

# Inflammatory Effect of Intradermal Administration of Soluble Phospholipase A<sub>2</sub> in Rabbits

Waldemar Pruzanski, M.D., F.R.C.P.(C), Peter Vadaš, M.D., Ph.D.,  
and Victor Fornasier, M.D., F.R.C.P.(C)

Immunology Diagnostic and Research Centre and Department of Pathology, University of Toronto, The Wellesley Hospital, Toronto, Ontario, Canada

Extracellular phospholipase A<sub>2</sub> (PLA<sub>2</sub>) has been found in association with inflamed sites in experimental animals and in humans. The tissue effects of soluble PLA<sub>2</sub> have not been defined. We studied the development of inflammatory changes in rabbit skin subsequent to intradermal injection of active and inactivated venom and pancreatic PLA<sub>2</sub>, over a broad concentration range. PLA<sub>2</sub>, at concentrations en-

countered in human disease, caused acute inflammatory changes characterized grossly by erythema and induration, and histologically by inflammatory cell infiltration, vascular and tissue damage, and abscess formation. Extracellular PLA<sub>2</sub> may be considered as one of the pathogenic factors in inflammatory reaction. *J Invest Dermatol* 86:380-383, 1986

**P**hagocytic cells, both mononuclear and polymorphonuclear, respond to specific stimuli with the extracellular release of lysosomal enzyme phospholipase A<sub>2</sub> (PLA<sub>2</sub>) [1-5]. Extracellular PLA<sub>2</sub> has been identified in experimental delayed-type hypersensitivity [4], allergic uveitis [6], glycogen-induced peritonitis [7], and endotoxin shock [8]. Inflammatory diseases in humans, such as acute pancreatitis [9], gram-negative septic shock [10], and rheumatoid arthritis [11,12], have been associated with high extracellular PLA<sub>2</sub> activity. It was proposed that lysosomal PLA<sub>2</sub>, released into the extracellular milieu, cleaves phospholipids of contiguous plasma membranes, giving rise to strongly proinflammatory fatty acids and lysophosphatides [4,11].

In fact, high levels of PLA<sub>2</sub> were documented in lesion-free epidermis of patients with psoriasis [13,14]. The highest specific activity of PLA<sub>2</sub> was noted in patients with pustular psoriasis, lesions of which are characterized by massive polymorphonuclear leukocyte infiltration [14].

Enhanced PLA<sub>2</sub> activities in both rheumatoid arthritis and psoriasis may be the result of autoimmune deregulation of this enzyme [12,14], allowing for the increased production of proinflammatory products. However, the proinflammatory effect of PLA<sub>2</sub> has been demonstrated only indirectly. The intratracheal instillation of cobra venom factor contaminated with PLA<sub>2</sub> caused a marked pneumonitis in rabbits which was abolished by prior inactivation of the PLA<sub>2</sub> contaminant [15]. Soluble PLA<sub>2</sub> has been identified in rheumatoid synovial fluids at levels greater than 21,000 units/ml, and in serum in acute hemorrhagic pancreatitis and septic shock at levels greater than 60,000 units/ml (Pruzanski, Vadaš, Fornasier, unpublished observations). The tissue effect of purified PLA<sub>2</sub>, at concentrations comparable to those found in

humans in rheumatoid arthritis and septic shock, have not been defined. Herein, we describe the acute inflammatory effects of exogenous, soluble PLA<sub>2</sub>.

## MATERIALS AND METHODS

New Zealand white rabbits, 2.5-3 kg, of either sex were used. *Naja naja* venom PLA<sub>2</sub> and porcine pancreatic PLA<sub>2</sub> (Sigma) were dissolved in sterile, nonpyrogenic 0.9% saline. Enzyme activity was determined as described previously [1]. One unit of PLA<sub>2</sub> activity is defined as the hydrolysis of 56 pmol of *Escherichia coli* phospholipid in 15 min at 37°C.

**Dose-Response Relationship** The backs of rabbits were shaved and injected intradermally with 0.1-ml aliquots of PLA<sub>2</sub> in concentrations ranging 1,500-60,000 units/0.1 ml. The diameter of resultant erythema and increase in skin thickness (induration) were measured immediately before sacrifice. Animals were sacrificed 24 h after injection and the injection sites were biopsied and fixed in buffered formalin.

**Time-Course of Inflammation** The time course of development of lesions after PLA<sub>2</sub> injection was investigated. PLA<sub>2</sub>, in concentrations of either 1,500 units/0.1 ml or 60,000 units/0.1 ml, was injected at times ranging from 0-48 h before sacrifice. The diameters of erythema and skin thickness of lesions were determined prior to sacrifice, and biopsies of injection sites were submitted for histologic examination. Sterile saline (0.9%) served as the control.

Aliquots of venom PLA<sub>2</sub> (60,000 units/0.1 ml) were inactivated with the active site-directed histidine reagent, *p*-bromophenacyl bromide (*p*BPB) as described elsewhere [16]. Briefly, PLA<sub>2</sub> was incubated with 10<sup>-4</sup> M *p*BPB for 60 min at 22°C. The reaction mixture was dialyzed overnight at 4°C against 0.15 M NaCl. This removed unreacted *p*BPB, and resulted in greater than 99% inactivation of PLA<sub>2</sub>. Skin sites injected with inactivated PLA<sub>2</sub> were biopsied 3 h and 24 h after injection.

**Histologic Examination** Skin biopsies were coded (for single-blind interpretation) and stained with WHO stain (hematoxylin, phloxine, saffron, and Alcian green). The following inflammatory variables were assessed: edema, hemorrhage, histiocytic and polymorphonuclear infiltrate, muscle and fat cell necrosis, collagen degeneration, and inflammatory changes in epidermis, der-

Manuscript received July 22, 1985; accepted for publication November 29, 1985.

Sponsored by a grant-in-aid from the Arthritis Society.

Reprint requests to: Waldemar Pruzanski, M.D., Room 104 Jones Building, The Wellesley Hospital, 160 Wellesley Street East, Toronto, Ontario M4Y 1J3, Canada.

Abbreviations:

*p*BPB: *p*-bromophenacyl bromide

PLA<sub>2</sub>: phospholipase A<sub>2</sub>

mal appendages, and blood vessels. A histologic grade of 0 to 4 was assigned in each category, depending upon the degree of change observed, with grade 0 being normal and grade 4 the most severe [17]. The histologic score represents the arithmetic sum of the grades assigned to each category. Results are expressed as mean  $\pm$  SEM for  $n = 3$  rabbits per experiment.

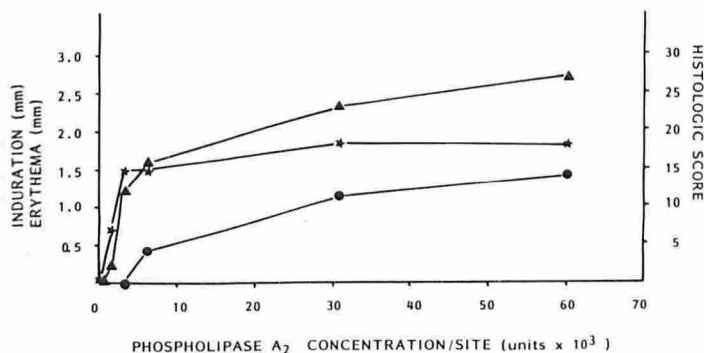
## RESULTS

Intradermal injection of venom PLA<sub>2</sub> in rabbits produced a profound inflammatory reaction characterized grossly by erythema and induration, and microscopically by inflammatory cell infiltration, abscess formation, vascular changes, and muscle and fat necrosis. These changes were both dose- and time-dependent.

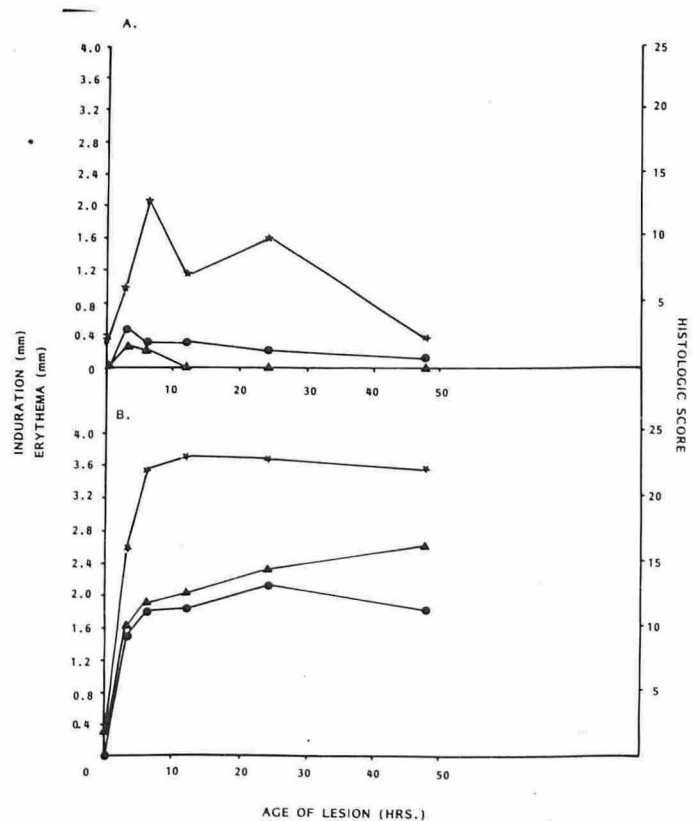
**Dose-Response Relationship** In 24-h-old lesions, skin sites injected with increasing concentrations of venom PLA<sub>2</sub> demonstrated increasing histologic scores up to 6,000 units PLA<sub>2</sub> per injection site with little further increase (Fig 1). The diameters of erythema and induration continued to increase directly with the dose. The cutaneous connective tissue remained intact. Histologically, beginning with the 3,000-U dose, vascular distention with progressive edema of dermal tissue was detectable. Edema and acute inflammatory changes increased with PLA<sub>2</sub> concentrations of up to 30,000 U/site with little progression in the 60,000-U injection sites. No thrombosis of vessels was noted but focal hemorrhages were present in response to the 2 highest doses. The lesions were characterized by necrosis of muscle with associated inflammation. Loss of integrity of individual muscle fibers was seen in sites injected with 1,500 U PLA<sub>2</sub>. The outline of muscle fibers was irregular and fragments of muscle fibers were enveloped by polymorphonuclear leukocytes. As the enzyme concentrations increased, the extent of myonecrosis increased. At the highest dose used, almost all muscle fibers were necrotic with basophilia, fragmentation, and separation by a massive infiltration of acute inflammatory cells. Similarly, fatty necrosis was evident in sites injected with 1,500 U and prominent at concentrations above 6,000 U. None of these changes was seen in the control sections.

**Time-Course of Inflammation** At relatively low concentrations (1,500 U per site), venom PLA<sub>2</sub> caused only transient changes, expressed mainly in induration and erythema which showed a rapid onset with maximal change at 3 h after injection, followed by a gradual resolution (Fig 2A). Histologic score was also greatest early after injection, with maximal microscopic inflammatory changes in 6-h-old lesions. Microscopic changes were still evident 48 h after injection.

Higher concentrations of venom PLA<sub>2</sub> (60,000 units per site) evoked a brisk, severe inflammatory response (Fig 2B). Induration and erythema increased rapidly over the first 6 h after injection, and these changes were sustained over 48 h. Histologic



**Figure 1.** Dose-response relationship of PLA<sub>2</sub> and inflammatory changes in skin in rabbits. SE were less than 5% of the mean,  $n = 6$ . Induration, ●—●; erythema, ▲—▲; histologic score, ★—★.

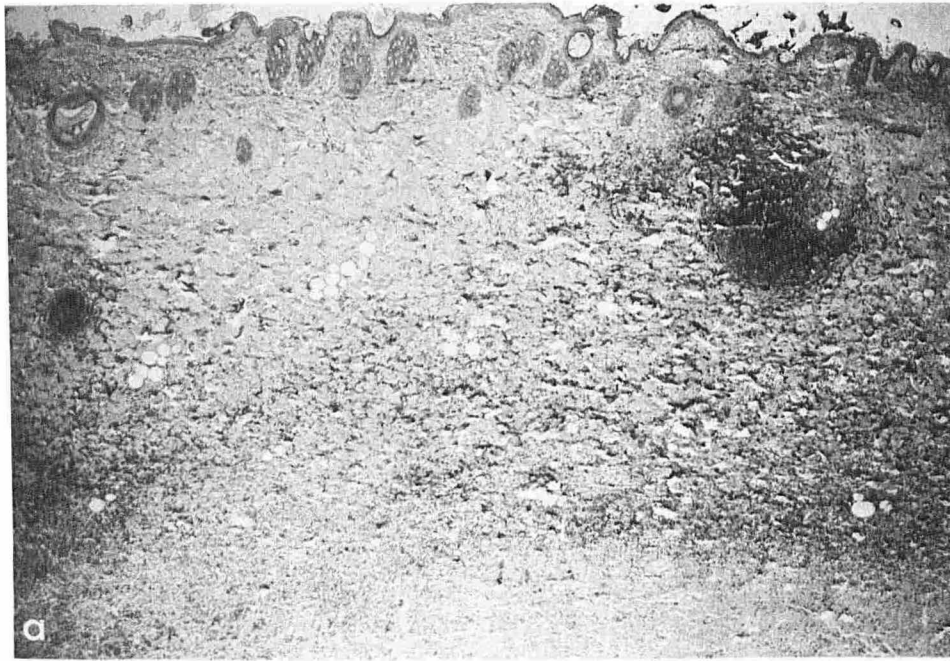


**Figure 2.** Time course of development of lesions after injection of PLA<sub>2</sub>. A, Injection of 1,500 units/site. B, Injection of 60,000 units/site. SE were less than 5% of the mean,  $n = 6$ . Induration, ●—●; erythema, ▲—▲; histologic score, ★—★.

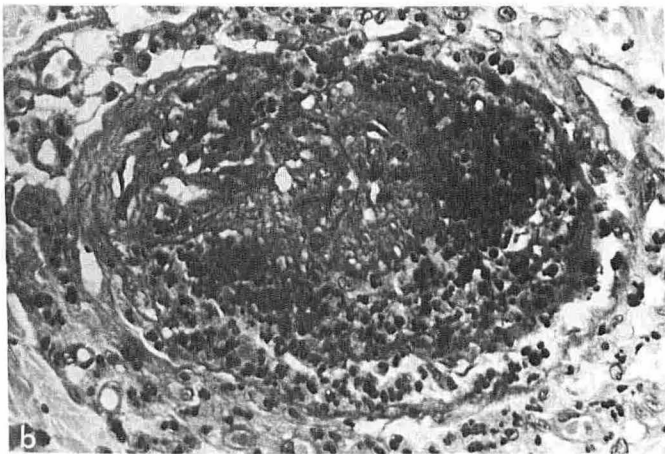
score also increased rapidly, with the highest scores noted 6–12 h after injection of PLA<sub>2</sub>.

Microscopically, the lower dosage showed only vascular engorgement and distention at 3 h. By 6 h, edema and separation of collagen fibers were evident, with polymorphonuclear leukocyte infiltration leading to involvement of the dermal connective tissue on the superficial side of the muscle. Muscle necrosis was extensive, with polymorphonuclear leukocyte infiltration in the dermis and around dead muscle fibers. Massive necrosis of muscle and the related acute inflammation appeared to have reached their maximum by 3 h, in sites injected with 60,000 units PLA<sub>2</sub>. A wider extent of inflammation in the dermis and up to the epidermis was maximal at 6 h and persisted unabated in the 24-h-old lesions (Fig 3). It appeared that injection of PLA<sub>2</sub> caused an accelerated inflammatory response (by 6 h), with little further progression with time (up to 24–48 h). The response was dose related, with the more severe reaction occurring at the higher dose. The lack of progression suggested again an immediate reaction to the injection, far ahead of the usual response to necrosis or injury in which the polymorphonuclear infiltrate is maximal 24 h after injury. This observation suggests a possible chemotactic effect along with an immediate change in vascular permeability, with endothelial swelling occurring in the latter stages (24 h) of the experiment.

The purity of both venom and pancreatic phospholipases was assessed by sodium dodecyl sulfate-polyacrylamide gel electrophoresis. Minor high-molecular-weight contaminants were identified in *Naja naja* venom PLA<sub>2</sub>, while porcine pancreatic PLA<sub>2</sub> was electrophoretically homogeneous. Inactivation of venom PLA<sub>2</sub> with the active site-directed histidine reagent, pBPB, resulted in significant reduction in the resultant inflammatory response in lesions of 3 and 24 h. However, residual inflammatory changes involving dermal appendages were apparent after injection of the



**Figure 3.** Photomicrograph of histologic appearance of site 24 h after injection of venom phospholipase A<sub>2</sub>. *a*, Overview of skin with injection site at right upper corner associated with localized necrotizing inflammation. Diffuse inflammation extends throughout dermis. At left [enlarged in (*b*)] is a focus of necrotizing angitis.  $\times 25$ . *b*, Blood vessel with central core of prominent endothelial cells surrounded by acute necrotizing inflammation. Polymorphonuclear leukocytes infiltrating the connective tissue at the periphery of the blood vessel are representative of the acute inflammation present in the dermis.  $\times 400$ .



molar equivalent of 60,000 units of inactivated PLA<sub>2</sub> protein. It is unlikely that low-molecular-weight proinflammatory components were inadvertently removed by dialysis, since the control preparation of venom PLA<sub>2</sub>, incubated and dialysed without *p*BPB, retained full proinflammatory activity.

Pancreatic PLA<sub>2</sub> also evoked an inflammatory response whose dose dependence and kinetics were similar to those observed for venom PLA<sub>2</sub>. However, while the response to venom PLA<sub>2</sub> was characterized by microabscess formation, myonecrosis, and edema, the response to pancreatic PLA<sub>2</sub> was comparatively attenuated, characterized only by an acute inflammatory infiltrate in the 3- to 6-h-old lesions, which had largely resolved by 24 h. The polymorphonuclear infiltration of dermal connective tissue and dermal appendages did not progress to microabscess formation. Blood vessels remained intact, as did muscle fibers. At the concentrations tested, there was no accompanying erythema or induration.

#### DISCUSSION

The intradermal injection of soluble venom PLA<sub>2</sub> at levels comparable to those found in synovial fluids aspirated from actively inflamed joints of patients with rheumatoid arthritis evoked a brisk inflammatory response. The severity of the resultant lesions was both time- and dose-dependent. The reactions were char-

acterized by early polymorphonuclear cell infiltration, followed by a preponderance of histiocytes. Higher concentrations of PLA<sub>2</sub> caused frank abscess formation with central liquefaction. Blood vessels were engorged but intact in early lesions, corresponding with the grossly erythematous appearance *in vivo*. The kinetics of development of erythema resembled those for hyperemia induced by PLA<sub>2</sub>, as quantitated with radiolabeled microspheres [2,18]. Later lesions were increasingly hemorrhagic, reflecting ongoing vascular damage, concomitant with the appearance of fat and striated muscle necrosis. The findings were similar to those observed in the lungs following exposure to exogenous venom PLA<sub>2</sub> [15].

Interestingly, although the inflammatory response evoked by pancreatic PLA<sub>2</sub> was definite, it was considerably less prominent than that caused by venom PLA<sub>2</sub>. Unlike venom and epidermal PLA<sub>2</sub> activities [14], pancreatic PLA<sub>2</sub> is not inhibited *in vitro* by albumin [19]. Therefore, it is unlikely that serous exudate contributes to the reduced inflammatory response evoked by the pancreatic enzyme. The attenuated response is, however, consistent with the relative inability of pancreatic PLA<sub>2</sub> to cleave intact tissue as compared with the venom enzyme [20,21]. Pancreatic PLA<sub>2</sub> hydrolyzes phospholipids of intact tissue at only one-tenth the rate of venom PLA<sub>2</sub> in the absence of bile salts [20]. Significantly, the resultant inflammatory responses seen in our study were similar quantitatively and with respect to kinetics, and differed only in magnitude.

Hydrolysis of phospholipid substrate by PLA<sub>2</sub> results in the generation of 2 potent biologically active lipids, free fatty acids and lysophosphatides. These lipids are surface-active, cytotoxic, chemotactic for neutrophils, increase vascular permeability, and enhance phagocytosis [11,22]. The proinflammatory effects seen subsequent to injection of PLA<sub>2</sub> may reside in the activities of one or both reaction products of PLA<sub>2</sub>. However, PLA<sub>2</sub> may induce inflammation independently of its enzyme activity, perhaps as a consequence of its net cationic charge [23]. In fact, several studies have demonstrated a dissociation between the enzymatic activity of PLA<sub>2</sub>, and its pharmacologic actions [23,24].

Soluble PLA<sub>2</sub> is a powerful proinflammatory enzyme capable of inducing acute inflammatory changes upon intradermal injection. The association of high levels of endogenous PLA<sub>2</sub> with inflamed sites *in vivo*, and the proinflammatory effect of exog-

enous PLA<sub>2</sub> suggest that the release and accumulation of extracellular PLA<sub>2</sub> in sequestered sites, such as joints and skin, may contribute to the resultant inflammatory changes.

## REFERENCES

1. Vadas P, Hay JB: The release of phospholipase A<sub>2</sub> from aggregated platelets and stimulated macrophages of sheep. *Life Sci* 26:1721-1729, 1980
2. Vadas P, Wasi S, Movat HZ, Hay JB: Extracellular phospholipase A<sub>2</sub> mediates inflammatory hyperemia. *Nature* 295:583-585, 1981
3. Traynor JR, Authi KS: Phospholipase A<sub>2</sub> activity of lysosomal origin secreted by polymorphonuclear leukocytes during phagocytosis or on treatment with calcium. *Biochim Biophys Acta* 665:571-577, 1981
4. Vadas P, Hay JB: The appearance and significance of phospholipase A<sub>2</sub> in lymph draining tuberculin reactions. *Am J Pathol* 107:285-291, 1982
5. Wightman PD, Dahlgren ME, Davies P, Bonney RJ: The selective release of phospholipase A<sub>2</sub> by resident mouse peritoneal macrophages. *Biochem J* 200:441-444, 1981
6. Secchi AG, Fregona I, D'ermo F: Lysophosphatidylcholine in the aqueous humour during ocular inflammation. *Br J Ophthalmol* 63:768-770, 1979
7. Franson R, Dobrow R, Weiss J, Elsbach P, Weglicki WB: Isolation and characterization of a phospholipase A<sub>2</sub> from an inflammatory exudate. *J Lipid Res* 19:18-23, 1978
8. Vadas P, Hay JB: Involvement of circulating phospholipase A<sub>2</sub> in the pathogenesis of the hemodynamic changes in endotoxin shock in rabbits. *Can J Physiol Pharmacol* 61:561-566, 1983
9. Nevalainen TJ: The role of phospholipase A in acute pancreatitis. *Scand J Gastroenterol* 15:641-650, 1980
10. Vadas P: Plasma phospholipase A<sub>2</sub> levels correlate with the hemodynamic and pulmonary changes in gram negative septic shock in man. *J Lab Clin Med* 104:873-881, 1984
11. Vadas P, Pruzanski W: Role of extracellular phospholipase A<sub>2</sub> in inflammation. *Adv Inflam Res* 7:51-59, 1984
12. Pruzanski W, Vadas P, Stefanski E, Urowitz M: Phospholipase A<sub>2</sub> activity in sera and synovial fluids in rheumatoid arthritis and osteoarthritis. Its possible role as a pro-inflammatory enzyme. *J Rheumatol* 12:211-216, 1985
13. Forster S, Ilderton E, Summerly R, Yardley H: Epidermal phospholipase A<sub>2</sub> activity is raised in the uninvolved skin of psoriasis. *Br J Dermatol* 109 (suppl 25):30-35, 1983
14. Forster S, Ilderton E, Norris J, Summerly R, Yardley H: Characterization and activity of phospholipase A<sub>2</sub> in normal human epidermis and in lesion-free epidermis of patients with psoriasis or eczema. *Br J Dermatol* 112:135-147, 1985
15. Shaw J, Roberts M, Ulevitch R, Henson P, Dennis E: Phospholipase A<sub>2</sub> contamination of cobra venom factor preparations. *Am J Pathol* 91:571-580, 1978
16. Vadas P, Stefanski E, Pruzanski W: Characterization of extracellular phospholipase A<sub>2</sub> in rheumatoid synovial fluid. *Life Sci* 36:579-588, 1985
17. Keystone EC, Taylor-Robinson D, Ling L, Pope C, Metcalfe A, Furr P, Formasier V: Enhanced resistance of mice to *Mycoplasma pulmonis*-induced arthritis by administration of killed *Corynebacterium parvum*. *Clin Exp Immunol* 46:355-362, 1981
18. Vadas P, Hay JB: Cutaneous blood flow measurements: a standardization of the microsphere assay for vasoactive agents. *Agents Actions* 8:504-508, 1978
19. Vadas P, Stefanski E, Pruzanski W: Comparative analysis of assays for phospholipase A<sub>2</sub> activity using bio-membrane associated and micellar substrates. The influence of albumin. *Inflammation*, in press
20. Ibrahim S, Sanders H, Thompson R: The action of phospholipase A on purified phospholipids, plasma and tissue preparations. *Biochem J* 93:588-594, 1964
21. Nishijima J, Okamoto M, Yamano T, Nakaguchi K, Ogawa M: Hydrolytic activities of human pancreatic phospholipase A<sub>2</sub> and endotoxin-stimulated endogenous phospholipase A<sub>2</sub> toward membrane phospholipids of erythrocytes. *J Surg Res* 38:237-245, 1985
22. Blackwell GH, Flower JR: Inhibition of phospholipase. *Br Med Bull* 39:260-264, 1983
23. Rosenberg P, Condrea E, Rapuano B, Soons K, Yang C-C: Dissociation of pharmacological and enzymatic activities of snake venom phospholipases A<sub>2</sub> by modification of carboxylate groups. *Biochem Pharmacol* 32:3525-3530, 1983
24. Barrington P, Yang C-C, Rosenberg P: Cardiotoxic effects of *Naja nigricollis* venom phospholipase A<sub>2</sub> are not due to phospholipid hydrolytic products. *Life Sci* 35:987-995, 1984