

Brief review

Animal models of pediatric chronic kidney disease. Is adenine intake an appropriate model?

Débora Claramunt^a, Helena Gil-Peña^b, Rocío Fuente^a, Olaya Hernández-Frías^a, Fernando Santos^{a,b,*}

^a University of Oviedo, Spain

^b Hospital Universitario Central de Asturias, Spain

ARTICLE INFO

Article history:

Received 29 January 2015

Accepted 6 August 2015

Available online 29 October 2015

Keywords:

CKD

Growth

Experimental models

Uremia

5/6 nephrectomy

Adenine

ABSTRACT

Pediatric chronic kidney disease (CKD) has peculiar features. In particular, growth impairment is a major clinical manifestation of CKD that debuts in pediatric age because it presents in a large proportion of infants and children with CKD and has a profound impact on the self-esteem and social integration of the stunted patients. Several factors associated with CKD may lead to growth retardation by interfering with the normal physiology of growth plate, the organ where longitudinal growth rate takes place. The study of growth plate is hardly possible in humans and justifies the use of animal models. Young rats made uremic by 5/6 nephrectomy have been widely used as a model to investigate growth retardation in CKD. This article examines the characteristics of this model and analyzes the utilization of CKD induced by high adenine diet as an alternative research protocol.

© 2015 Sociedad Española de Nefrología. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Modelos animales de la enfermedad renal crónica en edad pediátrica. ¿La ingesta de adenina es un modelo apropiado?

RESUMEN

La enfermedad renal crónica (ERC) tiene características específicas. De manera especial, el retraso del crecimiento es una manifestación clínica importante de la ERC que se inicia en la infancia ya que se presenta en un gran número de lactantes y niños con ERC, y repercute profundamente en la autoestima e integración social de los pacientes afectados. Varios factores asociados con la ERC pueden provocar retraso del crecimiento por interferencia con la fisiología normal de la placa de crecimiento, el órgano donde se produce el ritmo de crecimiento longitudinal. Apenas es posible estudiar la placa de crecimiento en seres humanos y ello justifica el uso de modelos animales. El modelo más utilizado para investigar el retraso del

Palabras clave:

ERC

Crecimiento

Modelos experimentales

Uremia

Nefrectomía 5/6

Adenina

* Corresponding author.

E-mail address: fsantos@uniovi.es (F. Santos).

<http://dx.doi.org/10.1016/j.nefro.2015.08.004>

0211-6995/© 2015 Sociedad Española de Nefrología. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

crecimiento en la ERC son ratas jóvenes que se convierten en urémicas por nefrectomía 5/6. Este artículo revisa las características de este modelo y analiza el uso de la ERC inducida por una dieta con elevado contenido de adenina como protocolo de investigación alternativo.

© 2015 Sociedad Española de Nefrología. Publicado por Elsevier España, S.L.U. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Impact of chronic kidney disease

Chronic kidney disease (CKD) is a global public health problem because of its associated adverse health outcomes and high healthcare costs. The Disease Fact Sheet 2014 published by the Centers for Disease Control and Prevention (CDC), estimates that more than 10% of adults in USA, which accounts for more than 20 million people only in this country, have CKD with variable disease seriousness. Diabetes and hypertension are responsible for 7 out of 10 new cases of end stage renal disease (ESRD) in USA. From 1990 to 2010, deaths from CKD raised by about 82% worldwide accounting for the third largest increase among the top 25 causes of death, after acquired immunodeficiency syndrome (39%) and diabetes (93%).¹ Cardiovascular complications are the leading cause of mortality in adults with CKD so that these patients are more prone to die from cardiovascular events than to reach ESRD.

Peculiarities of pediatric chronic kidney disease

As adults represent the vast majority of CKD patients, most publications on CKD focus on adult population. However, CKD that presents at pediatric age, although less prevalent in absolute terms, has important and distinct peculiarities, as briefly commented below.

Demography

Whereas the prevalence of CKD in adults is well known, there are scarce reliable data in children. In North America, 11 cases per million of diagnosed CKD in children between 0 and 19 years have been reported, the prevalence being higher in males and blacks. In the European Union the incidence stands around 11–12 cases per million of the age-related population and the prevalence 56–75 cases per million of the age-related population, according to several national registries.^{2,3}

Causes

The causes of CKD in pediatric population also differ from those of adults. Congenital anomalies of the kidney and urinary tract (CAKUT) are the leading cause and account for approximately 50% of cases. By contrast, acquired glomerulonephritis are the cause of CKD only in 5–14% of children although this percentage is higher in adolescents and ESRD cases.⁴ In European registries, the proportion of CAKUT (58–59%) was slightly higher, while the proportion of glomerulonephritis was lower (5–7%) than in the NAPRTCS database.^{5,6}

Clinical manifestations

The key role of interstitial nephropathies as responsible for pediatric CKD explains that the majority of children who have CKD, and even ESRD, are polyuric or have preserved diuresis, unlike adults. This facilitates the management of these patients but also entails peculiar therapeutic implications because of the poorly regulated loss of water and electrolytes, sodium in particular. It is also of note that CKD is often present since the first months of life and lasts until adulthood.

Although the final height of CKD patients has improved over the last decades,⁷ the North American Pediatric Trials and Collaborative Studies (NAPRTCS) 2008 Annual Report showed that the mean height of 7037 pediatric CKD patients was -1.44 SD score (SDS) and 35% of children had a height below -2 SD.⁴ Similar data shows the Spanish national registry of children with CKD (REPIR II), where the mean height of 605 patients was -1.03 SD and 25% of the children had a height below -1.88 SD.⁸

On the other hand, a variety of neurocognitive deficits occur in children with CKD.⁹ Thus, pediatric CKD has a profound impact on somatic growth, bone metabolism, and neurocognitive development.¹⁰ The CKD-related effects and its long-term sequelae are to a large extent different from those found in adult patients.

Treatment

An adequate metabolic control, optimal nutritional management, appropriate hormonal therapy, intensive dialysis and early renal transplantation are the best remedies to improve growth and neurological development of children with CKD.¹¹

Animal models

Animal models have long been used as a research strategy to increase the knowledge of diseases that affect humans, particularly to better understand pathophysiological mechanisms and to test new therapies. There is wide agreement that the best experimental models for the study of human disease should mimic the entity under consideration in terms of anatomy, physiology, and course of the disease. Thus, useful experimental models should facilitate studies both as the disease evolves and in stable chronic disease. Further, a useful animal model must adhere to current animal welfare regulations and needs to be technically feasible and financially sustainable. Therefore, it should be reproducible, simple, with brief experimental time and easy interpretation of the results. Even if the animal model meets these criteria, it is of note that findings derived from animal models must be taken with

caution at the moment of establishing a translation of the results to clinical practice.¹²

Several types of animal models can be used. Over the last few decades the use of genetically modified mice has progressively increased mostly to figure out the function of a gene or a protein and, in turn, to clarify the pathogenesis and/or the pathophysiological mechanism of a given disease. Non-genetic models utilize infusions of drugs/substances, manipulation of diet or environment, or surgical procedures to induce the target disease. Depending upon the aims of the intended research, such models may be important and necessary. The use of inbred animals, which has been common for over 50 years,¹³ is of special interest to preserve particular traits and to study a hereditary disease in young animals.

Rats and mice are most frequently used as animal models of human disease because these species are easy to handle, relatively cheap and, as a result of their quick life cycle, the effects of manipulations or therapeutic trials can usually be appreciated in a short period of time. Moreover, the rodent animal model is able to reproduce the metabolic disorders and bone disease that patients with CKD develop during the clinical course of the disease.¹⁴⁻¹⁶

Models of CKD

Table 1 shows CKD rat models used to clarify the pathogenesis and pathophysiology of kidney disease and the response to several types of therapeutic strategies. **Table 1** informs on the first publication describing the model, the latest publication based on the model at the time of writing this manuscript, methodology, mechanism by which renal failure is reached and characteristic features.

This manuscript focuses on the 5/6 nephrectomy uremic model, because it has been the most widely used to analyze several aspects of CKD in young rats, particularly those related with longitudinal growth and bone metabolism, and on the model of CKD induced by adenine and described in adult rats, because this model might be useful as animal model of pediatric CKD and presents significant advantages on the subtotal nephrectomy.

The 5/6 nephrectomy model

Subtotal nephrectomy in two stages, usually 4–7 days apart, to induce CKD was first described by Chanutin and Ferris¹⁴ in 1932. In the first stage, the upper and lower poles of the left kidney are functionally suppressed either by surgical scission, by ligation of the renal artery branches or by electrocauterization of renal cortex. In the second stage, the entire right kidney is removed giving rise to the final 5/6 elimination of renal mass, approximately. After the two stages, animals become immediately uremic. According to the original description¹⁴ urea blood concentration decreases to a constant level during the first 10 days followed by a gradual increment when animals initiate polyuria with early dehydration and loss of weight. Posterior investigations showed loss of weight in uremic rats compared to sham operated animals.³³ In 1974, Chantler et al.,¹⁵ used 5/6 nephrectomy as a model of growth

retardation secondary to CKD and described anorexia as a characteristic feature of the experimental CKD so that when glomerular filtration rate drops to below 50% of normal values, uremic rats eat 30% less than control group. This finding justified the need of using sham-operated, normal renal function rats pair fed with the uremic ones to distinguish between the effects of uremia per se from those induced by the associated malnutrition.

The adenine model

The deficit of adenine phosphoribosyltransferase is a rare inherited metabolic disorder that impairs the formation of 2,8-dihydroxyadenine (DHA) in urine.³⁴ Low solubility of DHA results in recurrent urolithiasis and nephropathy secondary to crystal precipitation into renal parenchyma.³⁵ Adenine is more extensively retained in body tissues than the other dietary purines orally consumed (guanine, hypoxantine and xantine).³⁶ High oral administration of adenine is immediately metabolized to DHA, which precipitates and forms crystals in the microvilli and the apical epithelial region of the proximal tubule in just 2 days after the adenine intake.³⁷ In 1982, Yokowaza et al.¹⁶ proposed a new renal failure model based on excessive intake of adenine. This model induces metabolic alterations reproducing CKD characterized by crystalline deposits, foreign body granulomas formation in the renal tubules, and interstitium, and marked fibrosis leading to tubule-interstitial disease, as usually occurs in many pediatric patients with CKD. Moreover, these animals develop hyperphosphatemia, secondary hyperparathyroidism, bone mineral disease, and vascular calcification.³⁸ Recent data indicate that some chemokines (CCR5 and CCL3) play an important role in this CKD model by exacerbating the insult and promoting fibrosis of the tubule-interstitial lesions.³⁹

Different degrees of uremia can be achieved by varying the concentration of adenine in diet. Most publications in adult rats use 0.75% adenine diet to develop this rat model. However, dietary protocols have not been well established, either in terms of the adenine concentration in chow, ranging from 0.075 to 0.75%, or the duration of adenine administration, up to 25 weeks.⁴⁰ Observational periods and therapeutic strategies have also varied across studies.

The rat's gender influences on the response to the adenine protocol. In 2006, Origima et al.⁴¹ concluded that adenine diet concentration needs to be higher in female rats than in male rats to induce the same degree of renal failure. Moreover, they found that male rats fed with adenine diet presented lower levels of testosterone and reduced bone mineral density than those receiving normal chow probably because of a mechanism independent of the severity of renal failure. This could be explained by the fact that adenosine triphosphate and other nucleotides can stimulate directly the formation and resorptive activity of osteoclasts, as well as inhibit bone mineralization by osteoblasts. Thus, high doses of adenine lead to an increase in adenosine, which might have direct effects on the function of osteoblasts and osteoclasts.⁴² There are no differences between both genders with regard to increased blood pressure, ventricular stiffness or cardiac fibrosis.

Table 1 – Models of chronic kidney disease (CKD) developed in rats.^{14,16-32}

Initial description ^a	Latest published reference ^b	Procedure	Mechanism	Distinct features
Lindemann ¹⁷	Ehrich ¹⁸	Heterologous antikidney serum	Nephrotoxin organ specific	-Proper dosage of serum resembles lipid nephrosis in man -Evident renal failure 84–313 days after injection
Schwentker ¹⁹	Peng ²⁰	Injection of bacterial toxins with homologous kidney serum	1. Induced by <i>Streptococcus</i> , <i>Staphylococcus</i> , <i>Escherichia coli</i> or meningococcus toxines. 2. Development of complex antikidney-antibodies. 3. Two types of antibodies: -specific for kidney -non-specific for kidney.	-Clinical and pathological appearance of acute hemorrhagic nephritis or cortical necrosis
Vernier ²¹	Petrovic-Djergovic ²²	Injection of purine	6-Aminonucleoside of puromycin	-Physiological, biochemical, and histological changes similar to those observed in human nephrotic syndrome -Chronic administration with unilateral nephrectomy reproduces focal sclerosis -Mesangiolysis and subsequent hypercellularity -Mesangial proliferative nephritis
Yamamoto ²³	Chen ²⁴	Anti-Thy antibody	Polyclonal or a complement-fixing monoclonal antibody directed against the Thy 1 antigen of mesangial cells.	-Mesangiolysis and subsequent hypercellularity -Mesangial proliferative nephritis
Mackay ²⁵	Muirhead ²⁶	Total bilateral nephrectomy Nephrectomy in one stage Nephrectomy in 2 stages	Nephrectomy in one stage Nephrectomy in 2 stages	-Rapid onset of acute renal failure. Survival about 3–5 days -Two stage surgery with 7–10 days of interval -Survival longer than one stage nephrectomy
Leconte ²⁷	Krim ²⁸	Chemical toxins	Vinylamine, chromates, uranium nitrate.	-Achieve chronic nephritis while coagulated necrosis -Glomerular and tubular changes
Nagle ²⁹	Lopes ³⁰	Chronic unilateral ureteral obstruction	Ureter ligation	-Progressive renal fibrosis -It requires functional suppression of contralateral kidney -Possibility of reversing obstruction -Glomerular sclerosis -Tubule-interstitial nephropathy
Chanutin ¹⁴ Yokowaza ¹⁶	Tóthová ³¹ Yamada ³²	5/6 nephrectomy Oral adenine	Loss of renal mass Crystal deposits in tubular lumen and interstitium	-Glomerular sclerosis -Tubule-interstitial nephropathy

^a First author and year of publication.^b At the time of writing the manuscript.

The adenine model has been used for testing few treatments. Thus, Ali et al.⁴³ demonstrated in a model of CKD caused by adenine the usefulness of Gum acacia (a variety of plant) to treat the anemia of CKD. In a posterior study, and using a 0.75% adenine model, they reported that gum acacia improved histological kidney alterations by reducing adenine deposits and reduced hypertrophy of myocardium.⁴⁴ Tong et al.⁴⁵ demonstrated a slight amelioration of renal function by administration of Chinese herbs or glibenclamide using a protocol of rats fed with adenine.

5/6 nephrectomy vs adenine models

Some reports have shown that CKD induced by adenine intake results in a greater degree and extent of cardiovascular lesions

in the intima-media of the carotid artery than CKD caused by 5/6 nephrectomy, for the same time duration of renal failure.⁴⁶ Likewise, CKD rats receiving adenine develop significant bone mineral alterations earlier than 5/6 nephrectomized rats.⁴⁷ In the adenine model, vascular calcification and left ventricular mass increased progressively with the duration of CKD in association with increased serum fibroblast growth factor 23 (FGF23) and high pulse pressure.⁴⁸

Ferrari et al.⁴⁹ recently reported the first comparative study between the adenine and the subtotal nephrectomy models in a 9 weeks' protocol. Animals were submitted to a 0.75% adenine diet for the first 2 weeks and, subsequently, to a 0.5% adenine diet. The two groups were comparable in terms of circulating concentrations of ionized calcium, phosphate, parathyroid hormone, and FGF23. Despite the similar serum phosphate levels, fractional excretion of phosphorus

Table 2 – Characteristics of rat models of chronic kidney disease CKD induced by subtotal nephrectomy o adenine intake.^{14,16,33,37,38,42,46–49}

	5/6 nephrectomy model	Adenine model
Renal manifestations		
Intervention	Surgery in two stages ¹⁴	Adenine intake ¹⁶
Renal damage	Loss of renal mass ¹⁴	Tubulo-interstitial deposit of 2,8-dihydroxyadenine ¹⁶
Renal failure course	Acute renal failure followed by CKD ¹⁴	Progressive CKD ¹⁶
Biochemical and clinical characteristics associated to CKD	Proteinuria, hypertension and cardiac hypertrophy ⁴⁹	Reversible after the first four weeks ³⁷
Kidney histological findings	Progressive glomerular sclerosis ⁴⁹	Acidosis, polyuria and hyperphosphaturia ⁴⁹
Extrarenal manifestations		
Bone	CKB-BMD ⁴⁹	CKD-BMD. High-turnover bone disease by possible direct effect of adenine ^{38,42}
Heart and vessels	Hypertrophy associated with hypertension ⁴⁹	Extensive calcification of great vessels ^{46,48}
Growth	Growth retardation and reduction of endochondral ossification at growth plate ³³	Unknown
Drawbacks and limitations	Surgery-related mortality ¹⁴ Variability in the degree of renal failure ¹⁴ Irreversibility ¹⁴	Need to adjust the amount of adenine how to give and duration ⁴⁸ Long term studies are difficult because progressive malnutrition and severe CKD ⁴⁷

CKD, chronic kidney disease; CKB-BMD, CKD-bone mineral disorders.

was higher in the adenine group. These animals developed a more severe form of bone disease than rats with subtotal nephrectomy.

It is of note that when the 5/6 nephrectomy is carried out by excision of one kidney and infarction of the two poles of the remaining kidney by arterial ligation, there is a marked increased risk of renovascular hypertension which must be taken into account at the time of assessing the cardiovascular effects of this CKD model.⁵⁰

Table 2 summarizes the main characteristics of 5/6 nephrectomy and adenine models of CKD.

Adenine model useful to study pediatric CKD

The adenine model of CKD is easy to do and reproducible. It does not require invasive interventions and results in tubule-interstitial lesions similar to those of CKD that debuts in infancy and early-childhood. It is associated with the 100% of animal survival when the adenine diet concentration is adjusted and the induced CKD may be reversible at not very high adenine doses. In spite of these characteristics, there are no published studies of young rats made uremic by adenine administration.

We propose that administration of adenine to prepuberal rats might be a suitable procedure for the study of CKD in young individuals. Further studies are needed to characterize this model in young rats.

Funding

This study has been supported partially by Fondo de Investigación Sanitaria, Instituto de Salud Carlos III (PI12/00987 y PI15/02122), National Plan 2013–2016, Fondos FEDER, University of Oviedo and Fundación Nutrición y Crecimiento.

Conflict of interest

The authors declare no conflict of interest.

REFERENCES

- Lozano R, Naghavi M, Foreman K, Lim S, Shibuya K, et al. Global and regional mortality from 235 causes of death for 20 age groups in 1990 and 2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet*. 2012;380(9859):2095–128.
- Coresh J, Byrd-Holt D, Astor BC, Briggs JP, Eggers PW, et al. Chronic kidney disease awareness, prevalence, and trends among U.S. adults, 1999 to 2000. *J Am Soc Nephrol*. 2005;16(1):180–8.
- Harambat J, Van Stralen KJ, Kim JJ, Tizard EJ. Epidemiology of chronic kidney disease in children. *Pediatr Nephrol*. 2012;27(3):363–73.
- NAPRTCS. NAPRTCS 2008 Annual Report; 2008. Available at: <http://spitfire.emmes.com/study/ped/index.htm> (accessed 25.11.14).
- Ardissino G, Daccò V, Testa S, Bonaudo R, Claris-Appiani AT, et al. Epidemiology of chronic renal failure in children: data from the ItalKid project. *Pediatrics*. 2003;111:e382–7.
- Mong Hiep TT, Ismaili K, Collart F, Van Damme-Lombaerts R, Godefroid N, et al. Clinical characteristics and outcomes of children with stage 3–5 chronic kidney disease. *Pediatr Nephrol*. 2010;25:935–40.
- Ingulli EG, Mak RH. Growth in children with chronic kidney disease: role of nutrition growth hormone, dialysis, and steroids. *Curr Opin Pediatr*. 2014;26(2):187–92.
- Areses Trapote R, Sanahuja Ibáñez MJ, Navarro M, Investigadores Centros Participantes en el REPIR II. Epidemiology of chronic kidney disease in Spanish pediatric population. REPIR II Project. *Nefrologia*. 2010;30(5):508–17.
- Rotundo A, Nevins TE, Lipton M, Lockman LA, Mauer SM, et al. Progressive encephalopathy in children with chronic renal insufficiency in infancy. *Kidney Int*. 1982;21(3):486–91.

10. Greenbaum LA, Warady BA, Furth SL. Current advances in chronic kidney disease in children: growth, cardiovascular, and neurocognitive risk factors. *Semin Nephrol.* 2009;29(4):425-34.
11. Smith JM, Stablein DM, Munoz R, Hebert D, McDonald RA. Contributions of the transplant registry: the 2006 annual report of the North American Pediatric Renal Trials and Collaborative Studies (NAPRTCS). *Pediatr Transpl.* 2007;11(4):366-73.
12. Merino J, Merino R. Contribution of animal models to the study and treatment of systemic autoimmune disease. *Reumatol Clin.* 2008;4(1):5-10.
13. Young FG. Growth and diabetes in normal animals treated with pituitary (anterior lobe) diabetogenic extract. *Biochem J.* 1945;39(5):515-36.
14. Chanutin A, Ferris EB. Experimental renal insufficiency produced by partial nephrectomy. *Arch Intern Med.* 1932;49:767-87.
15. Chantler C, Lieberman E, Holliday MA. A rat model for the study of growth failure in uremia. *Pediatr Res.* 1974;8:109-13.
16. Yokozawa T, Oura T, Okada T. Metabolic effects of dietary purine in rats. *J Nutr Sci Vitaminol.* 1982;28:519-26.
17. Lindemann W. Ann Inst Pasteur. 1900;14:49.
18. Ehrich WE, Forman CW, Seifer J. Diffuse glomerular nephritis and lipid nephrosis: correlation of clinical, morphological, and experimental observations. *AMA Arch Pathol.* 1952;54(5):463-503.
19. Schwentker FF, Comploier FC. The production of kidney antibodies by injection of homologous kidney plus bacterial toxins. *J Exp Med.* 1939;70(3):223-30.
20. Peng W, Liu ZR. Comparison of two rat models of IgA nephropathy. *Nan Fang Yi Ke Da Xue Xue Bao.* 2008;28(10):1842-5.
21. Vernier RL, Papermaster BW, Good RA. Aminonucleoside nephrosis. I. Electron microscopic study of the renal lesion in rats. *J Exp Med.* 1959;109(1):115-26.
22. Petrovic-Djergovic D, Popovic M, Chittiprol S, Cortado H, Ransom RF, et al. CXCL10 induces the recruitment of monocyte derived macrophages into kidney, which aggravate puromycin aminonucleoside nephrosis. *Clin Exp Immunol.* 2015;180(2):305-15.
23. Yamamoto T, Wilson CB. Quantitative and qualitative studies of antibody-induced mesangial cell damage in the rat. *Kidney Int.* 1987;32:514-25.
24. Chen D, Li Y, Mei Y, Geng W, Yang J, et al. miR-34a regulates mesangial cell proliferation via the PDGFR- β /Ras-MAPK signaling pathway. *Cell Mol Life Sci.* 2014;71(20):4027-42.
25. Mackay EM, Mackay LL. Enlargement of the adrenal cortex in experimental uremia. *J Exp Med.* 1927;46(3):429-42.
26. Muirhead EE, Stirman JA, Jones F. Further observations on the potentiation of postnephrectomy hypertension of the dog by dietary protein. *Circ Res.* 1959;7(1):68-78.
27. Leconte C. Gaz Méd. 1854;ix:488.
28. Krim M, Messaadia A, Maida I, Aouacheri O, Saka S. Protective effect of ginger against toxicity induced by chromate in rats. *Ann Biol Clin.* 2013;71(2):165-73.
29. Nagle RB, Bulger RE. Unilateral obstructive nephropathy in the rabbit. II. Late morphologic changes. *Lab Invest.* 1978;38(3):270-8.
30. Lopes RI, Dénes FT, Bartolamei MG, Reis S, Sanches TR, et al. Serum and urinary values of CA 19-9 and TGF β 1 in a rat model of partial or complete ureteral obstruction. *Eur J Pediatr Surg.* 2015, <http://dx.doi.org/10.1055/s-0034-1395263>.
31. Tóthová L, Bábičková J, Borbelyová V, Filová B, Sebeková K, et al. Chronic renal insufficiency does not induce behavioral and cognitive alteration in rats. *Physiol Behav.* 2015;138:133-40.
32. Yamada S, Tatsumoto N, Tokumoto M, Noguchi H, Ooboshi H, et al. Phosphate binders prevent phosphate-induced cellular senescence of vascular smooth muscle cells and vascular calcification in a modified adenine-based uremic rat model. *Calcif Tissue Int.* 2015;96(4):347-58.
33. Souhami RL, Smith J, Bradfield JW. The effect of uremia on organ graft survival in the rat. *Br J Exp Path.* 1973;54:183-91.
34. Kelley WN, Levy RI, Rosenblom FM, Henderson JE, Seegmiller JE. Adenine phosphoribosyltransferase deficiency: a previously undescribed genetic defect in man. *J Clin Invest.* 1968;47(10):2281-9.
35. Cartier P, Hamet M, Hamburger J. A new metabolic disease: the complete deficit of adenine phosphoribosyltransferase and lithiasis of 2,8-dihydroxyadenine. *C R Acad Sci Hebd Séances Acad Sci D.* 1974;279(10):883-6.
36. Savaiano DA, Clifford AJ. Absorption tissue incorporation and excretion of free purine bases in the rat. *Nutr Rep Int.* 1978;170:551-6.
37. Koeda T, Wakaki K, Koizumi F, Yokozawa T, Oura H. Early changes of proximal tubules in the kidney of adenine-ingesting rats, with special reference to biochemical and electron microscopic studies. *Nippon Jinzo Gakkai Shi.* 1988;30:239-46.
38. Katsumata K, Kusano K, Hirata M, Tsunemi K, Nagano N, et al. Sevelamer hydrochloride prevents ectopic calcification and renal osteodystrophy in chronic renal failure rats. *Kidney Int.* 2003;64:441-50.
39. Correa-Costa M, Braga TT, Felizardo RJ, Andrade-Oliveira V, Perez KR, et al. Macrophage trafficking as key mediator of adenine-induced kidney injury. *Mediat Inflamm.* 2014;291024.
40. Damment S, Secker R, Shen V, Lorenzo V, Rodriguez M. Long-term treatment with lanthanum carbonate reduces mineral and bone abnormalities in rats with chronic renal failure. *Nephrol Dial Transpl.* 2011;26(6):1803-12.
41. Origima T, Tano K, Kanehara M, Gao M, Wang X, et al. Sex difference of adenine effects in rats: renal function, bone mineral density and sex steroidogenesis. *Endocr J.* 2006;53(3):407-13.
42. Orris I, Burnstock G, Arnett T. Purinergic signaling and bone remodeling. *Curr Opin Pharmacol.* 2010;10(3):322-30.
43. Ali BH, Al Za'abi M, Ramkumar A, Yasin J, Nemmar A. Anemia in adenine-induced chronic renal failure and the influence of treatment with gum acacia thereon. *Physiol Res.* 2014;63(3):351-8.
44. Ali BH, Inuwa I, Al Za'abi M, Al Bahlani S, Al Issaei H, et al. Renal and myocardial histopathology and morphometry in rats with adenine-induced chronic renal failure: influence of gum acacia. *Cell Physiol Biochem.* 2014;34(3):818-28.
45. Tong Y, Han B, Guo H, Liu Y. Protection of Chinese herbs against adenine-induced chronic renal. *Afr J Tradit Complement Altern Med.* 2010;7(4):331-8.
46. Price PA, Roublack AM, Williamson MK. Artery calcification in uremic rats is increased by a low protein diet and prevented by treatment with ibandronate. *Kidney Int.* 2006;70:1577-83.
47. Tamagaki K, Yuan Q, Ohkawa H, Imazeki I, Moriguchi Y, et al. Severe hyperparathyroidism with bone abnormalities and metastatic calcification in rats with adenine-induced uremia. *Nephrol Dial Transpl.* 2006;21:651-9.
48. Shobeiri N, Panga J, Adams MA, Holden RM. Cardiovascular disease in an adenine-induced model of chronic kidney disease: the temporal link between vascular calcification and haemodynamic consequences. *J Hypertens.* 2013;31:160-8.
49. Ferrari GO, Ferreira JC, Cavallari RT, Neves KR, dos Reis LM, et al. Mineral bone disorder in chronic kidney disease: head-to-head comparison of the 5/6 nephrectomy and adenine models. *BMC Nephrol.* 2014;15:69.
50. Nguy L, Nilsson H, Lundgren J, Johansson ME, Teerlink T, et al. Vascular function in rats with adenine-induced chronic renal failure. *Am J Physiol Regul Integr Comp Physiol.* 2012;302:R1426-35.