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Liver tonics: review of plants used in Iranian traditional medicine

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PEER REVIEW

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Comments

This is a valuable review. It introduced plants which have been traditionally used as liver tonics in Iran. This paper will promote the utilization of natural and traditional resources for contemporary health care. Herbal medicines have an extremely valuable, rich, lengthy and extensive practical history.

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ABSTRACT

Considering the fact that liver is one of the most important organs in our body, it deserves special attention and protection. Among various recommended supplements, complementary and alternative medicines particularly herbal remedies have received much attention owing to their truly healing properties. This review profits from Iranian traditional medicine and presents advantageous herbal guide directions for liver protection. According to credible Iranian medical literature such as Al Qanun Fil Tibb, Al-Havi and Makhzan-al-Aadvia, a wide spectrum of plants have been found to be useful for cleansing and protecting the liver. Some herbs such as ghafes (Agrimonia eupatoria), kasni (Cichorium intybus), anar (Punica granatum), darchin (Cinnamomum zeylanicum), za'feran (Crocus sativus), gole-sorkh (Rosa damascena) and zereshk (Berberis vulgaris) appeared to get strong consideration and were well documented as outstanding liver tonics. We conducted a comprehensive review of available Iranian medical resources such as scientific information database and medical sciences databases which cover all in vitro and in vivo studies of medicinal plants as liver tonics and hepatoprotective candidates. Literature survey was accomplished using multiple databases including PubMed, ISI web of knowledge, and Google Scholar.

KEYWORDS

Iranian traditional medicine, Liver tonic, Hepatoprotective agents, Herbal medicine

1. Introduction

The use of medicinal properties of plants in the prevention and treatment of diseases goes back to thousands years ago, and recently, it has received lots of attention due to the available scientific evidences[1]. Now, traditional medicine systems continue to play a fundamental role in health care. It should not be forgotten that about 80% of the world's population relies mainly on complementary and alternative medicines especially herbal therapies for their primary health care[2-4]. From antiquity to now, nature has been the center

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of attention as it is the richest source of medicinal agents. It has provided important opportunities for the scrutinized recognition of diseases and the related preventions[5]. In this regard, isolation and identification of herbal active ingredients have been common strategies in traditional medicine[1,6].

Liver is one of the most important organs which plays a crucial role in the daily functions of our body[7-11]. It is the main site for carbohydrates, proteins and lipid metabolism, synthesis of essential materials, and detoxifying harmful substances. Furthermore, expulsion of waste metabolites, detoxification, blood coagulation, homeostatic activities, storage of vitamins, finally excretion of bile, hormones, and drugs are other significant functions of liver.

Iranian traditional medicine (ITM) includes a wide range of medical experiences used in the prevention, diagnosis, and treatment of diseases based on the humor theory of temperament in which liver is one of the most important organs in the body. Persian scientists such as Sina[12] and Razi[13] believed that liver is one of the three essential organs (liver, heart and brain) in the body. In canon of medicine, the liver was described as a blood factory, refinery and distributor[12]. It was supposed that the liver receives all blood coming from stomach and bowel through portal vein, manufactures nutrients, and then distributes them to the rest of the body.

ITM believes that proper liver function can improve other organs performance and liver failure would lead to the inefficiency of other organs which causes various diseases such as uterine and ovarian diseases, loss of mental ability, chronic fatigue syndrome, *etc*. At this juncture, ITM has focused not only on liver diseases and treatments but also special attention to liver protection.

Liver tonics include principles of liver health protection, nutraceuticals, medications with nutritional implication and effective drugs. According to ITM, the most important principle to keep the liver healthy is to avoid eating different kinds of foods together or immediately after each other. Also, drinking too much water along with a meal, eating different incompatible foods with liver, eating large amount of sweets, and drinking cold water especially during or after exercise are important reasons leading to liver failure.

Functions of liver from both ITM and modern medicine points of view prove that a healthy liver is central to maintaining a healthy body. Liver impairment is one of the serious threats to general health around the world[14]. It is mainly caused by some toxicants such as chemotherapeutic agents (some antibiotics, large doses of acetaminophen, carbon tetrachloride, thioacetamide, peroxidised oil, aflatoxin, *etc.*), chronic consumption of alcohol, infections, and autoimmune disorders[15]. It seems that strengthening the liver against the aforementioned factors is one of the most reliable ways to prevent liver damage. For this purpose, various liver tonics have been designed and developed. Because of the increasing demand for the efficient liver tonics and hepatoprotective agents with minimal side effects[16-19], complementary and alternative therapies including nontoxic, natural and inexpensive products have attracted lots of consideration. Herein, in order to emphasize on the efficacy of ITM,

we introduced a well-documented collection of therapeutic herbs acting as liver tonics.

2. Methodology

In this study, related information was obtained from available ancient sources such as Al Qanun Fil Tibb, Al-Havi, and Makhzan-al-Aadvia. Accordingly, a wide spectrum of plants was found to be useful for cleansing and protecting the liver. Finally, the obtained data was compared with those of reported in modern medicinal databases covering all *in vitro* and *in vivo* hepatoprotective investigations. In the present paper, the literature review was performed by using PubMed, ISI web of knowledge, scientific information database and Google Scholar focusing on the following keywords: "liver tonic" and "hepatoprotective".

3. Modern proofs for the efficacy of medicinal plants used as liver tonics in ITM

3.1. Agrimonia eupatoria (A. eupatoria)

A. eupatoria known as ghafes in ITM has been repeatedly utilized in order to strengthen the liver[12,20]. The study of Yoon et al. showed that A. eupatoria decreased the toxic effects of chronic ethanol consumption on rat liver, aspartate aminotransferase (AST) and alanine aminotransferase (ALT) levels[21]. In addition, oral ingestion of aqueous extract of aerial parts of A. eupatoria in the experimental animals which were treated by tetrachloride carbon (CCl₄) decreased AST and ALT levels[22] (Table 1).

3.2. Vitis vinifera L. (V. vinifera)

The fruits of V. vinifera known as maveez munaqqa in ITM have been widely used as liver tonic[12]. The hepatoprotective effects of ethanolic extract of V. vinifera leaves were investigated against CCl₄-induced acute hepatotoxicity in rats. The AST and ALT levels were reduced in rats pretreated by V. vinifera leaves extract[23]. Effects of V. vinifera leaf extract on alcohol-induced oxidative stress in rats were investigated by Pari and Suresh. Grape leaf extract at a dose of 100 mg/kg was highly effective than 25 and 50 mg/kg body weight. In addition, it significantly reduced the levels of lipid peroxidation level and restored the enzymatic and non-enzymatic antioxidants level in liver and kidney of alcohol administration rats[24]. In another study reported by Liu et al., the hepatoprotective effect of total triterpenoids (VTT) and total flavonoids (VTF) from V. vinifera against immunological liver injury (ILI) in mice was investigated. The hepatoprotective effects of Vitis VTT and VTF from V. vinifera were evaluated in bacille-Calmette-Guérin (BCG) plus lipopolysaccharide (LPS)-induced ILI in mice. Moreover, the increased Bax/Bcl-2 ratio was significantly down which was regulated by VTT and VTF in liver tissue of ILI mice. These results

Table 1
Plants used in ITM as liver tonics.

Iranian name	Species	Family	Part (s) used	Type of assay/hepatotoxic agent	References
Ghafes	A. eupatoria	Rosaceae	Aerial part	In vivo/ethanol	[21]
	***	5.7°.		In vivo/CCl ₄	[22]
Maveez	V. vinifera	Vitaceae	Leaves	In vivo/CCl ₄	[23]
			=	In vivo/ethanol	[24]
			Fruit	In vivo/BCG-LPS	[25]
			Grape seed	In vivo/ethanol	[26]
			Root	In vivo/CCl ₄	[27]
Za'feran	C. sativus	Iridaceae	Stigma	In vivo/RIF	[28]
				In vivo/cisplatin	[29]
				In vivo/vitamin A	[30]
				In vivo/streptozotocin	[31]
Afsantin	A. absinthium	Asteraceae	Flowering aerial parts	In vivo/CCl ₄ and BCG-LPS	[32]
				In vivo/CCl ₄ and acetaminophen	[33]
Anar	P. granatum	Lythraceae	Edible portion (seedcoats and juice)	In vivo/INH and rifampin RIF	[34]
			Peel	In vivo/acetaminophen	[35]
			Leaves	In vivo/CCl ₄	[36]
			Flower infusion	In vivo/trichloroacetic acid	[37]
Rivande chini	R. palmatum	Polygonaceae	Aerial part	In vivo/CCl ₄	[38]
			Dried roots and rhizome	In vivo/CCl ₄	[39]
			Rhizome	In vivo/CCl ₄	[40]
Karafs	A. graveolens	Apiaceae	Seeds	In vivo/-	[41]
				In vivo/CCl ₄	[42]
			Roots and leaves	In vitro and in vivo/CCl ₄	[43]
			Seeds	In vivo/DEN	[44]
				In vivo/acetaminophen/thioacetamide	[45]
			Leaves	In vivo/acetaminophen	[46]
Basbaseh	M. fragnans	Myristicaceae	Mace	In vivo/LPS/D-galactosamine	[47]
	, 0	Ť		In vivo/-	[49]
				In vivol-	[50]
				In vitro/t-BHP	[51]
Mastaki	P. lentiscus	Anacardiaceae	Gums	In vivo/CCl ₄	[52]
Wastaki	1. termisens	7 Inacar araceae	Leaves	In vivo/CCl ₄	[53]
Zereshk	B. vulgaris	Berberidaceae		In vivo/CCl ₄	[55]
	B. vargaris	Derberidaeeae	Whole plant	In vitro and in vivo/t-BHP	[56]
			Root	In vivo/CCl ₄	[57]
			Fruit	In vivo/CCl ₄	[58]
Shahtareh	F. parviflora	Papaveraceae	Aerial parts	In vivo/acetaminophen	[59]
Silalitatell	r. parvijiora	1 apaveraceae	Acriai parts	In vivo/CCl ₄	[60]
			W/l114	In vivo/CCl ₄	[61]
D 11	C1	T	Whole plant	In vitro/nimesulide	[62]
Darchin	C. zeylanicum	Lauraceae	Inner bark	in vivo/CCl ₄	[64,65]
C 1 11	D 1	D	Pl	In vivo/gamma irradiation	[66]
Gole-sorkh	R. damascena	Rosaceae	Flower	In vivo/acetaminophen	[67]
	<i>a</i> .	G 11.		In vivo/CCl ₄	[68]
Kadooye shirin	C. maxima	Cucurbitaceae	•	In vivo/CCl ₄	[69]
Ameleh	D 111	F1 11 1	Seeds	In vivo/acetaminophen	[71]
	P. emblica	Phyllanthaceae	Fruits	In vivo/acetaminophen	[72]
				In vivo/DEN	[73]
			Stem and fruit	In vivo/RIF, INH and pyrazinamide	[74]
			Fruits	In vivo/ethanol	[75]
				In vivo/thioacetamide	[76]
Barhang	P. major	Plantaginaceae	Seeds	In vivo/CCl ₄	[77]
				In vivo/CCl ₄	[78]
			Not mentioned	In vivo/7,12-dimethylbenz(a)anthracene	[80]
Salikheh	C. cassia	Lauraceae	Bark	In vivo/dimethylnitrosamine	[84]
				In vivo/alcohol	[86]
Balasan	C. opobalsamum	Burseraceae	Aerial parts	In vivo/CCl ₄	[87]
Kasni	C. intybus	Asteraceae	Leaves	In vivo/CCl ₄	[88]
			Aerial parts	In vivo/CCl ₄	[89]
			Seeds	In vivo/CCl ₄	[90]
				In vivo/thioacetamide	[91]

Table 1, continuedPlants used in ITM as liver tonics.

Iranian name	Species	Family	Part (s) used	Type of assay/hepatotoxic agent	References
			Leaves	In vivo/acetaminophen	[92]
				In vivo/nimesulide	[93]
Annab	Z. vulgaris	Rhamnaceae	Aerial parts	In vivo/CCl ₄	[94]
Fandogh	C. avellana	Betulaceae	Leaves	In vivo/CCl ₄ and acetaminophen	[95]
Foloos	C. fistula	Fabaceae	Leaves	In vivo/DEN	[97]
				In vivo/DEN	[99]
			Fruit pulp	In vivo/CCl ₄	[101]
Zanjabil	Z. officinale	Zingiberaceae	Rhizomes	In vivo/acetaminophen	[102,103]
				In vivo/CCl ₄ and acetaminophen	[104]
				In vivo/ADR	[106]
				In vivo/atorvastatin	[107]
				In vivo/Hg, Pb and Cd	[108]

C. sativus: Crocus sativus; A. absinthium: Artemisia absinthium; P. granatum: Punica granatum; R. palmatum: Rheum palmatum; A. graveolens: Apium graveolens; M. fragnans: Myristica fragnans; P. lentiscus: Pistacia lentiscus; B. vulgaris: Berberis vulgaris; F. parviflora: Fumaria parviflora; C. zeylanicum: Cinnamomum zeylanicum; R. damascena: Rosa damascena; C. maxima: Cucurbita maxima; P. emblica: Phyllanthus emblica; P. major: Plantago major; C. cassia: Cinnamomum cassia; C. opobalsamum: Commiphora opobalsamum; C. intybus: Cichorium intybus; Z. vulgaris: Zyziphus vulgaris; C. avellana: Corylus avellana; C. fistula: Cassia fistula; Z. officinale: Zingiber officinale. RIF: Rifampin; INH: Isoniazid; DEN: Diethylnitrosamine; t-BHP: Tert-butyl hydroperoxide; ADR: Adriamycin.

are comparable to those of biphenyl dicarboxylate (distributed data base, the reference hepatoprotective agent) and suggest that VTT and VTF play a protective role against ILI, which may have important implications for our understanding of the immunoregulatory mechanisms of this plant[25]. In the next study by Dogan and Celik, hepatoprotective and antioxidant activities of grape seeds against ethanol-induced oxidative stress in rats were evaluated. The results indicated that grape seeds could be as important as diet-derived antioxidants in preventing oxidative damage in the tissues by reducing the lipid oxidation or inhibiting the production of ethanol-induced free radicals in rats[26]. Sharma *et al.* studied the ethanolic extract of the root of *V. vinifera* for its hepatoprotective activity in rats with liver damage induced by CCl₄[27]. The activity of extract was also comparable to that of silymarin, a known hepatoprotective drug.

3.3. C. sativus

Stigma of *C. sativus* known as Za'feran in ITM is another natural product used as liver tonic[12,13]. Hepatoprotective effects of *C. sativus* stigma against RIF in compare with silymarin have been investigated by Mohajeri *et al.* The results demonstrated that ethanolic extract of *C. sativus* has the same protection as silymarin[28]. In addition, hepatoprotective effects of saffron stigma against cisplatin hepatotoxicity were evaluated by the same group[29]. In the work of Mokhtari *et al.*, the protective and antioxidant effects of hydro-alcoholic extract of saffron on liver enzymes following vitamin A toxicity were investigated[29]. It was found that saffron extracts protected hepatocytes against oxidative stress which was caused by hyper-vitaminosis A[30]. Also, protective effects of ethanolic extract of *C. sativus* on hepatic tissue damage in streptozocin-induced diabetic rats was investigated by Rahbani *et*

al. It was depicted that saffron had beneficial effects on antioxidant defense system of diabetic rats[31].

3.4. A. absinthium

A. absinthium known as Afsantin in ITM has been traditionally used as liver tonic in Iran[12]. Amat et al. evaluated in vivo hepatoprotective activity of the aqueous extract of A. absinthium. It was demonstrated that the pretreatment with A. absinthium dose-dependently chemically or immunologically induced increase in serum levels of hepatic enzymes. Furthermore, it has significantly reduced the lipid peroxidation in the liver tissue[32]. Effect of hydro alcoholic extract of A. absinthium against acetaminophen and CCl₄-induced hepatic damage was investigated by Gilani et al. This study indicated that the crude extract of A. absinthium exhibited hepatoprotective property and confirmed the traditional use of plant in hepatic damage[33].

3.5. P. granatum

P. granatum known as anar in ITM is a famous fruit in all around the world. This amazing fruit has been traditionally consumed in Iran as liver tonic[12]. Antihepatotoxic effects of acetone extract of P. granatum agains INH and RIF and induced hepatotoxicity in rats were studied by Yogeeta et al. In addition, its hepatoprotective property on tissue defense systems in rat was well established[34]. The hepatoprotective activity of P. granatum aqueous extract has been evaluated by Khalil. The acute elevation of AST, ALT, lactate dehydrogenase, and liver damage reduced in pretreated group by pomegranate mix with acetaminophen[35]. In another report by Rao and Dama, hepatoprotective activity of aqueous and alcoholic extract of P. granatum leaves using CCl₄-induced liver damage in rats was

proved. It was revealed that biochemical changes produced by CCl₄ were restored to normal by aqueous and alcoholic extracts of *P. granatum* leaves[36]. Celik *et al.* investigated the hepatoprotective and antioxidant effects of *P. granatum* beverage against trichloroacetic acid-exposure in rats[37].

3.6. R. palmatum

R. palmatum known as Rivande chini in ITM has been widely used as liver protectant by Persians[12]. Investigations of Guo et al. revealed that rhein could protect hepatocyte from injury and prevent the progress of hepatic fibrosis in rats. It may be associated with the fact that rhein plays a vital role in antioxidation and antiinflammation, inhibiting the expression of transforming growth factor-beta1, and suppressing the activation of hepatic stellate cells. Also it can inhibit liver fibrosis induced by CCl₄/ethanol in rats[38]. Wang et al. investigated the protective effect of R. palmatum on CCl₄-treated rats. The curative effect of administering the two lowest dosages of R. palmatum to CCl₄-treated rats was mainly expressed as a decrease in the extent of cellular injury. The hepatoprotective mechanism of R. palmatum might be related to its antioxidant effect that the antagonism of the free radical damage to hepatocytes caused by CCl₄. By contrast, the liver damage induced by R. palmatum was mainly expressed as a significant increase in the amount of fibrosis in both normal rats at all dosage levels and CCl₄-treated rats at the two highest dosage levels[39]. Tseng et al. detailed prevention of hepatic oxidative injury by R. palmatum in combination with some other herbal medicines. The results suggested that this formulation has noteworthy antioxidant activity and hepatic protection potential[40].

3.7. A. graveolens

Various products from A. graveolens known as Karafs in ITM have been used to relieve some of liver dysfunctions in Persian traditional therapies[19]. The effect of volatile oil of A. graveolens seeds on some hepatic enzymes including ALT, AST and alkaline phosphatase (ALP) in rats was examined. The results demonstrated that the active ingredients of A. graveolens may act as an antioxidant or to decrease the production of free radicals, causing stabilization of hepatocyte membrane and decreasing the release of enzymes into the blood[41]. In the study of Ahmed et al., various extracts of A. graveolens seeds were tested for their hepatoprotective activity against CCl4-induced hepatotoxicity in albino rats. Treatment of rats using different extracts of A. graveolens at dose of 250 mg/ kg markedly prevented CCl4-induced elevation of serum glutamicoxaloacetic transaminase (GOT), glutamic-pyruvic transaminase (GPT) and ALP, and increased the level of total protein and albumin[42]. Efficacy of A. graveolens leaves and roots extracts as antioxidant in CCl_a-treated rats was examined by Popović et al. It was concluded that the examined extracts showed promising protective effects. All the *n*-butanol extracts exhibited the highest protective effect[43]. The chemopreventive activity of methanolic extract of A. graveolens seeds was investigated against Solt-Farber protocol of hepatocarcinogenesis, oxidative stress, and induction of gamma-glutamyl transpeptidase-positive foci in the liver of Wistar rats by Sultana et al. According to their results, A. graveolens is a potent plant against experimentally induced hepatocarcinogenesis in Wistar rats[44]. The antihepatotoxic effects of methanolic extract of A. graveolens seeds was studied by Singh and Handa on rat liver damage induced by a single dose of acetaminophen or thioacetamide by monitoring several liver function tests. This study confirmed remarkable hepatoprotective activity of the methanolic extract of celery[45]. The hepatoprotective effects of celery leaves on acetaminophen-induced toxicity in a freshwater fish was demonstrated by Shivashri et al. It was clear that the abnormalities associated with acetaminophen exposure were reversed by treatment with A. graveolens[46].

3.8. M. fragrans

Mace of M. fragrans known as basbaseh in ITM has been broadly used as liver tonic in Iran[12,20]. It has been recognized as chemoprevention of chemically induced carcinogenesis[47]. Myristicin extraction from nutmeg has exhibited important hepatoprotective effects[48]. It has also been reported that the mace modulates glutathione (GSH) S-transferase activity in liver of mouse[49]. Chhabra and Rao examined the possible transfer of active principles of mace through the transmammary route and its ability to modulate hepatic xenobiotic-metabolizing enzymes in mice. Active principles which presented in the aqueous extract of mace were effective in transmammary modulation of hepatic xenobioticmetabolizing enzymes in the liver of mouse pups[50]. Sohn et al. investigated the protective effect of macelignan, isolated from M. fragrans against t-BHP-induced cytotoxicity. The results showed that macelignan intensively reduced the cell growth inhibition and necrosis caused by t-BHP. The results strongly suggested that macelignan has significant protective ability against oxidative damage caused by reactive intermediates[51].

3.9. P. lentiscus

P. lentiscus known as Mastaki in ITM has been used as liver tonic affirmed by Sina[12]. Mavridis *et al.* investigated total extracts of *P. lentiscus* to protect liver from CCl₄-induced damage in Wistar rats. Animals treated with mastic and silimarin alone showed a decrease in AST, ALT and malondialdehyde levels either in control or in CCl₄-treated rats. The result suggested that mastic gum extracts have a strong inhibitory effect against lipid peroxidation in rat liver. The CCl₄-treated group showed a remarkable reduced GSH level but the pre-treatment with mastic gum and silimarin inhibited GSH depletion caused by CCl₄. Mastic gum increased the content of GSH in the rat liver compared to control rats. Similar results have been

observed in activities of GSH-related enzymes, superoxide dismutase and catalase. It was concluded that total extracts of mastic gum could protect liver cells from CCl₄-induced oxidative damage[52]. The hepatoprotective effect of the boiled and non-boiled aqueous extracts of *P. lentiscus*, *Phillyrea latifolia* and *Nicotiana glauca* was evaluated *in vivo* using CCl₄ intoxicated rats as an experimental model by Janakat and Al-Merie *et al.* At the end, aqueous extract of *P. lentiscus* (both boiled and non-boiled) showed satisfactory hepatotoxic activities against CCl₄ by reducing the activity of the three enzymes and the level of bilirubin. The effect of the non-boiled aqueous extract was more noticeable than that of the boiled[53].

3.10. B. vulgaris

B. vulgaris known as zereshk in ITM has been widely used as liver protectant from the past to till now in Iran[12,13,54]. The capacity of formulated B. vulgaris extract/β-cyclodextrin to protect liver against CCl₄-induced hepatotoxicity in mice was investigated by Hermenean et al. Their results showed that B. vulgaris/β-cyclodextrin treatment prevented hepatic injury induced by CCl₄ and could be considered for further nutraceutical studies[55]. Inhibitory effect study of berberine on t-BHP-induced oxidative damage in rat liver was continued by Hwang et al. Berberine had a dose-dependent ability to quench free radicals in diphenylpicrylhydrazyl test. Furthermore, the in vivo study showed that pretreatment with berberine for 5 d before a single dose of t-BHP significantly lowered the serum levels of hepatic enzyme markers (ALT and AST) and reduced oxidative stress in the liver. The histopathological evaluation of the liver revealed that berberine reduced the incidence of liver lesions induced by t-BHP. It was conceived that berberine may play a chemopreventive role via reducing oxidative stress in living systems[56]. The effects of B. vulgaris root extracts at the different doses in CCl₄-induced liver toxicity in rats was investigated by Huseini et al. B. vulgaris reduced the serum ALP levels significantly as compared to control group. The liver micro-vesicular steatosis was considerably inhibited as well[57]. Likewise, the efficacy of methanolic extract of B. vulgaris fruits in CCl₄-induced liver injury was detailed by Eidi et al[58].

3.11. F. parviflora

F. parviflora known as shahtareh in ITM has been traditionally used to support liver in Iran[12]. The hepatoprotective activity of aqueous and methanolic extract of *F. parviflora* was investigated against acetaminophen- and CCl₄-induced hepatic damage by Gilani *et al.* Pretreatment of rats with plant extract prevented the acetaminophen-induced rise in serum enzymes ALP, AST and ALT. Posttreatment with the extract also restricted the acetaminophen-induced hepatic damage. It is conceivable that *F. parviflora* extract exhibits protective effect against acetaminophen-induced hepatotoxicity[59]. Protective effect of hydroalcoholic extract of *F. parviflora* was determined in a CCl₄-induced hepatotoxicity model in male rats

by Jamshidzadeh and Nikmahad, in which the liver function test and hystopathological observations were performed[60]. The data from this study outlined that the hydroalcoholic extract of F. parviflora with doses higher than 100 mg/kg prevented CCl₄induced liver damage. In the study of Alqasoumi et al., the efficacy of F. parviflora on CCl₄- induced liver injury in rats was evaluated. The results indicated a good protection of the extracts from CCl₄ hepatotoxicity[61]. Tripathi et al. conducted complete studies on nimesulide (a non-steroidal anti-inflammatory drug with serious hepatotoxicity)-induced cell death in primary rat hepatocyte cultures to explicate the effect of F. parviflora. It was indicated that F. parviflora extract modulates critical events and pro- and antiapoptotic proteins in mitochondria-dependent apoptosis which were induced by nimesulide[62]. In vivo studies were conducted to explore the hepatoprotective potential of F. parviflora extract against nimesulide-induced hepatotoxicity by the same group. Pretreatment with F. parviflora extract for 5 d significantly reduced the impact of nimesulide-induced toxicity as evident from the serum biomarkers of liver damage and histopathology[63].

3.12. C. zeylanicum

C. zeylanicum known as darchin in ITM has been frequently consumed in Iran as liver tonic[12,20]. Hepatoprotective effect of cinnamon extracts against CCl₄-induced oxidative stress and liver injury in rats was demonstrated by Moselhy and Ali[64]. Similary, Eidi et al. evaluated the protective effect of cinnamon bark extract against CCl₄-induced liver damage in male Wistar rats, which was reported as a potent hepatoprotective[65]. Rezk studied the protective effects of cinnamon against tissue injuries which were induced by gamma irradiation. It was found that taking adequate amount of aqueous extract of cinnamon would protect hepatic and cardiac tissues from gamma radiation-induced damage for a long time[66].

3.13. R. damascena

Aqueous extract of *R. damascena* known as gole-sorkh in ITM has been well known and popular in Iran[19]. Saxena *et al.* confirmed hepatoprotective activity of the aqueous extract of *R. damascena* flowers at different oral dose levels (250, 500, and 1000 mg/kg body weight) on acetaminophen-induced toxicity in rats[67]. It was assumed that it is likely to be mediated through its antioxidant activities. Achuthan *et al.* evaluated the antioxidant activity of the partially purified acetone fraction of *R. damascena*. Also it was proved that *R. damascena* protected against CCl₄-induced hepatotoxicity[68].

3.14. C. maxima

C. maxima known as kadooye-shirin in ITM was nominated as one of the liver tonics[19]. The hepatoprotective activity of *C. maxima*

against CCl_4 -induced hepatotoxicity has been evaluated. It was shown that methanol extract of aerial parts of C. maxima possessed significant hepatoprotective activity[69,70]. In another study conducted by Nidhi and Pathak, methanolic extract of C. maxima seeds showed strong hepatoprotective activity against acetaminophen toxicity[71].

3.15. Emblica officinalis (E. officinalis) or P. emblica

P. emblica known as ameleh in ITM has been used as liver tonic in Iran[19]. The efficacy of the P. emblica for the prevention of acetaminophen-induced hepatotoxicity in rats was studied by Malar and Bai. The results clearly confirmed the hepatoprotective effect of aqueous extract of P. emblica fruits[72]. Jeena et al. demonstrated that extract of E. officinalis inhibited hepatocarcinogenesis induced by DEN in a dose-dependent manner[73]. The hepatoprotective effects of two medicinal plants (Tinospora cordifolia and P. emblica) and their combination were investigated in a rat model of INH, RIF and pyrazinamide-induced hepatic damage by Panchabhai et al. This study proved the synergistic protective effects which were exerted by the combination of Tinospora cordifolia and P. emblica when co-administered with mentioned drugs[74]. The protective effects of P. emblica extract on ethanol-induced rat hepatic injury were reported by Pramyothin et al. The results which were confirmed by histopathological studies showed that the treatment of rats with P. emblica with ethanol enhanced liver cell recovery by bringing the levels of AST, ALT and interleukin-1beta back to normal[75]. Sultana et al. found that E. officinalis fruit extracts inhibit thioacetamideinduced oxidative stress and hyper-proliferation in rat liver[76].

3.16. P. major

P. major known as barhang in ITM have been extensively applied as liver tonic in Iran[12,20]. Türel et al. studied hepatoprotective effect of P. major in CCl₄-induced hepatotoxic rats. The results showed that *P. major* has a considerable hepatoprotective activity[77]. In a similar study by Atta et al., methanol extract of P. major was evaluated for its potential hepatoprotective effect against CCl₄induced hepatic damage. The applied treatment significantly decreased the elevated AST and gamma-glutamyl transpeptidase activities and blood triglyceride level. The effect was nearly similar to that of silymarin[78]. Mello et al. indicated that P. major was able to prevent oxidative mitochondrial damage, which contributed to the understanding of its hepatoprotective action against reactive oxygen species-mediated toxicity[79]. The results in the work of Oto et al., suggested that the preventive effects of P. major on 7,12dimethylbenz(a)anthracene-induced oxidative damage in Wistar albino rats might be due to the decrease of free radical generation. The antioxidant activity of this plant was established by Oto et al. as well[80]. It was also proved that some flavonoids which derived from P. major possessed hepatoprotective effects of baicalein against CCl₄-induced liver injury in rats[81,82]. Furthermore, aucubin was found to be potential antidote for poisonous amanita mushrooms in mice to protect against liver damage induced by amanitin[83].

3.17. C. cassia

The bark of *C. cassia* known as salikheh in ITM has been well-known as liver tonic in Iran[12,13]. The protective effects of *C. cassia* in the fibrogenesis of activated hepatic stellate cells-T6 cells and dimethylnitrosamine-induced acute liver injury in rats were successfully demonstrated by Lim *et al.* The results were significantly protected by *C. cassia* powder in the serum total protein, albumin, total bilirubin, direct-reacting bilirubin, GOT, GPT, and ALP[84]. Bansode reported that *C. cassia* inhibited fibrogenesis, followed by hepatic stellate cells-T6 cell activation, and increased restoration of liver function, which ultimately resulted in acute liver injury[85]. According to the results reported by Kanuri *et al.*, cinnamon extract protects against acute alcohol-induced liver steatosis in mice[86].

3.18. C. opobalsamum

Fruits of *C. opobalsamum* known as balasan in ITM have been the focus of Iranians[20]. Its applications as an efficient liver tonic were frequently emphasized by Sina[12]. Al-Howiriny *et al.* investigated the hepatoprotective activity of the ethanolic extract of *C. opobalsamum* in rats by inducing hepatotoxicity with CCl₄. This extract has been shown to possess significant protective effect by lowering serum transaminase levels of GOT, GPT, ALP and bilirubin. This study suggested that the plant *C. opobalsamum* may act as an antioxidant agent as it possessed hepatoprotective effect[87].

3.19. C. intybus

C. intybus known as kasni in ITM is one of the most important liver tonics[12]. Hepatoprotective activity of *C. intybus* leaves extract against CCl4-induced toxicity was investigated by Jamshidzadeh et al. It was worth to mention that high concentration of the plant extract was hepatotoxic[88]. In a similar study, the hepatoprotective activity of hydroalcholic extract of C. intybus was investigated on CCl₄-induced liver injury in rats. The results confirmed the hepatoprotective activity effects of the hydroalcholic extract of C. intybus[89]. Different fractions of alcoholic extract and phenolic compound from C. intybus seeds were screened for their antihepatotoxic activity on CCl₄-induced liver damage in albino rats by Ahmed et al. The histopathological study of the liver was also carried out, and the results showed almost complete normalization of the tissues as neither fatty accumulation nor necrosis[90]. In another study by Madani et al., the protective effects of polyphenolic extracts of Silybum marianum (S. marianum) and C. intybus on thioacethamide-induced hepatotoxicity in rat were investigated. Treatment with the polyphenolic extracts of *S. marianum* and *C. intybus* reduced the level of enzymes activities (ALT, AST and ALP) as well as total bilirubin, when comparing with thioacetamide group[91]. Butt *et al.* evaluated the hepatoprotective potential of *C. intybus* leaf extract on acetaminophen-induced liver damage in albino rats. Extract at a dose of 400 mg/kg body weight exhibited remarkable anti-hepatotoxic activity[92]. The phytochemical and hepatoprotective activities of hydroalcoholic extract of *C. intybus* leaves against nimesulide intoxicated albino rats was conducted by Mushtaq *et al.* It was ascertained that the leaves extracts of *C. intybus* possessed significant hepatoprotective activity[93].

3.20. Z. vulgaris

Z. vulgaris known as annab in ITM has been extensively used by Iranian scholars[20]. The protective effect of ethanolic extract of Z. vulgaris against hepatic injury induced by CCl₄ in rats has been investigated. Results revealed that although there was a significant decrease in liver enzymes of the treated groups, there were insignificant differences in protein and albumin concentrations between experimental groups. In addition, Z. vulgaris treatment reduced hepatic necrosis and portal inflammation compared with the control group. According to both serological and pathological investigations, Z. vulgaris showed hepatoprotective impact against CCl₄-induced liver injury[94].

3.21. C. avellana

C. avellana known as fandogh in ITM has been endorsed by Sina as an efficient liver tonic[12]. Effects of *C. avellana* in acetaminophenand CCl₄-induced toxicosis have been investigated by Rusu *et al. C. avellana* extract had some beneficial effects in CCl₄ toxicosis: it reduced hepatocytolysis as well as histological lesions and returned the activity of some enzymes to normal values[95,96].

3.22. C. fistula

C. fistula known as foloos in ITM was frequently utilized by Persians[12,13,20]. Pradeep et al. evaluated the hepatoprotective and antioxidant effect of C. fistula leaf extract on DEN-induced liver injury. It was observed that C. fistula protected the liver against DEN-induced hepatic injury in rats[97,98]. Bhakta et al. investigated the hepatoprotective activity of the n-heptane extract of C. fistula leaves. The extract at a dose of 400 mg/kg body weight exhibited significant protective effect by lowering serum levels of AST, ALT, bilirubin and ALP. The protective effect was comparable to that of a standard hepatoprotective agent[99,100]. Evaluation of hepatoprotective activity of aqueous extract of C. fistula fruit pulp against CCl₄-induced liver damage in albino rats was investigated by Das et al. Aqueous extract of fruit pulp of C. fistula possessed significant hepatoprotective activity[101].

3.23. Z. officinale

Z. officinale known as zanjabil in ITM has been widely used in ancient Iranian medicine[12,20]. The protective effects of the ethanolic extract of Z. officinale rhizome on acute hepatotoxicity induced by acetaminophen were studied in plasma and hepatic tissue samples. It showed protective effect against acetaminopheninduced hepatotoxicity. The most satisfactory results were obtained with high doses of plant extract[102,103]. In another study, the effect of ethanolic extract of Z. officinale rhizome was tested against CCl₄and acetaminophen-induced liver toxicities in rats. It was indicated that the rhizome oil of Z. officinale could be useful in preventing chemically induced acute liver injury[104,105]. The effect of ginger (Z. officinale) upon hepatotoxicity induced by the anticancer drug and ADR in albino rats, was studied by Sakr et al. The results depicted that ginger had protective effect against liver damage induced by ADR due to its antioxidant activities[106]. The combination therapy of atorvastatin and Z. officinale in rat liver was studied by Heeba and Abd-Elghany. Combination regimens containing Z. officinale and low dose of statins could be advantageous in treating hypercholesterolemia patients who are susceptible to liver failure[107]. The protective ability of Z. officinale against Hg, Pb and Cd accumulation in liver was examined. It was conceived that Z. officinale could affect the bioavailability, elimination and uptake of these metals in a time-dependent way with the highest beneficial reducing effect to Cd followed by Hg and the least protection to Pb in the liver[108].

4. Discussion

Despite the fact that people pay much more attention to their health, they have been involved in controversial health issues in today's society. Considering the fact that liver plays a crucial role in the body, liver failure is now the most common health issue. If it does not work properly, the whole body may encounter severe problems. Viral, autoimmune and metabolic diseases, wrong life style and dietary, cancers, drugs and herbal supplements abuse are some of the common causes of liver injuries. It seems that protecting and strengthening the liver against the above mentioned factors is essential. Meanwhile, finding effectual liver tonics has been emerged as a main issue.

ITM has offered efficient natural healing herbs with successful clinical trials for several diseases. In this review, a wide range of plants frequently used in ITM were presented as liver tonics which could make an immense impact on the preventive strategies in liver protection. It should be noted that all of them have shown satisfactory *in vitro* and *in vivo* hepatoprotective. Although the mechanisms of action have been understood to some extent, thoroughgoing studies would be more advantageous in discovery of drugs with low toxicity. Among various reported herbs in this review, ghafes (*A. eupatoria*), kasni (*C. intybus*), anar (*P. granatum*), darchin

(C. zeylanicum), za'feran (C. sativus), gole-sorkh (R. damascena), and zereshk (B. vulgaris) were found to be much more useful and extensively recommended in Persian medical literature.

In spite the fact that ITM possess a valuable source in medicine, current marketed hepatoprotective agents such as milk thistle (*S. marianum*) and general antioxidant supplements, are still prescribed by Iranian physicians. Recently, Iranian scientists have paid more attention to herbal remedies evolved from ITM to develop novel therapeutic agents in their drug discovery researches. In this context, there are various plants such as Gharanfal, Faranjamashk, Kholanjan, Kashoos, Zaranb, Oshnah, Gole-nasrin, Narmeshk, Poodineh koohi, *etc.* which have been traditionally used as liver tonics by Persians, but any investigation has not yet been conducted. We intend to investigate the therapeutic efficacy of these plants and hope to publish the results in our next research articles.

Conflict of interest statement

We declare that we have no conflict of interest.

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Comments

Background

Human liver is one of the most important organs in our body. It has a wide range of functions, including detoxification, protein synthesis, and production of biochemicals which are necessary for digestion. Because of wrong lifestyle and dietary habits, food/drinking contamination, chemical drug abuse, the incidence of liver diseases and/or liver function abnormalities are increasing in the world. Therefore, new hepatoprotective remedies derived from plants are urgent.

Research frontiers

The present manuscript reviewed the plants which have hepatoprotection or antihepatotoxicity ability as liver tonic in Iran.

Related reports

In the current study, authors conducted a comprehensive review of scientific information database and medical sciences databases covering all *in vitro* and *in vivo* studies of medicinal plants as liver tonics and hepatoprotective candidates.

Applications

From the literature survey, it has been found that plants used in Iran are safe to humans. They may be used as an adjuvant for the treatment and prevention of liver injury.

Peer review

This is a valuable review. It introduced plants which have been traditionally used as liver tonics in Iran. This paper will promote the utilization of natural and traditional resources for contemporary health care. Herbal medicines have an extremely valuable, rich, lengthy and extensive practical history.

References

- [1] Balunas MJ, Kinghorn AD. Drug discovery from medicinal plants. *Life Sci* 2005; **78**(5): 431-441.
- [2] Farnsworth NR, Akerele O, Bingel AS, Soejarto DD, Guo Z. Medicinal plants in therapy. *Bull World Health Organ* 1985; **63**(6): 965-981.
- [3] Bai RL, Paull KD, Herald CL, Malspeis L, Pettit GR, Hamel E. Halichondrin B and homohalichondrin B, marine natural products binding in the vinca domain of tubulin. Discovery of tubulin-based mechanism of action by analysis of differential cytotoxicity data. *J Biol Chem* 1991; 266(24): 15882-15889.
- [4] Newman DJ, Cragg GM, Snader KM. The influence of natural products upon drug discovery. *Nat Prod Rep* 2000; **17**(3): 215-234.
- [5] Cragg GM, Newman DJ. International collaboration in drug discovery and development from natural sources. *Pure Appl Chem* 2005; 77(11): 1923-1942.
- [6] Kinghorn AD. Pharmacognosy in the 21st century. J Pharm Pharmacol 2001; 53(2): 135-148.
- [7] Shahani S. Evaluation of hepatoprotective efficacy of APCL-A poly herbal formulation in vivo in rats. Indian Drugs 1999; 36: 628-631.
- [8] Subramoniam A, Pushpangadan P. Development of phytomedicine for liver diseases. *Indian J Pharmacol* 1999; 31(3): 166-175.
- [9] Adewusi EA, Afolayan AJ. A review of natural products with hepatoprotective activity. J Med Plants Res 2010; 4(13): 1318-1334.
- [10] Ahsan R, Islam KM, Musaddik A, Haque E. Hepatoprotective activity of methanol extract of some medicinal plants against carbon tetrachloride induced hepatotoxicity in albino rats. *Global J Pharmacol* 2009; 3(3): 116-122.
- [11] Bodakhe SH, Ram A. Hepatoprotective properties of *Bauhinia variegata* bark extract. *Yakugaku Zasshi* 2007; **127**(9): 1503-1507.
- [12] Sina I. [The canon of medicine]. Tehran: Soroush Press; 2005. Persian.
- [13] Razi M. [Alhavi fi-al-tib]. Beirut: Dar Al_Kotob Al-ilmiyah; 2000. p. Volumes 20, 21. [Online] Available from: http://www.almaktabah.eu/images/images_books [Accessed on 2nd December, 2014] Arabic.
- [14] Asha VV, Pushpangadan P. Preliminary evaluation of the antihepatotoxic activity of *Phyllanthus kozhikodianus*, *P. maderaspatensis* and *Solanum indicum*. *Fitoterapia* 1998; 69(3): 255-259.
- [15] Saleem TSM, Chetty CM, Ramkanth S, Rajan VST, Kumar KM, Gauthaman K. Hepatoprotective herbs-a review. *Int J Res Pharm Sci* 2010; 1(1): 1-5.
- [16] Samojlik I, Laki N, Mimica-Duki N, akovic-Svajcer K, Bozin B. Antioxidant and hepatoprotective potential of essential oils of coriander (Coriandrum sativum L.) and caraway (Carum carvi L.)(Apiaceae). J Agric Food Chem 2010; 58(15): 8848-8853.

- [17] Langmead L, Rampton DS. Review article: herbal treatment in gastrointestinal and liver disease-benefits and dangers. *Aliment Pharmacol Ther* 2001; 15(99): 1239-1252.
- [18] Kumar CH, Ramesh A, Kumar JNS, Ishaq BM. A review on hepatoprotective activity of medicinal plants. *Int J Pharm Sci Res* 2011; 2(3): 501-515.
- [19] Farghali H, Kmonícková E, Lotková H, Martínek J. Evaluation of calcium channel blockers as potential hepatoprotective agents in oxidative stress injury of perfused hepatocytes. *Physiol Res* 2000; 49(2): 261-268.
- [20] Aghili MH. [*Makhzan-al-Advia*]. Tehran: Tehran University of Medical Sciences; 2009. Persian.
- [21] Yoon SJ, Koh EJ, Kim CS, Zee OP, Kwak JH, Jeong WJ, et al. *Agrimonia eupatoria* protects against chronic ethanol-induced liver injury in rats. *Food Chem Toxicol* 2012; **50**(7): 2335-2341.
- [22] Kang SC, Kawk JH, Lee MH, Oh JS, Zee OP, Lee CM, et al. [Hepatoprotective effects of aqueous extract from aerial part of Agrimmony]. Korean J Pharmacogn 2006; 37: 28-32. Korean.
- [23] Orhan DD, Orhan N, Ergun E, Ergun F. Hepatoprotective effect of Vitis vinifera L. leaves on carbon tetrachloride-induced acute liver damage in rats. J Ethnopharmacol 2007; 112(1): 145-151.
- [24] Pari L, Suresh A. Effect of grape (Vitis vinifera L.) leaf extract on alcohol induced oxidative stress in rats. Food Chem Toxicol 2008; 46(5): 1627-1634.
- [25] Liu T, Zhao J, Ma L, Ding Y, Su D. Hepatoprotective effects of total triterpenoids and total flavonoids from *Vitis vinifera* L. against immunological liver injury in mice. *Evid Based Complement Alternat Med* 2012; doi: 10.1155/2012/969386.
- [26] Dogan A, Celik I. Hepatoprotective and antioxidant activities of grapeseeds against ethanol-induced oxidative stress in rats. Br J Nutr 2012; 107(1): 45-51.
- [27] Sharma SK, Suman, Vasudeva N. Hepatoprotective activity of Vitis vinifera root extract against carbon tetrachloride-induced liver damage in rats. Acta Pol Pharm 2012; 69(5): 933-937.
- [28] Mohajeri D, Doustar Y, Rezaei A, Mesgari-Abbasi M. Hepatoprotective effect of ethanolic extract of *Crocus sativus* L. (Saffron) stigma in comparison with silymarin against rifampin induced hepatotoxicity in rats. *Zahedan J Res Med Sci* 2011; 12(5): 53-59.
- [29] Mohajeri D, Doustar Y. [Protective effect of ethanolic extract of *Crocus sativus* L. (Saffron) stigma against cisplatin induced hepatotoxicity in rats]. *Med Sci J Islam Azad Univ* 2012; 21(4): 251-261. Persian.
- [30] Mokhtari M, Shariati M, Ajdari D. [Protective effect of hydroalcoholic extracts of saffron on liver enzymes (AST, ALT, ALP) by hypervitaminosis A in male rat]. J Sabzevar Univ Med Sci 2013; 20(2): 133-141. Persian.
- [31] Rahbani M, Mohajeri D, Rezaie A, Nazeri M. Protective effect of ethanolic extract of saffron (dried stigmas of *Crocus sativus L.*) on hepatic tissue injury in streptozotocin-induced diabetic rats. *J Anim Vet Adv* 2012; 11(12): 1985-1994.
- [32] Amat N, Upur H, Blazekovi B. *In vivo* hepatoprotective activity of the aqueous extract of *Artemisia absinthium* L. against chemically and

- immunologically induced liver injuries in mice. *J Ethnopharmacol* 2010; **131**(2): 478-484.
- [33] Gilani AH, Janbaz KH. Preventive and curative effects of Artemisia absinthium on acetaminophen and CCl₄-induced hepatotoxicity. Gen Pharmacol 1995; 26(2): 309-315.
- [34] Yogeeta S, Ragavender HRB, Devaki T. Antihepatotoxic effect of *Punica granatum*. Acetone extract against isoniazid- and rifampicin-induced hepatotoxicity. *Pharm Biol* 2007; 45(8): 631-637.
- [35] Khalil EAM. A hepatoprotective effect of an aqueous extract of pomegranate (*Punica granatum* L.) rind against acetaminop hen treated rats. *Egypt J Hosp Med* 2004; 16: 112-118.
- [36] Bhanoji Rao ME, Dama GY. Evaluation of hepatoprotective activity of *Punica granatum* leaves on carbon tetrachloride induced hepatotoxicity in rats. *Int J Univers Pharm Life Sci* 2011; 1: 23-36.
- [37] Celik I, Temur A, Isik I. Hepatoprotective role and antioxidant capacity of pomegranate *Punica granatum* flowers infusion against trichloroacetic acid-exposed in rats. *Food Chem Toxicol* 2009; 47(1): 145-149.
- [38] Guo MZ, Li XS, Xu HR, Mei ZC, Shen W, Ye XF. Rhein inhibits liver fibrosis induced by carbon tetrachloride in rats. *Acta Pharmacol Sin* 2002; **23**(8): 739-744.
- [39] Wang JB, Zhao HP, Zhao YL, Jin C, Liu DJ, Kong WJ, et al. Hepatotoxicity or hepatoprotection? Pattern recognition for the paradoxical effect of the Chinese herb *Rheum palmatum L*. in treating rat liver injury. *PloS One* 2011; doi: 10.1371/journal.pone.0024498.
- [40] Tseng SH, Chien TY, Tzeng CF, Lin YH, Wu CH, Wang CC. Prevention of hepatic oxidative injury by Xiao-Chen-Chi-Tang in mice. J Ethnopharmacol 2007; 111(2): 232-239.
- [41] Taher M, Ghannadi A, Karmiyan R. [Effects of volatile oil extracts of *Anethum graveolens* L. and *Apium graveolens* L. seeds on activity of liver enzymes in rat]. *J Qazvin Univ Med Sci* 2007; **11**(2): 8-12. Persian.
- [42] Ahmed B, Alam T, Varshney M, Khan SA. Hepatoprotective activity of two plants belonging to the Apiaceae and the Euphorbiaceae family. J Ethnopharmacol 2002; 79(3): 313-316.
- [43] Popović M, Kaurinović B, Trivić S, Mimica-Dukić N, Bursać M. Effect of celery (*Apium graveolens*) extracts on some biochemical parameters of oxidative stress in mice treated with carbon tetrachloride. *Phytother Res* 2006; **20**(7): 531-537.
- [44] Sultana S, Ahmed S, Jahangir T, Sharma S. Inhibitory effect of celery seeds extract on chemically induced hepatocarcinogenesis: modulation of cell proliferation, metabolism and altered hepatic foci development. *Cancer let* 2005; 221(1): 11-20.
- [45] Singh A, Handa SS. Hepatoprotective activity of *Apium graveolens* and *Hygrophila auriculata* against paracetamol and thioacetamide intoxication in rats. *J Ethnopharmacol* 1995; **49**(3): 119-126.
- [46] Shivashri C, Rajarajeshwari T, Rajasekar P. Hepatoprotective action of celery (*Apium graveolens*) leaves in acetaminophen-fed freshwater fish (*Pangasius sutchi*). Fish physiol biochem 2013; 39(5): 1057-1069.
- [47] Morita T, Jinno K, Kawagishi H, Arimoto Y, Suganuma H, Inakuma T, et al. Hepatoprotective effect of myristicin from nutmeg *Myristica fragrans* on lipopolysaccharide/d-galactosamine-induced liver injury.

- J Agric Food Chem 2003; 51(6): 1560-1665.
- [48] Latha PG, Sindhu PG, Suja SR, Geetha BS, Pushpangadan P, Rajasekharan S. Pharmacology and chemistry of *Myristica fragrans* Houtt.-a review. *J Spices Arom Crops* 2005; 14(2): 94-101.
- [49] Singh A, Rao AR. Modulatory effect of *Areca nut* on the action of mace (*Myristica fragrans*, Houtt) on the hepatic detoxification system in mice. *Food Chem Toxicol* 1993; **31**(7): 517-521.
- [50] Chhabra SK, Rao AR. Transmammary modulation of xenobiotic metabolizing enzymes in liver of mouse pups by mace (*Myristica fragrans* Houtt.). *J Ethnopharmacol* 1994; **42**(3): 169-177.
- [51] Sohn JH, Han KL, Choo JH, Hwang JK. Macelignan protects HepG2 cells against *tert*-butylhydroperoxide-induced oxidative damage. *Biofactors* 2007; 29(1): 1-10.
- [52] Mavridis SK, Gortzi O, Lalas S, Paraschos S, Skaltsounis AL, Pappas IS. Hepatoprotective effect of *Pistacia lentiscus* var. *Chia* total extract against carbon tetrachloride-induced liver damage in rats. *Planta Med* 2008; doi: 10.1055/s-0028-1084336.
- [53] Janakat S, Al-Merie H. Evaluation of hepatoprotective effect of Pistacia lentiscus, Phillyrea latifolia and Nicotiana glauca. J Ethnopharmacol 2002; 83(1): 135-138.
- [54] Tonkaboni MM. [*Tohfeh al-Momenin*]. Tehran: Shahid Beheshti University of Medical Sciences; 2007. Persian.
- [55] Hermenean A, Popescu C, Ardelean A, Stan M, Hadaruga N, Mihali CV, et al. Hepatoprotective effects of *Berberis vulgaris* L. extract/β-cyclodextrin on carbon tetrachloride-induced acute toxicity in mice. *Int J Mol Sci* 2012; 13(7): 9014-9034.
- [56] Hwang JM, Wang CJ, Chou FP, Tseng TH, Hsieh YS, Lin WL, et al. Inhibitory effect of berberine on *tert*-butyl hydroperoxide-induced oxidative damage in rat liver. *Arch Toxicol* 2002; 76(11): 664-670.
- [57] Fallah Huseini H, Zareei Mahmoudabady A, Ziaei SA, Mehrazma M, Alavian SM, Kianbakht S, et al. The effects of *Taraxacum officinale* L. and *Berberis vulgaris* L. root extracts on carbon tetrachloride induced liver toxicity in rats. *J Med Plants* 2010; 9(Suppl 6): 45-52.
- [58] Eidi A, Zarin Ghalam J, Rezazade S, Adeli R. [Hepatoprotective effect of *Berberis vulgaris* L. extract on CCl₄-induced toxicity in rats]. *Kowsar Med J* 2011; 16: 169-173. Persian.
- [59] Gilani AH, Janbaz KH, Akhtar MS. Selective protective effect of an extract from *Fumaria parviflora* on paracetamol-induced hepatotoxicity. *Gen Pharmacol* 1996; 27(6): 979-983.
- [60] Jamshidzadeh A, Nikmahad H. Hepatoprotective effects of *Fumaria parviflora* L. on CCl₄-induced hepatotoxicity. *J Med Plants* 2006; 5: 34-39.
- [61] Alqasoumi SI, Al-Dosari MS, Al-Sheikh AM, Abdel-Kader MS. Evaluation of the hepatoprotective effect of *Fumaria parviflora* and *Momordica balsamina* from Saudi folk medicine against experimentally induced liver injury in rats. *Res J Med Plants* 2009; 3: 9-15.
- [62] Tripathi M, Singh BK, Mishra C, Raisuddin S, Kakkar P. Involvement of mitochondria mediated pathways in hepatoprotection conferred by *Fumaria parviflora* Lam. extract against nimesulide induced apoptosis in vitro. Toxicol In Vitro 2010; 24(2): 495-508.

- [63] Tripathi M, Singh BK, Raisuddin S, Kakkar P. Abrogation of nimesulide induced oxidative stress and mitochondria mediated apoptosis by Fumaria parviflora Lam. extract. J Ethnopharmacol 2011; 136(1): 94-102
- [64] Moselhy SS, Ali HK. Hepatoprotective effect of cinnamon extracts against carbon tetrachloride induced oxidative stress and liver injury in rats. *Biol Res* 2009; 42(1): 93-98.
- [65] Eidi A, Mortazavi P, Bazargan M, Zaringhalam J. Hepatoprotective activity of cinnamon ethanolic extract against CCl₄-induced liver injury in rats. Excli J 2012; 11: 495-507.
- [66] Rezk RG. Cinnamon (Cinnamomum zeylanicum N) attenuates hepatic and cardiac tissues injury induced by gamma radiation in male albino rats. Arab J Nucl Sci Appl 2013; 46(2): 356-362.
- [67] Saxena M, Shakya AK, Sharma N, Shrivastava S, Shukla S. Therapeutic efficacy of *Rosa damascena* Mill. on acetaminophen-induced oxidative stress in albino rats. *J Environ Pathol Toxicol Oncol* 2012; 31(3): 193-201.
- [68] Achuthan CR, Babu BH, Padikkala J. Antioxidant and hepatoprotective effects of *Rosa damascena*. *Pharm Biol* 2003; **41**(5): 357-361.
- [69] Saha P, Mazumder UK, Haldar PK, Bala A, Kar B, Naskar S. Evaluation of hepatoprotective activity of *Cucurbita maxima* aerial parts. *J Herb Med Toxicol* 2011; 5(1): 17-22.
- [70] Dubey SD. Overview on *Cucurbita maxima*. Int J Phyto Pharm 2012; **2**(3): 68-71.
- [71] Nidhi J, Pathak AK. Hepatoprotective effect of methanolic extract of Cucurbita maxima and Lagenaria siceraria seeds. Int J Pharm Chem Biol Sci 2012; 2(2): 151-154.
- [72] Malar HLV, Bai SSM. Hepatoprotective activity of *Phyllanthus emblica* against paracetamol induced hepatic damage in Wister albino rats. *Afr J Basic Appl Sci* 2009; 1(1-2): 21-25.
- [73] Jeena KJ, Joy KL, Kuttan R. Effect of Emblica officinalis, Phyllanthus amarus and Picrorrhiza kurroa on N-nitrosodiethylamine induced hepatocarcinogenesis. Cancer Let 1999; 136(1): 11-16.
- [74] Panchabhai TS, Ambarkhane SV, Joshi AS, Samant BD, Rege NN. Protective effect of *Tinospora cordifolia*, *Phyllanthus emblica* and their combination against antitubercular drugs induced hepatic damage: an experimental study. *Phytother Res* 2008; 22(5): 646-650.
- [75] Pramyothin P, Samosorn P, Poungshompoo S, Chaichantipyuth C. The protective effects of *Phyllanthus emblica* Linn. extract on ethanol induced rat hepatic injury. *J Ethnopharmacol* 2006; **107**(3): 361-364.
- [76] Sultana S, Ahmed S, Sharma S, Jahangir T. Emblica officinalis reverses thioacetamide-induced oxidative stress and early promotional events of primary hepato carcinogenesis. J Pharm Pharmacol 2004; 56(12): 1573-1579.
- [77] Türel I, Ozbek H, Erten R, Oner AC, Cengiz N, Yilmaz O. Hepatoprotective and anti-inflammatory activities of *Plantago major L. Indian J Pharmacol* 2009; 41(3): 120-124.
- [78] Atta AH, Nasr SM, Mouneir SM. Potential protective effect of some plant extracts against carbon tetrachloride-induced hepatotoxicity. Afr J Tradit Complement Altern Med 2006; 3(3): 1-9.

- [79] Mello JC, Guimarães NS, Gonzalez MV, Paiva JS, Prieto T, Nascimento OR. Hydroxyl scavenging activity accounts for differential antioxidant protection of *Plantago major* against oxidative toxicity in isolated rat liver mitochondria. *J Pharm Pharmacol* 2012; 64(8): 1177-1187.
- [80] Oto G, Ekin S, Ozdemir H, Demir H, Yasar S, Levent A, et al. *Plantago major* protective effects on antioxidant status after administration of 7,12-dimethylbenz(a)anthracene in rats. *Asian Pac J Cancer Prev* 2011; 12(2): 531-535.
- [81] Aziz SA, Yik OP, Hanafiah SN, Mustafa MS. Antioxidant properties of *Plantago major* (ekor anjing) and *Pithecellobium jiringa* (jering). *J Herbal Med Toxicol* 2012; **6**: 147-152.
- [82] Samuelsen AB. The traditional uses, chemical constituents and biological activities of *Plantago major* L. A review. *J Ethnopharmacol* 2000; 71(1-2): 1-21.
- [83] Chang LM, Yun HS, Kim YS, Ahn JW. Aucubin: potential antidote for alpha-amanitin poisoning. J Toxicol Clin Toxicol 1984; 22(1): 77-85.
- [84] Lim CS, Kim EY, Lee HS, Soh Y, Sohn Y, Kim SY, et al. Protective effects of *Cinnamomum cassia* Blume in the fibrogenesis of activated HSC-T6 cells and dimethylnitrosamine-induced acute liver injury in SD rats. *Biosci Biotechnol Biochem* 2010; 74(3): 477-483.
- [85] Bansode VJ. A review on pharmacological activities of *Cinnamomum cassia* Blume. *Int J Green Pharm* 2012; **6**(2): 102-108.
- [86] Kanuri G, Weber S, Volynets V, Spruss A, Bischoff SC, Bergheim I. Cinnamon extract protects against acute alcohol-induced liver steatosis in mice. *J Nutr* 2009: 139(3): 482-487.
- [87] Al-Howiriny TA, Al-Sohaibani MO, Al-Said MS, El-Tahir KH, Rafatullah S. Hepatoprotective properties of *Commiphora opobalsamum* ("balessan"), a traditional medicinal plant of Saudi Arabia. *Drugs Exp Clin Res* 2004; 30(5-6): 213-220.
- [88] Jamshidzadeh A, Khoshnood MJ, Dehghani Z, Niknahad H. Hepatoprotective activity of *Cichorium intybus* L. leaves extract against carbon tetrachloride induced toxicity. *Iran J Pharm Res* 2006; 1: 41-46.
- [89] Heibatollah S, Reza NM, Izadpanah G, Sohailla S. Hepatoprotective effect of *Cichorium intybus* on CCl₄-induced liver damage in rats. *Afr J Biochem Res* 2008; 2: 141-144.
- [90] Ahmed B, Al-Howiriny TA, Siddiqui AB. Antihepatotoxic activity of seeds of *Cichorium intybus*. J Ethnopharmacol 2003; 87(2-3): 237-240.
- [91] Madani H, Talebolhosseini M, Asgary S, Naderi GH. Hepatoprotective activity of *Silybum marianum* and *Cichorium intybus* against thioacetamide in rat. *Pak J Nutr* 2008; 7(1): 172-176.
- [92] Butt K, Yunas S, Sheikh RM. Hepatoprotective effect of *Cichorium intybus* on paracetamol induced liver damage in albino rats. *Libyan Agric Res Cen J Int* 2012; **3**(2): 60-63.
- [93] Mushtaq A, Ahmad M, Jabeen Q. Pharmacological role of *Cichorium intybus* as a hepatoprotective agent on the elevated serum marker enzymes level in albino rats intoxicated with nimesulide. *Int J Curr Pharm Res* 2013; 5(3): 25-30.

- [94] Ebrahimi S, Ashkani-Esfahani S, Emami Y, Riazifar S. Hepatoprotective effect of *Zizyphus vulgaris* on carbon tetrachloride (CCl₄) induced liver damage in rats as animal models. *Galen Med J* 2013; **2**(3): 88-94.
- [95] Rusu MA, Bucur N, Puică C, Tămaş M. Effects of Corylus avellana in acetaminophen and CCl₄ induced toxicosis. Phytother Res 1999; 13(2): 120-123.
- [96] Sharma A, Makwana M, Rathore HS. Will herbal-paracetamol combination drug prevent both liver and kidney disease?-results and possibilities. *Ethnobot Leaflets* 2008; 12: 286-298.
- [97] Pradeep K, Mohan CV, Gobianand K, Karthikeyan S. Effect of *Cassia fistula* Linn. leaf extract on diethylnitrosamine induced hepatic injury in rats. *Chem Biol Intract* 2007; 167: 12-18.
- [98] Danish M, Singh P, Mishra G, Srivastava S, Jha KK, Khosa RL. Cassia fistula Linn. (Amulthus) an important medicinal plant: a review of its traditional uses, phytochemistry and pharmacological properties. J Nat Prod Plant Resour 2011; 1(1): 101-118.
- [99] Bhakta T, Banerjee S, Mandal SC, Maity TK, Saha BP, Pal M. Hepatoprotective activity of *Cassia fistula* leaf extract. *Phytomedicine* 2001; 8(3): 220-224.
- [100]Bhakta T, Mukherjee PK, Mukherjee K, Banerjee S, Mandal SC, Maity T K, et al. Evaluation of hepatoprotective activity of *Cassia fistula* leaf extract. *J Ethnopharmacol* 1999; 66(3): 277-282.
- [101]Das S, Sarma G, Barman S. Hepatoprotective activity of aqueous extract of fruit pulp of *Cassia fistula* (AFCF) against carbon tetrachloride (CCl₄) induced liver damage in albino rats. *J Clin Diagn Res* 2008; **2**: 1133-1138.
- [102]Abdullah N, Saat NZM, Hasan HA, Budin SB, Kamaralzaman S. Protective effect of the ethanol extract of *Zingiber officinale* Roscoe on paracetamol induced hepatotoxicity in rats. *J Sains Kesihatan Malaysia* 2004; 2(2): 85-95.
- [103] Ajith TA, Hema U, Aswathy MS. Zingiber officinale Roscoe prevents acetaminophen-induced acute hepatotoxicity by enhancing hepatic antioxidant status. Food Chem Toxicol 2007; 45(11): 2267-2272.
- [104]Yemitan OK, Izegbu MC. Protective effects of Zingiber officinale (Zingiberaceae) against carbon tetrachloride and acetaminopheninduced hepatotoxicity in rats. Phytother Res 2006; 20(11): 997-1002.
- [105] Ezeonu CS, Egbuna PAC, Ezeanyika LUS, Nkwonta CG, Idoko ND. Antihepatotoxicity studies of crude extract of *Zingiber officinale* on CCl₄ induced toxicity and comparison of the extract's fraction D hepatoprotective capacity. *Res J Med Sci* 2011; 5(2): 102-107.
- [106]Sakr SA, Mahran HA, Lamfon HA. Protective effect of ginger (*Zingiber officinale*) on adriamycin-induced hepatotoxicity in albino rats. *J Med Plant Res* 2011; 5(1): 133-140.
- [107]Heeba GH, Abd-Elghany MI. Effect of combined administration of ginger (*Zingiber officinale* Roscoe) and atorvastatin on the liver of rats. *Phytomedicine* 2010; **17**(14): 1076-1081.
- [108]Nwokocha CR, Owu DU, Nwokocha MI, Ufearo CS, Iwuala MO. Comparative study on the hepatoprotection to heavy metals of *Zingiber officinale. Pharmacognosy Res* 2012; **4**(4): 208-213.