

*Osteoarthritis and Cartilage* (2008) 16, 638–646

© 2008 Osteoarthritis Research Society International. Published by Elsevier Ltd. All rights reserved.

doi:10.1016/j.joca.2008.01.014

# Osteoarthritis and Cartilage

**International  
Cartilage  
Repair  
Society**

## Review

### Should subchondral bone turnover be targeted when treating osteoarthritis?

M. A. Karsdal M.Sc., Ph.D.<sup>†\*</sup>, D. J. Leeming M.Sc.<sup>‡</sup>, E. B. Dam M.Sc., Ph.D.<sup>‡</sup>,  
K. Henriksen M.Sc., Ph.D.<sup>‡</sup>, P. Alexandersen M.Sc., Ph.D.<sup>‡</sup>, P. Pastoureau Ph.D.<sup>§</sup>,  
R. D. Altman M.D.<sup>||</sup> and C. Christiansen M.D.<sup>†</sup>

<sup>†</sup> *Nordic Bioscience A/S, Herlev, Denmark*

<sup>‡</sup> *Center for Clinical and Basic Research, Ballerup Byvej 222, Ballerup, Denmark*

<sup>§</sup> *Institut de Recherche Servier (IdRS), Paris, France*

<sup>||</sup> *David Geffen School of Medicine, University of California-Los Angeles, Los Angeles, USA*

## Summary

**Objective:** Osteoarthritis (OA) is the most common form of arthritic disease, and it is a major cause of disability and impaired quality of life in the elderly. OA is a complex disease of the entire joint, including bone and cartilage, thereby presenting alternative approaches for treatment. This review summarizes emerging observations from cell biology to preliminary clinical trials, describing interactions between the bone and cartilage components. We speculate whether a treatment for OA would be possible without targeting the bone compartment?

**Methods:** Peer-reviewed articles found using pre-defined search criteria and published in the PubMed database until June 2007 are summarized. In addition, abstracts from the OsteoArthritis Research Society International (OARS) conferences in the time period 2000–2007 were included.

**Results:** Bone and cartilage health seem to be tightly associated. Ample evidence is found for bone changes during progression of OA, including, but not limited to, increased turnover in the subchondral bone, thinning of the trabecular structure, osteophytes, bone marrow lesions and sclerosis of the subchondral plate. In addition, a range of investigations has described secondary positive effects on cartilage health when bone resorption was suppressed, or deterioration of the cartilage when resorption is increased.

**Conclusion:** An optimal treatment for OA might include targeting both the bone and cartilage compartments. Hence, as several cell systems are to be targeted in a safe manner, limited options seem possible.

© 2008 Osteoarthritis Research Society International. Published by Elsevier Ltd. All rights reserved.

**Key words:** Osteoarthritis, Osteoclasts, Chondrocytes, Bone, Cartilage, Turnover, Sclerosis, Subchondral, Phenotype.

## Introduction

Osteoarthritis (OA) is the most common form of arthritis<sup>1</sup>. A hallmark of the disease is progressive degeneration of articular cartilage and subsequent joint space narrowing (JSN).

Experimental and clinical observations suggest that the structural integrity of articular cartilage is dependent on normal subchondral bone turnover, intact chondrocyte function and ordinary biomechanical stresses<sup>2,3</sup>. An increasing line of evidence suggests that there are strong inter-relationships between the subchondral bone and the articular cartilage. Therefore, in the face of normal biomechanical stresses, an ideal therapeutic agent might logically be directed at regulating the metabolic activity of both bone and cartilage.

In the majority of patients, the etiology of OA is not known. Among the known risk factors of OA are age,

significant traumata, obesity, altered gait, altered biomechanics (e.g., varus or valgus deformity), and excessive loading<sup>4–7</sup>. Several lines of data indicate that OA could result from a metabolic component as evidenced by the finding that the decreased sex hormone levels in both animal models and women are associated with increased cartilage degradation, in addition to the well described increase in bone turnover<sup>8–12</sup>. Furthermore, the increased bone turnover leads to altered composition and biomechanical properties, which could exuberate the pathogenesis of OA<sup>13–18</sup>.

Most of these studies were realized in humans and focused on late and end-stage disease, which reveals little about early deregulation of cartilage and bone turnover. However, a biochemical investigation of the circulating levels of macromolecules released from cartilage and bone in serum, during the early stage of OA in humans, suggests that pathological processes in cartilage and subchondral bone coincide in OA<sup>19</sup>. From this, there is an obvious interest in studying early bone and cartilage events in animal models and in clinical settings.

The use of biochemical markers has recently been proposed as a supplement to classical imaging techniques for improved diagnosis, and for monitoring of disease

\*Address correspondence and reprint requests to: Dr Morten A. Karsdal, Nordic Bioscience A/S, Herlev Hovedgade 207, DK-2730 Herlev, Denmark. Tel: 45-4452-5216; Fax: 45-4452-5251; E-mail: [mk@nordicbioscience.com](mailto:mk@nordicbioscience.com)

Received 10 October 2007; revision accepted 18 January 2008.

progression in studies of OA<sup>20</sup>. These provide dynamic measures, in contrast to imaging techniques that offer “snap-shots” of the current disease state. These novel techniques have aided in understanding the dynamics of bone and cartilage turnover, and could reveal possible metabolic processes in OA.

In this review we have focused on the role of the bone compartment in OA, and the apparent coupling between cartilage and bone turnover. We have taken into account experimental observations from pre-clinical and clinical studies, including interventions affecting both the bone and cartilage compartments.

Based on this evidence, the question becomes: “Is it possible to inhibit cartilage health deterioration without targeting the joint as a whole organ, thereby involving multiple cell types, but most importantly both osteoclasts and chondrocytes?” Having this difficult scene set for finding a possible disease modifying drug for OA, targeting several cell systems in a safe manner, only a limited number of options seem possible.

## Methods

The PubMed database until September 2007 was searched, using the following search terms: cartilage, OA, subchondral, sclerosis, bone, biomechanical. In addition, articles were reviewed for other relevant references. Additional studies were found in a review of abstracts from the OsteoArthritis Research Society International (OARSI) conferences in the time period 2000–2006.

## Results

### THE ROLE OF SUBCHONDRAL BONE TURNOVER AND STRUCTURE IN OA

The relationship between bone and cartilage degradation in OA is complex, although an apparent co-existence between the two processes is present at a macroscopic observation scale<sup>2</sup>.

Examinations of peri-articular bone in knees and hips of patients with OA have confirmed that the subchondral bone is abnormal in OA joints, with altered trabecular structure and sclerosis of the subchondral plate<sup>21,22</sup>. Bone scintigraphy has revealed localization of the nuclide to sites of increased subchondral bone turnover<sup>23</sup>. Cross-sectional studies have also established that women with advanced

knee or hip OA have higher bone mineral densities (BMDs) near, or at the site of joint OA<sup>24</sup>. In fact, subchondral bone turnover has been shown to be as much as 20-fold increased compared to that of normal bone turnover<sup>14</sup>.

Studies using anterior cruciate ligament transection (ACLT) in dogs as well as meniscectomy (MNX) and ACLT in rat have provided instrumental information on the role of subchondral bone changes in OA<sup>25–27</sup>. During progression of OA the subchondral bone gradually becomes sclerotic, with bone formation in the periphery of the articular cartilage plate, both laterally and medially, resulting in osteophytes<sup>2</sup>. In addition, trabecular bone in the subchondral regions is thinned and elasticity is lost<sup>25,27,28</sup>. Findings, which have been speculated to be important for OA progression<sup>25,27,28</sup>. This combination of changes in the bone matrix are important features in the pathology of OA, although they are thought to be secondary effects caused by the loss of articular cartilage<sup>2,3,13–17,26,29–33</sup>. This is sought illustrated in Fig. 1.

Subchondral bone is separated from the articular cartilage only by a layer of calcified cartilage<sup>34</sup>. This allows for several possibilities of transmitting signals from one compartment to the other. Among these are increased vascularization, and the development of microcracks occurring in the bone matrix, both phenomena which have been strongly implicated in initiation of bone remodeling, as well as increased degradation of the calcified cartilage<sup>34,35</sup>. In alignment, in race horses, micro-crack propagation in the calcified cartilage and subchondral bones, was speculated to be involved in the initiation of articular cartilage breakdown, and thereby initiation and progression of OA<sup>36–38</sup>.

Subchondral bone turnover may logically depend on osteoclastic activities. Osteoclasts have been shown to play important roles in the degradation of cartilage in rheumatoid arthritis (RA)<sup>39–42</sup>, whereas the role of osteoclasts in the pathogenesis of OA still needs more attention. Support for the important role of osteoclast function and bone turnover in the pathogenesis of OA, recently came from an animal model of OA, where extensive inhibition of bone resorption resulted in a 50% decrease in cartilage pathology score, using the golden standard Mankin score<sup>3,26</sup>. In addition, a bone densitometric evaluation in the meniscectomized guinea pig model of OA revealed typical variations of bone metabolism in the initiation of the OA pathology with an early bone loss at the subchondral level followed

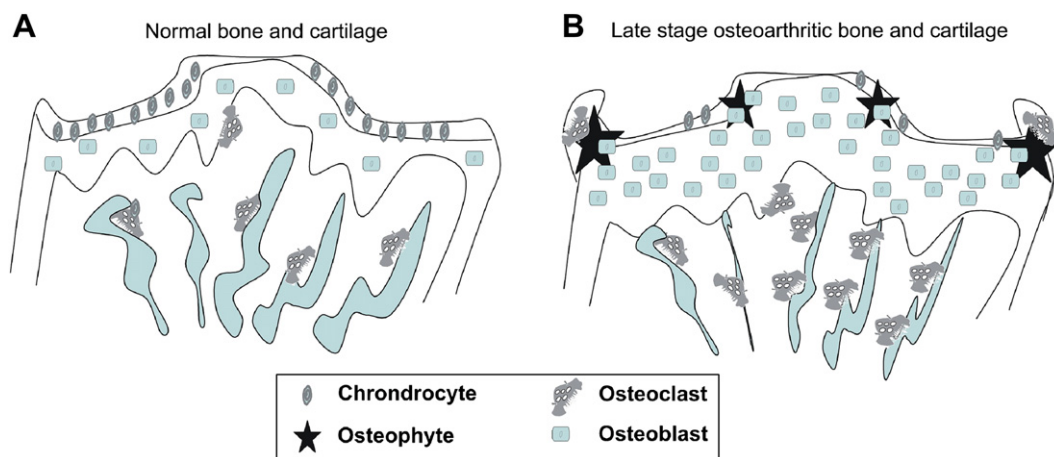


Fig. 1. Schematic illustration of the alterations in the subchondral area. Right panel A: normal tibial plateau. Left panel B: osteoarthritic tibial plateau, showing loss of cartilage, increased bone turnover, sclerosis of the subchondral plate, thinning of the trabeculae, and osteophytes.

by an increased bone density<sup>43</sup>. This finding corroborates previous extensive findings of altered subchondral turnover in the pathogenesis of OA<sup>13–17,29,44</sup>. In alignment, accelerated bone turnover, in both traumatic and estrogen deficiency models [ovariectomy (OVX)], in which increased bone resorption alone results in increased articular damage<sup>31</sup> has been shown to augment articular cartilage erosion<sup>9–12,30</sup>. Finally, evidence from the Dunkin–Hartley guinea pig model indicates that subchondral changes precede the changes in the cartilage<sup>45,46</sup>, thereby further supporting a role for bone remodeling in the development of OA.

Taken together, bone turnover and osteoclast functions are important elements in the pathogenesis of OA, in which both osteoclastic bone resorption and signals coming from osteoclasts may play important roles<sup>47</sup> in the different sclerotic and osteoporotic and sub-compartments of bone.

#### ALTERATIONS IN THE OSTEOBLAST, OSTEOCLAST AND BONE PHENOTYPES IN THE PATHOGENESIS OF OA

Whereas on a macroscopic observational scale there is an almost obvious coupling of bone and cartilage processes<sup>13–17,29</sup>, the cellular and molecular mechanisms remain to be further investigated and identified.

Recently, TGF- $\beta$  signaling which is known to play several important roles in bone and cartilage, was suggested to be involved in the coupling between bone and cartilage turnover<sup>3,13,29</sup>. Furthermore, interleukin (IL)-6 has also been implicated in the cross-talk between bone and cartilage, and studies have shown that IL-6 in combination with other cytokines can switch osteoblasts from a normal phenotype to a sclerotic phenotype<sup>48</sup>, as well as altering expression of different chemokines, such as regulated on activation, normal T cell expressed and secreted (RANTES) and monocyte chemoattractant protein (MCP-1)<sup>49</sup>. In addition, male IL-6 deficient mice shown increased age-related OA<sup>50</sup>, further indicating that IL-6 plays a role in OA. However, the present understanding remains on an observational level, in which an increasing range of experimental evidence is emerging.

With respect to changes at the molecular level, osteoarthritic bone matrix in the subchondral plate has been shown to have alterations in collagen turnover and structure, cytokine expression, as well as mineralization<sup>17,51</sup>. In this, OA results in increased bone formation, assessed by Alkaline Phosphatase and levels of C-terminal pro-peptide of type I collagen (PICP)<sup>29</sup> and associated hypomineralized bone, indicating compromised structural integrity<sup>29</sup>. The compromised bone integrity was corroborated by Bailey *et al.* who demonstrated that the composition of collagen trimers was altered towards homotrimers of the alpha1 chain, which are associated with lower tensile strength<sup>52</sup>. In addition, the hydroxylation of the lysines was increased leading to increased cross-linking, further contorting the collagen structure<sup>52</sup>, and not surprisingly the bone matrix was demonstrated to have impaired mechanical strength<sup>53</sup>, despite the increased density.

With regards to the cellular phenotype, only little information is available. Experiments performed using isolated osteoblasts from OA patients have shown increased proteoglycan degradation in cartilage, in contrast to healthy controls<sup>54</sup>. This has been suggested to be a result of increased matrix metalloproteinase (MMP)-2 expression of these bones<sup>29</sup>. This may indicate a role for altered osteoblast phenotypes during disease development, resulting in the above-described alterations in matrix function and composition. In addition, a group has shown that osteoblasts

from the sclerotic regions of bone isolated from OA patients could be involved in induction of chondrocyte hypertrophy<sup>55</sup>. Furthermore, the same group also demonstrated that the sclerotic osteoblasts were capable of switching the profile of chondrocytes towards cartilage degradation, as illustrated by a reduction in aggrecan production, but an upregulation of MMP production<sup>48</sup>. Finally, a study showed that the bone remodeling rates in healthy individuals correlate to receptor activator of nuclear factor-kappaB ligand (RANKL) expression levels, whereas in OA patients this correlation is no longer existing<sup>56</sup>. As osteoblasts are a major source of RANKL<sup>57,58</sup>, this further supports the view that changes in osteoblasts are involved in OA pathology and progression.

With respect to the phenotype of osteoclasts in the pathogenesis of OA, only sparse information is available. Recently, a study showed that a powerful anti-resorptive, the bisphosphonate alendronate, suppressed cartilage degradation and formation of osteophytes in a rat model, an effect most likely mediated *via* a reduction in osteoclast numbers and activity<sup>3</sup>. In this, alendronate treatment reduced the release of TGF- $\beta$ <sup>3</sup>, a cytokine which is present at abnormally high levels in OA<sup>59</sup>, and speculated to be important in the coupling of cartilage and bone turnover<sup>13</sup>. The exact role and phenotype of osteoclasts in the development of OA needs attention. It is of major interest to investigate whether the osteoclasts involved in the development of OA, indeed are authentic osteoclasts or chondroclasts, i.e., whether osteoclasts localized on calcified cartilage in the subchondral bone are different from those on normal bone matrix, as these may not be identical<sup>60,61</sup>. Drawing a parallel to rheumatoid arthritis (RA), which is characterized by excessive osteoclast function, and in models of RA, anti-osteoclastic treatments, such as anti-RANKL approaches, reduce the extent of the disease<sup>42,62</sup>.

Taken together, it appears that osteoclastic bone resorption, bone formation by the osteoblasts, and signals coming from both osteoclasts and osteoblasts may play important roles in the mal-metabolism of the subchondral bone during pathogenesis of OA<sup>13,47</sup>.

#### A POSSIBLE BIOMECHANICAL COUPLING OF BONE AND CARTILAGE

The possible coupling between bone and cartilage has also been investigated using novel magnetic resonance imaging (MRI) techniques. MRI has been validated for analysis of trabecular structure by comparison to  $\mu$ CT, in which in particular 3 T imaging has demonstrated a high correlation between MRI and  $\mu$ CT based measures<sup>63,64</sup>. Blumenkrantz *et al.* undertook MRI analyses, which were applied to compare longitudinal changes in bone density and quality, assessed by apparent trabecular structure, which were correlated to changes in cartilage volume and quality<sup>63</sup>. They concluded that there was a positive correlation between cartilage loss and localized bone changes closest to the joint line, specifically; the cartilage loss and the decrease in apparent trabecular count were correlated in both the medial ( $r=0.36$ ,  $P<0.05$ ) and the lateral ( $r=0.41$ ,  $P<0.05$ ) tibial chondyles. Furthermore, negative correlations were observed with the bone changes farthest from the joint line (e.g., the medial tibial cartilage volume was negatively correlated with the apparent bone volume fraction of the tibia,  $r=-0.53$ ,  $P<0.05$ ). They hypothesized that cartilage degeneration coincides locally with subchondral bone sclerosis, which then causes subchondral

bone osteopenia due to decreased load transmission<sup>64</sup>. Reactive bone formation may then occur farther away from the cartilage abnormality in order to compensate for the localized bone loss.

Applying the same methodology, Lindsey and colleagues found that cartilage loss in the medial compartment directly correlated to bone formation in the medial chondyle, and to bone resorption in the lateral compartment. Specifically, the medial tibial cartilage volume was positively correlated to lateral apparent trabecular count ( $r=0.33$ ,  $P=0.005$ )<sup>65</sup>. This relationship is illustrated in Fig. 2. Furthermore, by analyzing differences between lateral/medial measurements, they hypothesized that these cross-compartmental relationships are consistent with biomechanical changes in load distribution caused by varus mal-alignment. Using conventional radiographs Messent *et al.* presented similar findings. From knee macro-radiographs of 40 patients, they demonstrated that JSN was associated with a decrease of the subchondral trabecular number<sup>66</sup>. They hypothesized that thickening of the cortical plate; occurring before JSN, caused a load redistribution that reduced the subarticular stress leading to osteoporosis (OP).

These studies support a biomechanical coupling relationship between cartilage and bone in OA, although they do not provide clear indications of the causal relationship. The observation that cortical thickening occurs before JSN does not necessarily imply that bone changes are prior to cartilage changes, since JSN occurs relatively late in the evolution of OA. However, these studies do provide additional support for the close spatial and temporal relationship between bone and cartilage.

#### METABOLIC OA

Metabolic OA, showing a systemic increased turnover of cartilage, is found in postmenopausal women in whom OA is more pronounced, than in the pre-menopausal population<sup>8,67-70</sup>. An epidemiological study supports the concept that women, in addition to age, are at a higher risk than men<sup>71</sup>. This indirectly supports the hypothesis that sex hormones are related to the incidence of OA. In addition, experimental studies in monkeys<sup>30,31</sup> and rats<sup>72</sup> have shown that estrogen depletion results in increased bone turnover, and accelerated cartilage breakdown. These observations are somewhat complicated by the fact that chondrocytes have been shown to express estrogen receptors<sup>73-75</sup>, and respond to estrogen intervention<sup>76</sup>, however,

it is not believed that this accounts for the entire detrimental effect of estrogen loss on cartilage health<sup>77</sup>.

A relationship between OP and OA has been demonstrated in female rabbits<sup>9</sup>. Here induction of OA was performed by anterior cruciate section and partial medial MNX in 12 animals, and OP by OVX in six of these prior to OA induction. OP increased the severity of OA compared to animals without OP and an inverse correlation was observed between BMD at the lumbar spine and cartilage damage defined as increased Mankin scoring. These data indicate that a higher bone turnover exacerbates the OA characteristics. In alignment, development of OA was followed in mice having destabilization of the medial meniscus (DMM). Half of these animals were orchidectomized (ORX, male mice) or OVX (female mice)<sup>10</sup>. OA severity was markedly higher in DMM male mice than DMM female mice. However, OVX female mice developed more severe OA compared to controls, whereas ORX male mice develop significant less severe OA than control mice.

Taken together, this compilation of data indicates that increases in bone turnover, due to sex hormone deficiency, play a critical role in the progression of OA in both mouse models and humans, even though potential direct hormone effects on the chondrocytes cannot be ruled out.

#### BIOCHEMICAL MARKERS OF BONE AND CARTILAGE TURNOVER

Bone and cartilage degradation can be measured by biochemical markers, such as bone resorption by cross-linked C-terminal type I collagen fragments (CTX-I), reflecting type I collagen<sup>78</sup> and cartilage degradation by CTX-II, reflecting type II collagen<sup>79</sup>. These markers have been used extensively for the evaluation of these parameters in basic, experimental and clinical studies<sup>20</sup>.

In a subgroup of the OFELY (Os des Femmes de Lyon) study, 435 healthy untreated younger postmenopausal women were followed for 5 years to predict the risk of fracture<sup>80</sup>. Women in the highest quartile of baseline bone marker level were found to have a relative risk of fracture of 2.1, emphasizing the relevance and importance of the biochemical markers of bone turnover. With respect to cartilage degradation and the use of biochemical markers, a cross-sectional and longitudinal study of 1235 men and women from Rotterdam examined the association between CTX-II and the radiographic prevalence and progression of OA in the knee and hip<sup>81</sup>. Subjects with CTX-II levels in the highest quartile at baseline had a 4.2-fold increased risk of

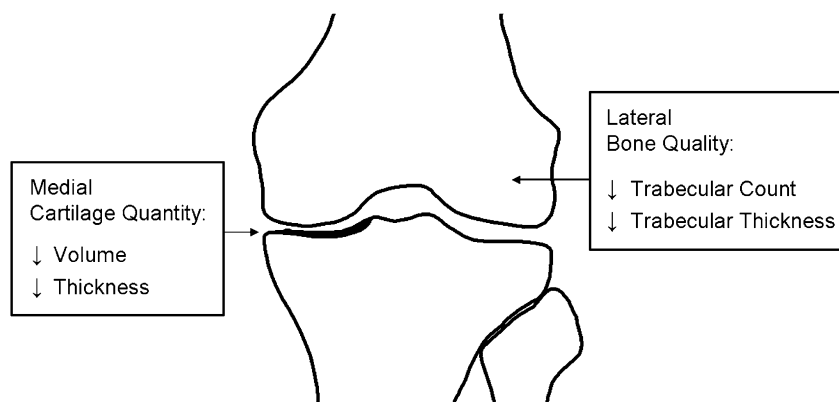


Fig. 2. Biomechanical coupling: medial cartilage loss is correlated to loss of lateral trabecular structure<sup>65</sup> revealing a potential biomechanical coupling between bone and cartilage. Illustration inspired by Lindsey *et al.*<sup>65</sup>



having radiographic OA of the knee and hip, as compared to subjects in the lowest quartile. The risk of disease progression was 6.0-fold increased in the knee and 8.4-fold increased in the hip of the same patients determined at follow-up. Thus, the highest risk of OA and progression was found in patients with the highest baseline levels of CTX-II. A similar finding, namely that those individual with the highest CTX-II levels had the highest odds ratio for having OA was validated in the genetics, arthrosis and progression (GARP) study of sibling pairs, in which symptomatic OA and the levels of a number of biochemical markers were assessed<sup>82</sup>. Results showed a direct correlation between CTX-II and radiographic signs of OA in signals joints ( $P < 0.001$ )<sup>48</sup>, and that those with the highest CTX-II levels had a 7.7 increased odd ratio for having OA.

Taken together, biochemical markers of bone and cartilage destruction may be valuable for the interpretations of the biological processes and for determining the sequence of bone and cartilage changes, leading to a better understanding of disease progression in OA.

#### ANTI-RESORPTIVE STRATEGIES IN OA: NON-CLINICAL STUDIES AND ANTI-RESORPTIVE TREATMENTS

An alternative approach to investigate the coupling between bone and cartilage turnover and deterioration in the pathogenesis in OA is through analysis of the response to various treatments in both pre-clinical and clinical studies. A 3-year study examined the effect of anti-resorptive estrogen replacement therapy (ERT) for the prevention of OA in 180 female cynomolgus monkeys<sup>30,31</sup>. Ovariectomized adult monkeys were divided into groups receiving ERT or receiving no treatment. Significantly fewer cartilage lesions of OA were seen in the ERT group compared to the control group, as evaluated by quantitative histology. The authors suggested that high bone turnover co-equals high risk for cartilage degradation and that exogenous estrogens may confer protection against the development of OA. As mentioned previously, Duong and colleagues investigated the effects of bone turnover and development of OA by the use of intravenous (i.v.) bisphosphonate treatment in a traumatic model of OA, the ACLT model<sup>3,26</sup>. Bisphosphonate treatment resulted in a 50% decrease in disease severity scores OA, further emphasizing the importance and coupling between bone and cartilage health.

Additional support for the role of the subchondral bone was found in studies examining the effect of calcitonin in ACLT dogs. Calcitonin significantly affected trabecular structure and prevented subchondral bone resorption and trabecular thinning, which was speculated to a major factor in the reduced cartilage degradation<sup>83–85</sup>. The mode of action of calcitonin may be different compared to that of other anti-resorptives, as calcitonin was demonstrated to have both direct and indirect actions on articular cartilage<sup>83–95</sup>. In golden standard traumatic and non-traumatic animal models of OA, i.e., the dog ACLT model, calcitonin has been demonstrated to have positive effects on articular cartilage surface erosion and bone structure as well as increased proteoglycan content of the articular cartilage<sup>84,86,95,96</sup>.

Taken together, this increasing line of evidence, points toward the fact that that some anti-resorptive treatments, such as bisphosphonates, calcitonin, estrogen or selective estrogen-receptor modulators (SERMs) have positive effects on both cartilage and bone degradation, possibly due to a tight coupling between these compartments. Some of these possible intervention options may, in addition to the effects on osteoclasts and bone turnover,

provide additional benefits by targeting chondrocytes directly.

#### ANTI-RESORPTIVE STRATEGIES IN OA: CLINICAL STUDIES

A number of studies have investigated the coupling between bone and cartilage degradation. We have previously demonstrated the therapeutic benefit of a SERM, levormeloxifene, in 301 postmenopausal women from a phase II trial, for the prevention of both bone loss and cartilage degradation by restoring levels to that of the pre-menopausal level during a 12-month period<sup>12</sup>. Both cartilage and bone degradation assessed by CTX-II and CTX-I were decreased by approximately 50% in the treatment groups compared to baseline. After treatment cessation CTX-II reversed back to baseline values, indicating a short-term effect of the SERM on the cartilage as compared to that on bone. Similarly, another group found that cartilage degradation in 384 postmenopausal women was significantly lower in women using HRT compared to women not receiving hormone replacement therapy (HRT)<sup>8</sup>. Additionally, groups receiving between 4 and 10 years of HRT had significant lower CTX-II than those receiving less than 4 years of treatment. It was observed that degradation was significantly higher in postmenopausal women compared to an age-matched group of pre-menopausal women.

Effects of calcitonin on both bone and cartilage have raised interest in using it as a therapy for OA. Calcitonin was demonstrated to have chondroprotective effects in two different clinical trials<sup>93,96</sup>. In these, calcitonin showed both effects on pain scores, and biochemical markers of cartilage degradation. However, additional clinical studies are needed to further investigate and validate these findings.

In some instances administration of anti-resorptive drugs to patients cause a decrease in both cartilage degradation and bone resorption<sup>8,12,97</sup>. In therapy of Paget's disease, one dose intravenously administered zoledronate resulted in a reduction of CTX-II and CTX-I at 1 month<sup>97</sup>. At baseline, CTX-II levels were not elevated compared to a group of age-matched healthy controls. This was in contrast to a ninefold increase in CTX-I compared to controls. Five days after a single 4-mg dose of zoledronate, CTX-II decreased by a median of 25% and then increased to pre-treatment levels 10 days after injection. CTX-I decreased a maximum of 51% at day 10 and levels remained suppressed during the 2 months of the study indicating that zoledronate has a long-term effect on bone resorption as well as a short-term effect on cartilage. This is in contrast of the effect of oral bisphosphonates in which a long-term effect on both and cartilage is observed<sup>98,99</sup>. Thus, different anti-resorptive agents, and even routes of administration, appear to have different effects on cartilage.

## Discussion

The potential coupling and uncoupling of bone and cartilage turnover are illustrated in Fig. 3. Under physiological conditions, bone and cartilage turnover are coupled. Increased bone resorption can exist together with increased cartilage degradation, but within the normal biological variation. Under pathological conditions of extremely high turnover, such as bone metastasis, bone resorption is highly elevated, although cartilage degradation is only minutely affected, thus these processes are uncoupled.

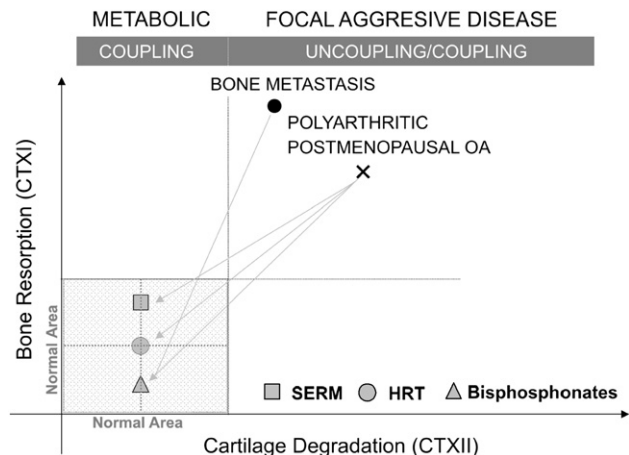


Fig. 3. Schematic figure showing the coupling/uncoupling relationship between cartilage and bone. Under normal physiological conditions, bone and cartilage turnover are coupled (lower left quadrant). In bone metastasis where pathologically increased bone turnover is observed, cartilage turnover is only minimally affected. In contrast, in postmenopausal OA both bone and cartilage are increased, probably due to increased metabolic activity in subchondral bone. Some anti-resorptive agents, such as estrogens, SERMs, bisphosphonates, and calcitonin inhibit increased bone turnover and also the increased cartilage turnover, in part through direct interaction of attenuating subchondral bone turnover, and possibly by direct action receptors activated action on chondrocytes. Additional data on anti-RANKL and strontium ranelate are needed in order to correctly incorporate them into this model.

By treatment with a bisphosphonate the levels of bone resorption can be restored to normal levels, whereas cartilage degradation is only vaguely affected<sup>97</sup>.

In contrast, during postmenopausal increased high turnover, both bone resorption and bone formation can be restored back to normal levels. HRT will restore both levels to pre-menopausal levels, whereas a SERM will have lower efficacy on bone. In alignment, treatment with bisphosphonates will suppress bone resorption to lower levels than cartilage degradation.

Presence and activation of osteoclasts seem to be involved in the pathogenesis of OA and other cartilage degenerative diseases<sup>41,42</sup>. Increased expression of osteoclast differentiation factors RANKL is known to occur in the synovial membrane of chronically OA joints, leading to secondary bone resorption<sup>39,100</sup>. In similar context, another group recently investigated the expression of bone resorption genes in OA and OP, in relation to differences in RANKL regulation of proteases<sup>101</sup>. In this, cathepsin K, MMP-9 and tartrate resistant acid phosphatase (TRACP) mRNA were transcribed at higher levels per given RANKL expression in the OA group compared to that in the fracture (OP) group<sup>101</sup>. This may indicate that blockade of osteoclast formation may constitute a potential target in preventing the structural damage seen in OA. Novel monoclonal antibodies neutralizing the RANKL may prove interesting in preventing both OP and OA-associated bone loss, but at present clinical data are sparse. In alignment, the recently approved anti-resorptive strategy strontium ranelate, might in addition have beneficial effects on cartilage health, although the anti-resorptive effect compared to that of anti-RANKL, SERMs, calcitonin and bisphosphonates is lower<sup>102</sup>.

This compilation of evidence suggests that bone and cartilage turnover are tightly associated, and that some

selected anti-resorptive treatments may have additional benefits on cartilage health. Does this suggest that stimulation of bone formation may have adverse effects? Bone sclerosis is an important part of the pathogenesis of OA<sup>103–107</sup>, that leads to deformation of the articular surfaces and absorption of local stresses producing an effect similar to stress-shielding that may augment OA progression. Recently wnt signaling was shown to be essential for osteophyte generation<sup>40</sup>, and bone sclerosis in an animal model of cartilage destruction. In addition, upregulation of genes in the wnt family has also been observed<sup>108</sup>. Finally, there is evidence indicating that low-density lipoprotein receptor-related protein (LRP)5 haplotypes are associated with OA<sup>109,110</sup>. An increasing amount of attention is focused on bone anabolic drugs, with special emphasis on wnt enhanced signaling. Will these interventions be associated with augmented OA symptoms and disease progression? These important questions of safety need to be addressed in future studies specifically designed for that purpose.

Due to the unmet medical need and the widespread nature of OA, it is urgent to develop safe and effective disease modifying treatments for patients with OA. Ample evidence is found for a tight local coupling between bone and cartilage. Thus, an optimal treatment option for OA would entail both bone and cartilage protective effects. However, with this information limited options seem possible. Some estrogens compounds and calcitonin may provide this dual actions approach, however, still more clinical research is needed.

## Conflict of interest

All authors declare that the affiliation declares full disclosure. In addition, MAK and CC own stock in Nordic Bioscience.

## References

- Abramson SB, Attur M, Yazici Y. Prospects for disease modification in osteoarthritis. *Nat Clin Pract Rheumatol* 2006;2:304–12.
- Felson DT, Neogi T. Osteoarthritis: is it a disease of cartilage or of bone? *Arthritis Rheum* 2004;50:341–4.
- Hayami T, Pickarski M, Wesolowski GA, McLane J, Bone A, Destefano J, *et al*. The role of subchondral bone remodeling in osteoarthritis: reduction of cartilage degeneration and prevention of osteophyte formation by alendronate in the rat anterior cruciate ligament transection model. *Arthritis Rheum* 2004;50:1193–206.
- Sun BH, Wu CW, Kalunian KC. New developments in osteoarthritis. *Rheum Dis Clin North Am* 2007;33:135–48.
- Richette P, Corvol M, Bardin T. Estrogens, cartilage, and osteoarthritis. *Joint Bone Spine* 2003;70:257–62.
- Gelber AC, Hochberg MC, Mead LA, Wang NY, Wigley FM, Klag MJ. Joint injury in young adults and risk for subsequent knee and hip osteoarthritis. *Ann Intern Med* 2000;133:321–8.
- Bijlsma JW, Knahr K. Strategies for the prevention and management of osteoarthritis of the hip and knee. *Best Pract Res Clin Rheumatol* 2007;21:59–76.
- Mouritzen U, Christgau S, Lehmann HJ, Tanko LB, Christiansen C. Cartilage turnover assessed with a newly developed assay measuring collagen type II degradation products: influence of age, sex, menopause, hormone replacement therapy, and body mass index. *Ann Rheum Dis* 2003;62:332–6.
- Calvo E, Castaneda S, Largo R, Fernandez-Valle ME, Rodriguez-Salvanes F, Herrero-Beaumont G. Osteoporosis increases the severity of cartilage damage in an experimental model of osteoarthritis in rabbits. *Osteoarthritis Cartilage* 2007;15:69–77.
- Ma HL, Blanchet TJ, Peluso D, Hopkins B, Morris EA, Glasson SS. Osteoarthritis severity is sex dependent in a surgical mouse model. *Osteoarthritis Cartilage* 2007;15:695–700.
- Hoegh-Andersen P, Tanko LB, Andersen TL, Lundberg CV, Mo JA, Heegaard AM, *et al*. Ovariectomized rats as a model of

- postmenopausal osteoarthritis: validation and application. *Arthritis Res Ther* 2004;6:R169–80.
12. Christgau S, Tanko LB, Cloos PA, Mouritzen U, Christiansen C, Delaisse JM, *et al.* Suppression of elevated cartilage turnover in postmenopausal women and in ovariectomized rats by estrogen and a selective estrogen-receptor modulator (SERM). *Menopause* 2004;11:508–18.
  13. Mansell JP, Collins C, Bailey AJ. Bone, not cartilage, should be the major focus in osteoarthritis. *Nat Clin Pract Rheumatol* 2007;3:306–7.
  14. Bailey AJ, Mansell JP, Sims TJ, Banse X. Biochemical and mechanical properties of subchondral bone in osteoarthritis. *Biorheology* 2004;41:349–58.
  15. Mansell JP, Bailey AJ. Increased metabolism of bone collagen in post-menopausal female osteoporotic femoral heads. *Int J Biochem Cell Biol* 2003;35:522–9.
  16. Bailey AJ, Mansell JP. Do subchondral bone changes exacerbate or precede articular cartilage destruction in osteoarthritis of the elderly? *Gerontology* 1997;43:296–304.
  17. Mansell JP, Tarlton JF, Bailey AJ. Biochemical evidence for altered subchondral bone collagen metabolism in osteoarthritis of the hip. *Br J Rheumatol* 1997;36:16–9.
  18. Davis MA, Ettinger WH, Neuhaus JM, Cho SA, Hauck WW. The association of knee injury and obesity with unilateral and bilateral osteoarthritis of the knee. *Am J Epidemiol* 1989;130:278–88.
  19. Petersson IF, Boegard T, Svensson B, Heinegard D, Saxne T. Changes in cartilage and bone metabolism identified by serum markers in early osteoarthritis of the knee joint. *Br J Rheumatol* 1998;37:46–50.
  20. Schaller S, Henriksen K, Hoegh-Andersen P, Sondergaard BC, Sumer EU, Tanko LB, *et al.* *In vitro*, *ex vivo*, and *in vivo* methodological approaches for studying therapeutic targets of osteoporosis and degenerative joint diseases: how biomarkers can assist? *Assay Drug Dev Technol* 2005;3:553–80.
  21. Hunter DJ, Hart D, Snieder H, Bettica P, Swaminathan R, Spector TD. Evidence of altered bone turnover, vitamin D and calcium regulation with knee osteoarthritis in female twins. *Rheumatology (Oxford)* 2003;42:1311–6.
  22. Hunter DJ, Spector TD. The role of bone metabolism in osteoarthritis. *Curr Rheumatol Rep* 2003;5:15–9.
  23. Dieppe P, Cushnaghan J, Young P, Kirwan J. Prediction of the progression of joint space narrowing in osteoarthritis of the knee by bone scintigraphy. *Ann Rheum Dis* 1993;52:557–63.
  24. Arden N, Nevitt MC. Osteoarthritis: epidemiology. *Best Pract Res Clin Rheumatol* 2006;20:3–25.
  25. Wohl GR, Shymkiw RC, Matyas JR, Kloiber R, Zernicke RF. Periarticular cancellous bone changes following anterior cruciate ligament injury. *J Appl Physiol* 2001;91:336–42.
  26. Hayami T, Pickarski M, Zhuo Y, Wesolowski GA, Rodan GA, Duong LT. Characterization of articular cartilage and subchondral bone changes in the rat anterior cruciate ligament transection and meniscectomized models of osteoarthritis. *Bone* 2005.
  27. Boyd SK, Matyas JR, Wohl GR, Kantzas A, Zernicke RF. Early regional adaptation of periarticular bone mineral density after anterior cruciate ligament injury. *J Appl Physiol* 2000;89:2359–64.
  28. Boyd SK, Muller R, Zernicke RF. Mechanical and architectural bone adaptation in early stage experimental osteoarthritis. *J Bone Miner Res* 2002;17:687–94.
  29. Mansell JP, Bailey AJ. Abnormal cancellous bone collagen metabolism in osteoarthritis. *J Clin Invest* 1998;101:1596–603.
  30. Ham KD, Loeser RF, Lindgren BR, Carlson CS. Effects of long-term estrogen replacement therapy on osteoarthritis severity in cynomolgus monkeys. *Arthritis Rheum* 2002;46:1956–64.
  31. Ham KD, Carlson CS. Effects of estrogen replacement therapy on bone turnover in subchondral bone and epiphyseal metaphyseal cancellous bone of ovariectomized cynomolgus monkeys. *J Bone Miner Res* 2004;19:823–9.
  32. Carbone LD, Nevitt MC, Wildy K, Barrow KD, Harris F, Felson D, *et al.* The relationship of antiresorptive drug use to structural findings and symptoms of knee osteoarthritis. *Arthritis Rheum* 2004;50:3516–25.
  33. Garnero P, Peterfy C, Zaim S, Schoenharting M. Bone marrow abnormalities on magnetic resonance imaging are associated with type II collagen degradation in knee osteoarthritis: a three-month longitudinal study. *Arthritis Rheum* 2005;52:2822–9.
  34. Burr DB, Radin EL. Microfractures and microcracks in subchondral bone: are they relevant to osteoarthritis? *Rheum Dis Clin North Am* 2003;29:675–85.
  35. Lajeunesse D, Reboul P. Subchondral bone in osteoarthritis: a biologic link with articular cartilage leading to abnormal remodeling. *Curr Opin Rheumatol* 2003;15:628–33.
  36. Sokoloff L. Microcracks in the calcified layer of articular cartilage. *Arch Pathol Lab Med* 1993;117:191–5.
  37. Muir P, McCarthy J, Radtke CL, Markel MD, Santschi EM, Scollay MC, *et al.* Role of endochondral ossification of articular cartilage and functional adaptation of the subchondral plate in the development of fatigue microcracking of joints. *Bone* 2006;38:342–9.
  38. Norrdin RW, Stover SM. Subchondral bone failure in overload arthritis: a scanning electron microscopic study in horses. *J Musculoskelet Neuronal Interact* 2006;6:251–7.
  39. Schett G, Hayer S, Zwerina J, Redlich K, Smolen JS. Mechanisms of disease: the link between RANKL and arthritic bone disease. *Nat Clin Pract Rheumatol* 2005;1:47–54.
  40. Diarra D, Stolina M, Polzer K, Zwerina J, Ominsky MS, Dwyer D, *et al.* Dickkopf-1 is a master regulator of joint remodeling. *Nat Med* 2007;13:156–63.
  41. Schett G, Stolina M, Bolon B, Middleton S, Adlam M, Brown H, *et al.* Analysis of the kinetics of osteoclastogenesis in arthritic rats. *Arthritis Rheum* 2005;52:3192–201.
  42. Stolina M, Adamu S, Ominsky M, Dwyer D, Asuncion F, Geng Z, *et al.* RANKL is a marker and mediator of local and systemic bone loss in two rat models of inflammatory arthritis. *J Bone Miner Res* 2005;20:1756–65.
  43. Pastoureaux PC, Chomel AC, Bonnet J. Evidence of early subchondral bone changes in the meniscectomized guinea pig. A densitometric study using dual-energy X-ray absorptiometry subregional analysis. *Osteoarthritis Cartilage* 1999;7:466–73.
  44. Sassi ML, Eriksen H, Risteli L, Niemi S, Mansell J, Gowen M, *et al.* Immunohistochemical characterization of assay for carboxyterminal telopeptide of human type I collagen: loss of antigenicity by treatment with cathepsin K. *Bone* 2000;26:367–73.
  45. Quasnichka HL, Anderson-MacKenzie JM, Bailey AJ. Subchondral bone and ligament changes precede cartilage degradation in guinea pig osteoarthritis. *Biorheology* 2006;43:389–97.
  46. Anderson-MacKenzie JM, Quasnichka HL, Starr RL, Lewis EJ, Billingham ME, Bailey AJ. Fundamental subchondral bone changes in spontaneous knee osteoarthritis. *Int J Biochem Cell Biol* 2005;37:224–36.
  47. Karsdal MA, Martin TJ, Bollerslev J, Christiansen C, Henriksen K. Are nonresorbing osteoclasts sources of bone anabolic activity? *J Bone Miner Res* 2007;22:487–94.
  48. Sanchez C, Deberg MA, Piccardi N, Msika P, Reginster JY, Henrotin YE. Osteoblasts from the sclerotic subchondral bone down-regulate aggrecan but upregulate metalloproteinases expression by chondrocytes. This effect is mimicked by interleukin-6, -1beta and oncostatin M pre-treated non-sclerotic osteoblasts. *Osteoarthritis Cartilage* 2005;13:979–87.
  49. Lisignoli G, Toneguzzi S, Grassi F, Piacentini A, Tschon M, Cristino S, *et al.* Different chemokines are expressed in human arthritic bone biopsies: IFN-gamma and IL-6 differentially modulate IL-8, MCP-1 and rantes production by arthritic osteoblasts. *Cytokine* 2002;20:231–8.
  50. de Hooge AS, van de Loo FA, Bennink MB, Arntz OJ, de Hooge P, Van den Berg WB. Male IL-6 gene knock out mice developed more advanced osteoarthritis upon aging. *Osteoarthritis Cartilage* 2005;13:66–73.
  51. Neilson M, White A, Malik U, Morrison E, McGill PE, McDonald SW. Changes in bone architecture in the femoral head and neck in osteoarthritis. *Clin Anat* 2004;17:378–91.
  52. Bailey AJ, Sims TJ, Knott L. Phenotypic expression of osteoblast collagen in osteoarthritic bone: production of type I homotrimer. *Int J Biochem Cell Biol* 2002;34:176–82.
  53. Li B, Aspden RM. Composition and mechanical properties of cancellous bone from the femoral head of patients with osteoporosis or osteoarthritis. *J Bone Miner Res* 1997;12:641–51.
  54. Westacott CI, Webb GR, Warnock MG, Sims JV, Elson CJ. Alteration of cartilage metabolism by cells from osteoarthritic bone. *Arthritis Rheum* 1997;40:1282–91.
  55. Sanchez C, Deberg MA, Piccardi N, Msika P, Reginster JY, Henrotin YE. Subchondral bone osteoblasts induce phenotypic changes in human osteoarthritic chondrocytes. *Osteoarthritis Cartilage* 2005;13:988–97.
  56. Fazzalari NL, Kuliwaba JS, Atkins GJ, Forwood MR, Findlay DM. The ratio of messenger RNA levels of receptor activator of nuclear factor kappaB ligand to osteoprotegerin correlates with bone remodeling indices in normal human cancellous bone but not in osteoarthritis. *J Bone Miner Res* 2001;16:1015–27.
  57. Lacey DL, Timms E, Tan HL, Kelley MJ, Dunstan CR, Burgess T, *et al.* Osteoprotegerin ligand is a cytokine that regulates osteoclast differentiation and activation. *Cell* 1998;93:165–76.
  58. Boyle WJ, Simonet WS, Lacey DL. Osteoclast differentiation and activation. *Nature* 2003;423:337–42.
  59. Massicotte F, Lajeunesse D, Benderdour M, Pelletier JP, Hilal G, Duval N, *et al.* Can altered production of interleukin-1beta, interleukin-6, transforming growth factor-beta and prostaglandin E(2) by isolated human subchondral osteoblasts identify two subgroups of osteoarthritic patients. *Osteoarthritis Cartilage* 2002;10:491–500.
  60. Engsig MT, Chen QJ, Vu TH, Pedersen AC, Therkidsen B, Lund LR, *et al.* Matrix metalloproteinase 9 and vascular endothelial growth factor are essential for osteoclast recruitment into developing long bones. *J Cell Biol* 2000;151:879–89.

61. Ortega N, Behonick D, Stickens D, Werb Z. How proteases regulate bone morphogenesis. *Ann N Y Acad Sci* 2003;995:109–16.
62. Schett G, Middleton S, Bolon B, Stolina M, Brown H, Zhu L, *et al.* Additive bone-protective effects of anabolic treatment when used in conjunction with RANKL and tumor necrosis factor inhibition in two rat arthritis models. *Arthritis Rheum* 2005;52:1604–11.
63. Phan CM, Matsuura M, Bauer JS, Dunn TC, Newitt D, Lochmueller EM, *et al.* Trabecular bone structure of the calcaneus: comparison of MR imaging at 3.0 and 1.5 T with micro-CT as the standard of reference. *Radiology* 2006;239:488–96.
64. Blumenkrantz G, Lindsey CT, Dunn TC, Jin H, Ries MD, Link TM, *et al.* A pilot, two-year longitudinal study of the interrelationship between trabecular bone and articular cartilage in the osteoarthritic knee. *Osteoarthritis Cartilage* 2004;12:997–1005.
65. Lindsey CT, Narasimhan A, Adolfo JM, Jin H, Steinbach LS, Link T, *et al.* Magnetic resonance evaluation of the interrelationship between articular cartilage and trabecular bone of the osteoarthritic knee. *Osteoarthritis Cartilage* 2004;12:86–96.
66. Messent EA, Ward RJ, Tonkin CJ, Buckland-Wright C. Tibial cancellous bone changes in patients with knee osteoarthritis: a short-term longitudinal study using Fractal Signature Analysis. *Osteoarthritis Cartilage* 2005;13:463–70.
67. Oliveria SA, Felson DT, Reed JI, Cirillo PA, Walker AM. Incidence of symptomatic hand, hip, and knee osteoarthritis among patients in a health maintenance organization. *Arthritis Rheum* 1995;38:1134–41.
68. Cauley JA, Kwok CK, Egeland G, Nevitt MC, Cooperstein L, Rohay J, *et al.* Serum sex hormones and severity of osteoarthritis of the hand. *J Rheumatol* 1993;20:1170–5.
69. Spector TD, Perry LA, Jubb RW. Endogenous sex steroid levels in women with generalised osteoarthritis. *Clin Rheumatol* 1991;10:316–9.
70. Sowers MF, Hochberg M, Crabbe JP, Muhich A, Crutchfield M, Updike S. Association of bone mineral density and sex hormone levels with osteoarthritis of the hand and knee in premenopausal women. *Am J Epidemiol* 1996;143:38–47.
71. Cole AA, Kuettner KE. Molecular basis for differences between human joints. *Cell Mol Life Sci* 2002;59:19–26.
72. Pampena DA, Robertson KA, Litvinova O, Lajoie G, Goldberg HA, Hunter GK. Inhibition of hydroxyapatite formation by osteopontin phosphopeptides. *Biochem J* 2004;378:1083–7.
73. Claassen H, Hassenpflug J, Schunke M, Sierralta W, Thole H, Kurz B. Immunohistochemical detection of estrogen receptor alpha in articular chondrocytes from cows, pigs and humans: *in situ* and *in vitro* results. *Ann Anat* 2001;183:223–7.
74. Richmond RS, Carlson CS, Register TC, Shanker G, Loeser RF. Functional estrogen receptors in adult articular cartilage: estrogen replacement therapy increases chondrocyte synthesis of proteoglycans and insulin-like growth factor binding protein 2. *Arthritis Rheum* 2000;43:2081–90.
75. Ushiyama T, Ueyama H, Inoue K, Ohkubo I, Hukuda S. Expression of genes for estrogen receptors alpha and beta in human articular chondrocytes. *Osteoarthritis Cartilage* 1999;7:560–6.
76. Oestergaard S, Sondergaard BC, Hoegh-Andersen P, Henriksen K, Qvist P, Christiansen C, *et al.* Effects of ovariectomy and estrogen therapy on type II collagen degradation and structural integrity of articular cartilage in rats: implications of the time of initiation. *Arthritis Rheum* 2006;54:2441–51.
77. Sondergaard BC, Oestergaard S, Christiansen C, Karsdal MA. The effect of oral calcitonin on cartilage turnover and surface erosion in the ovariectomized rat model. *Arthritis Rheum* 2007;56:2674–8.
78. Bonde M, Qvist P, Fledelius C, Riis BJ, Christiansen C. Immunoassay for quantifying type I collagen degradation products in urine evaluated. *Clin Chem* 1994;40:2022–5.
79. Christgau S, Garnero P, Fledelius C, Moniz C, Ensig M, Gineyts E, *et al.* Collagen type II C-telopeptide fragments as an index of cartilage degradation. *Bone* 2001;29:209–15.
80. Garnero P, Sornay-Rendu E, Claustrat B, Delmas PD. Biochemical markers of bone turnover, endogenous hormones and the risk of fractures in postmenopausal women: the OFELY study. *J Bone Miner Res* 2000;15:1526–36.
81. Reijman M, Hazes JM, Bierma-Zeinstra SM, Koes BW, Christgau S, Christiansen C, *et al.* A new marker for osteoarthritis: cross-sectional and longitudinal approach. *Arthritis Rheum* 2004;50:2471–8.
82. Meulenbelt I, Kloppenburg M, Kroon HM, Houwing-Duistermaat JJ, Garnero P, Heliö-Le Graverand MP, *et al.* Clusters of biochemical markers are associated with radiographic subtypes of osteoarthritis (OA) in subject with familial OA at multiple sites. The GARP study. *Osteoarthritis Cartilage* 2007;15:379–85.
83. Behets C, Williams JM, Chappard D, Devogelaer JP, Manicourt DH. Effects of calcitonin on subchondral trabecular bone changes and on osteoarthritic cartilage lesions after acute anterior cruciate ligament deficiency. *J Bone Miner Res* 2004;19:1821–6.
84. Manicourt DH, Altman RD, Williams JM, Devogelaer JP, Druetz-Van Egeren A, Lenz ME, *et al.* Treatment with calcitonin suppresses the responses of bone, cartilage, and synovium in the early stages of canine experimental osteoarthritis and significantly reduces the severity of the cartilage lesions. *Arthritis Rheum* 1999;42:1159–67.
85. Karsdal MA, Tanko LB, Riis BJ, Sondergaard BC, Henriksen K, Altman RD, *et al.* Calcitonin is involved in cartilage homeostasis: is calcitonin a treatment for OA? *Osteoarthritis Cartilage* 2006;14:617–24.
86. Wronski TJ, Yen CF, Burton KW, Mehta RC, Newman PS, Soltis EE, *et al.* Skeletal effects of calcitonin in ovariectomized rats. *Endocrinology* 1991;129:2246–50.
87. Martin TJ, Harris GS, Melick RA, Fraser JR. Effect of calcitonin on glycosaminoglycan synthesis by embryo calf bone cells *in vitro*. *Experientia* 1969;25:375–6.
88. Baxter E, Fraser JR, Harris GS, Martin TJ, Melick RA. Stimulation of glycosaminoglycan synthesis by thyrocalcitonin preparations. *Med J Aust* 1968;1:216–7.
89. Tanko LB, Bagger YZ, Alexandersen P, Devogelaer JP, Reginster JY, Chick R, *et al.* Safety and efficacy of a novel salmon calcitonin (sCT) technology-based oral formulation in healthy postmenopausal women: acute and 3-month effects on biomarkers of bone turnover. *J Bone Miner Res* 2004;19:1531–8.
90. Heliö MP, Peschard MJ, Cohen C, Richard M, Vignon E. Calcitonin inhibits phospholipase A2 and collagenase activity of human osteoarthritic chondrocytes. *Osteoarthritis Cartilage* 1997;5:121–8.
91. Franchimont P, Bassleer C, Henrotin Y, Gysen P, Bassleer R. Effects of human and salmon calcitonin on human articular chondrocytes cultivated in clusters. *J Clin Endocrinol Metab* 1989;69:259–66.
92. Manicourt DH, Devogelaer JP, Azria M, Silverman S. Rationale for the potential use of calcitonin in osteoarthritis. *J Musculoskelet Neuronal Interact* 2005;5:285–93.
93. Bagger YZ, Tanko LB, Alexandersen P, Karsdal MA, Olson M, Mindeholm L, *et al.* Oral salmon calcitonin induced suppression of urinary collagen type II degradation in postmenopausal women: a new potential treatment of osteoarthritis. *Bone* 2005;37:425–30.
94. Khaldi L, Karachalios T, Galanos A, Lyritis GP. Morphometric changes in the epiphyseal plate of the growing and young adult male rat after long-term salmon calcitonin administration. *Calcif Tissue Int* 2005;76:426–32.
95. El Hajjaji H, Williams JM, Devogelaer JP, Lenz ME, Thonar EJ, Manicourt DH. Treatment with calcitonin prevents the net loss of collagen, hyaluronan and proteoglycan aggregates from cartilage in the early stages of canine experimental osteoarthritis. *Osteoarthritis Cartilage* 2004;12:904–11.
96. Manicourt DH, Azria M, Mindeholm L, Thonar EJ, Devogelaer JP. Oral salmon calcitonin reduces Lequesne's algofunctional index scores and decreases urinary and serum levels of biomarkers of joint metabolism in knee osteoarthritis. *Arthritis Rheum* 2006;54:3205–11.
97. Garnero P, Christgau S, Delmas PD. The bisphosphonate zoledronate decreases type II collagen breakdown in patients with Paget's disease of bone. *Bone* 2001;28:461–4.
98. Spector TD, Conaghan PG, Buckland-Wright JC, Garnero P, Cline GA, Beary JF, *et al.* Effect of risedronate on joint structure and symptoms of knee osteoarthritis: results of the BRISK randomized, controlled trial [SRCTN01928173]. *Arthritis Res Ther* 2005;7:R625–33.
99. Bingham CO III, Buckland-Wright JC, Garnero P, Cohen SB, Dougados M, Adami S, *et al.* Risedronate decreases biochemical markers of cartilage degradation but does not decrease symptoms or slow radiographic progression in patients with medial compartment osteoarthritis of the knee: results of the two-year multinational knee osteoarthritis structural arthritis study. *Arthritis Rheum* 2006;54:3494–507.
100. Andersson MK, Lundberg P, Ohlin A, Perry MJ, Lie A, Stark A, *et al.* Effects on osteoclast and osteoblast activities in cultured mouse calvarial bones by synovial fluids from patients with a loose joint prosthesis and from osteoarthritis patients. *Arthritis Res Ther* 2007;9:R18.
101. Logar DB, Komadina R, Prezelj J, Ostanek B, Trost Z, Marc J. Expression of bone resorption genes in osteoarthritis and in osteoporosis. *J Bone Miner Metab* 2007;25:219–25.
102. Karsdal MA, Qvist P, Christiansen C, Tanko LB. Optimising antiresorptive therapies in postmenopausal women: why do we need to give due consideration to the degree of suppression? *Drugs* 2006;66:1909–18.
103. Lajeunesse D. The role of bone in the treatment of osteoarthritis. *Osteoarthritis Cartilage* 2004;12(Suppl A):S34–8.
104. Buckland-Wright JC, Lynch JA, Dave B. Early radiographic features in patients with anterior cruciate ligament rupture. *Ann Rheum Dis* 2000;59:641–6.
105. Wachsmuth L, Engelke K. High-resolution imaging of osteoarthritis using microcomputed tomography. *Methods Mol Med* 2004;101:231–48.
106. Cake MA, Read RA, Guillou B, Ghosh P. Modification of articular cartilage and subchondral bone pathology in an ovine meniscectomy



- model of osteoarthritis by avocado and soya unsaponifiables (ASU). *Osteoarthritis Cartilage* 2000;8:404–11.
107. Buckland-Wright C. Subchondral bone changes in hand and knee osteoarthritis detected by radiography. *Osteoarthritis Cartilage* 2004; 12(Suppl A):S10–9.
108. Hopwood B, Tsykin A, Findlay DM, Fazzalari NL. Microarray gene expression profiling of human osteoarthritic bone suggests altered bone remodelling, WNT and TGF beta/BMP signalling. *Arthritis Res Ther* 2007;9:R100.
109. Urano T, Shiraki M, Narusawa K, Usui T, Sasaki N, Hosoi T, *et al.* Q89R polymorphism in the LDL receptor-related protein 5 gene is associated with spinal osteoarthritis in postmenopausal Japanese women. *Spine* 2007;32:25–9.
110. Smith AJ, Gidley J, Sandy JR, Perry MJ, Elson CJ, Kirwan JR, *et al.* Haplotypes of the low-density lipoprotein receptor-related protein 5 (LRP5) gene: are they a risk factor in osteoarthritis? *Osteoarthritis Cartilage* 2005;13:608–13.
-