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Two double-stranded RNA viruses exist as permanent persistent infections of the yeast *Saccharomyces cerevisiae*: ScVL1 and ScVL_a. Both belong to the Totiviridae, which include a number of fungal and protozoan double-stranded RNA viruses. Although ScVL1 and ScVL_a share the same genomic organization and mode of expression and coexist in the same cells, they show no evidence of recombination: with one limited exception, sequence conservation is detectable only in regions conserved in all totiviruses. Both have two open reading frames on their single essential RNAs: *cap* (encoding a capsid polypeptide) and *pol* (encoding an RNA-dependent RNA polymerase). The ScVL_a virus, like ScVL1, appears to express its Pol domain by a -1 translational frameshift. © 1996 Academic Press, Inc.

We have shown that a group of double-stranded RNA (dsRNA) viruses of lower eucaryotes are more closely related to each other than they are to any other viruses (7). These are members of the Totiviridae, which have a single essential dsRNA segment; most are noninfectious. At least in the fungi, it appears to be quite common for a given species to be a carrier of several such viruses simultaneously. This was first conclusively demonstrated for the yeast *Saccharomyces cerevisiae*, most laboratory strains of which have at least two such viruses (2). We originally named their genomic RNAs L1 and La (2). In an average laboratory strain of yeast, the *S. cerevisiae* La virus (ScVL_a) is present at about 5% the level of the L1 virus (ScVL1) (3). La is also known as L-BC and L1 as L-A (4).

The entire sequence of the essential segment of the ScVL1 virus (L1) has been known for some time (5, 6). We have cloned and sequenced cDNAs representing the entire sequence of La, which has a genomic organization similar to that of L1. La dsRNA was extracted from strain 299 (*MAT a lys1 mak3-1 [KIL-O] KIL-R⁺ La*) from the Yeast Genetics Stock Center. dsRNA was isolated by phenol extraction from cells disrupted by glass beads, followed by CF11 chromatography (7). The original La cDNA clone was derived by the RNase H method (8) by Martin Nemeroff. Subsequent clones were made by the same method, using primers derived from the known sequence for the first strand. Sequencing was by the dideoxy method (9). The set of six cDNA clones used for sequencing La and the sequencing runs performed, as well as the genomic organization of La, are outlined in Fig. 1. Sequencing of

both strands is complete, except for the very small regions at the ends which were determined by RNA sequencing. All overlapping regions of independently isolated clones were identical in sequence. The entire sequence of the viral plus strand of 4615 bases (given as DNA here) is shown in Fig. 2.

The 3' end sequences of both strands of total L dsRNA were previously determined by ³²pCp labeling and direct RNA sequencing (10). It is now clear that the 3' T1 oligonucleotides of the plus strands of both L1 and La are (G)CA, while the 3' T1 oligonucleotides of the minus strands are (G)AAAAUUUUUCA (L1) and (G)AAAAU-UCA (La). This last is the minor oligonucleotide 6 of the T1 digest of ³²pCp-labeled total L dsRNA (10). Both L1 and La therefore have 3' terminal A residues not encoded in the template strands.

The genomic organization of La is identical to that of

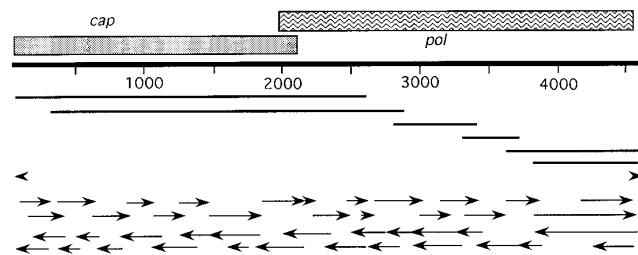


FIG. 1. Cloning and sequencing strategy for La. The two open reading frames are labeled above a scale (in bases on the plus strand). Below the scale the individual cDNA clones sequenced are indicated as straight lines. The orientation of sequencing runs is indicated by arrows. The two short RNA sequencing runs are above and the DNA sequencing runs below. All regions (with the exception of short regions at the 5' and 3' ends of the plus strand) were sequenced on both strands, usually with multiple runs.

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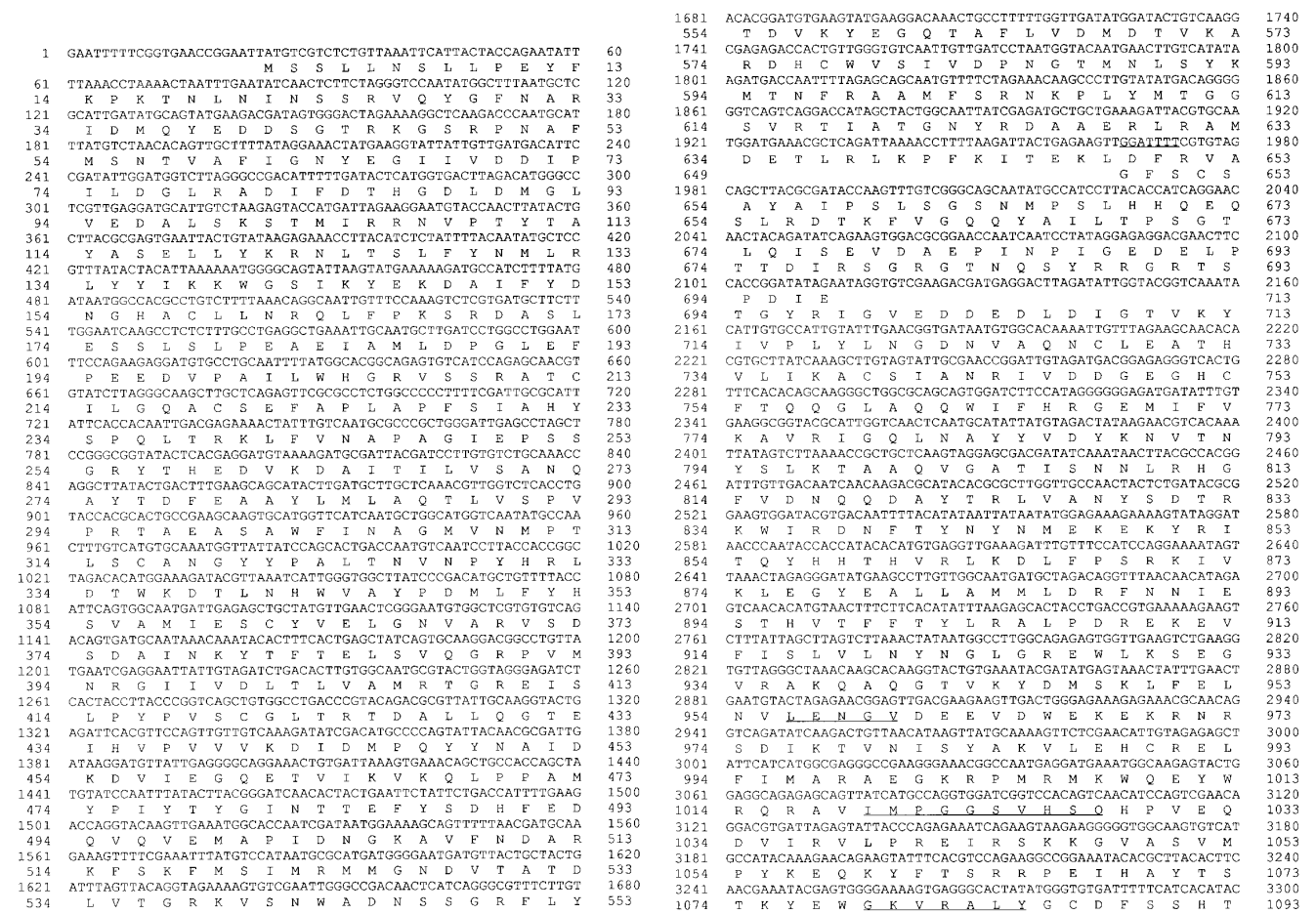


FIG. 2. Sequence of La. The complete sequence of the cDNA plus strand is shown, along with the predicted plus strand in the DNA sequence, the slippery site is underlined. In the protein sequence, the La sequences that correspond to the eight conserved motifs of the totivirus RDRPs are underlined. This cDNA sequence is deposited in Genbank under Accession No. U01060.

L1 (Fig. 1). There are two large overlapping reading frames, *cap* and *pol*, which in this case overlap for 153 bases. The *cap* ORF (bases 24–2114) predicts a protein (Cap) of 78.3 kDa and 697 amino acids in length. This ORF begins with the first AUG in the viral plus strand. The predicted size of La Cap (78.3 kDa) is quite close to that determined by SDS–PAGE, by which it has been estimated at 77 (11) and 80 kDa (3). The major *in vitro* translation product of denatured La dsRNA is a protein that comigrates with La Cap, as expected (3).

A *cap*–*pol* frameshift fusion protein of 1512 amino acids or 171.5 kDa is predicted, with a frameshift at a GGAUUUU slippery sequence starting at base 1967. Again, this is similar to the L1 Cap–Pol fusion protein of 171 kDa, which is generated by a frameshift at the slippery sequence starting at base 1957 (5, 6, 12).

The L1 packaging signal, two stems separated by an unpaired A residue, with several conserved bases in the loop, was not found in the La plus strand. This is consistent with the inability of ScVL1 particles to package La plus strands (13).

As described previously (1), the La and L1 RDRPs are

38% identical through the portion containing the eight conserved motifs in Pol. Overall, the Pol domains are 29% identical in predicted amino acid sequence, while the Cap domains are only 21% identical overall (below statistical significance).

Unexpectedly, there is an additional region of high conservation of sequence within the *cap* ORF (Fig. 3). This region of 60 amino acids is 37.7% identical in amino acid sequence in the two proteins. We previously noted a region of L1 Cap with some similarity to the picornavirus capsid polypeptides (14). The 60-amino-acid region of similarity between the L1 and La Cap proteins maps in the middle of the previously observed region of similarity between the L1 Cap and the picornavirus vp3. This lends further support to speculation that this region is involved in the protein–protein contacts necessary to assemble an icosahedron. Viral interference experiments are also consistent with this hypothesis (15).

In contrast, comparison of the RNA sequences of L1 and La detects only very limited similarities, confined to the most conserved region of *pol* (results not shown).

3301 AATGGCTGATTTGGATTGTTACAATGCGAGGATACATTCCCGGGCITTTGTACCAACAGG 3360
 1094 M A D F G L L Q C E D T F P G F V P T G 1113
 3361 GTCITACGCCAATGAGGATTATGTCAGGACCAGAATTGCTGGGACTCACTCATGTATGCCC 3420
 1114 S Y A N E D Y V R T R I A G T H S L I P 1133
 3421 TTTCTGTTACGATTTTCAGATTTTCAACAGGCAACATTAAGGAAGCCATGCAAGCAGT 3480
 1134 F C Y D F D D F N S O H S K E A M Q A V 1153
 3481 GATTGATCGATGGATACTGCTCTATCAGGATAAGTTAACAGATGACCAGATGAGGCGGCG 3540
 1154 I D A W I S V Y H D K L T D D Q I E A A 1173
 3541 AAAGTGGACACGAACCGGTAGATAGAATTGGTCCTCACCACCTAACACTGGTGAGAC 3600
 1174 K W T R N S V D R M V A H Q P N T G E T 1193
 3601 TTAATGATGTTAAAGGGCACTGTTAGTGGCTGGCGATTAACAACATTTTCAATACGGC 3660
 1194 Y D V K G T L F S G W R L T T F F N T A 1213
 3661 GTTGACTATTGCTACCTGGCTAATGCAGGTAATAACTCACTAGTGCCAACGAGTCTCCA 3720
 1214 L N Y C Y L A N A G I N S L V P T S L H 1233
 3721 TAATGGTATGATGATTTTTCAGGGATAAGGACAATAGCTGACGGTATTTCTTGATCAA 3780
 1234 N S D D V F A G I R T I A D G I S L I K 1253
 3781 AAACCGCCAGCCCGGAGTTCGCCCTAATCAACAATAAATGAACATTTGGTACGATAGC 3840
 1254 N A A A T G V R A N T T K M N T I G T T A 1273
 3841 AGAGTTTGTAGAGTIGATATCGTGCAAAAATAGTACTGGCAGCAGTATTTAACAAAG 3900
 1274 E F L E V D M R A K N S T G S Q Y L T B 1293
 3901 AGGGATTGCTACTCTCACGCACAGTAGGGTGTAGCTGATGCAACCACTGACATTGGCGCA 3960
 1294 G I A T F T H S R V E S D A P L T L R N 1313
 3961 TCTATGATCTGCTTACAAAACCAGATATGACGAGATTTTGTAGCTCGTGGCGCAGGATCGA 4020
 1314 L V S A Y K T R Y D E I L A R G A S I D 1333
 4021 TAACATGAAGCCACTCTATCGTAAGCAATTTATTTTTTCTAGAAGTGTCTCAATGTCGA 4080
 1334 N M K P L Y R K Q L F F A R K L F N V E 1353
 4081 GAAGGACATTTGTCACAACTCTGATAACGATGGACATATCATGTGGCGTGTGAAGKAA 4140
 1354 K D I V D N L I D K G V G M S G L Q E K 1373
 4141 GSGTAGGTTATCAGAGATGGTTTACAGGAGGTTGACATTAAGAATATAGATAGTATAG 4200
 1374 G R V S E M V L Q E V D I E N I D S Y R 1393
 4201 GAAGACAAGGATGATCCCAAACTGATTGACAGGGGGTGGCCATTATATCATCTCTCT 4260
 1394 K T R M I A V R H A W K G M S G L H I V 1413
 4261 GAAACTAATTTCCGAGATAGCTGATCTATCACAAAGAGACACGGTAGAGTACAGT 4320
 1414 K T N F S E I A D A I T R E T R V E S V 1433
 4321 GACCAAGGCTTATAATGTTAAGAAGAAACGGTCCGTACGCCGGTTTAGGCACTAAGCGC 4380
 1434 T K A Y N V K K T V V R A F R D L S A 1453
 4381 AGCATATCATGAAAGGCGGTGAGACATGCTTGGAAAGGGATGAGTGGACTACACATAGT 4440
 1454 A Y H E R A V R H A W K G M S G L H I V 1473
 4441 CAACAGGATTCGATGGGAGTGAGCAACTTAGTATGGTGTGTAGCAAAAATCAATCCTGC 4500
 1474 N R I R M G V S N L V M V V S K I N F A 1493
 4501 AAAAGCTAATGTGCTAGCCAAATCAGGAGATCTCAAAAATGGCTGCAGTCCCTACATG 4560
 1494 K A N V L A K S G D P T K W L A V L T 1512
 4561 ATATACAGGCAACCACATAAGACCTGAGAACAAAGGTACATACGATACTACGCA 4615

FIG. 2—Continued

TABLE 1

Frameshift Activity of La Minimal Region in Yeast

Region present in construct	Mutation	Activity	% read-through vector
Read-through vector	None	11,343	100
1957–2001	None	104.8	0.92
1957–2004	None	210	1.85
1957–2004	GGATTTT to GcATTTT	34.4	0.30
1957–2004	GGATTTT to GGATgTT	33.4	0.29

without a triplet of identical bases at the 5' end (20). We have placed this region (the slippery site and adjacent sequences) in a β-gal reporter vector (pG4LacZ), which was derived from pG4 (15), by inserting LacZ as a NcoI flush end, KpnI sticky end fragment from p3p (21) into a SalI flush-ended–KpnI sticky-ended pG4. This allows us to detect frameshifting in yeast strain T120 (15) by the synthesis of β-gal in vivo (21). β-Galactosidase assays were performed as described (17) on whole cell lysates and expressed as arbitrary units per cell (22). All assays were performed on three independent transformants simultaneously. Standard deviations of measurements were usually less than 10% of the mean.

A minimal region of 48 bp does function, with a frameshift efficiency of about 2% with respect to the read-through vector (Table 1). We analyzed mutants (23) in which alterations of the slippery site prevent frameshifting (Table 1). The sixfold decrease in frameshift efficiency observed in these two slippery site mutants, in which a single base is altered in the first or second codon, is similar to that observed for two-base changes in the L1 slippery site (18). In short, the La pol ORF seems to be read by a translational frameshift, just as with L1.

ACKNOWLEDGMENTS

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275 ALRKYVNRNRLYNQFYTAQLLQIMMKPVNCAEGYAWLMHDALVNIPIK 324
 |: .|. |. |. |. |. |. |||.:.: |||. |. |. |. |.:.: |. |.
 264 AITILVSNQAYTDFEAAYLMLAQTLVSPVPRTEASAWFINAGVMNMPT 313
 325 FGSIRGRYPFL 335
 ::: .| . | | |
 314 LSCANGIYAL 324

FIG. 3. The 60-amino-acid region of Cap in La and L1 that is similar (37.7% identity). Vertical lines indicate identity, double dots, similarity. This alignment was generated by GAP (24, 25). The La Cap sequence is the lower and the L1 Cap sequence the upper line.

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