

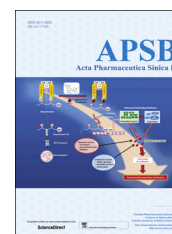
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REVIEW

The antiviral and antimicrobial activities of licorice, a widely-used Chinese herb



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Abstract Licorice is a common herb which has been used in traditional Chinese medicine for centuries. More than 20 triterpenoids and nearly 300 flavonoids have been isolated from licorice. Recent studies have shown that these metabolites possess many pharmacological activities, such as antiviral, antimicrobial, anti-inflammatory, antitumor and other activities. This paper provides a summary of the antiviral and antimicrobial activities of licorice. The active components and the possible mechanisms for these activities are summarized in detail. This review will be helpful for the further studies of licorice for its potential therapeutic effects as an antiviral or an antimicrobial agent.

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Abbreviations: CCEC, cerebral capillary vessel endothelial; CCL5, chemokine (C-C motif) ligand 5; CVA16, coxsackievirus A16; CVB3, coxsackievirus B3; CXCL10, chemokine (C-X-C motif) ligand 10; DGC, dehydroglyasperin C; DHV, duck hepatitis virus; EV71, enterovirus 71; GA, 18 β -glycyrrhetic acid; GATS, glycyrrhizic acid trisodium salt; GL, glycyrrhizin; GLD, glabridin; HBV, hepatitis B virus; HCV, hepatitis C virus; HIV, human immunodeficiency virus; HMGB1, high-mobility-group box1; HRSV, human respiratory syncytial virus; HSV, herpes simplex virus; HSV1, herpes simplex virus type 1; IFN, interferon; IL-6, interleukin-6; LCA, licochalcone A; LCE, licochalcone E; ISL, isoliquiritigenin; LTG, liquiritigenin; MgIG, magnesium isoglycyrrhizinate; MRSA, methicillin-resistant *Staphylococcus aureus*; MSSA, methicillin-sensitive *Staphylococcus aureus*; PMN, polymorph nuclear; PrV, pseudorabies virus; TCM, traditional Chinese medicine

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1. Introduction

Licorice is a very well known herb in traditional Chinese medicine (TCM). In China, it is called “gancao” (meaning “sweet grass”) and has been recorded in the *Shennong's Classic of Materia Medica* around 2100 BC. In this book, licorice was supposed to have life-enhancing properties. During the following thousands of years licorice has been present in most of Chinese traditional prescriptions. It was believed to have the functions of nourishing *qi*, alleviating pain, tonifying spleen and stomach, eliminating phlegm, and relieving coughing¹.

Glycyrrhiza uralensis Fisch., *Glycyrrhiza inflata* Bat. and *Glycyrrhiza glabra* L. were prescribed as licorice in *Chinese pharmacopoeia*². They are widespread in Inner Mongolia, Gansu, Heilongjiang, Ningxia, Qinghai and many other provinces in China³. The roots and rhizomes are the main medicinal parts of licorice. Numerous studies have revealed many pharmacological activities of licorice, such as antiviral^{4,5}, anti-inflammatory^{6,7}, antitumor^{8,9}, antimicrobial^{10,11} and many other activities^{12,13}. Among the pharmacological activities of licorice mentioned above, the antiviral and antimicrobial activities have been most commonly reported. Viral and other microbial infections play a critical role in many highly prevalent diseases, especially in developing countries. The development of safe and effective antiviral or antimicrobial agents is very important, and licorice deserves more attention for its outstanding activities.

Licorice contains more than 20 triterpenoids and nearly 300 flavonoids. Among them, glycyrrhizin (GL), 18 β -glycyrrhetic acid (GA), liquiritigenin (LTG), licochalcone A (LCA), licochalcone E (LCE) and glabridin (GLD) are the main active components which possess antiviral and antimicrobial activities. Their chemical structures are listed in Fig. 1.

2. The antiviral active components and their possible mechanisms

Among the components isolated from licorice, 73 bioactive components and 91 potential targets have been identified to date^{14,15}. Many studies have demonstrated that two triterpenoids, GL^{16,17} and GA¹⁸, are responsible for the antiviral activity. The possible mechanisms for virus prevention of GL and GA, and the viral types are listed in Table 1.

2.1. GL

GL is one of the major compounds isolated from the roots of licorice. In recent years, many studies have confirmed the antiviral activity of GL. Matsumoto et al.¹⁶ reported that GL targeted the release step in which infectious anti-hepatitis C virus (HCV) particles were infecting cells. These findings indicated possible novel roles for GL to treat patients suffering from chronic hepatitis C. In another study, researchers also found that GL treatment inhibited HCV titer and caused 50% reduction of HCV at the concentration of $14 \pm 2 \mu\text{g/mL}$ by inhibiting HCV full length viral particles and their core gene expression¹⁹.

Previous studies showed that intercellular adhesion molecules played an important role in some viral infections, such as human immunodeficiency virus (HIV)²⁰. Huang et al.⁵ found that the adhesion force and stress between cerebral capillary vessel endothelial (CCEC) cells and polymorph nuclear (PMN) leukocytes were clearly increased in HSV infection; GL perfusion significantly reduced adhesion force and stress between CCEC and PMN.

Zhang's study²¹ reported that GL showed a significant improvement of coxsackievirus B3 (CVB3)-induced myocarditis by improving weight loss profile, reducing serological levels of cardiac enzymes and increasing survival rate. This effect was evidenced by significantly reduced expression of proinflammatory cytokines, such as nuclear factor- κ B, interleukin-1 β and interleukin-6. The inhibition of CVB3-induced nuclear factor- κ B activity blocks the degradation of nuclear factor- κ B inhibitor I κ B. All these data suggested that GL had an effect on CVB3-induced myocarditis and may present as a new therapeutic approach for the treatment of viral myocarditis.

Soufy et al.²² found that GL had excellent immunostimulant properties and induced a synergistic effect to duck hepatitis virus (DHV) vaccine by activating T lymphocyte proliferation. Four groups, control, GL treated, vaccinated with live attenuated DHV vaccine and GL treated and vaccinated, were investigated. Among them, treatment with GL alone or with DHV vaccine showed good immune stimulant and antiviral effects against DHV. GL combined with DHV vaccine produced higher antibody titers against DHV than by the use of DHV vaccine alone.

Several studies have demonstrated that GL showed a significant inhibiting effect to influenza virus. At a concentration of 100 $\mu\text{g/mL}$ (a therapeutically achievable concentration), GL weakened

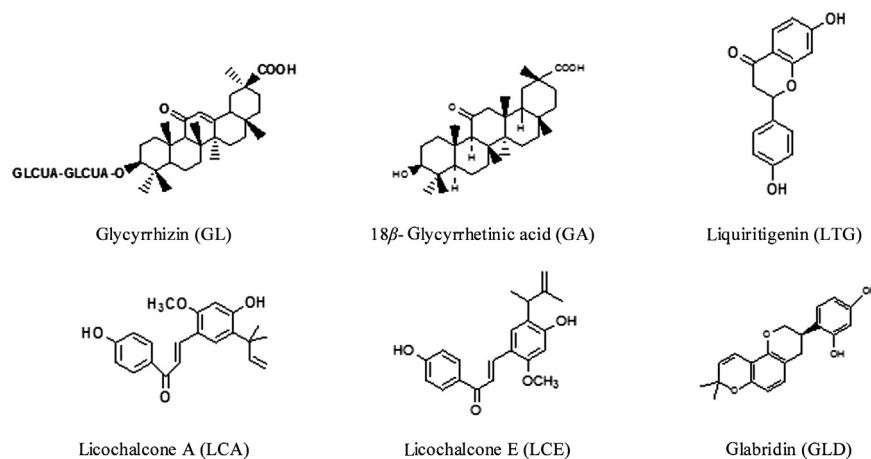


Figure 1 The chemical structures of the antiviral activity or antimicrobial components in licorice.

Table 1 The antiviral active components and their possible mechanisms for virus prevention.

Component	Antiviral mechanism	Viral type
GL	Affect release step while infectious HCV particles are infecting cells.	HCV
	Inhibit HCV full length viral particles and HCV core gene expression.	
	Reduce adhesion force and stress between CCEC and PMN.	HSV
	Block the degradation of nuclear factor κ B inhibitor I κ B.	CVB3
	Activate T lymphocyte proliferation.	DHV
	Weaken H5N1-induced production of CXCL10, IL-6 and CCL5, and suppress H5N1-induced apoptosis.	H5N1
	Reduce HMGB1 binding to DNA, and inhibit influenza virus polymerase activity.	Influenza virus
GA	Inactivate CVA16 directly, while the effect of anti-EV71 is associated with an event(s) during the virus cell entry.	CVA16 EV71
	Establish a resistance state to HSV1 replication.	HSV1
	Reduce the levels of viral proteins VP2, VP6 and NSP2 at a step or steps subsequent to virus entry.	Rotavirus
	Prevent viral attachment, internalization and stimulate IFN secretion.	HRSV

H5N1-induced production of chemokine (C-X-C motif) ligand 10 (CXCL10), interleukin 6 (IL-6) and chemokine (C-C motif) ligand 5 (CCL5), and suppressed H5N1-induced apoptosis²³. The high-mobility-group box1 (HMGB1) DNA-binding site was indicated to enhance influenza virus replication. GL could reduce HMGB1 binding to DNA, which inhibited influenza virus polymerase activity²⁴. Smirnov's study²⁵ indicated that GL could be considered a promising agent for the treatment of influenza.

Wang's study¹⁷ revealed that GL was an antiviral component in licorice against enterovirus 71 (EV71) and coxsackievirus A16 (CVA16) infection with defined mechanisms. It activated CVA16 directly, while the effect of anti-EV71 was associated with an event during the cell entry for virus.

GL was also a strong inducer of the autophagy activator Beclin 1. After 24 h of treatment, Beclin 1 production induced by GL was more than two fold higher than that was induced by rapamycin, the reference compound. GL was a strong inducer of Beclin 1, which inhibited the replication of herpes simplex virus type 1 (HSV1)²⁶. Therefore, GL possessed its anti-HSV1 activity by establishing a resistant state to HSV1 replication.

Above all, GL is an effective antiviral compound against HCV, HIV, CVB3, DHV, EV71, CVA16, HSV and H5N1 by weakening virus activity, such as inhibiting virus gene expression and replication, reducing adhesion force and stress, and reducing HMGB1 binding to DNA. The compound also enhances host cell activity, *e.g.*, by blocking the degradation of I κ B, activating T lymphocyte proliferation and/or suppressing host cell apoptosis.

2.2. GA

Compared with GL, studies of the antiviral activity of GA are limited. GA treatment inhibited rotavirus replication, which likely occurred at steps subsequent to virus entry. GA reduced rotavirus yields by 99% when it was added to infected cultures post-viral adsorption. The levels of viral proteins VP2, VP6 and NSP2 were substantially reduced²⁷. GA also showed potent anti-human respiratory syncytial virus (HRSV) activity. It inhibited HRSV mainly by internalization, stimulating interferon (IFN) secretion, and preventing viral attachment¹⁸.

There is a difference between the antiviral profiles of GA and GL. GA has activity against rotavirus and HRSV. However, the antiviral mechanisms of these compounds are similar. GA exerts its antiviral activity also by inhibiting virus replication, preventing viral attachment or enhancing host cell activity.

3. The antimicrobial active components and their possible mechanisms

Increasing antibiotic resistance has resulted in an urgent need for alternative therapies to treat diseases. In recent years, many studies have shown that licorice aqueous extract²⁸, ethanol extract²⁹ and supercritical fluid extract³⁰ have potent effects in inhibiting the activities of Gram-positive bacteria and Gram-negative bacteria, such as *Staphylococcus aureus*³¹, *Escherichia coli*³², *Pseudomonas aeruginosa*³³, *Candida albicans* and *Bacillus subtilis*³⁴. These extracts are also being considered as potential alternatives to synthetic fungicides, or as lead compounds for new classes of synthetic fungicides. Based on the above inhibitory activities against bacteria, licorice may serve as an alternative therapy for treating dental caries, periodontal disease, digestive anabrosis and tuberculosis. The possible mechanisms for antimicrobial effects of the active components and the microorganism types were listed in Table 2.

3.1. GA

Methicillin-resistant *S. aureus* (MRSA) has become a main source of infection in both hospitals and the community. Increasing antibiotic resistance in *S. aureus* strains has created a need for other therapies to treat disease. GA showed bactericidal activity to destroy MRSA by decreasing the expression of *SaeR* and *Hla*, the key virulence genes of MRSA³¹. Studies also indicated that GA produced a better Th1 immune response than Th2 response. This Th1-immunological adjuvant activity would be helpful in the treatment of Th1-related disease caused by *C. albicans*³⁵.

3.2. Chalcones

Zhou et al.³⁶ suggested that licochalcone E (LCE) could be used for chemical synthesis of novel anti-*S. aureus* compounds which could reduce the production of α -toxin in both methicillin-sensitive *S. aureus* (MSSA) and MRSA. Licochalcone A (LCA) and glabridin (GLD) showed antifungal activity on *C. albicans*. They were both potent antifungal agents against *C. albicans*. LCA (0.2 μ g/mL) inhibited biofilm formation by 35%–60% and both LCA and GLD had strong inhibitory effects (>80%) in preventing yeast-hyphal transition in *C. albicans*³⁷.

Table 2 The antimicrobial active components and their possible mechanisms for microbe prevention.

Component	Antimicrobial mechanism	Microbial type
GA	Decrease the expression of <i>SaeR</i> and <i>Hla</i> , which are the key virulence genes of MRSA. Exert the Th1-immunological adjuvant activity.	<i>S. aureus</i> <i>C. albicans</i>
LCA	Inhibit the biofilm formation and prevent yeast-hyphal transition.	<i>C. albicans</i>
LCE	Reduce the production of α -toxin.	<i>S. aureus</i>
GLD	Prevent yeast-hyphal transition.	<i>C. albicans</i>
LTG	Decrease the production of α -hemolysin.	<i>S. aureus</i>

3.3. Liquiritigenin

α -Hemolysin is an important exotoxin in the pathogenesis of *S. aureus* infections. Such infections are associated with a broad spectrum of diseases ranging from endocarditis to minor skin infections, toxinoses, and lethal pneumonia. Liquiritigenin (LTG), one of the most significant active components in licorice, can prevent human lung cells (A549) from α -hemolysin-mediated injury by decreasing α -hemolysin production³⁸. Such data suggest that LTG is potentially useful in developing drugs which target staphylococcal α -hemolysin.

In summary, one triterpene (GA) and four flavones (LCA, LCE, GLD and LTG) seem to account for much of the antimicrobial activity in licorice. These compounds can decrease the expression of microbe genes, inhibit microbe growth and reduce the production of microbe toxin.

4. Discussion

Presently we have summarized the antiviral and antimicrobial activities of licorice. Many studies found that several components were responsible for the antiviral and antimicrobial activities through different mechanisms. Licorice contains more than 20 triterpenoids and nearly 300 flavonoids. Among them, only two triterpenes, GL and GA have been reported to have antiviral effects. They can weaken virus activities by inhibiting virus gene expression and replication, reducing adhesion force and stress, and reducing HMGB1 binding to DNA. They can also enhance host cell activities by blocking the degradation of I κ B, activating T lymphocyte proliferation and suppressing host cell apoptosis. In contrast, flavonoids, especially chalcones, play an important role in the treatment of bacterial infection by decreasing expression of bacterial genes, inhibiting bacterial growth and reducing the production of bacterial toxin.

In addition, many studies have reported that the six active compounds listed in this paper, GL, GA, LCA, LCE, GLD and LTG, possess other activities. For example, GL and GA also have antitumor^{39,40}, anti-inflammatory^{41,42}, and immunoregulatory activities^{12,43,44}. LCA, LCE, LTG and GLD also have inhibitory effects on diabetes^{45–48}. All of these reports demonstrate potentially broad applications for these agents. In addition, there are many other compounds isolated from licorice with different pharmacological activities. For example, isoliquiritigenin (ISL) shows effective immunoregulatory activity⁴⁹, glabrol has an inhibitory effect on diabetes⁵⁰, and dehydroglyasperin C (DGC) has hepatoprotective activity⁵¹.

Among the six compounds listed in this paper, only GL has been clinically developed as a drug. As the most important marker component in licorice, the development of GL preparations has a

long history in China, from GL tablets to ammonium glycyrrhizinate, diammonium glycyrrhizinate and magnesium isoglycyrrhizinate (MgIG). All of the above GL preparations possess antiviral and antimicrobial activities. Diammonium glycyrrhizinate inhibits cell infection by pseudorabies virus (PrV) and decreases cell apoptosis during PrV infection⁵². Compared with diammonium glycyrrhizinate, the fourth generation GL preparation, MgIG, has better lipophilic properties, higher targeting activity and fewer adverse reactions. It has been used in treating liver disease^{53–55}, pulmonary fibrosis⁵⁶ and testicular injuries⁵⁷. However, reports about mechanisms of antiviral and antimicrobial activities of MgIG are still very limited. The development of new licorice preparations will improve the safety and efficacy of licorice-related products.

In many African countries with poorly developed health care systems, viruses and bacteria are significant sources of disease. More than 2 billion people have been exposed to HBV over the world, and the situation in some areas of Africa is much more serious⁵⁸. The development of effective and affordable licorice-related medicines could introduce dramatic improvements in treating the many prevalent diseases of third world populations. It is hoped that the present work will facilitate the development of improved licorice preparations with antiviral and antimicrobial activities.

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References

- Zeng L, Li SH, Lou ZC. Morphological and histological studies of Chinese licorice. *Acta Pharm Sin* 1988;23:200–8.
- The state Pharmacopoeia Committee of China. *The pharmacopoeia of the people's republic of China. Part I*. Beijing: China Medical Science Press; 80–1.
- Delectis Florae Reipublicae Popularis Sinicae Agendae Academiae Sinicae Edita. *Flora reipublicae popularis sinicae*, Vol. 42. Beijing: Science Press; 169.
- Adianti M, Aoki C, Komoto M, Deng L, Shoji I, Wahyuni TS, et al. Anti-hepatitis C virus compounds obtained from *Glycyrrhiza uralensis* and other *Glycyrrhiza* species. *Microbiol Immunol* 2014;58:180–7.
- Huang W, Chen X, Li Q, Li P, Zhao GN, Xu MM, et al. Inhibition of intercellular adhesion in herpes simplex virus infection by glycyrrhizin. *Cell Biochem Biophys* 2012;62:137–40.
- Chandrasekaran CV, Deepak HB, Thiyagarajan P, Kathiresan S, Sangli GK, Deepak M, et al. Dual inhibitory effect of *Glycyrrhiza glabra* (GutGard™) on COX and LOX products. *Phytomedicine* 2011;18:278–84.
- Wu TY, Khor TO, Saw CLL, Loh SC, Chen AI, Lim SS, et al. Anti-inflammatory/anti-oxidative stress activities and differential regulation of Nrf2-mediated genes by non-polar fractions of tea *Chrysanthemum zawadskii* and licorice *Glycyrrhiza uralensis*. *AAPS J* 2011;13:1–13.

8. Choi AY, Choi JH, Hwang KY, Jeong YJ, Choe W, Yoon KS, et al. Licochalcone A induces apoptosis through endoplasmic reticulum stress via a phospholipase C γ 1-, Ca²⁺-, and reactive oxygen species-dependent pathway in HepG2 human hepatocellular carcinoma cells. *Apoptosis* 2014;**19**:682–97.
9. Khan R, Khan AQ, Lateef A, Rehman MU, Tahir M, Ali F, et al. Glycyrrhizic acid suppresses the development of precancerous lesions via regulating the hyperproliferation, inflammation, angiogenesis and apoptosis in the colon of Wistar rats. *PLoS One* 2013;**8**:e56020.
10. Ahn SJ, Cho EJ, Kim HJ, Park SN, Lim YK, Kook JK. The antimicrobial effects of deglycyrrhizinated licorice root extract on *Streptococcus mutans* UA159 in both planktonic and biofilm cultures. *Anaerobe* 2012;**18**:590–6.
11. Treutwein J, Cergel S, Runte J, Nowak A, Konstantinidou-Doltsinis S, Kleeberg H, et al. Efficacy of *Glycyrrhiza glabra* extract fractions against phytopathogenic fungi. *Julius-Kühn-Archiv* 2010;**428**:82.
12. Bordbar N, Karimi MH, Amirhofran Z. Phenotypic and functional maturation of murine dendritic cells induced by 18 alpha- and beta-glycyrrhetic acid. *Immunopharm Immunot* 2013;**36**:52–60.
13. Hong YK, Wu HT, Ma T, Liu WJ, He XJ. Effects of *Glycyrrhiza glabra* polysaccharides on immune and antioxidant activities in high-fat mice. *Int J Biol Macromol* 2009;**45**:61–4.
14. Li YJ, Chen J, Li Y, Li Q, Zheng YF, Fu Y, et al. Screening and characterization of natural antioxidants in four *Glycyrrhiza* species by liquid chromatography coupled with electrospray ionization quadrupole time-of-flight tandem mass spectrometry. *J Chromatogr A* 2011;**1218**:8181–91.
15. Liu H, Wang JN, Zhou W, Wang YH, Yang L. Systems approaches and polypharmacology for drug discovery from herbal medicines: an example using licorice. *J Ethnopharmacol* 2013;**146**:773–93.
16. Matsumoto Y, Matsuura T, Aoyagi H, Matsuda M, Hmwe SS, Date T, et al. Antiviral activity of glycyrrhizin against hepatitis C virus *in vitro*. *PLoS One* 2013;**8**:e68992.
17. Wang JJ, Chen XQ, Wang W, Zhang YT, Yang ZY, Jin Y, et al. Glycyrrhizic acid as the antiviral component of *Glycyrrhiza uralensis* Fisch. against coxsackievirus A16 and enterovirus 71 of hand foot and mouth disease. *J Ethnopharmacol* 2013;**147**:114–21.
18. Yeh CF, Wang KC, Chiang LC, Shieh DE, Yen MH, Chang JS. Water extract of licorice had anti-viral activity against human respiratory syncytial virus in human respiratory tract cell lines. *J Ethnopharmacol* 2013;**148**:466–73.
19. Ashfaq UA, Masoud MS, Nawaz Z, Riazuddin S. Glycyrrhizin as antiviral agent against Hepatitis C Virus. *J Transl Med* 2011;**9**:112.
20. Wang JH, Kwas C, Wu L. Intercellular adhesion molecule 1 (ICAM-1), but not ICAM-2 and -3, is important for dendritic cell-mediated human immunodeficiency virus type 1 transmission. *J Virol* 2009;**83**:4195–4204.
21. Zhang HC, Song YX, Zhang ZC. Glycyrrhizin administration ameliorates coxsackievirus B3-induced myocarditis in mice. *Am J Med Sci* 2012;**344**:206–10.
22. Soufy H, Yassein S, Ahmed AR, Khodier MH, Kutkat MA, Nasr SM, et al. Antiviral and immune stimulant activities of glycyrrhizin against duck hepatitis virus. *Afr J Tradit Complement Altern Med* 2012;**9**:389–95.
23. Michaelis M, Geiler J, Naczek P, Sithisarn P, Ogbomo H, Altenbrandt B, et al. Glycyrrhizin inhibits highly pathogenic H5N1 influenza A virus-induced pro-inflammatory cytokine and chemokine expression in human macrophages. *Med Microbiol Immunol* 2010;**199**:291–7.
24. Moisy D, Avilov SV, Jacob Y, Laoide BM, Ge XY, Baudin F, et al. HMGB1 protein binds to influenza virus nucleoprotein and promotes viral replication. *J Virol* 2012;**86**:9122–33.
25. Smirnov VS, Zarubaev VV, Anfimov PM, Shtro AA. Effect of a combination of glutamyl-tryptophan and glycyrrhizic acid on the course of acute infection caused by influenza (H3N2) virus in mice. *Vopr Virusol* 2012;**57**:23–7.
26. Laconi S, Madeddu MA, Pompei R. Autophagy activation and antiviral activity by a licorice triterpene. *Phytother Res* 2014;**28**:1890–1892.
27. Hardy ME, Hendricks JM, Paulson JM, Faunce NR. 18 β -glycyrrhetic acid inhibits rotavirus replication in culture. *Virol J* 2012;**9**:96.
28. Al-Turki AI, El-Ziney MG, Abdel-Salam AM. Chemical and antibacterial characterization of aqueous extracts of oregano, marjoram, sage and licorice and their application in milk and labneh. *J Food Agric Environ* 2008;**6**:39–44.
29. Park IK, Kim J, Lee YS, Shin SC. *In vivo* fungicidal activity of medicinal plant extracts against six phytopathogenic fungi. *Int J Pest Manag* 2008;**54**:63–8.
30. Bodet C, La VD, Gafner S, Bergeron C, Grenier D. A licorice extract reduces lipopolysaccharide-induced proinflammatory cytokine secretion by macrophages and whole blood. *J Periodontol* 2008;**79**:1752–61.
31. Long DR, Mead J, Hendricks JM, Hardy ME, Voyich JM. 18 β -Glycyrrhetic acid inhibits methicillin-resistant *Staphylococcus aureus* survival and attenuates virulence gene expression. *Antimicrob Agents Chemother* 2013;**57**:241–7.
32. Awandkar SP, Mendhe MS, Badukale DM, Kulkarni MB. Antimicrobial action of cold aqueous and cold ethanolic extracts of *Glycyrrhiza glabra* against bovine mammary pathogens. *Anim Sci Report* 2012;**6**:88–91.
33. Yoshida T, Yoshida S, Kobayashi M, Herndon DN, Suzuki F. Pivotal advance: glycyrrhizin restores the impaired production of β -defensins in tissues surrounding the burn area and improves the resistance of burn mice to *Pseudomonas aeruginosa* wound infection. *J Leukoc Biol* 2010;**87**:35–41.
34. Irani M, Sarmadi M, Bernard F, Ebrahimi PGH, Shaker BH. Leaves antimicrobial activity of *Glycyrrhiza glabra* L. *Iran J Pharm Res* 2010;**9**:425–8.
35. Kim J, Joo I, Kim H, Han Y. 18 β -Glycyrrhetic acid induces immunological adjuvant activity of Th1 against *Candida albicans* surface mannan extract. *Phytomedicine* 2013;**20**:951–5.
36. Zhou TZ, Deng XM, Qiu JZ. Antimicrobial activity of licochalcone E against *Staphylococcus aureus* and its impact on the production of staphylococcal alpha-toxin. *J Microbiol Biotechnol* 2012;**22**:800–5.
37. Messier C, Grenier D. Effect of licorice compounds licochalcone A, glabridin and glycyrrhizic acid on growth and virulence properties of *Candida albicans*. *Mycoses* 2011;**54**:e801–6.
38. Dai XH, Li HE, Lu CJ, Wang JF, Dong J, Wei JY, et al. Liquiritigenin prevents *Staphylococcus aureus*-mediated lung cell injury via inhibiting the production of α -hemolysin. *J Asian Nat Prod Res* 2013;**15**:390–399.
39. Kim HJ, Seo JY, Suh HJ, Lim SS, Kim JS. Antioxidant activities of licorice-derived prenylflavonoids. *Nutr Res Pract* 2012;**6**:491–8.
40. Yasukawa K. Inhibitory effect of a combined treatment of glycyrrhizin and caffeine on tumor promotion by 12-*O*-tetradecanoylphorbol-13-acetate in two-stage carcinogenesis in mouse skin. *J Pharm Nutr Sci* 2013;**3**:202–5.
41. Wang CY, Kao TC, Lo WH, Yen GC. Glycyrrhizic acid and 18 β -glycyrrhetic acid modulate lipopolysaccharide-induced inflammatory response by suppression of NF- κ B through PI3K p110 δ and p110 γ inhibitions. *J Agric Food Chem* 2011;**59**:7726–33.
42. Imai K, Takagi Y, Iwazaki A, Nakanishi K. Radical scavenging ability of glycyrrhizin. *Free Radic Antioxid* 2013;**3**:40–2.
43. Li W, Li JH, Sama AE, Wang HC. Carbenoxolone blocks endotoxin-induced protein kinase R (PKR) activation and high mobility group box 1 (HMGB1) release. *Mol Med* 2013;**19**:203–11.
44. Kim ME, Kim HK, Kim DH, Yoon JH, Lee JS. 18 β -Glycyrrhetic acid from licorice root impairs dendritic cells maturation and Th1 immune responses. *Immunopharmacol Immunotoxicol* 2013;**35**:329–35.
45. Yao K, Chen HY, Lee MH, Li HT, Ma WY, Peng C, et al. Licochalcone A, a natural inhibitor of c-Jun N-terminal kinase 1. *Cancer Prev Res* 2014;**7**:139–49.
46. Park HG, Bak EJ, Woo GH, Kim JM, Quan ZJ, Kim JM, et al. Licochalcone E has an antidiabetic effect. *J Nutr Biochem* 2012;**23**:759–767.
47. Gaur R, Yadav KS, Verma RK, Yadav NP, Bhakuni RS. *In vivo* antidiabetic activity of derivatives of isoliquiritigenin and liquiritigenin. *Phytomedicine* 2014;**21**:415–22.

48. Wu FH, Jin ZG, Jin J. Hypoglycemic effects of glabridin, a polyphenolic flavonoid from licorice, in an animal model of diabetes mellitus. *Mol Med Rep* 2013;**7**:1278–82.
49. Park SJ, Song HY, Youn HS. Suppression of the TRIF-dependent signaling pathway of toll-like receptors by isoliquiritigenin in RAW264.7 macrophages. *Mol Cells* 2009;**28**:365–8.
50. Choi JH, Choi JN, Lee SY, Lee SJ, Kim K, Kim YK. Inhibitory activity of diacylglycerol acyltransferase by glabrol isolated from the roots of licorice. *Arch Pharm Res* 2010;**33**:237–42.
51. Seo JY, Han JH, Kim YJ, Lim SS, Kim JS. Protective effects of dehydroglyasperin c against carbon tetrachloride-induced liver damage in mice. *Food Sci Biotechnol* 2014;**23**:547–53.
52. Sui XW, Yin JC, Ren XF. Antiviral effect of diammonium glycyrrhizinate and lithium chloride on cell infection by pseudorabies herpesvirus. *Antiviral Res* 2010;**85**:346–53.
53. Huang XL, Qin JJ, Lu S. Magnesium isoglycyrrhizinate protects hepatic L02 cells from ischemia/reperfusion induced injury. *Int J Clin Exp Pathol* 2014;**7**:4755–64.
54. Chen KJ, Chen WY, Chen X, Jia YM, Peng GQ, Chen L. Increased elimination of paclitaxel by magnesium isoglycyrrhizinate in epithelial ovarian cancer patients treated with paclitaxel plus cisplatin: a pilot clinical study. *Eur J Drug Metab Pharmacokinet* 2014;**39**:25–31.
55. Cheng Y, Zhang J, Shang J, Zhang LY. Prevention of free fatty acid-induced hepatic lipotoxicity in HepG2 cells by magnesium isoglycyrrhizinate *in vitro*. *Pharmacology* 2009;**84**:183–90.
56. Xiao ZW, Zhang W, Ma L, Qiu ZW. Therapeutic effect of magnesium isoglycyrrhizinate in rats on lung injury induced by paraquat poisoning. *Eur Rev Med Pharmacol Sci* 2014;**18**:311–20.
57. He YQ, Zeng FQ, Liu Q, Ju W, Fu HJ, Hao H, et al. Protective effect of magnesium isoglycyrrhizinate on ethanol-induced testicular injuries in mice. *J Biomed Res* 2010;**24**:153–60.
58. Rey-Cuille MA, Njouom R, Bekondi C, Seck A, Gody C, Bata P, et al. Hepatitis B virus exposure during childhood in Cameroon, Central African Republic and Senegal after the integration of HBV vaccine in the expanded program on immunization. *Pediatr Infect Dis J* 2013;**32**: 1110–1115.