

Eur J Vasc Endovasc Surg 23, 3–10 (2002)

doi:10.1053/ejvs.2001.1526, available online at <http://www.idealibrary.com> on IDEAL®

REVIEW ARTICLE

Homocysteine in Myointimal Hyperplasia

M. Hansrani¹, J. I. Gillespie² and G. Stansby^{*3}

Departments of ¹Surgery, ²Human Physiology and ³Vascular Surgery, University of Newcastle upon Tyne

Introduction: homocysteine, a sulphur-containing non-essential amino acid, appears to play a role in the pathophysiology of atherosclerosis. However, its role in myointimal hyperplasia, the cause of almost 30% of failures of interventional therapeutic procedures, is much less clear.

Methods: a review of the published scientific data concerning the role of homocysteine in myointimal hyperplasia was performed using MEDLINE and other on-line databases. Evidence was sought from cell culture experiments, animal models and clinical studies.

Results: several clinical studies have recently been published linking plasma homocysteine levels to restenosis in coronary and peripheral arterial disease. However, several contradictory studies also exist making the role of homocysteine unclear. There are currently no published randomised trials. Cell culture and animal model experiments have elucidated several potential mechanisms by which may stimulate myointimal hyperplasia. Possible mechanisms include endothelial cell activation with the enhanced release of inflammatory cytokines and growth factors and a direct effect on vascular smooth muscle cell migration and proliferation.

Conclusion: further studies are required before the true role of homocysteine in the pathogenesis of myointimal hyperplasia can be clearly evaluated. If evidence does confirm a role, the ease with which homocysteine levels can be normalised makes it an attractive alternative therapeutic target for intervention.

Key Words: Homocysteine; Myointimal hyperplasia; Restenosis; Review.

Introduction

Balloon angioplasty and surgical bypass techniques have revolutionised the management of patients with coronary and peripheral vascular disease. However, approximately a third of these patients will develop restenosis^{1,2} mainly due to the development of myointimal hyperplasia (MIH). Recently the non-essential amino acid homocysteine (HCy) has been implicated as a possible risk factor in premature graft and angioplasty restenosis.^{3–5} This has generated particular interest as plasma levels of HCy can be normalised simply and cheaply by the intake of folic acid and the vitamins B₆ and B₁₂.^{6,7} However, the precise role of HCy in the development of MIH remains unclear and will be the subject of this review.

Myointimal Hyperplasia

MIH can be considered an exaggerated healing response to injury. Activated endothelial cells initiate the inflammatory response via platelet activation, or via the direct release of cytokines and growth hormones. This leads to a cascade of effects which result in: migration of vascular smooth muscle cells (VSMC) from the media into the intima,^{8,9} VSMC proliferation and phenotypic change to a more secretory fibroblastic cell type¹⁰ and continued production of extracellular matrix (ECM), in particular collagen, in excess of degradation,¹¹ which eventually leads to stenosis of the increasingly non-compliant vessel wall. Continued trauma from the presence of sutures; shear stresses;^{12,13} or low flow of blood in areas of turbulence perhaps leading to a local concentration of harmful irritants in the blood, may then act to maintain the endothelium in an activated state. This leads to continuing platelet and macrophage activation even after the endothelial monolayer has regenerated.

* Please address all correspondence to: G. Stansby, Department of Surgery, University of Newcastle upon Tyne, Framlington Place, Newcastle upon Tyne, NE2 4HH.

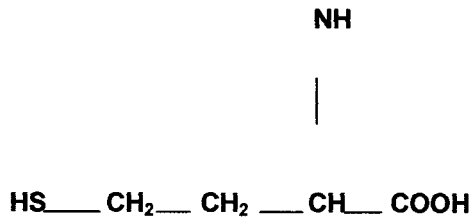


Fig. 1. Structure of homocysteine.

Homocysteine

HCy is a non-essential, non-protein forming, sulphur-containing amino acid, produced from the metabolism of dietary methionine (Fig. 1). The metabolic pathways involving HCy are illustrated in Figure 2. It is found in the serum as protein-bound HCy (70%), HCy-HCy and HCy-Cysteine mixed disulphides (30%) or as free HCy thiolactone (1%). Elevations of plasma total HCy have a multi-factorial basis, resulting from a combination of increased dietary intake of methionine, inheritable genetic defects and vitamin deficiencies (Fig. 2).¹⁴ Severe homocysteinaemia (>100 µmol/l) results from rare homozygous genetic defects such as deficiency of cystathionine β synthase (CβS), methylenetetrahydrofolate reductase (MTHFR) or of enzymes involved in methyl-B₁₂ synthesis and homocysteine methylation. Mild (15–30 µmol/l) to moderate (30–100 µmol/l) homocysteinaemia is due to vitamin deficiencies, heterozygous CβS defect, or MTHFR thermolability. These later genetic defects have been found to affect a third of some populations, whilst vitamin deficiencies play a large role in the elevated levels of HCy found in the elderly population.^{15–17}

Free HCy and HCy-th are highly reactive molecules, forming non-covalent and covalent bonds with a variety of molecules, in particular cysteine-rich or other thiol moieties.^{18,19} This can lead to loss of electrical charge, conformational changes or even precipitation of the protein, resulting in loss or degradation of the biological function of multiple enzymes, receptors, growth factors and structural proteins.^{20,21} HCy is also known to activate and inactivate specific mRNA transcription and stimulate DNA synthesis directly perhaps via a specific receptor, which initiates calcium fluxes in an as yet unclear mechanism.^{22,23}

The role of the endothelial cell (EC) is central to the aetiology of MIH.²⁴ Endothelial cells have a lower basal intracellular HCy concentration and appear to be more influenced by extracellular concentrations of the amino-acid, suggesting an increased sensitivity/decreased ability to metabolise it than other cells.²⁵

At physiological concentrations HCy is able to inhibit EC cell proliferation at the same time as stimulating VSMC proliferation.^{26,27} Although, studies have also shown increased EC turnover and aging in response to elevated plasma HCy.^{24,28} HCy activates and damages endothelial cells by the generation of reactive oxygen species²⁹ combined with the removal of EC protective antioxidant mechanisms such as nitric oxide (NO)³⁰ and glutathione³¹ thus potentiating the injurious effect and increasing activation. In VSMC (see Figure 3) HCy stimulates proliferation through a variety of means probably via a specific NMDA-like receptor(s).³² These initiate several cellular cascades:

- The induction of cyclin D1 and A in a dose-dependant manner stimulates the re-entry of the normally quiescent cells into the cell cycle²⁶ via cyclin-dependant kinase,³³
- Activation of phospholipases such as C and D, the synthesis of diacylglycerol (DAG) and inositol triphosphate (IP3) leads to an increase in intracellular calcium ions and activation of protein kinase C (PKC). Both of these intermediaries are capable of activating mitogen activating protein (MAP) kinase which leads to activation of DNA transcription factors and cellular proliferation,^{22,32}
- Redox-specific and dose-dependant PKC activation leads to transcription of c-myb and c-fos mRNA which code for proteins that combine with others to form AP-1 which in turn initiates transcription of genes involved in cellular proliferation,³⁴
- Potentiation of the mitogenic effect of angiotensin II³⁵ and possibly other growth factors.

HCy directly inhibits the production and extracellular release of specific proteins. This stimulates a decrease in the release of heparin-like compounds, which are potent inhibitors of VSMC proliferation,³⁶ as well as producing a highly procoagulant milieu through the modulation of elements of the coagulation and fibrinolytic pathways.^{37–40} The coagulation/fibrinolytic pathways are further modulated through chemical interactions with other cysteine moieties present in the protein structure of many proteins involved in these pathways. HCy is in addition able to modulate the activity of matrix metalloproteinases (MMPs) and their inhibitors (TIMPs) leading to elastin degradation and accumulation of collagen within the intima and media in a dose-dependant manner.^{41,42}

Recent studies have shown that, *in vitro* and *in vivo*, HCy has a direct effect on vasodilatation selectively inhibiting endothelium-dependant vasodilatation, predominantly through inhibition of NO.^{43,44}

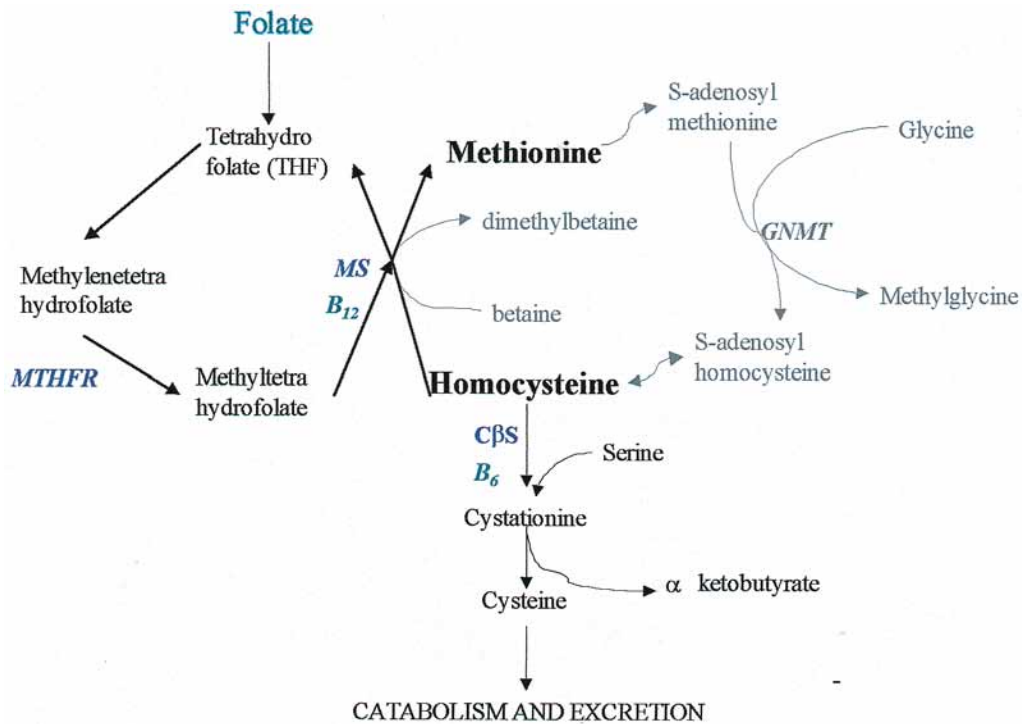


Fig. 2. Homocysteine metabolism in man. Key: MS=methionine synthase, MTHFR=methylene tetrahydrofolate reductase, CBS=cystathionine β synthase, GNMT=glycine N-methyltransferase.

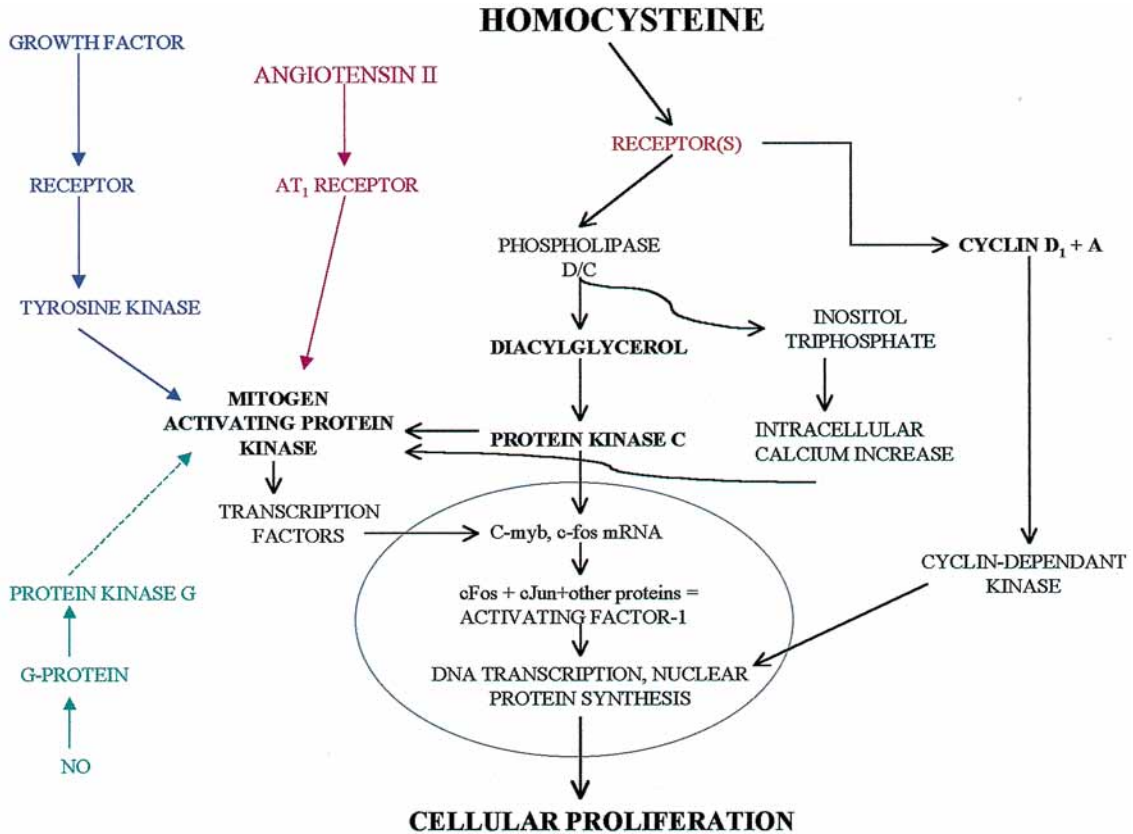


Fig. 3. Mechanisms by which cellular proliferation may be induced by homocysteine in vascular smooth cells.

Homocysteine and Clinical Restenosis

An association between elevated blood levels of homocysteine (HCy) and cardiovascular disease was originally postulated in 1969.⁴⁵ Thirty percent of patients presenting to vascular clinics have elevated plasma HCy, compared to 2–5% in the normal population.⁴⁶ There has been a deluge of studies assessing HCy as a risk factor for cardiovascular disease in the last 10 years. Boushey *et al.*⁴⁷ published a meta-analysis of 27 studies to 1995, looking at almost 4000 patients. They found that HCy was an independent, graded risk factor for atherosclerotic disease in coronary, cerebral and peripheral vessels. They calculated the odds ratio of a 5 $\mu\text{mol/l}$ increment of plasma HCy level for coronary artery disease (CAD) of 1.6, cerebrovascular disease (CVA) of 1.5 and for peripheral vascular disease (PVD) of 6.8 although this value is likely to be inaccurate due to the small number of trials involving PVD available for analysis. These results have been supported by a further 40 subsequent studies⁴⁸ and will not be considered further here. The evidence for the role of HCy in premature restenosis post therapeutic revascularisation by vein graft or angioplasty is, in comparison, much less clear. However, studies do exist in both coronary and peripheral vascular disease and will be reviewed. There are currently no published studies of the association between plasma HCy and restenosis in cerebrovascular disease.

Homocysteine and coronary restenosis

Iwama⁴ in a study of 40 patients who had undergone saphenous vein coronary artery bypass grafting (CABG) 1–13 years (mean 6.1 ± 3.1 years) previously, found that patients with MIH ($n=23$) in any graft (defined as angiographic stenosis $\geq 50\%$) had significantly elevated plasma HCy levels (median 15.1 vs 10.6 $\mu\text{mol/l}$) compared to those with no evidence of restenosis. Using multiple regression analysis they were able to show that elevated HCy appeared to be an independent risk factor for vein graft stenosis. However, this retrospective study may be misleading as it supposes that levels of HCy remain relatively unchanged over several years, which is as yet unestablished. Eritsland,⁴⁹ in a prospective cohort study of 565 patients undergoing CABG in Norway found no such association between the preoperative HCy levels and the frequency of graft occlusions (determined by angiography) at 1-year. Unfortunately they did not exclude stenoses occurring within the first 6 weeks, which are usually due to thrombosis

and unlikely to be due to MIH. Benoit⁵⁰ in a prospective trial of 222 patients undergoing coronary angioplasty in France who were followed up clinically for 6 months, found no significant differences in homocysteine levels between patients with multiple restenosis ($n=79$; diagnosed by coronary angiography in 55 cases, by myocardial scintigraphy in 23 cases and strongly suspected clinically in one patient) or those requiring revascularisation, and those without restenosis and not requiring revascularisation. Morita *et al.*⁵¹ consecutively enrolled 112 male patients who had undergone a successful elective coronary artery angioplasty and measured plasma HCy at follow-up angiography (3–6 months). They found that HCy levels were significantly higher (15.0 vs 13.0 $\mu\text{mol/l}$) in patients with restenosis (final stenosis $>50\%$ more than 3 months post angioplasty). Finally, HCy was found to be significantly higher (23.6 ± 7.8 versus 16.9 ± 7.1 $\mu\text{mol/l}$) in a case-control study of heart transplant patients who developed graft vasculopathy, defined as coronary stenosis $>25\%$ or aneurysms, as opposed to those who did not.⁵² Blood was sampled a mean of 52 weeks post transplant.

Homocysteine and restenosis in peripheral vascular disease

Irvine³ in a retrospective case-control study of 19 patients (median age 70 years at intervention) who had undergone infrainguinal vein bypass grafting and had developed stenosis between 6 weeks and 1 year post-operatively (defined as angiographically determined lumen reduction of $>50\%$ or with a persistent duplex abnormality (minimum of 2 scans) with a doubling of velocities) matched against similar patients ($n=19$) who had not, found plasma HCy to be significantly elevated in those who later developed stenoses (median 17.8 $\mu\text{mol/l}$ vs 13.8 $\mu\text{mol/l}$). Mireskandari⁵ performed a similar study prospectively in 62 limbs (median age 72 years) undergoing femorodistal vein bypass. They confirmed that elevated plasma HCy was a risk factor for restenosis at 12 months (median 14.4 $\mu\text{mol/l}$, range 9.6–32 vs 9.5 $\mu\text{mol/l}$). However, they do not state whether or not they excluded stenoses occurring within the first 6 weeks. Interestingly, Beattie *et al.*⁵³ found hyperhomocysteinaemia (>12.2 $\mu\text{mol/l}$ in women and >15.5 $\mu\text{mol/l}$ in men) in 57% of 57 patients undergoing infrainguinal vein bypass surgery. Nonetheless they found that plasma HCy levels were not associated with restenosis/outcome directly in these patients, but that it was significantly associated with the development of pre-existing MIH in the vein and this,

in turn, was associated with an increased risk of graft failure. Tsakiris *et al.*⁵⁴ also found no correlation between HCy levels and rates of restenosis in 71 patients (age 68 ± 13 years) undergoing percutaneous angioplasty of predominantly femoropopliteal segments of artery in patients with PVD (52% of whom had elevated HCy levels). Currie⁵⁵ reviewed 66 patients selected retrospectively and correlated HCy levels with failure of vascular intervention, which they defined as, return of symptoms with a fall in ankle-brachial pressure index, requirement for further revascularisation procedure in the same limb or amputation. Using multiple logistic regression they found hyperhomocysteinaemia to be an independent risk factor for the failure of all forms of peripheral vascular intervention, including angioplasty, endarterectomy and bypass grafting, along with young age at intervention and diabetes.

Unfortunately, these studies do not as yet give us a clear picture of the strength of the link between elevated HCy levels and MIH. Differences in the timing of blood taken for HCy levels, the length of time to follow-up, differences in outcomes assessed, possible publication biases and the relatively small number of studies overall mean no definitive conclusions can be reached.

Restenosis in animal models of hyperhomocysteinaemia

A number of mammals have been used as models for the effects of elevated HCy in man. Baboons continuously infused with HCy developed sustained aortic EC loss and regeneration, platelet consumption and intimal lesion formation typical of arteriosclerotic or preatherosclerotic intimal lesions composed of proliferating VSMC surrounded by collagen, elastin fibres, glycosaminoglycans and sometimes lipid.²⁴ Similar histological changes have been reproduced in rats,⁵⁶ minipigs⁵⁷ and rabbits⁵⁸ fed methionine-rich diets. Matthias further demonstrated the potentiation of HCy-induced damage by other atherogenic substances such as cholesterol, angiotensin II, cholestane, cholic acid and methylthiouracil in spontaneously hypertensive rats. Young⁵⁹ found an increase in malondialdehyde, a major secondary oxidation product of polyunsaturated lipids, in nitrous oxide-induced hyperhomocysteinaemic pigs, whilst increased markers of lipid peroxidation were also noted in hyperhomocysteinaemic rabbits.⁵⁸ However, plasma HCy levels were far in excess of that usually found in hyperhomocysteinaemic patients and in fact, *in vivo* lipid peroxidation has not been demonstrated in man.⁶⁰

Table 1. Therapeutic options in hyperhomocysteinaemia.

-
- Lifestyle changes
 - Folic acid
 - Pyroxidine (B₆)
 - Vitamin B₁₂
 - Betaine
 - Vitamin C/Vitamin E
-

Monkeys fed an "atherogenic diet" developed impaired EC-dependent vasodilatation in resistance vessels of the lower limb⁴⁴ and defects in the protein C anticoagulant pathway.⁶¹ These atherosclerotic hyperhomocysteinaemic monkeys when fed vitamin B supplemented atherosclerotic diet for 6 months demonstrated a significant fall in plasma HCy levels (12.8 ± 2.8 to 3.5 ± 0.3 $\mu\text{mol/l}$), though this did not normalise vascular function or prevent progression of structural lesions. This may be due to the fact that the elevation in HCy caused by the diet was small, however, it is more likely that this lack of effect was due to the persistent hypercholesterolaemia induced by the atherosclerotic diet, which was not affected by the vitamin supplementation. Zulli *et al.*⁶² demonstrated a synergistic effect of the combination of diet-induced hyperhomocysteinaemia and hypercholesterolaemia in rats as measured by aortic wall thickness and disintegration of the elastic lamina when compared against the effects of feeding with each agent alone.

Hyperhomocysteinaemia also induced procoagulant effects,^{63,64} increased prostaglandin synthesis,⁶⁵ increased diapedesis through mesenteric post capillary venules⁶⁶ and increased cyclin-dependant kinase in rats.³³

Although these studies demonstrate that HCy can initiate or maintain endothelial and vessel wall injury, there is at present only one study looking specifically at MIH in animals: Southern *et al.*⁶⁷ studied the effects of HCy on restenosis in carotid endarterectomised rats. They found a 4-fold increase in MIH in the diet-induced hyperhomocysteinaemic group (36.32 ± 15.28 $\mu\text{mol/l}$) when calculated as percent lumen stenosis and a 8-fold increase in MIH area when compared to normals. Using a linear regression model to compare HCy levels and percent stenosis they were able to demonstrate a linear relationship between the two.

Therapeutic Options in Hyperhomocysteinaemia (Table 1)

To date, therapeutic attempts at preventing MIH have been unsatisfactory. Homocysteine provides a particularly attractive therapeutic target as blood levels

can be easily normalised with the administration of cheap, patient-acceptable, oral vitamin treatment. The implementation of a "cardiovascular lifestyle" is particularly apt, with decreased red meat intake, increased fruit and fibre, low coffee consumption, smoking cessation and regular exercise which have all been shown to decrease plasma HCy levels.⁶⁸ Both Brattstrom *et al.*⁶ and Clarke *et al.*⁶⁹ in reviews of randomised controlled trials of the effects of folic acid \pm pyridoxine and B₁₂ administration on lowering plasma HCy levels, found that combined vitamin treatment could significantly reduce plasma HCy levels. Among these vitamins folic acid had the most potent effect: the higher the pretreatment plasma HCy and the lower the pretreatment plasma folic acid level the greater the effect. *In vitro* and *in vivo* studies have demonstrated empirical reduction in HCy-induced EC damage, VSMC proliferation and inhibition of vasodilatation when any of these vitamins were added.⁷⁰⁻⁷² There are currently no defined guidelines as to the recommended doses of these vitamins, or the level of HCy at which treatment should be initiated.^{6,68} Wilcken *et al.*⁷³ have also demonstrated that betaine therapy (6g per day) can reduce HCy levels to normal in patients with homocystinuria and were further able to show that normalising HCy levels led to a significant decrease in vascular events in these patients.⁷⁴ Vitamin C and vitamin E, natural antioxidants, appear to ameliorate the oxidative damage caused by hyperhomocysteinaemia and may provide another simple mode of treatment.⁷⁵

Vermeulen *et al.*⁷⁶ in a small, short-term trial of vitamin therapy, could only show an improvement in coronary artery risk, as assessed by exercise electrocardiography tests, with HCy lowering treatment. More recently, his group has reported results from a large prospective trial of hyperhomocysteinaemic patients presenting with premature PVD or cerebrovascular disease (before the age of 56 years). When treated with folic acid and B₆, these patients demonstrated a similar risk of future cardiovascular events as normohomocysteinaemic controls.^{77,78}

The results from large-scale, long-term randomised controlled trials of such regimens in people at high risk from cardiovascular events and those undergoing therapeutic intervention are currently awaited.

Conclusions

Restenosis is a common and serious problem encountered after arterial bypass or endoluminal treatment of atherosclerotic arterial disease and any

effective treatment could potentially have wide applications in both coronary and peripheral vascular disease. HCy, if proven to be a true primary risk factor, would be an attractive target for therapy since it is both easily measured and easily treated. To date the majority of epidemiological studies have confirmed that elevated levels of HCy are associated with PVD, though the association with the development of MIH is still tenuous. Animal and *in vitro* studies have demonstrated potential mechanisms by which HCy could produce endothelial injury, smooth muscle cell activation and other aspects of the pathophysiology of MIH, and support the hypothesis that it is an important risk factor for MIH. Though few prospective studies and no randomised trials of HCy lowering therapy have yet been published. Such studies are needed before therapy to lower HCy can be widely recommended.

References

- 1 TAYLOR LM, JR, DEFRANG RD, HARRIS EJ, JR, PORTER JM. The association of elevated plasma homocyst(e)ine with progression of symptomatic peripheral arterial disease. *J Vasc Surg* 1991; **13**: 128-136.
- 2 WHITTEMORE AD, CLOWES AW, COUCH NP, MANNICK JA. Secondary femoropopliteal reconstruction. *Ann Surg* 1981; **193**: 35-42.
- 3 IRVINE C, WILSON YG, CURRIE IC *et al.* Hyperhomocysteinaemia is a risk factor for vein graft stenosis. *Eur J Vasc Endovasc Surg* 1996; **12**: 304-309.
- 4 IWAMA Y, MOKUNO H, YOKOI H *et al.* Elevated levels of plasma homocysteine related to saphenous vein graft disease after coronary artery bypass graft surgery. *J Cardiol* 1998; **32**: 357-362.
- 5 MIRESKANDARI M, SCHACHTER M, TIMMS IGO, SEVER P, WOLFE JHN. Plasma homocysteine is an independent risk factor for vein graft stenosis. *Br J Surg* 1998; **85**: 27.
- 6 BRATTSTROM L. Vitamins as homocysteine-lowering agents. *J Nutr* 1996; **126**: 1276S-1280S.
- 7 FRANKEN DG, BOERS GH, BLOM HJ, TRIJBELS FJ, KLOPPENBORG PW. Treatment of mild hyperhomocysteinemia in vascular disease patients. *Arterioscler Thromb* 1994; **14**: 465-470.
- 8 BERGUER R, HIGGINS RF, REDDY DJ. Intimal hyperplasia. An experimental study. *Arch Surg* 1980; **115**: 332-335.
- 9 DILLEY RJ, MCGEACHIE JK, PRENDERGAST FJ. A review of the histologic changes in vein-to-artery grafts, with particular reference to intimal hyperplasia. *Arch Surg* 1988; **123**: 691-696.
- 10 ROSS R. The pathogenesis of atherosclerosis: a perspective for the 1990s. *Nature* 1993; **362**: 801-809.
- 11 TYAGI SC, MEYER L, SCHMALTZ RA, REDDY HK, VOELKER DJ. Proteinases and restenosis in the human coronary artery: extracellular matrix production exceeds the expression of proteolytic activity. *Atherosclerosis* 1995; **116**: 43-57.
- 12 SZILAGYI DE, ELLIOTT JP, HAGEMAN JH, SMITH RF, DALL'OLMO CA. Biological fate of autogenous vein implants as arterial substitutes. *Ann Surg* 1973; **178**: 232-244.
- 13 CAMPBELL PA, MCGEACHIE JK, PRENDERGAST FJ. Vein grafts for arterial repair: their success and reasons for failure. *Ann R Coll Surg Engl* 1981; **63**: 257-260.
- 14 MALINOW MR. Homocyst(e)ine, vitamins and genetic interactions in vascular disease. *Can J Cardiol* 1999; **15**: 31B-34B.

- 15 ROSENBERG IH. Homocysteine, vitamins and arterial occlusive disease: an overview. *J Nutr* 1996; **126**: 1235S–1237S.
- 16 SELHUB J, JACQUES PF, WILSON PW, RUSH D, ROSENBERG IH. Vitamin status and intake as primary determinants of homocysteinemia in an elderly population [see comments]. *JAMA* 1993; **270**: 2693–2698.
- 17 SELHUB J. Homocysteine metabolism. *Annu Rev Nutr* 1999; **19**: 217–246.
- 18 JAKUBOWSKI H. Protein homocysteinylation: possible mechanism underlying pathological consequences of elevated homocysteine levels. *FASEB J* 1999; **13**: 2277–2283.
- 19 JAKUBOWSKI H. Homocysteine thiolactone: Metabolic origin and protein homocysteinylation in humans. *J Nutr* 2000; **130**: 377S–381S.
- 20 KRUMDIECK CL, PRINCE CW. Mechanisms of homocysteine toxicity on connective tissues: implications for the morbidity of aging. *J Nutr* 2000; **130**: 365S–368S.
- 21 GAO XM, WORDSWORTH P, McMICHAEL AJ, KYAW MM, SEIFERT M, REES D *et al*. Homocysteine modification of HLA antigens and its immunological consequences. *Eur J Immunol* 1996; **26**: 1443–1450.
- 22 DALTON ML, GADSON PF, JR, WRENN RW, ROSENQUIST TH. Homocysteine signal cascade: production of phospholipids, activation of protein kinase C, and the induction of c-fos and c-myc in smooth muscle cells. *FASEB J* 1997; **11**: 703–711.
- 23 TANG L, MAMOTTE CD, VAN BOCKXMEER FM, TAYLOR RR. The effect of homocysteine on DNA synthesis in cultured human vascular smooth muscle. *Atherosclerosis* 1998; **136**: 169–173.
- 24 HARKER LA, ROSS R, SLICHTER SJ, SCOTT CR. Homocysteine-induced arteriosclerosis. The role of endothelial cell injury and platelet response in its genesis. *J Clin Invest* 1976; **58**: 731–741.
- 25 HULTBERG B, ANDERSSON A, ISAKSSON A. Higher export rate of homocysteine in a human endothelial cell line than in other human cell lines. *Biochim Biophys Acta* 1998; **1448**: 61–69.
- 26 TSAI JC, PERRELLA MA, YOSHIZUMI M *et al*. Promotion of vascular smooth muscle cell growth by homocysteine: a link to atherosclerosis. *Proc Natl Acad Sci U.S.A.* 1994; **91**: 6369–6373.
- 27 WANG H, YOSHIZUMI M, LAI K *et al*. Inhibition of growth and p21ras methylation in vascular endothelial cells by homocysteine but not cysteine. *J Biol Chem* 1997; **272**: 25380–25385.
- 28 XU D, NEVILLE R, FINKEL T. Homocysteine accelerates endothelial cell senescence. *FEBS Lett* 2000; **470**: 20–24.
- 29 HULTBERG B, ANDERSSON A, ISAKSSON A. The cell-damaging effects of low amounts of homocysteine and copper ions in human cell line cultures are caused by oxidative stress. *Toxicology* 1997; **123**: 33–40.
- 30 UPCHURCH GR, JR, WELCH GN, LOSCALZO J. Homocysteine, EDRF, and endothelial function. *J Nutr* 1996; **126**: 1290S–1294S.
- 31 LOSCALZO J. The oxidant stress of hyperhomocyst(e)inemia [editorial; comment]. *J Clin Invest* 1996; **98**: 5–7.
- 32 MUJUMDAR VS, HAYDEN MR, TYAGI SC. Homocyst(e)ine induces calcium second messenger in vascular smooth muscle cells. *J Cell Physiol* 2000; **183**: 28–36.
- 33 LUBEC B, LABUDOVA O, HOEGER H *et al*. Homocysteine increases cyclin-dependent kinase in aortic rat tissue. *Circulation* 1996; **94**: 2620–2625.
- 34 SUZUKI YJ, LORENZI MV, SHI SS, DAY RM, BLUMBERG JB. Homocysteine exerts cell type-specific inhibition of AP-1 transcription factor. *Free Radic Biol Med* 2000; **28**: 39–45.
- 35 BROWN JC, ROSENQUIST TH, MONAGHAN DT. ERK2 activation by homocysteine in vascular smooth muscle cells. *Biochem Biophys Res Commun* 1998; **251**: 669–676.
- 36 VADIVELOO PK, FILONZI EL, STANTON HR, HAMILTON JA. G1 phase arrest of human smooth muscle cells by heparin, IL-4 and cAMP is linked to repression of cyclin D1 and cdk2. *Atherosclerosis* 1997; **133**: 61–69.
- 37 NISHINAGA M, OZAWA T, SHIMADA K. Homocysteine, a thrombogenic agent, suppresses anticoagulant heparan sulfate expression in cultured porcine aortic endothelial cells. *J Clin Invest* 1993; **92**: 1381–1386.
- 38 HAJJAR KA. Homocysteine-induced modulation of tissue plasminogen activator binding to its endothelial cell membrane receptor. *J Clin Invest* 1993; **91**: 2873–2879.
- 39 LENTZ SR, SADLER JE. Inhibition of thrombomodulin surface expression and protein C activation by the thrombogenic agent homocysteine. *J Clin Invest* 1991; **88**: 1906–1914.
- 40 LENTZ SR, SADLER JE. Homocysteine inhibits von Willebrand factor processing and secretion by preventing transport from the endoplasmic reticulum. *Blood* 1993; **81**: 683–689.
- 41 MAJORS A, EHRHART LA, PEZACKA EH. Homocysteine as a risk factor for vascular disease. Enhanced collagen production and accumulation by smooth muscle cells. *Arterioscler Thromb Vasc Biol* 1997; **17**: 2074–2081.
- 42 TYAGI SC, SMILEY LM, MUJUMDAR VS, CLONTS B, PARKER JL. Reduction-oxidation (Redox) and vascular tissue level of homocyst(e)ine in human coronary atherosclerotic lesions and role in extracellular matrix remodeling and vascular tone. *Mol Cell Biochem* 1998; **181**: 107–116.
- 43 TAWAKOL A, OMLAND T, GERHARD M, WU JT, CREAGER MA. Hyperhomocyst(e)inemia is associated with impaired endothelium-dependent vasodilation in humans [published erratum appears in *Circulation* 2000 Mar 28;101(12):E116]. *Circulation* 1997; **95**: 1119–1121.
- 44 LENTZ SR, SOBEY CG, PIEGORS DJ *et al*. Vascular dysfunction in monkeys with diet-induced hyperhomocyst(e)inemia [see comments]. *J Clin Invest* 1996; **98**: 24–29.
- 45 McCULLY KS. Vascular pathology of homocysteinemia: implications for the pathogenesis of arteriosclerosis. *Am J Pathol* 1969; **56**: 111–128.
- 46 MOHAN IV, STANSBY G. Nutritional hyperhomocysteinemia. *BMJ* 1999; **318**: 1569–1570.
- 47 BOUSHEY CJ, BERESFORD SA, OMENN GS, MOTULSKY AG. A quantitative assessment of plasma homocysteine as a risk factor for vascular disease. Probable benefits of increasing folic acid intakes [see comments]. *JAMA* 1995; **274**: 1049–1057.
- 48 BERWANGER CS, JEREMY JY, STANSBY G. Homocysteine and vascular disease. *Br J Surg* 1995; **82**: 726–731.
- 49 ERITSLAND J, ARNESEN H, SELJEFLOT I *et al*. Influence of serum lipoprotein(a) and homocyst(e)ine levels on graft patency after coronary artery bypass grafting. *Am J Cardiol* 1994; **74**: 1099–1102.
- 50 BENOIT C, FURBER A, LE BOUIL A *et al*. [Plasma homocysteine is not a predictive factor of restenosis after coronary angioplasty]. *Arch Mal Coeur Vaiss* 1999; **92**: 1457–1460.
- 51 MORITA H, KURIHARA H, KUWAKI T *et al*. Homocysteine as a risk factor for restenosis after coronary angioplasty. *Thromb Haemost* 2000; **84**: 27–31.
- 52 AMBROSI P, GARCON D, RIBERI A *et al*. Association of mild hyperhomocysteinemia with cardiac graft vascular disease. *Atherosclerosis* 1998; **138**: 347–350.
- 53 BEATTIE DK, SIAN M, GREENHALGH RM, DAVIES AH. Influence of systemic factors on pre-existing intimal hyperplasia and their effect on the outcome of infrainguinal arterial reconstruction with vein. *Br J Surg* 1999; **86**: 1441–1447.
- 54 TSAKIRIS DA, TSOCHOPL M, JAGER K *et al*. Circulating cell adhesion molecules and endothelial markers before and after transluminal angioplasty in peripheral arterial occlusive disease. *Atherosclerosis* 1999; **142**: 193–200.
- 55 CURRIE IC, WILSON YG, SCOTT J *et al*. Homocysteine: an independent risk factor for the failure of vascular intervention. *Br J Surg* 1996; **83**: 1238–1241.
- 56 MATTHIAS D, BECKER CH, RIEZLER R, KINDLING PH. Homocysteine induced arteriosclerosis-like alterations of the aorta in normotensive and hypertensive rats following application of high doses of methionine. *Atherosclerosis* 1996; **122**: 201–216.
- 57 ROLLAND PH, FRIGGI A, BARLATIER A *et al*. Hyperhomocysteinemia-induced vascular damage in the minipig. Captopril-hydrochlorothiazide combination prevents elastic alterations. *Circulation* 1995; **91**: 1161–1174.
- 58 TOBOREK M, KOPIECZNA-GRZEBIENIAK E, DROZDZ M, WIECZOREK M. Increased lipid peroxidation as a mechanism of methionine-induced atherosclerosis in rabbits. *Atherosclerosis* 1995; **115**: 217–224.

- 59 YOUNG PB, KENNEDY S, MOLLOY AM *et al.* Lipid peroxidation induced *in vivo* by hyperhomocysteinaemia in pigs. *Atherosclerosis* 1997; **129**: 67–71.
- 60 BLOM HJ, KLEINVELD HA, BOERS GH *et al.* Lipid peroxidation and susceptibility of low-density lipoprotein to *in vitro* oxidation in hyperhomocysteinaemia. *Eur J Clin Invest* 1995; **25**: 149–154.
- 61 LENTZ SR, FERNANDEZ JA, GRIFFIN JH *et al.* Impaired anticoagulant response to infusion of thrombin in atherosclerotic monkeys associated with acquired defects in the protein C system. *Arterio Thromb & Vasc Biol* 1999; **19**: 1744–1750.
- 62 ZULLI A, BUXTON BF, DOOLAN L, LIU JJ. Augmented effects of methionine and cholesterol in decreasing the elastic lamina while thickening the aortic wall in the rat aorta. *Clin Sci* 1998; **95**: 589–593.
- 63 DURAND P, PROST M, BLACHE D. Pro-thrombotic effects of a folic acid deficient diet in rat platelets and macrophages related to elevated homocysteine and decreased n-3 polyunsaturated fatty acids. *Atherosclerosis* 1996; **121**: 231–243.
- 64 DURAND P, LUSSIER-CACAN S, BLACHE D. Acute methionine load-induced hyperhomocysteinemia enhances platelet aggregation, thromboxane biosynthesis, and macrophage-derived tissue factor activity in rats. *FASEB J* 1997; **11**: 1157–1168.
- 65 UNGVARI Z, SARKADI-NAGY E, BAGI Z, SZOLLAR L, KOLLER A. Simultaneously increased TXA(2) activity in isolated arterioles and platelets of rats with hyperhomocysteinemia. *Arterio Thromb & Vasc Biol* 2000; **20**: 1203–1208.
- 66 DUDMAN NP, TEMPLE SE, GUO XW, FU W, PERRY MA. Homocysteine enhances neutrophil-endothelial interactions in both cultured human cells and rats *In vivo*. *Circ Res* 1999; **84**: 409–416.
- 67 SOUTHERN FN, CRUZ N, FINK LM *et al.* Hyperhomocysteinemia increases intimal hyperplasia in a rat carotid endarterectomy model. *J Vasc Surg* 1998; **28**: 909–918.
- 68 REFSUM H, UELAND PM, NYGARD O, VOLLSET SE. Homocysteine and cardiovascular disease. *Ann Rev Med* 1998; **49**: 31–62.
- 69 CLARKE R, SMITH AD, JOBST KA *et al.* Folate, vitamin B12, and serum total homocysteine levels in confirmed Alzheimer disease [see comments]. *Arch Neurol* 1998; **55**: 1449–1455.
- 70 CARMODY BJ, ARORA S, AVENA R, COSBY K, STRAWY AN. Folic acid inhibits homocysteine-induced proliferation of human arterial smooth muscle cells. *J Vasc Surg* 1999; **30**: 1121–11288.
- 71 CONSTANS J, BLANN AD, RESPLANDY F *et al.* Three months supplementation of hyperhomocysteinaemic patients with folic acid and vitamin B6 improves biological markers of endothelial dysfunction. *Br J Haematol* 1999; **107**: 776–778.
- 72 WOO KS, CHOOK P, LOLIN YI *et al.* Folic acid improves arterial endothelial function in adults with hyperhomocystinemia. *J Am Coll Cardiol* 1999; **34**: 2002–2006.
- 73 WILCKEN DE, DUDMAN NP, TYRRELL PA. Homocystinuria due to cystathionine beta-synthase deficiency – the effects of betaine treatment in pyridoxine-responsive patients. *Metabolism* 1985; **34**: 1115–1121.
- 74 WILCKEN DE, WILCKEN B. B vitamins and homocysteine in cardiovascular disease and aging. *Ann NY Acad Sci* 1998; **854**: 361–370.
- 75 NAPPO F, DE ROSA N, MARFELLA R *et al.* Impairment of endothelial functions by acute hyperhomocysteinemia and reversal by antioxidant vitamins. *JAMA* 1999; **281**: 2113–2118.
- 76 VERMEULEN EG, STEHOUWER CD, TWISK JW *et al.* Effect of homocysteine-lowering treatment with folic acid plus vitamin B6 on progression of subclinical atherosclerosis: a randomised, placebo-controlled trial [see comments]. *Lancet* 2000; **355**: 517–522.
- 77 DE JONG SC, STEHOUWER CD, VAN DEN BERG M *et al.* Normohomocysteinaemia and vitamin-treated hyperhomocysteinaemia are associated with similar risks of cardiovascular events in patients with premature peripheral arterial occlusive disease. A prospective cohort study. *J Intern Med* 1999; **246**: 87–96.
- 78 VERMEULEN EG, RAUWERDA JA, ERIX P *et al.* Normohomocysteinaemia and vitamin-treated hyperhomocysteinaemia are associated with similar risks of cardiovascular events in patients with premature atherothrombotic cerebrovascular disease. A prospective cohort study. *Neth J Med* 2000; **56**: 138–146.

Accepted 4 October 2001