



## Medical Microbiology

# Isoflavone formononetin from red propolis acts as a fungicide against *Candida* sp



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## ABSTRACT

A bioassay-guided fractionation of two samples of Brazilian red propolis (from Igarassu, PE, Brazil, hereinafter propolis 1 and 2) was conducted in order to determine the components responsible for its antimicrobial activity, especially against *Candida* spp. Samples of both the crude powdered resin and the crude ethanolic extract of propolis from both locations inhibited the growth of all 12 tested *Candida* strains, with a minimum inhibitory concentration of 256  $\mu\text{g}/\text{mL}$ . The hexane, acetate and methanol fractions of propolis 1 also inhibited all strains with minimum inhibitory concentration values ranging from 128 to 512  $\mu\text{g}/\text{mL}$  for the six bacteria tested and from 32 to 1024  $\mu\text{g}/\text{mL}$  for the yeasts. Similarly, hexane and acetate fractions of propolis sample 2 inhibited all microorganisms tested, with minimum inhibitory concentration values of 512  $\mu\text{g}/\text{mL}$  for bacteria and 32  $\mu\text{g}/\text{mL}$  for yeasts. The extracts were analyzed by HPLC and their phenolic profile allowed us to identify and quantitate one phenolic acid and seven flavonoids in the crude ethanolic extract. Formononetin and pinocembrin were the major constituents amongst the identified compounds. Formononetin was detected in all extracts and fractions tested, except for the methanolic fraction of sample 2. The isolated isoflavone formononetin inhibited the growth of all the microorganisms tested, with a minimum inhibitory concentration of 200  $\mu\text{g}/\text{mL}$  for the six bacteria strains tested and 25  $\mu\text{g}/\text{mL}$  for the six yeasts. Formononetin also exhibited fungicidal activity against five of the six yeasts tested. Taken together our results demonstrate that the isoflavone formononetin is implicated in the reported antimicrobial activity of red propolis.

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## Introduction

Propolis is a resinous material collected by melliferous bees from various plant exudates, such as secretions of trees, leaves and flowers. This resin is used by bees in the protection of the hive against bacterial and fungal infections.<sup>1</sup>

The chemical composition of propolis samples is complex and varies according to its source. Amongst the compounds reported to occur in these samples, phenolic acids and flavonoids are particularly important since many of propolis' alleged biological activities are attributed to these substances.<sup>2</sup>

Brazilian propolis has been classified into 12 different chemotypes, according to its chemical profile as determined by UV/VIS spectrophotometry, thin layer and high performance liquid chromatography, as well as its antimicrobial and antioxidant activities.<sup>3</sup> Red propolis constitutes a new, separate chemotype occurring at mangrove regions of north-eastern Brazilian states and is characterized by its intense red color. This chemotype is the only propolis type whose botanical origin can be traced to a plant species (*Dalbergia ecastaphylum*) from the Leguminosae family and one of its characteristics is its high isoflavones content.<sup>4</sup> Red propolis has attracted considerable attention for its biological properties<sup>5</sup> which include antimicrobial,<sup>6</sup> antioxidant,<sup>7</sup> leishmanicidal,<sup>8</sup> and antileukemic<sup>6</sup> activities. To our knowledge there is no chemical or biological characterization reported for the propolis sample from Igarassu (Pernambuco, Brazil).

Considering that fractionation is the first step toward identification of new bioactive natural products,<sup>9</sup> the present work aimed at performing a bioassay-guided fractionation of red propolis samples from Igarassu (Pernambuco, Brazil) in order to determine the main constituents associated with its antimicrobial activity, especially against *Candida sp.*

## Material and methods

### Red propolis samples and chemical standards

Red propolis samples were obtained from collecting traps placed at the hives of *Apis mellifera* bees. Collection took place between April 2011 and May 2012 at two apiaries (7°50'37.05" S and 34°53'00.66" W elev. 7 m; 7°49'57.90" S and 34°56'02.38" W elev. 64 m), located in the municipality of Igarassu, Pernambuco state, Brazil. The samples were labeled as Sample 1 (apiary 1) and Sample 2 (apiary 2). After collection, samples were cleaned by removing any foreign material and then powdered in a shaker until a thin red powder was obtained. The sample was weighed and stored at –18 °C and will be referred to from now on as crude powdered propolis. The crude powdered propolis was submitted to chromatographic analysis, tested for antimicrobial activity and was used for the preparation of the crude ethanol extract and its fractions. Solvents used were HPLC-grade unless otherwise specified. The chemical standards ferulic acid, rutin, daidzein, quercetin, luteolin, formononetin, pinocembrin, and biochanin A (≥98–99% purity) were acquired from Sigma–Aldrich (São Paulo, Brasil).

### Preparation of red propolis extracts

The crude powdered propolis (100 g) was extracted with 500 mL of ethanol 96%. Extraction was carried out at room temperature (24 °C) in an ultrasonic bath for 30 min. The solution was then filtered through a cotton plug and the resulting crude ethanol extract left overnight in the freezer (–18 °C) in order to precipitate the waxes. The extract was then placed in a separating funnel for better separation from the waxes and the supernatant was evaporated to dryness in a rotary evaporator at 40 °C. This crude ethanolic extract was submitted to chromatographic analysis, tested for antimicrobial activity and used for preparation of the fractions.

### Fractionation of the crude ethanol extract and isolation of formononetin

The crude ethanol extract (50 g) was suspended in a mixture of 100 mL HPLC-grade methanol and 100 mL water (1:1, v/v) in a separating funnel and fractionated by liquid-liquid extraction with hexane (700 mL, 7 extractions for sample 1 and 400 mL, 4 extractions for sample 2) and ethyl acetate (500 mL, 5 extractions for sample 1 and for sample 2). The resulting fractions were evaporated to dryness to afford the hexane (27.86 g), acetate (8.34 g) and methanol (2.13 g) fractions for sample 1 and hexane (27.89 g), acetate (9.14 g) and methanol fractions (1.70 g) for sample 2. All fractions were submitted to chromatographic analysis and tested for antimicrobial activity. Since the acetate fraction was the most flavonoid-rich fraction obtained (see Fig. 2 and Table 1), it was also used for further purification. An acetate fraction of sample 1 (8.34 g) was dissolved in a mixture of CHCl<sub>3</sub>:MeOH (1:1, v/v). This sample was submitted to column chromatography with Sephadex® LH-20 and eluted isocratically with CHCl<sub>3</sub>:MeOH (1:1, v/v), yielding 48 fractions. These fractions were monitored by thin layer chromatography using silica gel plates, a solvent system of CHCl<sub>3</sub>:MeOH (9:1, v/v) and a visualization reagent PEG-NP (for phenolics). Fractions were grouped according to their TLC profile into four groups (F7-8, F9-14, F15-29 and F30-48).

### High performance liquid chromatography analysis

HPLC analysis was performed using a Shimadzu liquid chromatograph composed of a LC-10AD vp pump, FCV-10AL vp solvent mixer, DGU-14A degasser unit, CTO-10AS column oven, SIL-10AD vp autosampler, SP-10AV vp UV/Vis detector, and a SCL-10A vp system controller. Samples (20 µL) of the crude powdered propolis, ethanol extract, hexane fraction, acetate fraction and methanol fraction at a concentration of 5 mg/mL (dissolved in MeOH:H<sub>2</sub>O 8:2, v/v) or formononetin at 100 µg/mL were injected into the chromatograph. Separation was achieved using a Luna C-18 column (250 mm × 4.6 mm, 5 µm) from Phenomenex (Torrance, USA) and a C-18 guard column (Security guard®, Phenomenex, Torrance, USA). The mobile phase consisted of ultrapure water (A) and methanol (B) delivered at a flow rate of 0.8 mL/min. A gradient of 45–65% B from 0 to 90 min, 65–75% B from 90 to 120 min, 75–95% B from 120 to 170 min, and 95–45% B from 170 to 185 min was used. Detection was done at 254 nm and identification of compounds was done by comparing with the

**Table 1 – Content ( $\mu\text{g}/\text{mg}$ ) of phenolic compounds found in the analyzed samples as determined by HPLC.**

Compounds	Sample 1					Sample 2				
	Powdered sample	EtOH extract	Fr-Hex	Fr-AcOEt	Fr-MeOH	Powdered sample	EtOH extract	Fr-Hex	Fr-AcOEt	Fr-MeOH
Ferulic acid	1.53	0.96	0.29	2.37	1.47	0.82	0.12	0.21	4.36	20.47
Biochanin A	1.03	0.56	0.09	2.69	0.07	0.05	0.17	0.55	0.10	0.05
Daidzein	0.36	0.02	0.07	0.61	0.05	0.03	0.01	ND	0.93	0.07
Formononetin	12.38	2.86	3.84	44.14	2.13	2.04	1.71	1.11	16.12	ND
Luteolin	0.24	1.04	ND	0.50	1.05	1.07	1.08	1.20	0.22	1.22
Pinocembrin	4.86	2.07	1.63	7.31	5.66	1.79	2.31	3.03	14.51	0.37
Rutin	0.53	0.21	0.20	0.84	0.14	0.20	0.02	0.19	3.28	0.42
Quercetin	1.87	0.37	0.38	1.29	0.39	0.39	0.39	0.41	1.82	0.37

Fr-Hex, hexane fraction; Fr-AcOEt, acetate fraction; Fr-MeOH, methanol fraction. ND, not detected.

retention time of standards at two different mobile phase compositions. Calibration curves for each of the standards were obtained by dissolving an appropriate amount of each compound in HPLC-grade MeOH to produce a 1 mg/mL stock solution and then diluting these stock solutions with a mixture of MeOH:H<sub>2</sub>O (80:20 v/v) to produce the final standard concentrations (0.5–100  $\mu\text{g}/\text{mL}$ ). The concentration range for each standard was chosen so as to bracket the peak area of each compound detected in the samples in preliminary chromatographic runs. The calibration curves were used if the correlation coefficient was 0.99 or higher.

#### Antimicrobial activity

Determination of the antimicrobial activity of the crude powdered propolis, ethanol extract, fractions and formononetin was carried out by determining the minimum inhibitory concentration (MIC), minimum bactericidal concentration (MBC) and minimum fungicidal concentration (MFC). The following bacteria species were tested: *Staphylococcus aureus* ATCC 13150, *Staphylococcus aureus* ATCC 25923, *Staphylococcus epidermidis* ATCC 12228, *Pseudomonas aeruginosa* ATCC 9027, *Pseudomonas aeruginosa* ATCC P-12, *Pseudomonas aeruginosa* ATCC P-03; for antifungal tests: *Candida albicans* ATCC 76645, *Candida albicans* LM P-20, *Candida tropicalis* ATCC 13803, *Candida tropicalis* LM 6, *Cryptococcus neoformans* ICB 59, *Cryptococcus neoformans* LM 2601. The culture medium for tests was RPMI 1640 with L-glutamine and no bicarbonate (Sigma-Aldrich®) for antifungal activity assays and nutrient broth (Difco Laboratories/USA/France) for the antibacterial activity tests. The inoculum was prepared from colonies taken from recently grown cultures in appropriate media incubated at 35–37 °C for 24–48 h for bacteria and 24–72 h for yeast. The colonies were suspended in sterile 0.9% NaCl. The inoculum suspensions were shaken for 2 min and the inoculum density was adjusted to the turbidity of a 0.5 McFarland standard with sterile saline (equivalent to 1–5  $\times 10^6$  cfu/mL).<sup>10–13</sup>

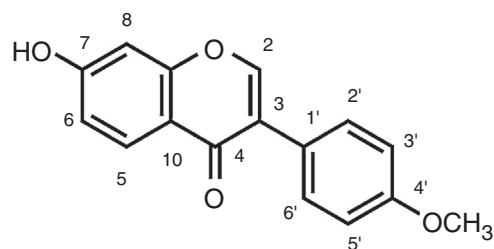
#### Determination of the minimum inhibitory concentration (MIC)

The determination of the minimum inhibitory concentration (MIC) was done in duplicate by the broth microdilution

method using U-shaped round-bottomed 96 well microplates. In each well, 100  $\mu\text{L}$  of the doubly concentrated medium was added and on the first row of each plate, 100  $\mu\text{L}$  of the tested substances were added to give final concentrations of 2048  $\mu\text{g}/\text{mL}$  for all samples except for isolated formononetin which was added to give a final concentration of 400  $\mu\text{g}/\text{mL}$ . Then, by serial dilution the test substances and formononetin were diluted up to 32  $\mu\text{g}/\text{mL}$  or 6.25  $\mu\text{g}/\text{mL}$  respectively. After the dilution step, 10  $\mu\text{L}$  of the microorganism inoculum was added in each well. For each microorganism, wells containing the following controls were also added: chloramphenicol 100  $\mu\text{g}/\text{mL}$  (on the plates used in the antibacterial tests), nystatin 100 UI/mL (on the plates used in the antifungal tests) and 1% DMSO (on all plates). Plates were incubated at 35–37 °C for 24–48 h for bacteria and 24–72 h for yeast. After the incubation period, 20  $\mu\text{L}$  of resazurin sodium salt solution at 0.01% (w/v) was added to the plates used in the antibacterial tests and 20  $\mu\text{L}$  of 0.5% triphenyl tetrazolium chloride (TTC) was added to the plates used for antifungal activity as indicators of cell viability. The MIC was defined as the lowest concentration capable of inhibiting the bacterial and fungal growth when compared with the control wells. Stock solutions of test substances were diluted in DMSO (final concentration of DMSO in each well was no higher than 0.1%).

#### Determination of the minimum bactericidal (MBC) and minimum fungicidal concentrations (MFC)

A modification of the microdilution method was used to estimate MBC and MFC values. Aliquots of 10  $\mu\text{L}$  from wells that did not present growth during the experiments for MIC determination (taken from those wells corresponding to 1 $\times$ , 2 $\times$  and 4 $\times$  the MIC value calculated for each tested substances) were transferred to sterile microtitration plates (DISPOPETRI, Brazil) containing 100  $\mu\text{L}$ /well of incubation medium for determination of MBC and MFC. Plates were incubated at 35–37 °C for 24–48 h for bacteria and 24–72 h for yeast. The MBC and MFC were considered as the lowest concentration of tested substances able to induce inactivation of the inoculum for bactericidal and fungicidal tests respectively.<sup>14,15</sup> All antimicrobial tests were performed in duplicate and the results were expressed as the geometric mean of the MIC, MBC, or MFC.<sup>11,16,17</sup> For interpretation of the results the guidelines



**Fig. 1 – Structure of isoflavone formononetin.**

adopted by Sartoratto et al.<sup>18</sup> were followed: high activity for values of MIC between 50 and 500  $\mu\text{g/mL}$ , average activity for MIC values between 600 and 1500  $\mu\text{g/mL}$  and low activity for MIC values above 1600  $\mu\text{g/mL}$ .

## Results

### HPLC profile of the samples and formononetin isolation

The chromatograms of samples are shown in Fig. 2. The chromatograms of the crude ethanolic extract of both propolis samples were remarkable similar considering the geographic separation of the apiaries from which the samples were obtained. The acetate fraction concentrates most of the constituents found in the ethanolic extract, while the hexane and methanol fractions are less abundant in compounds. The chromatogram of the methanol fraction displayed a single major peak with a retention time of 10 min was identified as ferulic acid.

After chromatography with Sephadex<sup>®</sup> LH-20, fractions 9–14 produced a precipitate that was filtered, dried and submitted to <sup>1</sup>H and <sup>13</sup>C NMR spectroscopy for structure elucidation. The spectral data confirmed the identity of the substance as the isoflavone formononetin (12.3 mg) (Fig. 1). <sup>1</sup>H NMR data (DMSO-d<sub>6</sub>, 300 MHz): 3.79 (3H, s, OCH<sub>3</sub>), 6.99 (2H, d, J = 9.0 Hz, H-3', 5'), 6.87 (1H, d, J = 2.2 Hz, H-8), 6.95 (1H, dd, J = 8.9, 2.2 Hz, H-6), 7.51 (2H, d, J = 8.7 Hz, H-2', 6'), 8.34 (1H, s, H-2), 7.98 (1H, d, J = 9 Hz, H-5).

The phenolic compounds identified and quantified in the samples are listed in Table 1. One phenolic acid and seven flavonoids were identified in the samples. Amongst the compounds identified, formononetin and pinocembrin were the most abundant compounds and their concentration in the samples varied from 220.72  $\mu\text{g/mL}$  in the acetate fraction of sample 1 to 5.56  $\mu\text{g/mL}$  in the hexane fraction of sample 2 for formononetin and from 72.54  $\mu\text{g/mL}$  in the acetate fraction of sample 2 to 1.84  $\mu\text{g/mL}$  in the methanol fraction of sample 2 for pinocembrin.

### Antimicrobial activity

#### Minimum inhibitory concentration (MIC)

The results of MIC determination for the tested materials are presented in Table 2. The powdered red propolis of both samples 1 and 2, and the ethanol extract of both samples inhibited the growth of all twelve microorganisms tested (bacteria and yeasts) with a MIC of 256  $\mu\text{g/mL}$ , which according to Sartoratto

et al.<sup>18</sup> should be considered a high activity. The hexane and acetate fractions from both samples, together with the methanol fraction of sample 1 inhibited all microorganisms tested with MIC values varying from 32  $\mu\text{g/mL}$  to 1024  $\mu\text{g/mL}$ . These fractions had a very strong antifungal activity with all of them exhibiting a MIC value of 32  $\mu\text{g/mL}$  for the six yeasts tested, except for the methanol fraction of sample 1 which had a MIC of 128  $\mu\text{g/mL}$  against *C. tropicalis* ATCC 13803 and the acetate fraction of sample 1, with a MIC of 1024  $\mu\text{g/mL}$  against *C. tropicalis* LM 6. The methanol fraction of sample 2 did not inhibit the growth of any of the bacteria strains tested and was active against five out of six yeasts tested. The isoflavone formononetin, isolated from the acetate fraction was active against all microorganisms tested, with a MIC of 200  $\mu\text{g/mL}$  for the six bacteria and a MIC of 25  $\mu\text{g/mL}$  for the six yeast strains.

#### Minimum bactericidal concentration (MBC) and minimum fungicidal concentration (MFC)

Only the acetate fraction of sample 2 presented bactericidal activity, with MBC of 1.024  $\mu\text{g/mL}$  against four of the six tested bacterial strains. It showed no bactericidal activity against *Staphylococcus aureus* ATCC 13150 and *Staphylococcus aureus* ATCC 25923. The results for MFC determination are presented in Table 3. The ethanol extract of sample 2 had fungicidal activity against all strains tested, with a MFC of 256  $\mu\text{g/mL}$ . Isolated formononetin exhibited fungicidal activity against five of a total of six strains tested with a MFC of 200  $\mu\text{g/mL}$ , and was inactive only against *C. tropicalis* LM 6. It is worth noting that from all samples tested, the acetate fraction of sample 1 was the one with the lowest MFC at 64  $\mu\text{g/mL}$ , and this was the fraction with the highest formononetin concentration, at 220.72  $\mu\text{g/mL}$ .

## Discussion

The antimicrobial activity of red propolis has been described before<sup>7,19</sup> but reports are scarce in comparison to other types of Brazilian propolis. The comparison of the antimicrobial activity of different propolis samples is difficult, especially when they are from different geographical origins, since their chemical composition and thus their antimicrobial activity can vary considerably under these circumstances.<sup>20</sup>

Junior et al.<sup>21</sup> studied the antimicrobial activity of red propolis from Alagoas state (Brazil). The ethanol extract presented antimicrobial activity against Gram-positive (100% of tested strains) and Gram negative bacteria (62.5% of tested strains) as well as antifungal activity (100% of strains). In our study the powdered sample and the ethanol crude propolis extract of samples 1 and 2 from Igarassu (Pernambuco state, Brazil) presented activity against all the Gram-positive and Gram-negative bacteria, as well as against all yeast strains.

Cabral et al.<sup>7</sup> studied the antibacterial activity of red propolis samples from the state of Alagoas (Brazil) and found that the antibacterial activity of the chloroform fraction and its subfractions were higher (MIC values ranging from 15.8 to 31.7  $\mu\text{g/mL}$ ) than that of the crude ethanol extract (MIC values ranging from 62.5 to 125  $\mu\text{g/mL}$ ). The authors thus suggested that the antimicrobial activity of red propolis is not due to

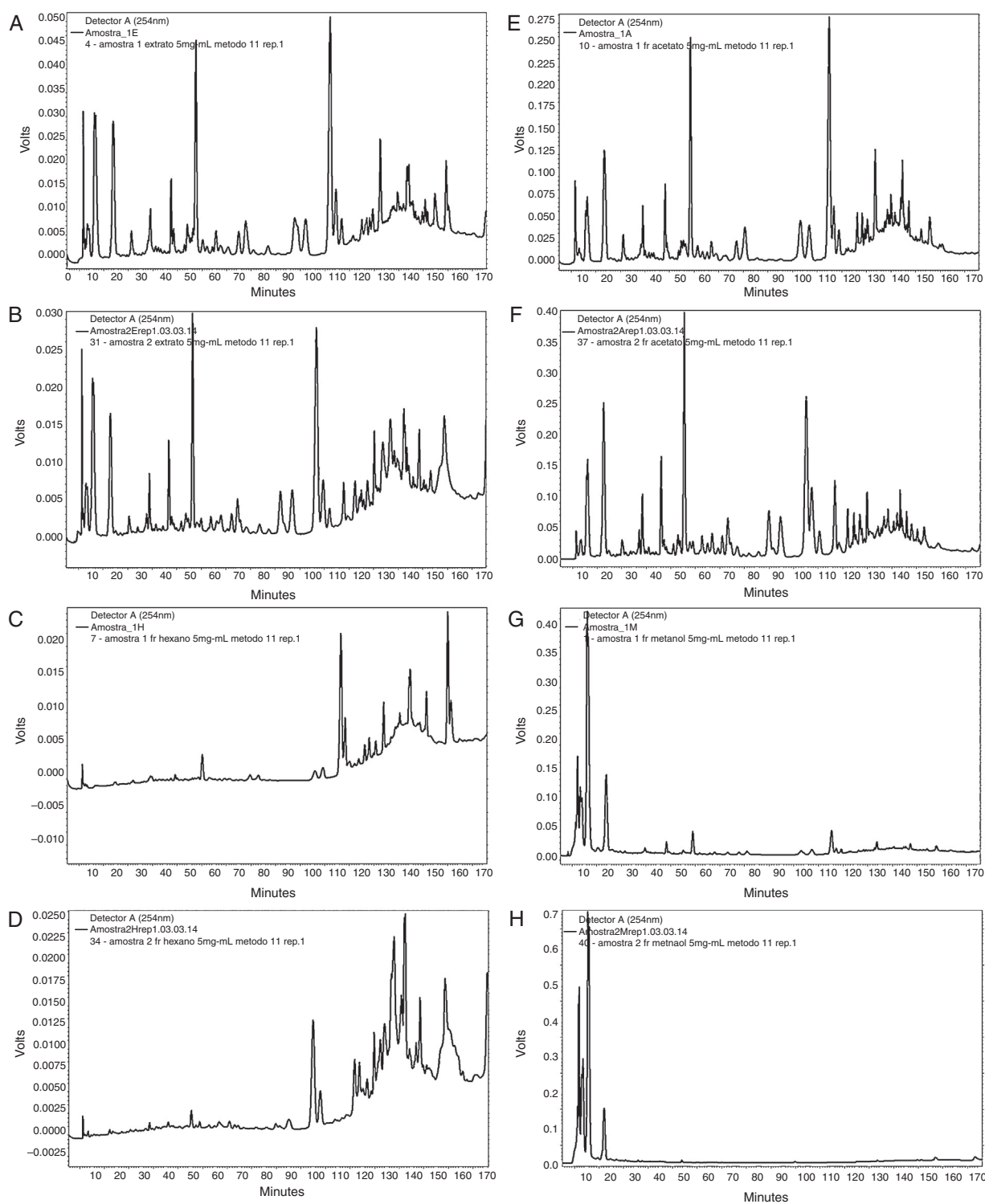
**Table 2 – Minimum inhibitory concentration – MIC ( $\mu\text{g/mL}$ ) of propolis samples 1 and 2, its fractions and extract+ (presence of microbial growth).**

Microorganisms	Sample 1					Sample 2					Formononetin	Chloramphenicol
	Powdered sample	Ethanol extract	<sup>a</sup> Fr-Hex	<sup>a</sup> Fr-AcOEt	<sup>a</sup> Fr-MeOH	Powdered sample	Ethanol extract	<sup>a</sup> Fr-Hex	<sup>a</sup> Fr-AcOEt	<sup>a</sup> Fr-MeOH		
<i>Gram-positive</i>												
<i>S. aureus</i> ATCC 13150	256	256	512	512	1024	256	256	512	512	+	200	32
<i>S. aureus</i> ATCC 25923	256	256	256	512	1024	256	256	512	512	+	200	32
<i>S. epidermidis</i> ATCC 12228	256	256	128	512	1024	256	256	512	512	+	200	64
<i>Gram-negative</i>												
<i>P. aeruginosa</i> ATCC 9027	256	256	128	512	1024	256	256	512	512	+	200	128
<i>P. aeruginosa</i> ATCC P-12	256	256	512	512	1024	256	256	512	512	+	200	128
<i>P. aeruginosa</i> ATCC P-03	256	256	256	128	1024	256	256	512	512	+	200	128
<i>Yeasts</i>												
<i>C. albicans</i> ATCC 76645	256	256	32	32	32	256	256	32	32	32	25	Nystatin 32
<i>C. albicans</i> LM P-20	256	256	32	32	32	256	256	32	32	32	25	32
<i>C. tropicalis</i> ATCC 13803	256	256	32	32	128	256	256	32	32	+	25	64
<i>C. tropicalis</i> LM 6	256	256	32	ND <sup>b</sup>	64	256	256	32	32	32	25	64
<i>C. neoformans</i> ICB 59	256	256	32	32	32	256	256	32	32	32	25	32
<i>C. neoformans</i> LM 2601	256	256	32	32	32	256	256	32	32	32	25	32

<sup>a</sup> Fr-Hex, hexane fraction; Fr-AcOEt, acetate fraction; Fr-MeOH, methanol fraction.

<sup>b</sup> ND, not determined.





**Fig. 2 – HPLC chromatographic profile of propolis extract and its fractions (at 5 mg/mL). A, C, E and G: ethanol extract, hexane fraction, acetate fraction, and methanol fraction of propolis sample 1 respectively; B, D, F and H: corresponding extract and fractions of propolis sample 2.**

a synergic effect of its components, but to individual constituents. In the same study, formononetin was the major compound identified in the ethanol extract and chloroform fraction. For the bacterial strains tested here, the acetate fraction showed a general trend to exhibit weaker antibacterial

activity than the ethanol fraction, but the antifungal activity against *Candida* strains of the acetate fraction was considerably higher than the one exhibited by the ethanol extract. In addition, formononetin, the isoflavone isolated from the samples exhibited good antifungal activity against all strains

**Table 3 – Minimum fungicidal concentration – MFC ( $\mu\text{g}/\text{ml}$ ) of propolis samples 1 and 2, its fractions and extract + (presence of microbial growth).**

Microorganisms	Sample 1						Sample 2					
	Powdered sample	Ethanol extract	<sup>a</sup> Fr-Hex	<sup>a</sup> Fr-AcOEt	<sup>a</sup> Fr-MeOH		Powdered sample	Ethanol extract	<sup>a</sup> Fr-Hex	<sup>a</sup> Fr-AcOEt	<sup>a</sup> Fr-MeOH	Formononetin
Yeasts												
<i>C. albicans</i> ATCC 76645	+	+	1.024	1.024	+	+	+	256	1.024	512	+	200
<i>C. albicans</i> LM P-20	+	+	+	64	+	+	+	256	1.024	512	+	200
<i>C. tropicalis</i> ATCC 13803	+	+	1.024	1.024	1.024	+	+	256	256	128	+	200
<i>C. tropicalis</i> LM 6	+	+	+	+	+	+	+	256	+	512	+	+
<i>C. neoformans</i> ICB 59	+	+	256	+	+	+	+	256	1.024	512	+	200
<i>C. neoformans</i> LM 2601	+	+	+	+	+	+	+	256	+	512	+	200

<sup>a</sup> Fr-Hex, hexane fraction; Fr-AcOEt, acetate fraction; Fr-MeOH, methanol fraction.

tested (MIC = 25  $\mu\text{g}/\text{ml}$ ), and it presented also fungicidal activity against all strains (MFC = 200  $\mu\text{g}/\text{ml}$ ), except for *C. tropicalis* LM 6. However, the antifungal activity of the acetate fraction cannot be attributed solely to formononetin, since the concentration of the isoflavone in the acetate fraction of sample 1 was determined to be 2.7 times higher than that of sample 2, and yet the fungicidal activity of the acetate fraction of sample 2 was higher (see Table 3). We also did not observe a trend of higher antibacterial potency of the fractions compared to the crude ethanol extract as observed by Cabral et al.<sup>7</sup> However, the MIC of the acetate fraction against all tested *Candida* strains was lower than the MIC of the crude ethanol fraction, demonstrating that formononetin is an important compound for the antifungal activity demonstrated by red propolis.

The botanical origin of propolis samples is difficult to ascertain on the basis of palynological analysis only, and a more definitive confirmation depends on analysis comparing the chemical profile of the samples with the chemical profile of resins and extracts from the plants found in close vicinity of the bee's hives. It should be stressed that red propolis has been suggested to be the only propolis type derived from a plant from the Leguminosae family (*D. ecastaphyllum*), rich in isoflavones such as genistein and formononetin.<sup>4</sup>

Although flavonoids exhibit pleiotropic activity affecting several different targets, and synergistic effects cannot be ruled out, our results suggest that the isoflavone formononetin is responsible at least partially for the antimicrobial activity of red propolis.

## Conflicts of interest

The authors declare no conflicts of interest.

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## REFERENCES

- Silva RA, Rodrigues AE, Ribeiro MCM, Custódio AR, Andrade NED, Pereira WE. Physicochemical characteristics and antimicrobial activity of the extracts propolis of the Paraíba, Brazil. *Cienc Rural*. 2006;36:1842–1848.
- Funari CS, Ferro VO. Análise de própolis. *Food Sci Technol (Campinas)*. 2006;26:171–178.
- Park YK, Alencar SM, Aguiar CL. Botanical origin and chemical composition of Brazilian propolis. *J Agric Food Chem*. 2002;50:2502–2506.
- Daugusch A, Moraes CS, Fort P, Park YK. Brazilian red propolis—chemical composition and botanical origin. *Evid Based Complement Altern Med*. 2008;5:435–441.
- Frozza COS, Garcia CSC, Gambato G, et al. Chemical characterization, antioxidant and cytotoxic activities of Brazilian red propolis. *Food Chem Toxicol*. 2013;52:137–142.
- Junior GCF, Moraes CS, Toreti VC, Daugsch A, Nowill AE, Park YK. Comparison of effects of the ethanolic extracts of Brazilian propolis in human leukemic cells as assessed with

- the MTT Assay. *Evid Based Complement Altern Med.* 2012, <http://dx.doi.org/10.1155/2012/91895>. Article ID 918956.
7. Cabral ISR, Oldoni TLC, Prado A, et al. Composição fenólica, atividade antibacteriana e antioxidante de própolis vermelha brasileira. *Quim Nova.* 2009;32:1523–1527.
  8. Ayres DC, Marcucci MC, Giorgio S. Effects of Brazilian propolis on *Leishmania amazonensis*. *Mem Inst Oswaldo Cruz.* 2007;102:215–220.
  9. Hayacibara MF, Koo H, Rosalen PL, et al. In vitro and in vivo effects of isolated fractions of Brazilian propolis on caries development. *J Ethnopharmacol.* 2005;101:110–115.
  10. Bauer AWMM, Kirby WM, Sherris JC, Turck M. Antibiotic susceptibility testing by a standardized single disk method. *Am J Clin Pathol.* 1966;45:493–496.
  11. Cleeland R, Squires E. Evaluation of new antimicrobials “in vitro” and in experimental animal infections. In: Lorian VMD, ed. *Antibiotics in Laboratory Medicine.* vol. 3. 1991:739–787.
  12. Hadacek F, Greger H. Testing of antifungal natural products: methodologies, comparability of results and assay choice. *Phytochem Anal.* 2000;11:137–147.
  13. NCCLS, NCCLS document M2-A7 *Performance Standards for Antimicrobial Disk Susceptibility Tests. Approved Standard.* 7th ed. Wayne, PA: NCCLS; 2000.
  14. Mann CM, Markham JL. A new method for determining the minimum inhibitory concentration of essential oils. *J Appl Microbiol.* 1998;84:538–544.
  15. Palomino JC, Martin A, Camacho M, Guerra H, Swings J, Portaels F. Resazurin microtiter assay plate: simple and inexpensive method for detection of drug resistance in *Mycobacterium tuberculosis*. *Antimicrob Agents Chemother.* 2002;46:2720–2722.
  16. Eloff JN. A sensitive and quick microplate method to determine the minimal inhibitory concentration of plant extracts for bacteria. *Planta Med.* 1998;64:711–713.
  17. Souza EL, Stamford TLM, Lima EO, Trajano VN. Effectiveness of *Origanum vulgare* L. essential oil to inhibit the growth of food spoiling yeasts. *Food Control.* 2007;18:409–413.
  18. Sartoratto A, Machado ALM, Delarmelina C, Figueira GM, Duarte NCT, Rehder VLG. Composition and antimicrobial activity of essential oils from aromatic plants used in Brazil. *Braz J Microbiol.* 2004;35:275–280.
  19. Righi AA, Alves TR, Negri G, Marques LM, Breyer H, Salatino A. Brazilian red propolis: unreported substances, antioxidant and antimicrobial activities. *J Sci Food Agric.* 2011;91:2363–2370.
  20. Seidel V, Peyfoon E, Watson DG, Fearnley J. Comparative study of the antibacterial activity of propolis from different geographical and climatic zones. *Phytother Res.* 2008;22:1256–1263.
  21. Junior WB, Miranda EO, Alvino V, Araujo B, Silva DW, Porfírio Z. Atividade antimicrobiana de frações da própolis vermelha de Alagoas, Brasil. *Semina.* 2012;33:03–10.