

Tulipa kolbintsevii Zonn., a new species from Eastern Kazakhstan

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Abstract The white-flowered *Tulipa kolbintsevii* (Liliaceae), section *Biflores*, occurs in SE Kazakhstan, Dzungarian Ala-Tau, Taskora valley. So far only *T. aff. altaica* and *T. brachystemon* (sect. *Kolpakowskianae*) both with yellow flowers, flushed greenish–violet or red on the outside, were known to be present there. It differs in that it is not only the easternmost species of the section *Biflores*, and has different placement of hairs on the tepals, but also has the lowest genome size of the species in this section with $2C = 48$ pg. The other 14 species have 51.5–59.4 pg. Flow cytometry supported morphological differences. This method is useful for identification of dormant bulbs or sterile plants and is therefore important for monitoring of the trade in bulbous species.

Keywords Flow cytometry · *Tulipa kolbintsevii* Zonn. sp. nov. · Kazakhstan · Genome size · $2C$ value · Taxonomy

Introduction

The genus *Tulipa* L. (Liliaceae) has at least 87 species (Zonneveld 2009). These are divided in four subgenera, *Tulipa*, *Clusianae* (Baker) Zonn., *Eriostemones* (Boiss.) van Raamsd., and *Orithyia* (D. Don) Baker, and twelve sections (Zonneveld 2009). Genome size and the presence of nearly complete crossing barriers between the sections

(Van Raamsdonk 1992; Van Raamsdonk and De Vries 1995) confirmed the close relationships of the species within the different sections. Tulips occur naturally in southern Europe, North Africa, the Middle East, and Central Asia, including Western China. The Tien Shan and Pamir-Alay mountain ranges in Central Asia are believed to be the primary gene centres for the species (Botschantzeva 1962), with the Caucasus as a secondary centre. Most species have the same basic chromosome number, $2n = 2x = 24$. However, the somatic DNA $2C$ value is shown to range from 32 to 69 picogram for the diploids (Zonneveld 2009). They are popular spring-flowering garden plants, millions of bulbs are sold annually and over 5000 cultivars have been registered (Van Scheepen 1996). Despite the existence of a large body of literature on *Tulipa*, its taxonomy is generally regarded as difficult, as stated in every taxonomic treatment. The main reason is that many character states are polythetic—there is hardly any that is not variable even within a species, only combinations define them: flowering time, the absence or presence and type of hairs on the inside of the bulb tunic, leaf/stem hairiness, flower colour, the absence or presence of a black blotch at the base of the tepals with or without a yellow edge, and whether hairs are present at the base of the filaments. In a previous study DNA $2C$ value (nuclear DNA content) was used for analysis of more than 400 different accessions representing nearly all recognized species (Zonneveld 2009).

Nuclear DNA content can conveniently be measured by flow cytometry using propidium iodide, a stoichiometric DNA stain that intercalates in the double helix. Where many species in a genus have the same chromosome numbers, differences in DNA $2C$ value have proved to be very effective in delimiting infrageneric divisions in a number of taxa (Ohri 1998; Zonneveld 2009). Moreover,

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Greilhuber (1998, 2005) has clearly shown that intraspecific variation of genome size is much less than assumed.

The smallest angiosperm genome size reported so far is for *Genlisea aurea* St.Hil. (received and published as *G. margaretae* Hutch; personal communication, J Greilhuber) (Lentibulariaceae) with $2C = 0.13$ pg (Greilhuber et al. 2006). The record holder for maximum genome size is now, for eudicots, *Viscum album* L. (Viscaceae) with $2C = 205.8$ pg. For monocots, for a short time, the hexaploid *Trillium hageae* Miyabe and Tatew. (Trilliaceae) with 264.9 pg (Zonneveld 2010a) was the record holder. This value was, a few months later, superseded by its relative, the octoploid *Paris japonica* Franch. (Trilliaceae) with $2C = 304.5$ pg (Pellicer et al. 2010). Flow cytometry has been successfully used to measure the $2C$ value for the genera *Agapanthus* L'Hér., *Eucomis* L' Hér., *Galanthus* L., *Helleborus* L., *Hepatica* Mill., *Hosta* Tratt., *Narcissus* L., *Nerine* Herb. and *Tulipa* L. by Zonneveld (2001, 2008, 2009, 2010a, b, c), Zonneveld and Van Iren (2001), Zonneveld and Duncan (2003, 2006), and Zonneveld et al. (2003). In these papers it was shown that many species can be distinguished by their genome size.

During investigations of the nuclear DNA content ($2C$ value) of species of *Tulipa* (Zonneveld 2009) several were encountered that had genome sizes differing from all other species of their section. They could not be described as new species at that time, because of incomplete material. *Tulipa lemmersii* Zonn., Peterse & J. de Groot from the Mashat canyon, Kazakhstan, was provisionally mentioned by Zonneveld (2009) and later validated by Veldkamp and Zonneveld (2012). Here, another interesting new species is reported.

Materials and methods

Plant material

Plant material was obtained from the collection of JJ de Groot, De Zilk, The Netherlands.

Flow-cytometric measurement of DNA $2C$ value

For isolation of nuclei, approximately 1 cm of leaf was chopped with a piece of *Clivia miniata* (Lindl.) Regel as internal standard. The material was chopped with a new razor blade in a Petri dish in 0.25 ml nuclei-isolation buffer to which 0.25 mg RNase/ml was added (Zonneveld and van Iren 2001). After adding 1.75 ml propidium iodide solution (50 mg PI/l in isolation buffer) the suspension with nuclei was filtered through a 30- μ m nylon filter. The fluorescence of the nuclei was measured 30 min and 1 h after addition of propidium iodide, by use of a Partec CA-II

flow cytometer. The optical path contained an HBO mercury lamp, filters KG1 and BG12, dichroic mirror TK500, filter OG570, and a Leitz 50×1 water-immersion objective. Results were analysed by means of DPAC software (Partec). The $2C$ DNA content of the sample was calculated as the sample peak mean, divided by the *Clivia* peak mean, and multiplied with the amount of DNA of the *Clivia* standard. At least three different samples, each with at least 5000 nuclei, were measured twice. For most histograms the coefficient of variation was $<5\%$. Fresh male human leucocytes ($2C = 7.0$ pg; 1 picogram = 10^{-12} g = 0.978×10^9 base pairs; Doležel et al. 2003) were chosen as primary standard (Tiersch et al. 1989). This yields $2C = 39.0$ pg for nuclei of *Clivia miniata*.

Results

Tulipa kolbintsevii Zonn. sp. nov. Figs. 1, 2, 3, 4.

Tulipae regelii similissima, sed foliis supra non porcatis differt. Orientalissima specierum 15 ceterarum sectionis *Biflorium*, tepala basi sparse pubescentia secus marginem, demum post meridiem florens.

Nucleo 48 pg tantum ponderenti contra 51.5–59.4 pg in sectionis speciebus ceteris.

Type: Taskora valley, 650 m Dzungarian Ala-Tau, Kazakhstan (2010). cult. JJ de Groot s.n. (Holotype L. barcode L 0821329).

Stature tulip erect in all stages, up to 15 cm.

Bulb globose, 20 mm diam.; tunic pale brown, extension 20 mm long, inside glabrous, at the neck with some hairs.

Stem erect, glabrous, single flowered, purplish–brown, underground part 10 cm, whitish, supraterranean part c. 12 cm.

Leaves two, canaliculate, greyish green, produced at ground level, glabrous and without cilia, the lower one 145×7 mm, the upper one 130×10 mm.



Fig. 1 *Tulipa kolbintsevii* growing in a garden in de Zilk, The Netherlands (all four pictures are from JJ de Groot)



Fig. 2 Bulbs of *Tulipa kolbintsevii*



Fig. 3 Details of the dissected flower of *Tulipa kolbintsevii*



Fig. 4 Details of the expanded flower of *Tulipa kolbintsevii*

Flower bud straight, obclavate, inner and outer tepals inside white with a whitish–yellow to yellow–orange blotch, c. 2 cm diam.

Outer tepals 26 × 7 mm, base margins sparsely hairy, outside greyish–green, in upper part flanked on both sides with a 1 mm white line and pinkish brown line 1 mm wide.

Inner tepals 28 × 8 mm, base margins hairy for 5 mm, outside with a narrow greenish vein, both sides in the upper part with a pinkish brown line.

Stamens longer than the ovary, unequal, 10 and 13 mm long. Filaments yellow, base hairy in the lower third. Anthers 7 mm long, yellow, pollen yellow.

Ovary 10 mm high, green, stigma sessile, yellow–white.

Distribution. Known from its type locality, Taskora valley and adjacent Kolasu valley, Dzungarian Ala-Tau, East Kazakhstan.

Habitat: Between low bushes, 650 m alt.

Phenology. Flowering March–April, both in nature and in the garden, opening only at midday similar to *T. patens* C. Agardh ex Schult f and *T. primulina* Baker. (sect. *Sylvestres* (Baker) Baker).

Discussion

The Taskora valley at 650 m. in the Dzungarian Ala-Tau, East Kazakhstan is poor in the number of tulip species. Only *T. aff. altaica* and *T. brachystemon* have been found so far (Pratov et al. 2006; Ivaschenko 2005). Both have yellow flowers, flushed red or greenish violet on the outside, and belong to section *Kolpakowskianae*. This new white flowered species belongs to section *Biflores* AD Hall ex Zonn. and Veldk. which contains 14 other species (Zonneveld 2009). *Tulipa kolbintsevii* is similar to *T. regelii* Elwes. in many respects including the bulb, but the latter has unique ridges on the upper side of the leaf. *Tulipa kolbintsevii* is diploid with a nuclear DNA content of 48.0 pg, lower than all the others of the section. These range from 51.5 to 59.4 pg (Zonneveld 2009). It is, moreover, also its easternmost species. The tepals have at the base a hairy edge, not a hairy band, as in the other species of section *Biflores*.

Etymology. This tulip is dedicated to Vladimir Kolbintsev who, as a guide, accompanied the expedition during which this species was found. He guided many other Dutch tulip expeditions and is an eminent connoisseur of the wild middle Asiatic (tulip) flora and fauna.

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