

Uremic toxins modulate the spontaneous apoptotic cell death and essential functions of neutrophils

GERALD COHEN, MICHAEL RUDNICKI, and WALTER H. HÖRL

Division of Nephrology, Department of Medicine, University of Vienna, Vienna, Austria

Uremic toxins modulate the spontaneous apoptotic cell death and essential functions of neutrophils. Clearance of neutrophils via apoptosis from the site of infection is crucial for the coordinated resolution of inflammation. The balance between stimulating and attenuating as well as between pro- and anti-apoptotic factors is necessary for maintenance of an effective immune response without the harmful side effects of neutrophil action. This article describes the effect of glucose-modified serum proteins and of free immunoglobulin light chains (IgLCs) on neutrophil functions and apoptosis. Both groups of proteins are found at elevated levels in sera of uremic patients. Glucose-modified proteins increase both the chemotactic movement of neutrophils and the activation of glucose uptake. Spontaneous neutrophil apoptosis is increased in the presence of these modified serum proteins. On the other hand, the presence of free IgLCs, previously shown to diminish neutrophil chemotaxis and the activation of glucose uptake, increase the percentage of viable neutrophils by inhibiting spontaneous apoptotic cell death. We conclude that both glucose-modified proteins and free IgLCs can be considered to be uremic toxins and both contribute to the disturbed immune function in uremic patients. Their concentrations as well as the microenvironment in which they are acting seem to be important for their actual effects.

Bacterial infections still represent a main cause for the increased morbidity and mortality among uremic patients [1, 2], mainly as a result of the altered functions of neutrophils [3]. Neutrophils are cells of the first-line unspecific immune defense. After the chemotactic movement to the site of infection, their main job is to destroy invading microorganisms after phagocytosis by secretion of proteolytic enzymes and the generation of reactive oxygen metabolites during the oxidative burst. Any modulation of one of these essential functions will lead to an increased risk for bacterial infections. Neutrophils isolated from uremic patients show functional changes such as a reduced chemotactic activity [4], a lower cellular response to phagocytic stimuli [5] and a diminished oxidative metabolism leading to disturbed intracellular killing [6]. This contributes to the higher risk of bacterial

infections. Factors responsible for the altered neutrophil functions have been described in the literature: anemia, malnutrition, iron overload, zinc deficiency, increased levels of intracellular calcium, and hemodialysis treatment per se [7]. Within the last few years the important role of uremic toxins in contributing to the diminished immune function in uremia has been recognized. A number of factors from hemodialysis ultrafiltrate and continuous ambulatory peritoneal dialysis (CAPD) effluents has already been isolated and characterized [8–14].

Killing of invading microbes is a precondition for a successful immune response. However, the discharge of neutrophil toxic products into the extracellular space leads to a prolonged inflammation and as a consequence to the destruction of the surrounding tissue [15]. Therefore, for a normal resolution of inflammation the clearance of neutrophils by macrophages from the site of infection is highly important when their plasma membrane is still intact. Recently, the importance of apoptosis, that is, programmed cell death, of neutrophils has been recognized. Human neutrophils have a short half-life and are already programmed to die via apoptosis when they enter circulation [16]. Apoptotic neutrophils that still have an undamaged cell membrane are recognized and taken up by macrophages and by semiprofessional phagocytes such as glomerular mesangial cells without the release of pro-inflammatory cytokines [17–19]. Apoptosis is an active process characterized by chromatin condensation, DNA cleavage at internucleosomal sites and loss of membrane asymmetry but not integrity. On the other hand, necrosis, the other main form of cell death, is a passive degenerative phenomenon characterized by the loss of the integrity of the cell membrane without changes in the morphology of the nuclei. The balance between neutrophil apoptosis and necrosis plays a role in the control of inflammation, and is important to avoid the development of inflammatory diseases [20].

The rate of neutrophil apoptosis seems to be regulated by death factors as well as by survival factors. Despite the importance of functioning neutrophil apoptosis at the site of inflammation, a general increase in neutrophil

Key words: immunoglobulin light chains, glucose uptake, uremia, bacterial infection, immune defence, chemotactic activity.

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Table 1. Effect of glucose-modified proteins on neutrophil chemotaxis (relative distance migrated) and relative activation of glucose uptake in the presence of buffer (Co), unmodified serum proteins (S-), serum proteins modified with glucose alone (S+) or with glucose in the presence of aminoguanidine (S+ AG), and in the presence of glycated proteins isolated from CAPD effluents using boronate affinity chromatography (boronate-binding proteins, BBP)

	Co	S-	S+	S+ AG	BBP
Chemotaxis	100 ± 0	107 ± 6	130 ± 11	101 ± 9	ND
<i>P</i>			= 0.01 vs. S-	= 0.01 vs. S+	
Glucose uptake	100 ± 0	99 ± 13	135 ± 13	144 ± 23	164 ± 7
<i>P</i>			<0.005 vs. S-	<0.01 vs. S-	<0.005 vs. S-

apoptosis will lead to a disturbed immune response. Recently, it could be demonstrated that neutrophils obtained from uremic patients undergo accelerated apoptosis under in vitro conditions [21] and that constitutive cellular factors contribute to this effect. On the other hand, uremic plasma has also been shown to accelerate neutrophil apoptosis [21].

Unlike for essential neutrophil functions, no uremic toxin with the potential to modulate neutrophil apoptosis has been described so far. Recently, we identified factors that accumulate in uremic sera, modulate essential neutrophil functions and have a significant influence on the spontaneous apoptotic cell death of neutrophils (G. Cohen et al, manuscript submitted for publication), namely glucose-modified proteins and free immunoglobulin light chains (IgLCs).

EFFECTS OF GLYCATED SERUM PROTEINS ON NEUTROPHIL FUNCTIONS

Serum proteins obtained from healthy donors were modified with glucose in vitro. Furthermore, we wanted to investigate whether early or late glycation products are responsible for the effects of the modified proteins. As the compound aminoguanidine inhibits the advanced glycation end product (AGE) formation [22], but does not prevent the formation the early glycation products, we incubated serum proteins in the presence of glucose with and without the addition of aminoguanidine to obtain early and a mixture of early and late glycation products, respectively.

The effect of the modified serum proteins on the chemotactic movement of neutrophils was tested using the under-agarose method. Neutrophil chemotaxis was higher in the presence of modified serum proteins (S+) as compared to the unmodified control samples (S-; Table 1). This effect was not observed when serum proteins modified with glucose were added in the presence of aminoguanidine (S+ AG; Table 1). Therefore, late glycation products are responsible for the stimulatory effect on neutrophil chemotaxis.

The uptake and accumulation of 2-deoxy-D-[1-³H]-glucose serves as a quantitative measurement of the state of activation of phagocytic cells [23]. The relative activa-

tion of the glucose uptake was significantly higher in the presence of the glucose-modified serum proteins (S+) than in the presence of unmodified proteins (S-; Table 1). Early and not late glycation products are responsible for this effect, because serum proteins incubated in the presence of glucose and aminoguanidine (S+ AG) showed the same effect on the activation of glucose uptake as proteins modified with glucose alone (Table 1).

EFFECTS OF GLYCATED SERUM PROTEINS ON SPONTANEOUS APOPTOTIC CELL DEATH OF NEUTROPHILS

We investigated the effect of the glucose-modified proteins on spontaneous neutrophil apoptosis by detecting the characteristic morphological changes and characteristic DNA-strand breaks. Apoptotic neutrophils are removed by phagocytes under in vivo conditions. Therefore, we did not distinguish between apoptotic and secondary necrotic cells, and show the percentage of viable cells in the results of the apoptosis assays. As shown in Table 2, neutrophils die faster in the presence of glucose-modified proteins as compared to the controls. We also used individual modified proteins, such as human serum albumin, in our apoptosis assays and could confirm the results obtained with whole serum proteins (Table 2). As observed for the effect on the uptake of glucose, early and not late glycation products are responsible for the effects observed, because proteins incubated in the presence of aminoguanidine show the same effect on spontaneous neutrophil apoptosis as proteins incubated in the presence of glucose alone (Table 2). There are high expectations towards aminoguanidine in terms of ameliorating AGE-related disturbances such as diabetes associated vascular hypertrophy [24]. However, the results from our in vitro experiments suggest that aminoguanidine is not able to prevent the increased neutrophil apoptosis caused by glycated proteins.

Our findings are also relevant for the status of the immune system in diabetic patients with increased plasma AGE concentrations. Interestingly, it has been suggested that AGEs play an important role in the development of microvascular complications in diabetes mellitus by inducing apoptosis and increasing pro-coagulant activity

Table 2. Effect of glucose modified proteins on neutrophil apoptosis

	Co	S-	S+	S+ AG	A-	A+	A+ AG	BBPs	S
% Viable neutrophils	32 ± 3	51 ± 4	44 ± 4	45 ± 4	42 ± 7	34 ± 5	32 ± 3	43 ± 3	65 ± 3
<i>P</i>			<0.005 vs. S-	<0.05 vs. S-		<0.05 vs. A-	<0.05 vs. A-	<0.0005 vs. S	

Percentage of viable cells after a 20-hour incubation in the presence of: buffer (Co), serum proteins (S-) or human serum albumin (A-) incubated without glucose; serum proteins (S+) or albumin (A+) modified with glucose alone or with glucose in the presence of aminoguanidine (S+ AG, A+ AG); glycated proteins isolated from CAPD effluents (boronate binding proteins, BBPs) and unmodified serum protein(s).

in umbilical vein endothelial cells [25]. Furthermore, it has been demonstrated that in diabetic patients there is a neutrophil activation and that this activation is correlated to glycated hemoglobin level [26].

As no AGE-binding proteins have been described for neutrophils to date, the mechanism of the effect of glycated proteins still remains to be elucidated. However, receptors for AGEs have been detected on monocytes [27] and endothelial cells [28] where they play a role in the removal of glycated proteins and in changes of gene expression [29]. For example, fluorescent AGE- β 2-microglobulin but not unmodified β 2-microglobulin exhibits biological activity with monocytes/macrophages [30].

Our results are consistent with the finding of Cendoroglo et al that neutrophil apoptosis is increased in uremia [21]. The same authors found diminished functions of apoptotic neutrophils. However, we investigated the immediate effect of protein samples on normal neutrophils, whereas Cendoroglo et al determined the functions of neutrophils that are already undergoing apoptotic cell death. They concluded that uremia in neutrophils enhances some functional activities such as oxidative metabolism at an early phase leading to functional impairment of neutrophils in a later phase [21]. This is also consistent with the observation that the dysregulation of the immune system in uremia is characterized by immune deficiency, even though the immune competent cells exist in a state of activation [31]. Another group of modified proteins, advanced oxidation protein products (AOPP), has a close correlation to AGEs [32]. Both AOPP and AGEs have significantly elevated plasma levels in uremic patients and both protein modifications confer features on proteins that lead to the stimulation of the oxidative burst of human monocytes in vitro [32]. It has also been shown that neutrophils from uremic patients are primed for an enhanced oxidative metabolism [33], and that this primed state could not be changed by hemodialysis or continuous ambulatory peritoneal dialysis (CAPD) treatment [34].

EFFECTS OF GLYCATED PROTEINS ISOLATED FROM UREMIC PATIENTS ON NEUTROPHIL FUNCTION AND SURVIVAL

We showed that the observed effect of glycated proteins is not restricted to in vitro modified samples, as glucose-modified proteins isolated from the effluent of CAPD

patients increase the relative activation of glucose uptake significantly (Table 1). Furthermore, we demonstrated that glucose-modified proteins that were isolated from CAPD effluents significantly reduce the number of surviving polymorphonuclear leukocytes (PMNL) as compared to the controls (Table 2). As it has been shown that the presence of proteins per se is able to modulate PMNL apoptosis, serum proteins of healthy donors at the same final concentration have been used as controls in this set of experiments.

INFLUENCE OF FREE IGLCs ON NEUTROPHIL FUNCTIONS

Plasma levels of free Ig light chains are elevated in patients with impaired kidney function. A significant increase in the levels of both κ and λ chains in anephric patients has been described by Solling [35]. Wakasugi et al reported on the increase in the concentration of free Ig light chains in sera after the start of hemodialysis therapy [36]. Using a newly developed assay based on SDS-PAGE, electrotransfer onto nitrocellulose membranes, and chemiluminescence detection, our group found increased levels of Ig light chains in uremic predialysis patients as well as in chronic renal failure patients undergoing hemodialysis treatment (Cohen et al, manuscript submitted). Furthermore, we showed that the light chain levels could not be significantly decreased by either low-flux or high-flux dialyzer membranes. We previously showed that free IGLCs isolated from hemodialysis and CAPD patients as well as commercially available Bence Jones proteins significantly inhibit chemotaxis and glucose uptake of neutrophils [10]. We conclude that free IGLCs are at least partly responsible for the diminished unspecific immune defense and consequently for a higher risk of infection in uremia.

INFLUENCE OF FREE IGLCs ON NEUTROPHIL APOPTOSIS

Using the same assays as described above for glucose-modified proteins, we could show that Ig light chains of both κ - and λ -type increase the percentage of viable

Table 3. Effect of free immunoglobulin light chains (IgLCs) on neutrophil apoptosis

Specific antibody	Co	κ	λ	A
Absent	12 \pm 1	38 \pm 8*	36 \pm 7*	21 \pm 3
Present	ND	12 \pm 4	17 \pm 3	17 \pm 6

Percentage of viable cells after a 20-hour incubation in the presence of buffer (Co), IgLCs of κ - or of λ -type and of human serum albumin (A) in the absence or presence of the respective specific antibody. * $P < 0.05$ versus Co.

neutrophils by diminishing apoptotic cell death (Table 3) in a concentration dependent manner (G. Cohen et al, manuscript submitted for publication). The effect of the IgLCs was specific as demonstrated by the fact that this effect can be abolished by specific antibodies against κ and λ light chains, respectively (Table 3). The presence of human serum per se is able to decrease neutrophil apoptosis [5]. However, there are three lines of evidence showing that our results are not solely based on a protein effect, but are specific for Ig light chains: (1) the addition of antibodies should increase and not abolish a protein effect; (2) the Ig light chain concentration used for the apoptosis experiments is much lower than the concentration of serum proteins leading to the same percentage of surviving neutrophils; and (3) in the presence of excess serum proteins κ and λ light chains inhibit neutrophil apoptosis as well, whereas albumin exerts no significant effect under the same experimental setup (G. Cohen et al, submitted for publication).

As discussed above, free IgLCs diminish essential neutrophil functions. This effect is at least partly caused by a prestimulation of neutrophils, for example an increased basal level of glucose uptake [10]. It is likely that IgLCs are able to contribute to the chronic state of inflammation found in end-stage renal disease patients [31] if they are also able to prolong the life of these pre-activated neutrophils that have shown to play an important role in tissue injury and the development of renal failure [15].

It is important to keep in mind that there seems to be a physiological balance between anti-apoptotic and counteracting pro-apoptotic factors, and that the micro-environment in which they are acting has to be considered. Freely circulating IgLCs, for example, will contribute to the fate of neutrophils in a different way than accumulated light chains as observed in the light chain deposition disease. Furthermore, it has been shown that neutrophils that transmigrated through endothelial monolayers have altered functional properties and have a different tendency to undergo apoptosis [37]. Reduced [38, 39] as well as increased neutrophil apoptosis has been described for PMNL after the migration across an endothelial monolayer. It also has been shown that extracellular matrix proteins regulate local inflammations by influencing apoptotic cell death in tumor necrosis factor- α

(TNF α)-activated neutrophils [40]. On the other hand it has been described that neutrophils are protected against apoptosis in circulation by red blood cells acting as scavengers of apoptosis promoting H₂O₂ [41]. In vitro experiments demonstrated that the extracellular pH modulates the rate of neutrophil apoptosis, alkaline conditions accelerating [42] and extracellular acidosis depressing [43] apoptosis.

In conclusion, uremic toxins accumulating in the plasma of patients with renal failure not only affect essential neutrophil functions and thereby the unspecific immune response, but also influence neutrophil survival by modulating the rate of apoptotic cell death. Identifying factors influencing both function and apoptosis of neutrophils is a first step in understanding the causes for the deranged immune system in uremia.

Reprint requests to Walter H. Hörl, M.D., Ph.D., FRCP, Department of Medicine, Medizinische Universitätsklinik III, Währinger Gürtel 18-20, A-1090 Wien, Austria.

REFERENCES

1. MAILLOUX LU, BELLUCCI AG, WILKES BM, NAPOLITANO B, MOSSEY RT, LESSER M, BLUESTONE PA: Mortality in dialysis patients: Analysis of the causes of death. *Am J Kidney Dis* 18:326-335, 1991
2. VANHOLDER R, VAN LOO A, DHONDT AM, DESMET R, RINGOIR S: Influence of uraemia and haemodialysis on host defence and infection. *Nephrol Dial Transplant* 11:593-598, 1996
3. HAAG-WEBER M, HÖRL WH: Dysfunction of polymorphonuclear leukocytes in uremia. *Semin Nephrol* 16:192-201, 1996
4. SIRIWATRATANANONTA P, SINSAKUL V, STERN K, SLAVIN RG: Defective chemotaxis in uremia. *J Lab Clin Med* 92:402-407, 1978
5. CANNISTRA SA, GRIFFIN JD: Regulation of the production and function of granulocytes and monocytes. *Semin Hematol* 25:173-188, 1988
6. RITCHEY EE, WALLIN JD, SHAH SV: Chemiluminescence and superoxide anion production by leukocytes from hemodialysis patients. *Kidney Int* 19:349-358, 1981
7. COHEN G, HAAG-WEBER M, HÖRL WH: Immune dysfunction in uremia. *Kidney Int* 52(Suppl 62):S79-S82, 1997
8. HÖRL WH, HAAG-WEBER M, GEORGOPOULOS A, BLOCK LH: Physicochemical characterization of a polypeptide present in uremic serum that inhibits the biological activity of polymorphonuclear cells. *Proc Natl Acad Sci USA* 87:6353-6357, 1990
9. HAAG-WEBER M, MAI B, HÖRL WH: Isolation of a granulocyte inhibitory protein from uraemic patients with homology of β_2 -microglobulin. *Nephrol Dial Transplant* 9:382-388, 1994
10. COHEN G, HAAG-WEBER M, MAI B, DEICHER R, HÖRL WH: Effect of immunoglobulin light chains from hemodialysis and continuous ambulatory peritoneal dialysis patients on polymorphonuclear leukocyte functions. *J Am Soc Nephrol* 6:1592-1599, 1995
11. TSCHESCHE H, KOPP C, HÖRL WH, HEMPELMANN U: Inhibition of degranulation of polymorphonuclear leukocytes by angiogenin and its tryptic fragment. *J Biol Chem* 269:30274-30280, 1994
12. BALKE N, HOLTkamp U, HÖRL WH, TSCHESCHE H: Inhibition of degranulation of human polymorphonuclear leukocytes by complement factor D. *FEBS Lett* 371:300-302, 1995
13. COHEN G, RUDNICKI M, HÖRL WH: Isolation of modified ubiquitin as a neutrophil chemotaxis inhibitor from uremic patients. *J Am Soc Nephrol* 9:451-456, 1998
14. VANHOLDER R, DE SMET R, WAATERLOOS MA, VAN LANDSCHOOT N, VOGELEERE P, HOSTE E, RINGOIR S: Mechanisms of uremic inhibition of phagocyte reactive species production. Characterization of the role of p-cresol. *Kidney Int* 47:510-517, 1995
15. HEINZELMANN M, MERCER-JONES MA, PASSMORE JC: Neutrophils and renal failure. *Am J Kidney Dis* 34:384-399, 1999

16. SENDO F, TSUCHIDA H, TAKEDA Y, GON S, TAKEI H, KATO T, HACHIYA O, WATANABE H: Regulation of neutrophil apoptosis—its biological significance in inflammation and the immune response. *Hum Cell* 9:215–222, 1996
17. SAVILL JS, WYLLIE AH, HENSON JE, WALPORT MJ, HENSON PM, HASLETT C: Macrophage phagocytosis of aging neutrophils in inflammation. Programmed cell death in the neutrophil leads to its recognition by macrophages. *J Clin Invest* 83:865–875, 1989
18. SAVILL J, FADOK V, HENSON P, HASLETT C: Phagocyte recognition of cells undergoing apoptosis. *Immunol Today* 14:131–136, 1993
19. HUGHES J, LIU Y, VAN DAMME J, SAVILL J: Human glomerular mesangial cell phagocytosis of apoptotic neutrophils: Mediation by a novel CD36-independent vitronectin receptor/thrombospondin recognition mechanism that is uncoupled from chemokine secretion. *Immunology* 158:4387–4397, 1997
20. HASLETT C: Resolution of acute inflammation and the role of apoptosis in the tissue fate of granulocytes. *Clin Sci* 83:639–648, 1992
21. CENDOROGLU M, JABER BL, BALAKRISHNAN VS, PERIANAYAGAM M, KING AJ, PEREIRA BJG: Neutrophil apoptosis and dysfunction in uremia. *J Am Soc Nephrol* 10:93–100, 1999
22. MIWA I, TSUGAWA T, KOYASU K, TERADA Y: Inhibition of advanced protein glycation by 8-quinolinecarboxylic hydrazide. *Pharmacology* 52:314–320, 1996
23. SEOW WK, SMITH SE, MCCORMACK JG, THONG YH: Uptake of ³H-deoxyglucose as a microassay of human neutrophil and monocyte activation. *J Immunol Methods* 98:113–118, 1987
24. RUMBLE JR, COOPER ME, SOULIS T, COX A, WU L, YOUSSEF S, JASIK M, JERUMS G, GILBERT RE: Vascular hypertrophy in experimental diabetes: Role of advanced glycation end products. *J Clin Invest* 99:1016–1027, 1997
25. MIN C, KANG E, YU SH, SHINN SH, KIM YS: Advanced glycation end products induce apoptosis and procoagulant activity in cultured human umbilical vein endothelial cells. *Diabetes Res Clin Pract* 46:197–202, 1999
26. PARLAPIANO C, DANESE C, MARANGI M, CAMPANA E, PANTONE P, GIOVANNIELLO T, ZAVATTARO E, SANGUINI S: The relationship between glycated hemoglobin and polymorphonuclear leukocyte leukotriene B4 release in people with diabetes mellitus. *Diabetes Res Clin Pract* 46:43–45, 1999
27. SCHMIDT AM, YAN S-D, BRETT J, MORA R, NOWYGRAD R, STERN D: Regulation of human mononuclear phagocyte migration by cell surface-binding proteins for advanced glycation end products. *J Clin Invest* 92:2155–2168, 1993
28. SCHMIDT AM, MORA R, CAO R, YAN SD, BRETT J, RAMAKRISHNAN R, TSANG TC, SIMIONESCU M, STERN D: The endothelial cell binding site for advanced glycation end products consists of a complex: An integral membrane protein and a lactoferrin-like polypeptide. *J Biol Chem* 269:9882–9888, 1994
29. SCHMIDT AM, HASU M, POPOV D, ZHANG JH, CHEN J, YAN SD, BRETT J, CAO R, KUWABARA K, COSTACHE G, SIMIONESCU N, SIMIONESCU M, STERN D: Receptor for advanced glycation end products (AGEs) has a central role in vessel wall interactions and gene activation in response to circulating AGE proteins. *Proc Natl Acad Sci USA* 91:8807–8811, 1994
30. MIYATA T, IIDA Y, UEDA Y, SHINZATO T, SEO H, MONNIER VM, MAEDA K, WADA Y: Monocyte/macrophage response to β 2-microglobulin modified with advanced glycation end products. *Kidney Int* 49:538–550, 1996
31. DESCAMPS-LATSCHA B: The immune system in end-stage renal disease. *Curr Opin Nephrol Hypertens* 2:883–891, 1993
32. WITKO-SARAT V, FRIEDLANDER M, NGUYEN-KHOA T, CAPELLÈRE-BLANDIN C, NGUYEN AT, CANTELOUP S, DAYER J-M, JUNGERS P, DRÛÈKE T, DESCAMPS-LATSCHA B: Advanced oxidation protein products as novel mediators of inflammation and monocyte activation in chronic renal failure. *J Immunol* 161:2524–2532, 1998
33. WARD RA, MCLEISH KR: Polymorphonuclear leukocyte oxidative burst is enhanced in patients with chronic renal insufficiency. *J Am Soc Nephrol* 5:1697–1702, 1995
34. KLEIN JB, MCLEISH KR, WARD RA: Transplantation, not dialysis, corrects azotemia-dependent priming of the neutrophil oxidative burst. *Am J Kidney Dis* 33:483–491, 1999
35. SOLLING K: Free light chains of immunoglobulins. *Scand J Clin Lab Invest* 157(Suppl):1–83, 1981
36. WAKASUGI K, SASAKI M, SUZUKI M, AZUMA N, NOBUTO T: Increased concentrations of free light chain lambda in sera from chronic hemodialysis patients. *Biomater Artif Cells Immobilization Biotechnol* 19:97–109, 1991
37. WATSON RW, ROTSTEIN OD, NATHENS AB, PARODO J, MARSHALL JC: Neutrophil apoptosis is modulated by endothelial transmigration and adhesion molecule engagement. *J Immunol* 158:945–953, 1997
38. SEELY AJ, SWARTZ DE, GIANNIAS B, CHRISTOU NV: Reduction in neutrophil cell surface expression of tumor necrosis factor receptors but not Fas after transmigration: Implications for the regulation of neutrophil apoptosis. *Arch Surg* 133:1305–1310, 1998
39. WATSON RW, ROTSTEIN OD, PARODO J, JIMENEZ M, SORIC I, BITAR R, MARSHALL JC: Impaired apoptotic death signaling in inflammatory lung neutrophils is associated with decreased expression of interleukin-1 beta converting enzyme family proteases (caspases). *Surgery* 122:163–172, 1997
40. KETTRITZ R, XU Y-X, KERREN T, QUASS P, KLEIN JB, LUFT FC, HALLER H: Extracellular matrix regulates apoptosis in human neutrophils. *Kidney Int* 55:562–571, 1999
41. AOSHIBA K, NAKAJIMA Y, YASUI S, TAMAOKI J, NAGAI A: Red blood cells inhibit apoptosis of human neutrophils. *Blood* 93:4006–4010, 1999
42. LEBLEBICIOGLU B, WALTERS J: Alkaline conditions accelerate polymorphonuclear leukocyte apoptosis in vitro. *Infect Immunol* 67:2019–2021, 1999
43. TREVANI AS, ANDONEGUI G, GIORDANO M, LOPEZ DH, GAMBERALE R, MINUCCI F, GEFFNER JR: Extracellular acidification induces human neutrophil activation. *J Immunol* 162:4849–4857, 1999