreover, their sequence was found to be 99.9% identical to ours. However, the few differences that exist presumably account for their failure to identify the HSV US1 homolog of MDV and the FPV ORF4 homolog (SORF2) in MDV. The US1 error has recently been corrected in a published erratum (*Virus Genes* 7, 109). A missing base in their sequence would lead to the premature termination of a smaller ORF, with only half of the homology depicted in our Fig. 3. Another missing base in their sequence would result in the premature termination of the US2 ORF near amino acid 95. Our US2 ORF is consistent with that of Cantello *et al.*, 1991. Also, recently Zelnick *et al.* (*J. Gen. Virol.* 74, 2151-2162) described the nucleotide sequence and gene organization of MDV's vaccine virus, HVT. HVT was found to contain a similar ORF organization, with the exception that it lacks MDV SORF1, SORF2, and SORF4 homologs.

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