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The Use of Tevagrastim (Biosimilar Filgrastim XMO2) for Hematopoietic Stem Cell Mobilization in HLA Matched Sibling Donors for Allogeneic Stem Cell Transplantation to AML/MDS Patients

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Introduction: G-CSF Filgrastim is widely used for the mobilization of CD34⁺ HSC. The experience with biosimilar G-CSF agents is limited. We initiated a prospective study assessing Tevagrastim (biosimilar Filgrastim XMO2) for mobilization of CD34⁺ PB HSC in MRD for alloSCT in 24 pts with AML/MDS (NCT01542944).

Materials and methods: The study was approved by the National Regulatory Authorities and both patients and donors signed an informed consent. The donors, median age 46 years (range, 25–64), F- 14; M- 10 received Tevagrastim (10 μ g/kg) BW s.c. BID for 4 days. On the morning of the 5th day they underwent conventional leukapheresis. The target yields of CD34 cell was 5 × 10⁶ CD34⁺ cells/kg BM of the recipient. The conditioning was myeloablative Bu/Cy (n=10), reduced toxicity Flu/Treo (n=7), Flu/Bu4 (n=3) or RIC Flu/Bu2 (n=4).

Results: Efficacy: 77-1982 × 10⁶ (median 749 × 10⁶) CD34⁺ were collected. The number of CD34⁺ cell per kg BW of the pts was 0.93-35.4 × 10⁶ (median 10.2 × 10⁶). Collections contained 144-709 × 10⁸ (median 299 × 10⁸) CD3⁺ T-cells, 1.74-11.6 × 10⁸ (median 4.4 × 10⁸) per kg BW of the pts and 0.3-11.2x10⁷/kg (median 2.3x10⁷/kg) CD3⁻ CD56⁺ CD16⁺ NK cells. The mean number of leukapheresis procedures was 1.3. Engraftment was: ANC >0.5× 10⁹/L and >1× 10⁹/L within a median of 13 days (range, 10–21) and 13.5 days (range, 10–22), respectively. PLT reached counts of >20× 10⁹/L and >50× 10⁹/L within a median of 16 (range, 12–33) and 17 (range, 12–33) days, respectively. The median days of isolation was 10 (range, 6-21). The median number of PC and PLT transfusions was 5 (range, 2-20) and 21 (range, 0-180), respectively. 20/24 (83.3%) pts showed full donor chimerism at 1 month after transplantation, respectively.

Safety: 12/24 donors reported transient arthralgias and 2 developed flu-like syndrome. The main side effects were mucositis (n-15, grade II -9, grade III-IV- 6), infections (n-20)

Bulk CTL after exposed to

and fluid retention (n-8). One pt suffered from VOD (Grade-I) and 5 pts developed aGVHD (grade II-III) which responded to conventional therapy. In total TRM was 1/24 at d 100. Total 5 pts died from leukemia progression during median 7 (range, 1-16) months; 2 of them died before d 100. 4 pts suffered from chronic, mild GVHD.

Conclusions: Our study with 24 AML/MDS pts, indicates that the G-CSF biosimilar XM02, Tevagrastim is safe and efficient for stem cells mobilization in MRD. The CD34 yield and post transplantation engraftment are similar to those achieved with the human recombinant G-CSF Filgrastim. We have not seen significant differences in the graft CD34⁺, CD3⁺ T and CD16⁺ NK cell count, the number of leukapheresis procedures and the regeneration of WBC, neutrophils and PLTs in comparison with our historical controls. All patients promptly engrafted, and the donors developed only expected side effects. Neither graft rejection nor side effects occurred more frequently than expected from the standard G-CSF.

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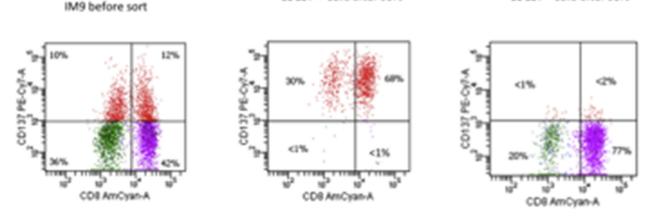
CD137 Expression Identifies Leukemia Specific CTL after in Vitro Priming of Cord Blood T Cells Previously Expanded By CD3/CD28 Co-Stimulation

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Introduction: We previously reported that cord blood T-cells (CBT), obtained from <3% of cord blood graft can be expanded, and primed against (lymphoid and myeloid leukemia cell lines in the presence of IL12, IL7 and IL15 (*Cancer Research*, 2010; 70(13): 5249). Recently, we demonstrated that IL15 alone can support the priming and expansion of CTL.

Hypothesis: CD137 (4-1BB) is a member of the TNFR- family, expressed on activated T cells, which augment the survival and proliferation. Here we tested the hypothesis that denovo CD137 expression may identify leukemia-specific CTL. **Methods:** IM9-specific CTL was generated from pre-expanded CBT cells using weekly stimulations with irradiated targets and IL15. After 3 weeks, cytotoxicity was tested, re-stimulated to sort CD137 expressing cells (BD Biosciences) and were re-tested for cytotoxicity and TCR β CDR3 spectratyping was done to identify their clonality. RT² profilerTM PCR array human Th1 & Th2 responses (SABiosciences) were employed to identify the differential gene expression.

CD137 - cells after sort



CD137 + cells after sort

Figure 1. Representative dot plot showing the expression of CD137 on CTL before and after sort