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ORIGINAL ARTICLE

In vitro effect of seven essential oils on the reproduction of the cattle tick *Rhipicephalus microplus*



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G R A P H I C A L A B S T R A C T



- Bovine tick infestation causes losses to producers

- *In vitro* test with essential oils is an alternative





Ticks treated with lemon gras (Cymbopogon citratus)

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ABSTRACT

The acaricidal effect of seven essential oils was examined *in vitro* against the cattle tick (*Rhipicephalus microplus*). Engorged female ticks were manually collected in farms of Southern Brazil and placed into petri dishes (*n* = 10) in order to test the following oils: juniper (*Juniperus communis*), palmarosa (*Cymbopogon martinii*), cedar (*Cedrus atlantica*), lemon grass (*Cymbopogon citratus*), ginger (*Zingiber officinale*), geranium (*Pelargonium graveolens*) and bergamot (*Citrus aurantium var bergamia*) at concentrations of 1%, 5%, and 10% each. A control group was used to validate the tests containing Triton X-100 only. Treatment effectiveness was measured considering inhibition of tick oviposition (partial or total), egg's weight, and hatchability. *C. martinii*, *C. citratus* and *C. atlantica* essential oils showed efficacy higher than 99% at all concentrations tested. In addition, *J. communis*, *Z. officinale*, *P. graveolens*, and *C. aurantium var bergamia* oils showed efficiency ranging from 73% to 95%, depending on the concentration tested, where higher concentrations showed greater efficacy. It was concluded that essential oils can affect tick reproduction *in vitro* by inhibiting oviposition and hatchability.

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The cattle tick *Rhipicephalus microplus* stands out as the most harmful pest for cattle, causing animal stress, lower growth, and poor performance, in addition to higher production costs due to constant anti-parasitic treatments [1,2]. The economic impact caused by cattle ticks in Brazil is of approximately \$3.24 billion dollars a year [1] since climatic conditions are favorable to their survival and development [3], increasing control costs with synthetic acaricides [4]. Moreover, restrictions on the use of insecticides and acaricides, such as organophosphates due to their effects on human and animal health [5], and the environment [6] have enhanced the development of effective alternatives for control, including essential oils.

The essential oils used in this study have exhibited several biological activities as previously described in the literature. Essential oils from Cymbopogon citratus (Poaceae family), Cymbopogon martinii (Gramineae family) and Juniperus communis (Cupressaceae family) have showed antioxidant [7], antimicrobial, antifungal and anthelmintic properties [8,9]. Cedrus atlantica (Pinaceae family) is the plant with fewer studies, even though its analgesic property has been described [10]. In vitro, Zingiber officinale (Zingiberaceae family) extract was able to reduce Streptococcus mutans and Streptococcus sanguinis growth with minimum inhibitory concentration of 0.02 mg/mL and 0.3 mg/mL, respectively [11]. The Pelargonium graveolens essential oil has been used due to its hypoglycemic and antioxidant [12] properties, and exhibits also antifungal and insecticidal activities against Rhizoctonia solani and Rhysopertha dominica, respectively [13]. The use of Citrus aurantium essential oil by Homa et al. [14] revealed the antifungal activity against different isolates of Fusarium keratitis, antibacterial activity against Vibrio species [15], as well as insecticidal activity against Aedes aegypti and Anopheles dirus [16]. As mentioned above, there are many properties of these essential oils, but there are only few studies on their acaricidal effects despite the great interest on finding alternative control methods. Therefore, this study aimed to evaluate the in vitro effect of essential oils (C. martinii, C. citratus, C. atlantica, J. communis, Z. officinale, P. graveolens, and Citrus aurantium var bergamia) on cattle tick R. microplus.

Material and methods

Essential oils

Seven essential oils were used to test the reproduction of engorged *R. microplus* females. The oils used were as follows: juniper (*J. communis*), palmarosa (*C. martinii*), cedar (*C. atlantica*), lemon grass (*C. citratus*), ginger (*Z. officinale*), geranium (*P. graveolens*), and bergamot (*C. aurantium var bergamia*). Three concentrations (1%, 5%, and 10%, i.e. 1v/99v, 5v/95v, and 10v/90v, respectively) were evaluated and Triton X-100 (Sigma Aldrich®, São Paulo, Brazil) was used as surfactant (1v/v), in addition to distilled water [17]. The essential oils of juniper, palmarosa, and lemon grass were acquired from BioEssencia® (São Paulo, Brazil), while essential oils of cedar, ginger, geranium, and bergamot were acquired from Phytoterápica® (São Paulo, Brazil).

Gas chromatography-flame ionization detector (GC-FID) of essential oils

The gas chromatography (GC) analyses were carried out using an 6890N GC-FID system equipped with DB-5 capillary column (30 m \times 0.25 mm; film thickness of 0.25 mm) (Agilent Technologies, Santa Clara, United States) connected to an FID detector. The injector and detector temperatures were set at 280 °C at a rate of 5 °C/min. The carrier gas was helium (> 99.2% purity) at a flow rate of 1.3 mL/min. All samples were analyzed in duplicate. Relative component concentrations were calculated based on GC peak areas without using correction factors [18].

Gas chromatography-mass spectrometry (GC-MS)

GC-MS analyses were performed on Agilent Technologies AutoSystem XL GC-MS operated in the EI mode at 70 eV (Hewlett Packard, Palo Alto, CA, USA) equipped with a splitless injector (250 °C). The transfer line temperature was 280 °C. Helium was used as the carrier gas (1.3 mL/min) and the capillary columns were DB-5 and HP5 MS (30 m × 0.25 mm; film thickness of 0.25 mm). Column temperature was programmed on 40–220 °C at 3 °C/min. The oils were diluted in hexane (1:5, v/v) and 1 μ L was injected.

Identification of the constituents was performed on the basis of retention index (RI) on DB-5 capillary column, determined in relation to homologous series of *n*-alkanes (C_7 - C_{30}) with those reported in the literature. Fragmentation patterns in the mass spectra library search (NIST and Wiley) were compared with those stored on databases [19]. The quantification of the compounds was performed on the basis of their relative peak areas on DB-5.

Ticks

The ticks were collected from dairy cows naturally infested in farms located in Quilombo city, Santa Catarina State, Southern Brazil. These animals did not receive any acaricidal treatment in the last 50 days prior to the beginning of the study in order to avoid any negative interference. The engorged female ticks were stored in plastic bottles, packed in a cooler ($\pm 15^{\circ}$ C), and immediately transported to the laboratory where the bioassays were conducted.

Bioassays

In the laboratory, engorged females ticks with similar weights were randomly distributed, placed into covered petri dishes during the incubation period. The experimental design was completely randomized with three replicates per oil concentration, and 10 ticks for each petri dish (total of 30 ticks per oil tested). The tests were performed according to the methodology described by Drummond et al. [20], where ticks were immersed for five minutes in the test solutions with essential oils at concentrations of 1%, 5%, and 10%. After that, they were dried and incubated under controlled conditions (25 °C: 75% relative humidity (RH)) for 14 days. Subsequently, oviposition was recorded as total, partial or absent and their eggs were weighted. Laid eggs were placed into glass tubes and incubated for 30 days in order to verify hatchability, which was measured considering the number of remaining eggs that did not hatch and the number of shells, versus the number of larvae (active or inactive) [21].

A control group containing only the diluents (water + Triton X-100) at concentration of 10% of Triton was used. The results were tabulated and reproductive efficiency (RE) and effectiveness of the treatment (ET) were calculated as described by Drummond et al. [20] [RE = egg weight \times % of hatchability \times 20,000/weight of engorged female ticks; ET = (RE control – RE treatment) \times 100/RE control].

Table 1 Mean and standard deviation of the weight of engorged tick, number of postures by treatment, egg weight, and hatchability after treatment with essential oils of juniper (*J. communis*), palmarosa (*C. martinii*), cedar (*C. atlantica*), lemon grass (*C. citratus*), ginger (*Z. officinale*), geranium (*P. graveolens*) and bergamot (*C. aurantium bergamia*).

Treatment	Engorged tick weight (g)	Number posture by treatment [*] ($n = 10$)	Weighing eggs per treatment (g)	Hatchability (%)
Control	0.190 ± 0.016	$10.0^{a} \pm 0.0$	$0.96^{a} \pm 0.03$	90
Juniper 1% Juniper 5% Juniper 10%	$\begin{array}{l} 0.198 \ \pm \ 0.021 \\ 0.201 \ \pm \ 0.011 \\ 0.187 \ \pm \ 0.018 \end{array}$	$\begin{array}{l} 8.0^{\rm b} \pm 1.1 \\ 7.0^{\rm bc} \pm 1.5 \\ 7.0^{\rm bc} \pm 0.2 \end{array}$	$\begin{array}{l} 0.35^{c} \pm 0.01 \\ 0.28^{d} \pm 0.03 \\ 0.25^{de} \pm 0.02 \end{array}$	38 10 08
Palmarosa 1% Palmarosa 5% Palmarosa 10%	$\begin{array}{l} 0.177 \pm 0.019 \\ 0.203 \pm 0.013 \\ 0.192 \pm 0.022 \end{array}$	$\begin{array}{l} 5.0^{\rm d} \ \pm \ 1.1 \\ 2.0^{\rm e} \ \pm \ 1.0 \\ 0.3^{\rm f} \ \pm \ 0.5 \end{array}$	$\begin{array}{l} 0.14^{\rm ef} \pm 0.01 \\ 0.06^{\rm g} \pm 0.01 \\ 0.06^{\rm g} \pm 0.01 \end{array}$	03 00 00
Cedar 1% Cedar 5% Cedar 10%	$\begin{array}{l} 0.196 \pm 0.016 \\ 0.184 \pm 0.020 \\ 0.188 \pm 0.012 \end{array}$	$\begin{array}{l} 8.7^{\rm ab} \pm 1.1 \\ 6.6^{\rm bc} \pm 0.5 \\ 4.6^{\rm d} \pm 1.1 \end{array}$	$\begin{array}{l} 0.51^{\rm b}\pm0.04\\ 0.35^{\rm c}\pm0.05\\ 0.06^{\rm g}\pm0.01 \end{array}$	00 00 00
Lemon grass 1% Lemon grass 5% Lemon grass 10%	$\begin{array}{l} 0.204 \ \pm \ 0.018 \\ 0.179 \ \pm \ 0.015 \\ 0.192 \ \pm \ 0.017 \end{array}$	$\begin{array}{l} 8.6^{\rm ab} \pm 1.1 \\ 5.6^{\rm cd} \pm 1.5 \\ 4.3^{\rm d} \pm 1.2 \end{array}$	$\begin{array}{l} 0.27^{\rm d} \ \pm \ 0.02 \\ 0.27^{\rm d} \ \pm \ 0.03 \\ 0.12^{\rm f} \ \pm \ 0.01 \end{array}$	00 00 00
Ginger 1% Ginger 5% Ginger 10%	$\begin{array}{l} 0.185 \pm 0.014 \\ 0.194 \pm 0.016 \\ 0.205 \pm 0.019 \end{array}$	$\begin{array}{l} 8.6^{ab} \pm 1.5 \\ 7.0^{bc} \pm 1.7 \\ 4.3^{d} \pm 0.6 \end{array}$	$\begin{array}{l} 0.42^{\rm bc}\pm0.06\\ 0.13^{\rm f}\pm0.04\\ 0.20^{\rm e}\pm0.01 \end{array}$	15 06 05
Geranium 1% Geranium 5% Geranium 10%	$\begin{array}{l} 0.191 \ \pm \ 0.013 \\ 0.188 \ \pm \ 0.017 \\ 0.199 \ \pm \ 0.015 \end{array}$	$\begin{array}{l} 9.0^{\rm ab} \pm 1.0 \\ 6.3^{\rm cd} \pm 2.0 \\ 5.3^{\rm d} \pm 1.2 \end{array}$	$\begin{array}{l} 0.42^{\rm bc}\pm0.04\\ 0.16^{\rm e}\pm0.02\\ 0.09^{\rm fg}\pm0.01 \end{array}$	13 09 05
Bergamot 1% Bergamot 5% Bergamot 10%	$\begin{array}{l} 0.178 \ \pm \ 0.010 \\ 0.197 \ \pm \ 0.014 \\ 0.180 \ \pm \ 0.013 \end{array}$	$\begin{array}{l} 7.3^{\rm bcd} \pm 1.1 \\ 6.3^{\rm cd} \pm 0.6 \\ 6.3^{\rm cd} \pm 1.5 \end{array}$	$\begin{array}{l} 0.36^{\rm c}\pm0.05\\ 0.26^{\rm d}\pm0.03\\ 0.29^{\rm cd}\pm0.04 \end{array}$	20 11 08

Note: Means followed by the same letter in the same column do not differ statistically among themselves, the significance level of 5% (P > 0.05).

* Number of engorged females (ticks) that perform posture (partial or total) per treatment, and "*n*" by repeating 10 specimens (test performed in triplicate).

Statistical analysis

The data collected were subjected to normality test which showed normal distribution. Then, the data were analyzed statistically by analysis of variance (one-way ANOVA) and Duncan's test. The results were considered significant when P < 0.05.

Results

In vitro test

The number of ticks that had partial or total oviposition, as well as egg weight, and percentage of hatched larvae is shown in Table 1. All results were compared to the control group that showed total oviposition and 86.3% of hatchability. The use of J. communis oil caused partial oviposition of smaller eggs (P = 0.0032) in all concentrations tested, even though it was unable to inhibit hatchability. On the contrary, the use of C. *martinii* oil (1%) led to lower egg hatchability (P = 0.0012), in addition to lower oviposition and egg weight, on a dosedependent effect. C. atlantica, C. citratus, Z. officinale, and P. graveolens essential oils tested at 1% were unable to reduce the number of ticks that showed oviposition, i.e. these oils did not cause any effect on reproduction (P = 0.142), which was not observed at concentrations of 5 and 10% (P = 0.092). C. atlantica and C. citratus oils were able to inhibit hatchability, an effect not seen for Z. officinale and P. graveolens oils. C. aurantium var bergamia oil was able to reduce the number of ticks that performed oviposition and the weight of eggs at all concentrations, but did not inhibit hatchability.

Data on tick reproductive efficiency and oil treatment efficacy are shown in Table 2. Oil treatment was able to significantly reduce tick reproductive efficiency compared to the control group (P = 0.0001). Regarding *C. atlantica* and *C. citratus* oils, all concentrations tested interfered with the reproduction of cattle ticks (100% efficacy) similar to *C. martinii* oil at 5% and 10%. The *C. aurantium var bergamia*, *Z. officinale*, *J. communis*, and *P. graveolens* oils at concentration 10%, exhibited an approximate efficiency of 90%, 94%, 96%, and 97%, respectively.

Oil composition

The major components found in each oil were as follows: linalool (*J.* communis; 18.07%), geraniol (*C.* martini; 35.27%), α -himachalene (*C. atlantica*; 19.74%), geranial (*C. citratus*; 46.51%), α -zingiberene (*Z. officinale*; 26.47%), citronellol (*P. graveolens*; 31.37%), and limonene (*C. aurantium var bergamia*; 30.17%) (Suppl. Table 1).

Discussion

In this study, it was observed that the *J. communis* oil was able to partially inhibit oviposition, and therefore, reduce tick reproductive efficiency. Carrol et al. [22] reported repellent action of juniper oil against two species of ticks (*Amblyomma americanum* and *Ixodes scapularis*). Studies conducted by Dietrich et al. [23] and Dolan et al. [24] have reported that the *J. communis* oil is a rich source of anti-tick compounds with

Table	2	Repr	roductive	effic	iency	and	effe	ective	ness	of	treat-
ment	of	seven	essential	oils	again	st ca	attle	tick	Rhip	ice	phalus
microplus.											

Treatment	Reproductive efficiency (%)	Treatment efficacy (%)
Control	81.0	0.0
Juniper 1%	23.3	73.8
Juniper 5%	4.2	95.2
Juniper 10%	3.6	96.3
Palmarosa 1%	1.8	99.7
Palmarosa 5%	0.0	100.0
Palmarosa 10%	0.0	100.0
Cedar 1%	0.0	100.0
Cedar 5%	0.0	100.0
Cedar 10%	0.0	100.0
Lemon grass 1%	0.0	100.0
Lemon grass 5%	0.0	100.0
Lemon grass 10%	0.0	100.0
Ginger 1%	5.7	85.7
Ginger 5%	2.9	92.6
Ginger 10%	2.5	94.0
Geranium 1%	5.6	85.9
Geranium 5%	3.3	91.6
Geranium 10%	1.2	97.0
Bergamot 1%	5.1	84.9
Bergamot 5%	5.3	86.6
Bergamot 10%	3.7	90.5

well-known repellent and insecticidal activities. Researchers also found 43.2% of repellent effect for juniper oil against *A. aegypti* after 210 min of application [25].

Additionally, *C. citratus* oil showed 100% efficacy against *R. microplus*, similar to those findings reported by other authors [26,27]. The effectiveness of *C. citratus* oil on ticks, according to Tchoumbougnang et al. [28] may be due to its geraniol content, measured as 47%. *C. martinii* oil at 5% and 10% showed 100% efficacy against adult ticks in this current study, and this oil has been studied for its repellent activity to insects [29,30] and antifungal actions [31], but it had not been tested on cattle ticks yet.

Z. Officinale belongs to Zingiberceae family, an aromatic plant used as spice and in medicine. According to the literature, the Z. officinale oil showed bactericidal effect on Staphylococcus aureus [32], repellent activity against mosquitoes of the species Culex quinquefasciatus [33], as well as repellent effect against Leptotrombidium deliense larvae, a species of mite [34], similar to the cattle tick used in this study.

The *C. aurantium var bergamia* oil negatively affected the reproduction of cattle tick. According to the literature, some compounds present in *Citrus* sp. essential oils showed repellant effect against mosquitoes and ticks [35]. Already, the *Cedrus deodara* oil demonstrated strong effect against cattle ticks [36], similar to what was observed in this study, even though a different kind of *C. atlantica* was used in this current study. Another study also reported efficacy to control cattle tick using a herbal preparation containing extracts of *C. deodara*, *Azadirachta indica*, and *Embelia ribes* [37], and according to

these authors, these extracts have acaricidal effect against larvae, nymphs, and adult stages of ticks.

The *P. graveolens* oil showed some effect on tick oviposition (inhibited or reduced), but it did not interfere on hatchability. Tabanca et al. [38] tested ten essential oils of *P. graveolens* and demonstrated repellent activities against nymphs of the medically important lone star tick, *A. americanum*. Researchers described that *P. graveolens* oil showed 100% repellency against host-seeking nymphs of *Ixodes ricinus* [39].

Conclusions

Based on these *in vitro* results it is possible to conclude that *C. martinii*, *C. citratus*, and *C. atlantica* oils may interfere on cattle tick reproduction. The essential oils of *J. communis*, *Z. officinale*, *P. graveolens*, and *C. aurantium var bergamia* also caused a negative effect on tick reproduction, but they were unable to inhibit hatchability. The use of essential oils in the control of *R. microplus* shows great potential for the future as an alternative method besides chemical products. Note that more tests, especially *in vivo*, are needed, in order to conclude whether such oils could be used as an alternative for the control of cattle ticks, and this is the main perspective of our research group.

Conflict of Interest

The authors have declared no conflict of interest.

Compliance with Ethics Requirements

This article does not contain any studies with human or animal subjects.

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Appendix A. Supplementary material

Supplementary data associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.jare.2016. 05.003.

References

- Grisi L, Leite RC, Martins JRS, Barros ATM, Andreotti R, Cançado PHD, et al. Reassessment of the potential economic impact of cattle parasites in Brazil. Rev Bras Parasitol Vet 2014;23(2):150–6.
- [2] Monteiro CMD, Prata MCD, Furlong J, Faza AP, Mendes AS, Andalo V, et al. *Heterorhabditis amazonensis* (Rhabditidae: Heterorhabditidae), strain RSC-5, for biological control of the catle tick *Rhipicephalus* (*Boophilus*) *microplus* (Acari: Ixodidae). Parasitol Res 2010;106:821–6.
- [3] Rocha CMBM. Caracterização da percepção dos produtores de leite do município de Divinolópis/MG sobre a importância do carrapato *Boophilus microplus* e fatores determinantes das

formas de combate utilizadas. 1995. 205f. Dissertação (Mestrado em Medicina veterinária preventiva e Epidemiologia) – Curso de pós-graduação em Medicina Veterinária, Universidade Federal de Minas Gerais; 1995.

- [4] Ellse L, Burden F, Wall R. Pyrethroid tolerance in the chewing louse *Bovicola* (*Werneckiella*) ocellatus. Vet Parasitol 2012;188:134–9.
- [5] Kolaczinski JH, Curtis CF. Chronic illness as a result of lowlevel exposure to synthetic pyrethroid insecticides: a review of the debate. Food Chem Toxicol 2004;4:697–706.
- [6] Ramwell CT, Sinclair CJ, Van Beinum GW, Bryning G. Management of the environmental inputs and risks of cypermethrin based sheep dips. Central Sci Lab Rep 2009;1:35–43.
- [7] Andrade BFMT, Braga CP, dos Santos KC, Barbosa LN, Rall VLM, Sforcin JM, et al. Effect of inhaling *Cymbopogon martinii* essential oil and geranial on serum biochemistry parameters and oxidative stress. Biochem Res Int 2014;2014:493183.
- [8] Katiti LM, Chagas AC, Bizzo HR, Ferreira AJ, Amarante AF. Anthelmintic activity of *Cymbopogon martinii*, *Cymbopogon schoenanthus and Mentha piperita* essential oils evaluated in four different *in vitro* tests. Vet Parasitol 2011;183:103–8.
- [9] Gemeda N, Woldeamanuel Y, Asrat D, Debella A. Effect of essential oils on *Aspergillus spore* germination, growth and mycotoxin production: a potential source of botanical food preservative. Asian Pac J Trop Biomed 2014;4:S373–81.
- [10] Martins DF, Emer AA, Paula Batisti A, Donatello N, Mazzardo-Martins L, Venzke D, et al. Inhalation of *Cedrus atlantica* essential oil Alleviates pain bevavior through activation of descending pain modulation Pathways in a mouse model of postoperative pain. J Ethnopharmacol 2015;175:30–8.
- [11] Azizi A, Aghayan S, Zaker S, Shakeri M, Entezari N, Lawaf S. In vitro effect of Zingiber officinale extract on growth of Streptococcus mutans and Streptococcus sanguinis. Int J Dent 2015;2015:489842.
- [12] Boukhris M, Bouaziz M, Feki I, Jemai H, El FekiA, Sayadi S. Hypoglycemic and antioxidant effects of leaf essential oil of *Pelargonium graveolens* L'Hér. in alloxan induce diabetic rats. Lipids Health Dis 2012;11:81.
- [13] Bouzenna H, Krichen L. Pelargonium graveolens L'Hér. and Artemisia arborescens L. essential oils: chemical composition, antifungal activity against *Rhizoctonia solani* and insecticidal activity against *Rhysopertha dominica*. Nat Prod Res 2013;27:841–6.
- [14] Homa M, Fekete IP, Boszorményi A, Singh YR, Selvam KP, Shobana CS, et al. Antifungal effect of essential oils against *Fusarium keratitis* isolates. Planta Med 2015;81:1277–84.
- [15] Tomotake H, Koga T, Yamato M, Kassu A, Ota F. Antibacterial activity of *Citrus fruit* juices against *Vibrio species*. J Nutrit Sci Vitaminol 2006;52:157–60.
- [16] Auysawasdi N, Chuntranuluck S, Phasomkusolsil S, Keeratinijakal V. Improving the effectiveness of three essential oils against *Aedes aegypti (Linn.)* and *Anopheles dirus* (Peyton and Harrison). Parasitol Res 2016;115(1):99–106.
- [17] Pazinato R, Klauck V, Volpato A, Tonin AA, Santos RC, Souza ME, et al. Influence of tea tree oil (*Melaleuca alternifolia*) on the cattle tick *Rhipicephalus microplus*. Exp Appl Acarol 2014;63:77–83.
- [18] Boligon AA, Kubiça TK, Mario DB, Brum TF, Piana M, Weiblen R, et al. Antimicrobial and antiviral activity-guided fractionation from *Scutia buxifolia* Reissek extracts. Acta Physiol Plant 2013;35:2229–39.
- [19] Adams RP. Identification of essential oil components by Gas Chromatography/Mass spectroscopy. Illinois (USA): Allured Publishing Corporation; 1995. p. 456p.
- [20] Drummond RO, Ernst SE, Trevino JL, Gladney WJ, Graham OH. *Boophilus annulatus* and *Boophilus microplus*: Laboratory tests of insecticides. J Econ Entomol 1973;66:130–3.

- [21] Buzzati A, Sprenger LK, Kucharsky T, Molento MB. Ação do óleo de nim frente à teleóginas de *Rhipicephalus (Boophilus) microplus* em testes *in vitro*. Arch Vet Sci 2013;18:7–12.
- [22] Carrol JF, Tabanca N, Kramer M, Elejalde NM, Wedge DE, Bemier UR, et al. Essential oils of *Cupressus funebris, Juniperus communis*, and *J. chinensis (Cupressaceae)* as repellents against ticks (Acari: Ixodidae) and mosquitoes (Diptera: Culicidae) and as toxicants against mosquitoes. J Vector Ecol 2011;2: 258–68.
- [23] Dietrich G, Dolan MC, Peralta-Cruz J, Schimidt J, Piesman J, Eisen RJ, et al. Repellent activity of fractioned compounds from *Chamaecyparis nootkatensis* essential oil against nymphal *Ixodes scapularis* (Acari: Ixodidae). J Med Entomol 2006;43:957–61.
- [24] Dolan MC, Jordan RA, Schulze TL, Schulze CJ, Manning MC, Ruffolo D, et al. Ability of two natural products, nootkatone and carvacrol, to suppress *Ixodes scapularis* and *Amblyomma americanum* (Acari: Ixodidae) in a Lyme disease endemic area of New Jersey. J Econ Entomol 2009;102:2316–24.
- [25] Amer A, Mehlkorn H. Repellency effect of forty one essential oils against *Aedes, Anopheles, and Culex mosquitoes*. Parasitol Res 2006;99:478–90.
- [26] Agnolin CA, Olivo CJ, Parra CLC. Efeito do óleo de capim limão (*Cymbopogon flexuosus Stapf*) no controle do carrapato dos bovinos. Rev Bras Pl Med 2014;16:77–82.
- [27] Silva WW, Athayde ACR, Rodrigues OG, Araújo GMB, Santos VD, Neto ABS, et al. Efeitos do neem (*Azadirachta indica A. Juss*) e do capim santo [*Cymbopogon citratus* (DC) Stapf] sobre os parâmetros reprodutivos de fêmeas ingurgitadas d *Boophilus microplus* e *Rhipicephalus sanguineus* (Acari: Ixodidae) no semiárido paraibano. Rev Bras Pl Med 2007;9:1–5.
- [28] Tchoumbougnang F, Amvam Zollo P, Dagne E, Mekonnen Y. In vivo antimalarial activity of essential oils from Cymbopogon citratus and Ocimum gratissimum on mice infected with Plasmodium berghei. Plant Med 2005;71:20–3.
- [29] Makhaik M, Narayana SN, Tewary D. Evaluation of antimosquito properties of essential oils. J Sci Ind Res 2005;64:129–33.

- [30] Kumar R, Srivastava M, Dubey NK. Evaluation of *Cymbopogon martinii* oil extract for control of postharvest insect deterioration in cereals and legumes. J Food Protect 2007;70:172–8.
- [31] Duarte MCT, Figueira GM, Sartoratto A, Rehder VL, Delarmelina C. Anti-*Candida* activity of Brazilian medicinal plants. J Ethnopharmacol 2005;97:305–11.
- [32] Silva WC, Marins JRS, Souza EM, Heinzen H, Cesio MV, Mato M, et al. Toxicity of piper aduncum L. (Piperales: Piperaceae) from the Amazon Forest for the cattle tick *Rhipicephalus* (*Boophilus*) microplus (Acari: Ixodidae). Vet Parasitol 2009;164:267–74.
- [33] Nerio LS, Olivero-Verbe J, Stashenko E. Repellet activity of essential oils: a review. Biores Technol 2010;101:372–8.
- [34] Hanifah AL, Ming HT, Narainasamy VV, Yusoff AT. Laboratory evaluation of six crude plant extracts as repellents against larval *Leptotrombidium deliense* (Acari: Trombiculidae). Asian Pacif J Trop Biomed 2012;2:257–9.
- [35] Weldon PJ, Carroll JF, Kramer M, Bedoukian RH, Coleman RE, Bernier UR. Anointing chemicals and hematophagous arthropods: responses by ticks and mosquitoes to *Citrus* (Rutaceae) peel exudates and monoterpene components. J Chem Ecol 2011;34:348–59.
- [36] Slathia PS, Bhagat GR, Singh S, Kher SK, Paul N. Traditional knowledge on utility of *Cedrus deodara* (Roxb.) Loud in Doda district of Jammu province. Indian J Trad Know 2007;6:518–20.
- [37] Maske DK, Bhilegaonkar NG, Jangde CR. Treatment of tick infestation in cattle with pestoban. J Indian Indig Med 1996;17:81–3.
- [38] Tabanca N, Wang M, Avonto C, Chittiboyina AG, Parcher JF, Carroll JF, et al. Bioactivity-guided investigation of geranium essential oils as natural tick repellents. J Agric Food Chem 2013;61:4101–7.
- [39] Jaenson TG, Garboui S, Palsson K. Repellency of oils of lemon eucalyptus, geranium, and lavender and the mosquito repellent MyggA natural to *Ixodes ricinus* (Acari: Ixodidae) in the laboratory and field. J Med Entomol 2006;4:731–6.