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Sequence of abductin, the molluscan 'rubber' protein

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The inner hinge ligament of bivalve molluscs opposes the action of the adductor muscle. This arrangement permits the opening and closing of the shell. In scallops, the apparatus has evolved for swimming [1]. By opening and closing their shells about four times per second, scallops swim a few meters at a time to escape slow-moving predators, such as starfish. The major component of all inner hinge ligaments is the protein abductin [2]. The scallop ligament is composed almost entirely of protein and its relatively low extent of mineralization compared to the ligaments of other molluscs is thought to contribute to its high resilience (96% recovered work), which is extraordinary even by molluscan standards [3].

Here, we report amino acid sequences of abductin deduced from the sequences of cDNAs obtained from *Argopecten* (the bay scallop). Degenerate primers based on the sequences of peptides obtained from digested inner hinge ligament were used to amplify *Argopecten* genomic DNA. The PCR products were in turn used to screen an *Argopecten* cDNA library, which yielded five full length cDNAs, Ap4, Ap5, Ap7, Ap9 and Ap12. The 136 amino acid open reading frames of Ap4 and Ap7 were identical, whereas the other three (Ap5, 126 residues; Ap9, 131 residues; Ap12, 132 residues) were very closely related (Figure 1). Analysis by northern blotting and RNase protection showed that abductin transcripts are present only in the mantle tissue. RNase protection experiments were carried out on RNA from *Argopecten* eggs and various larval stages through to

young adults at day 35 [4]. Weak transcription was noticeable at day 15 (prodissoconch, stage 20, 0.19 mm) and a strong signal was seen at day 35 (pre-adult scallop, stage 25, 1.67 mm). Expression of abductin continued in the adult.

Electron microscopy and X-ray diffraction experiments indicate that the inner hinge ligament is acellular and amorphous [2]. The physical properties of the ligament suggest that its resilience is a result of changes in entropy during deformation [5] and that abductin is lightly cross-linked [6,7], although the nature of the cross-links is a conspicuous problem. The abductin sequences can be divided into two domains. The first is the alanine-rich amino terminus (residues 1–20), which also contains the two conserved cysteines (residues 6 and 10) that might be involved in intermolecular or intramolecular disulfide formation. This domain also contains two of the three conserved tyrosines (residues 4 and 20), which could also be involved in cross-linking by forming 3,3'-methylene bistyrosine [6]. The first domain, however, is likely to form a signal sequence for secretion: indeed, the agreement between the amino acid analysis of the inner hinge ligament and the composition predicted from the cDNA sequences is increased when the amino-terminal residues are not included.

The second domain comprises an extraordinary glycine/methionine-rich sequence that stretches from residue 21 to the carboxyl terminus. The methionines are distributed throughout this domain and are included in the consensus sequence GGFGGMGGGX (single-letter amino acid code), which is found with the strongest compliance in the carboxy-terminal half of the polypeptide. Interestingly, it has been reported that most of the methionine in the inner hinge ligament of surf clams is actually methionine sulfoxide [8], which would modestly increase the polarity

Figure 1

	10	20	30	40	50
Ap4/7	MNAYICLAAC	LIAAVSAAGY	GGGAGSMGGT	GGMGGGMNAG	GFGGMGGGMG
Ap5	MNAYICLSAC	LIAAVSAAGY	GGGAGSMGGT	GGMGGGMNAG	GFGGMGGGMG
Ap9	MNAYICLAAC	LIAAVSAAGY	GGGAGSMGGT	GGMGGGMNAG	GFGGMGG-MG
Ap12	MNAYICLAAC	LIAVSAAGY	GGGAGSMGGT	GGMGGGMNAG	GFGGITGGGMG
	60	70	80	90	100
	GKGGFGGIG	GFGGMGGGMG	GGPGGFGGMG	GFGGMGGGKG	GFGGMGSGMG
	GKGGFGGGMG	GFGGMGGGMG	GGPGGFGGMG	GFGGMGGGKG	GFGGMGSGMG
	GKGGFGGIG	GF---GGMG	GGPGGFGGMG	GFGGMAA-KG	GFGGMGSGMG
	GKGGFGGGMG	GCPGGFGGIG	GGSGFGGMG	GFGGMGGGKG	GFGGMGSSMG
	110	120	130		
	GFGGMGGGNA	GFGGMGGGNA	GFGGMGGQGG	FGGKGY	
	GFGGMGGGNA	GFGGMGG---	-----QGG	FGGKGY	
	GFGGMGGGNA	GFGGMGGGNA	GFGGMGGQGG	FGGKGY	
	GFGGMGGGNA	GFGGMGG---	-QSGMGGQSG	FGGKGY	

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Amino acid sequences of abductin deduced from the sequences of five full-length cDNAs obtained from *Argopecten*. The open reading frames of Ap4 and Ap7 were identical, whereas the other three cDNAs were very closely related and may represent alleles from the population used to construct the library. Freshly dissected inner hinge ligament from adult animals was ground in 1.0 M aqueous acetic acid to extract traces of calcium carbonate. The insoluble residue was dried and subjected to amino acid analysis, which revealed an unusual composition rich in glycine (57.3% of residues) and with a high level of methionine (14.3% of residues), as noted previously [2,3]. The extracted tissue was also digested with 15% (w/v) CNBr in 70% formic acid at room temperature for up to 48 h or with 0.1 N HCl at 95°C for 3 h. The released peptides were separated by reverse-phase HPLC and subjected to

automated Edman degradation and MALDI-MS. The following sequences were obtained: NAGGFGGIGG (CNBr), GGGPGGFGGIG GSGGFGG(M) (CNBr), GGGLGGFGGI GGFGG(M) (CNBr), GFGG (HCl) and GFGGMGG (HCl), single-letter amino acid code where (M) is homoserine lactone. Degenerate primers based on the peptide sequences were used to amplify *Argopecten* genomic DNA. Two PCR products, ApG1 and ApG17, were used to screen an *Argopecten* oligo(dT)-primed cDNA library made from young animals (3–8 mm) in which the ligament is growing rapidly. The abductin sequences contained the Edman sequences, with the exception of GGGLGGFGGIGGGF G(M), which gave a single mismatch. The cDNA sequences are available through GenBank: accession numbers AFO26845–AFO26848 inclusive.

of the polypeptide. The high glycine content is in keeping with a lack of secondary structure, although the well conserved repeat argues that 'pattern implies structure' [9], as in the case of mammalian elastin, which also has a repetitive sequence albeit different from that of abductin [10]. Our sequence data therefore call for a re-examination of the structure of the inner hinge ligament using biophysical techniques. The second domain is punctuated by lysine residues at positions 53 and 89. The third conserved lysine is two residues from the carboxyl terminus. Finally, the third conserved tyrosine is at the extreme carboxyl terminus. If the first domain is removed by a signal

peptidase, these remaining lysine and tyrosine residues are probable candidates for sites of cross-linking.

Other proteins secreted by the mantle epithelium differ in composition from abductin. For example, shell proteins involved in mineralization are rich in acidic residues [11,12], whereas proteins of the periostracum (the outer surface of the shell) are rich in tyrosine [11,13]. Interestingly, two recently sequenced shell proteins contain long glycine-rich stretches of amino acids, but the characteristic abductin repeat is absent and the methionine content is low [12].

If the nature of the cross-links in abductin can be resolved, the

recombinant abductin protein could be converted into resilient materials with applications in biotechnology. Hybrid materials incorporating abductin sequences are likely to be more useful than materials that simply mimic the ligament.

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