

Review

Apoptotic pathways in pancreatic ductal adenocarcinoma

Rainer Hamacher, Roland M Schmid, Dieter Saur and Günter Schneider*

Address: II. Medizinische Klinik, Klinikum rechts der Isar, Technische Universität München, München, Germany

Email: Rainer Hamacher - rainer.hamacher@lrz.tu-muenchen.de; Roland M Schmid - roland.schmid@lrz.tu-muenchen.de;Dieter Saur - dieter.saur@lrz.tu-muenchen.de; Günter Schneider* - guenter.schneider@lrz.tum.de

* Corresponding author

Published: 24 July 2008

Received: 21 April 2008

Molecular Cancer 2008, **7**:64 doi:10.1186/1476-4598-7-64

Accepted: 24 July 2008

This article is available from: <http://www.molecular-cancer.com/content/7/1/64>

© 2008 Hamacher et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Pancreatic ductal adenocarcinoma (PDAC) is one of the most common causes of cancer related death. Despite the advances in understanding of the molecular pathogenesis, pancreatic cancer remains a major unsolved health problem. Overall, the 5-year survival rate is less than 5% demonstrating the insufficiency of current therapies.

Most cytotoxic therapies induce apoptosis and PDAC cells have evolved a plethora of molecular mechanisms to assure survival. We will present anti-apoptotic strategies working at the level of the death receptors, the mitochondria or involving the caspase inhibitors of the IAP family. Furthermore, the survival function of the phosphatidylinositol-3' kinase (PI3K)/AKT- and NF-kappaB-pathways are illustrated. A detailed molecular knowledge of the anti-apoptotic mechanisms of PDAC cells will help to improve therapies for this dismal disease and therapeutic strategies targeting the programmed cell death machinery are in early preclinical and clinical development.

Introduction

Pancreatic ductal adenocarcinoma (PDAC) is one of the most malignant tumor with an unfavorable prognosis. More than 30.000 people develop pancreatic adenocarcinoma each year in the United States, and almost all are expected to die from the disease. Although the incidence of pancreatic cancer is only about 10 in 10⁵, it is the fourth male and female leading cause of cancer-related death [1]. One reason for the poor prognosis of PDAC is the insensitivity to most therapies like chemotherapy, radiotherapy and immunotherapy. Therefore, surgical resection offers at the moment the only potential chance for cure. The 5-year survival rate of all patients is below 5%, and the median survival time after diagnosis is 6 months. Furthermore, only about 20% of patients curative resected survive longer than 5 years [2].

The hallmarks of nearly all cancers are deregulation of the cell cycle machinery, self-sufficiency in growth signals, insensitivity to growth inhibitory signals, evasion of apoptosis, tissue invasion, metastasis and sustained angiogenesis [2-4]. These characteristic changes can also be found in PDAC and are detailed discussed in excellent reviews from the groups of Depinho, Maitra, Hruban and Friess [5-7]. The demonstration that the fraction of apoptotic cells in PDAC predicts overall survival shows the important contribution of the apoptotic machinery towards the tumorbiology of PDAC [8].

Most chemotherapies act by induction of apoptosis. Therefore, evasion of apoptosis is mainly responsible for the insufficiency of current therapies [9]. Tumor cells use multiple pathways to escape apoptosis [10]. This review

will focus on the deregulation of apoptotic pathways in PDAC.

Apoptosis – An overview

Apoptosis or programmed cell death is a central regulator of normal tissue homeostasis. The physiological "cell suicide" program is essential for the elimination of redundant, damaged and infected cells [11,12]. Disturbed apoptosis is involved in the pathogenesis of multiple diseases, especially cancer.

Execution of apoptosis relies on a group of cysteine proteases, the caspases [13]. Caspases are synthesized as pro-forms and become activated by cleavage next to aspartate residues. Since caspases cleave and activate each other, an amplification mechanism through a protease cascade exists, assuring proper execution of apoptotic cell death [13]. In addition, caspases cleave numerous substrates, like nuclear lamins, inhibitors of DNase or cytoskeletal

proteins, ultimately leading to the typical morphological alterations of apoptosis [13].

There are two alternative pathways to initiate apoptosis and both finally activate the executioner caspases-3, -6 and -7 (figure 1):

The first pathway is called intrinsic or mitochondrial pathway, because the mitochondria takes the key position by initiating apoptosis. The exact mechanism of initiation by different apoptotic stimuli is still not entirely clear, but involves an imbalance of pro- and anti-apoptotic members of the BCL-2 protein family [14]. This imbalance finally leads to the activation of the pro-apoptotic BCL-2 family members BAX and/or BAK and the perturbation of the integrity of the outer mitochondrial membrane [14]. This induces the release of cytochrom c and other apoptotic regulators, like apoptosis-inducing factor (AIF), Smac (second mitochondria-derived activator of apopto-

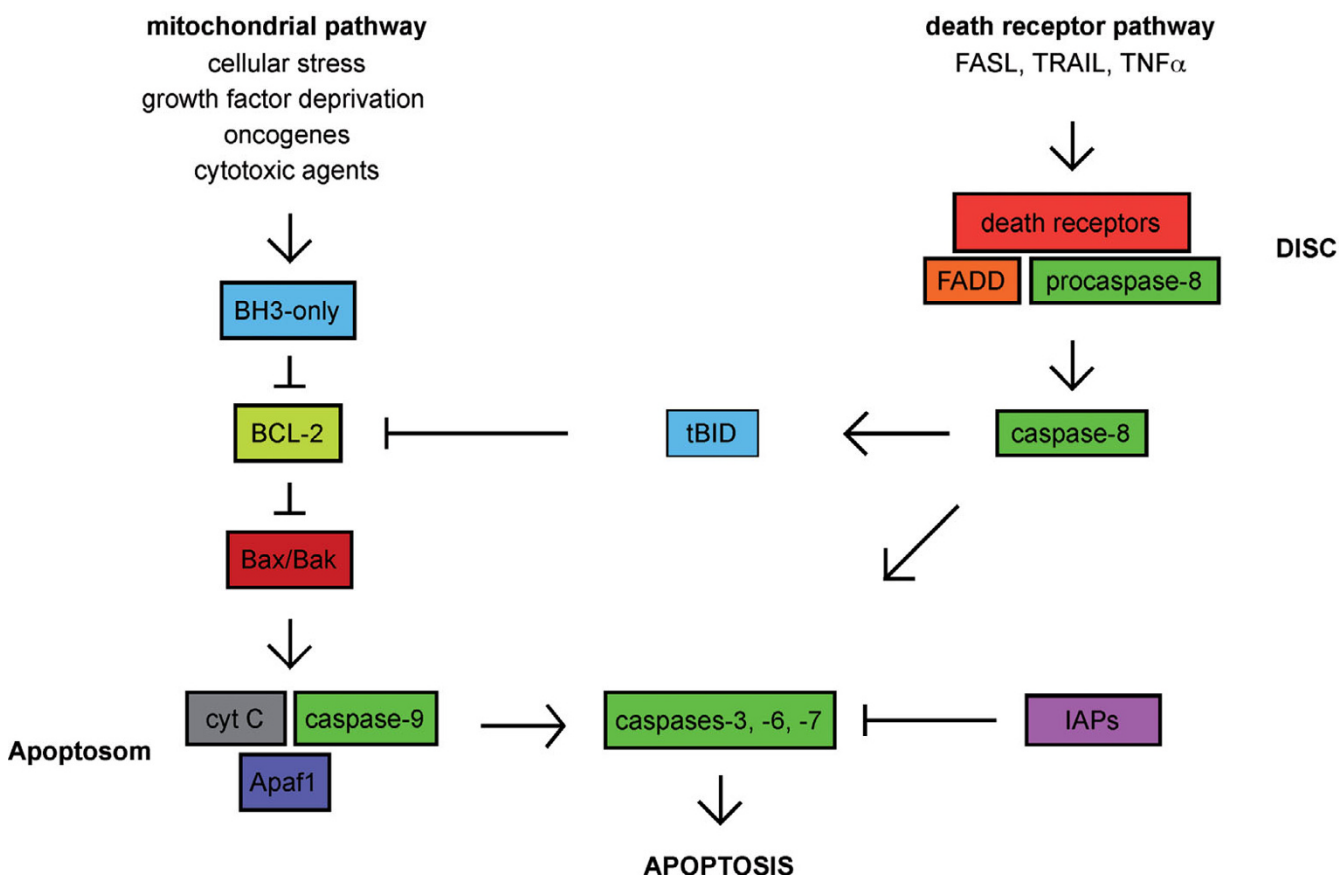


Figure 1 Pathways to Apoptosis. The mitochondrial pathway is activated by BH3-only proteins, which sense cellular stress and inactivate pro-survival BCL-2 family members. This leads to the permeabilization of the outer mitochondrial membrane and the release of cytochrom C, Apaf-I and caspase-9. The death receptor pathway is activated by the TNF family ligands. Caspase-8 is activated by adaptor proteins including FADD. In PDAC cells, the death receptor pathway is linked to the mitochondria by the BH3-only protein Bid that is cleaved by caspase-8.

sis)/DIABOLO (direct inhibitor of apoptosis protein (IAP)-binding protein with low PI), endonuclease G or Omi/HtrA2 from the intermembrane space of mitochondria [15]. In the cytosol, cytochrome c, APAF-1, ATP and the initiator procaspase-9 are forming the apoptosome multiprotein complex and activate the initiator caspase-9. This induces the cleavage of the executioner caspases, like caspase-3. Furthermore, the potent endogenous inhibitors of caspases, the inhibitor of apoptosis proteins (IAPs) are neutralized by Smac/DIABOLO or Omi/HtrA2 [16].

The second pathway is called the extrinsic pathway and is mediated by different death receptors on the cell surface [17-19]. These receptors are members of the tumor-necrosis factor (TNF) receptor superfamily, including the TNF-, FAS-(APO-1, CD95) and TRAIL-(TNF-related apoptosis inducing ligand) receptors. They share a common intracellular domain, which is called death domain. Activation of the receptors after extracellular binding of the specific ligands (TNF- α , FAS-L and TRAIL) initiates the recruitment of FADD (FAS-associated death domain protein), procaspase-8 and -10 to the death domain, which are forming the DISC (death inducing signaling complex) [20]. At the DISC, the initiator caspase-8 is activated. In type I cells the activated initiator caspases are sufficient to induce executioner caspases directly. In contrast type II cells need the signal enhancing-effect of mitochondria to induce apoptosis. Here, caspase-8 cleaves the pro-apoptotic BH3-only BCL-2 family member BID, which translocates to the mitochondrial membrane and induces the release of apoptogenic factors from the mitochondria [11]. Pancreatic carcinoma cells are type II cells [21,22]. The type I/II concept, is a good example of the crosstalk between extrinsic and intrinsic apoptotic signaling and demonstrates the complexity of apoptosis [23].

Apoptosis and Carcinogenesis of PDAC – an under-investigated field

Apoptosis is a fundamental anti-neoplastic mechanism in normal cells to prevent tumorigenesis. Nearly all neoplastic changes during the development of a normal cell to a cancer cell, like DNA-damage, oncogene activation or cell cycle deregulation, are potent inducers of the programmed cell death pathway. Therefore, overcoming the apoptotic failsafe is observed in many cancers as discussed by Lowe, Cepero and Evan [24].

In accordance with the multi-step theory of carcinogenesis, the natural history of PDAC seems to gradually evolve through precursor lesions, the so-called **pancreatic intraepithelial neoplasias** (PanIN) 1 to 3 to invasive PDAC [2,5-7]. Whereas the progressive accumulation of genetic somatic mutations, including mutations in the oncogene *K-RAS* and the tumor suppressors *CDKN2A*, *TP53*,

SMAD4/DPC4 and *BRCA2* are quite good understood, the contribution of apoptosis to carcinogenesis of PDAC is not well defined [3]. In PanIN 1 and 2 lesions, no apoptotic cells could be detected, arguing for the contribution of anti-apoptotic strategies early in the carcinogenesis of PDAC [25]. These anti-apoptotic strategies most likely enable cell survival under oncogenic stress conditions [25]. Recently, two members of the IAP (inhibitors of apoptosis) protein family, cIAP2 and Survivin were shown to be early overexpressed during carcinogenesis of PDAC. One anti-apoptotic function of the IAP family is dependent on the inhibition of caspase activity [16]. Early overexpression of cIAP2 was demonstrated in 30% of low-grade PanIN lesions, 50% of high grade PanIN lesions and 85% of PDAC [26]. In addition to cIAP2, Survivin was shown to be overexpressed already in PanIN1 lesions at the mRNA and protein level [27]. Although the observed overexpression of cIAP2 and Survivin, suggests early contribution in tumorigenesis of PDAC, molecular pathways engaged by these molecules in preneoplastic lesions of the pancreas are unknown.

Death receptor signaling – blocked and abused

Many PDAC cells are characterized by a high IC_{50} for death receptor mediated apoptosis, although they express the corresponding receptors and the execution machinery of apoptosis is intact [22,28-32]. PDAC cells have evolved resistance mechanisms, which are especially working at the death receptor level resulting in DISC insufficiency [33].

The protein tyrosine phosphatase FAP-1 (FAS-associated phosphatase-1) is overexpressed in PDAC and protects pancreatic carcinoma cells from FAS-mediated apoptosis by inhibiting the activation of caspase-8 [28,34]. It seems that FAP-1 interferes with the translocation of CD95 to the cell surface. This leads to a low receptor density on the cell surface and to a disruption of receptor trimerization, which is necessary to form the DISC. Furthermore a direct inhibition of caspase-8 by FAP-1 is discussed [34]. The resistance towards FAS-mediated apoptosis enables PDAC cells to express simultaneously FASL, which might contribute to escape the immunosurveillance. Compatibly, expression of FASL was observed in 6/6 investigated PDAC cell lines and co-culture of PDAC cells with T-cells results in T-cell death in a FasL-dependent fashion [28].

Another potent inhibitor of caspase-8 activation is c-FLIP_L, which is a structural homologue of caspase-8, lacking amino acid residues that are critical for caspase activity. c-FLIP_L competes with procaspase-8 for binding to FADD at the DISC [35,36]. Together with FAP-1, c-Flip_L is highly expressed in PDACs [35]. In addition to high c-FLIP_L expression, low amounts of FADD were shown to

contribute to resistance against CD95L- and TRAIL-induced apoptosis in PDAC cells [32].

A further important mechanism to escape death-receptor induced apoptosis is to favor pro-survival signaling after triggering the death receptor. In PDAC cells, especially the activation of NF- κ B signaling (see below) contributes to the resistance towards death receptor induced apoptosis [21]. One factor involved in the induction of pro-survival signaling is the death receptor adaptor TRAF2. TRAF2 belongs to the TRAF-family of adaptor proteins which are involved in the signal transduction pathways of the TNF-receptor family. Kalthoff's group observed TRAF2 overexpression in 94% of PDAC and the FASL-resistant cell lines PancTul, Panc89 and Panc1 [37]. Furthermore, ectopic expression of TRAF2 in the FASL-sensitive pancreatic cancer cell line Colo357 confers resistance towards FAS-mediated apoptosis, arguing for a pivotal role of TRAF2 in mediating FAS resistance of pancreatic cancer cells. Mechanistically TRAF2 increases as well as basal as FAS-induced NF- κ B activity [37]. In addition, TRAF2 overexpression increases the invasive properties of pancreatic cancer cells by inducing the expression of proteolytic enzymes, like matrix metalloproteinases 2 and 9 or the urokinase type plasminogen activator (uPA) [37].

Another strategy to evade death-receptor apoptosis is the overexpression of the decoy receptor 3 (DcR3). Decoy receptors are members of the TNF-receptor superfamily, but without an intact death domain. DcR3 competes with FAS for FASL binding and therefore interferes with FAS triggered apoptosis [35].

Since PDAC cells are type II cells, death receptor induced apoptosis is also blocked at the level of the mitochondria [22]. Furthermore, XIAP contributes to death receptor resistance in PDAC cells [36,38].

BCL-2 family and PDAC – In favor for survival

At the level of the mitochondria deregulated expression of members of the BCL-2 protein family causes apoptotic resistance in PDAC [2,39,40]. This protein family consists of two pro-apoptotic subgroups, the BAX-like group (BAX, BAK, BOK) and the BH3-only subgroup (BAD, BIK, BID, BIM, BMF, HRK, NOXA, PUMA) and the anti-apoptotic subgroup (BCL-2, BCL-X_L, BCL-W, MCL-1, A1) [11,14,41]. In most cases, the mitochondrial cell death pathway needs as well as the BAX-like group as the BH3-only group. Here, BH3-only proteins act as sensors of cellular stress, directly antagonizing anti-apoptotic BCL-2 members and activated BAX-like proteins are permeabilizing the outer mitochondrial membrane [14]. In contrast to various other human tumors, expression of the prototypical pro-survival protein BCL-2 is normal or even decreased in PDAC [39,40,42]. Faint BCL-2 expression

was detected especially in well-differentiated PDACs. Furthermore, an inverse correlation between BCL-2 and p53 immunoreactivity was observed [40].

Resistance towards apoptosis at the mitochondrial level in PDAC is conferred by at least two anti-apoptotic BCL-2 family members, BCL-X_L and MCL-1. Both proteins are known to be overexpressed [22,40,43,44]. BCL-X_L and MCL-1 are upregulated by MAPK signaling in PDAC cells [43]. Whether MCL-1 is also induced by the NF- κ B pathway in pancreatic cancer cells, as it has been shown for BCL-X_L, is unknown at the moment [43,45]. In a murine model of pancreatic cancer, the BCL-X_L promoter was shown to integrate the epidermal growth factor receptor (EGF-R) signal. Here, EGF-R signaling contributes to the activation of the transcription factors NF- κ B and STAT3, both activating the BCL-X_L gene [45]. In line, inhibition of both transcription factors is needed, to induce significant apoptosis in this model [45].

Functionally, MCL-1 was recently linked to the integrative stress response [46]. Therefore, high MCL-1 might protect pancreatic cancer cells from hypoxia and oxidative stress during tumorigenesis. From a therapeutically viewpoint, dependency of survival of pancreatic cancer cells onto MCL-1 has to be kept in mind, when molecular targeted therapies, like ABT-737 treatment, are considered. ABT-737 is a BAD homologous BH3-mimetic binding with high affinity to BCL-X_L, BCL-2 or BCL-w but not MCL-1 [47]. Hence, a molecular ABT-737 therapy is ineffective in cells expressing significant amounts of MCL-1 [48]. This applies also for pancreatic cancer cells, where only minor apoptosis was observed after ABT-737 treatment. In contrast, ABT-737 induces distinct apoptosis of pancreatic cancer cells after the depletion of MCL-1 [49]. A further approach to sensitize towards ABT-737 is activation of NOXA, a BH3-only selectively binding to and depleting MCL-1. Enforced NOXA expression in murine embryonic fibroblasts or the myelomonocytic cell line FDC-P1 increased sensitivity for ABT-737 up to 2000-fold [48]. Regulation of NOXA is not understood in pancreatic cancer, but strategies leading to NOXA activation are of value if a molecular ABT-737 therapy is planned.

Little is known about the contribution of the pro-death BCL-2 family members in PDAC. BAX expression is lowered in around 50% of PDACs [8,39]. In line with an essential function in tumor cell killing, the median survival of BAX positive patients after pancreatic resection is 19 months versus 11 months of BAX negative patients [8]. Since BH3-only activation is a potential therapeutic strategy, future studies have to address regulation of BH3-only expression in PDAC in detail.

Inhibitors of Apoptosis Proteins

The caspases are the central component of apoptosis. The expression levels of caspases are more or less normal in PDAC cells, but proper activation is impaired by potent inhibitors, called IAPs (inhibitor of apoptosis proteins) [29,33,50]. The IAP family, includes different members like cIAP1, cIAP2, XIAP, NAIP, ML-IAP, ILP2, Livin, Apollon and Survivin and one of the molecular mechanisms of these proteins is to bind caspases and inhibit their activity [51]. For PDAC the overexpression of cIAP2, XIAP and Survivin has been demonstrated [26,27,52-54]. A recent functional and systematic investigation of the IAP family in PDAC conducted by Lemoine's group provide evidence that cIAP2 and XIAP play key roles in anti-apoptotic signaling. Downregulation of cIAP2 and XIAP by RNA interference in PDAC cell lines efficiently induced sensitivity towards cisplatin, doxorubicin or paclitaxel [55]. Importantly, no synergism between downregulation of cIAP2 and XIAP was observed [55]. Furthermore, inhibition of XIAP was shown to sensitize PDAC cells for death receptor signaling, cytotoxic therapies and γ -irradiation [36,38,55-59]. Since small molecule inhibitors exist, which target XIAP function, the so-called XIAP antagonists (Xantags), XIAP is a promising target for novel treatment strategies of PDAC [36,59,60].

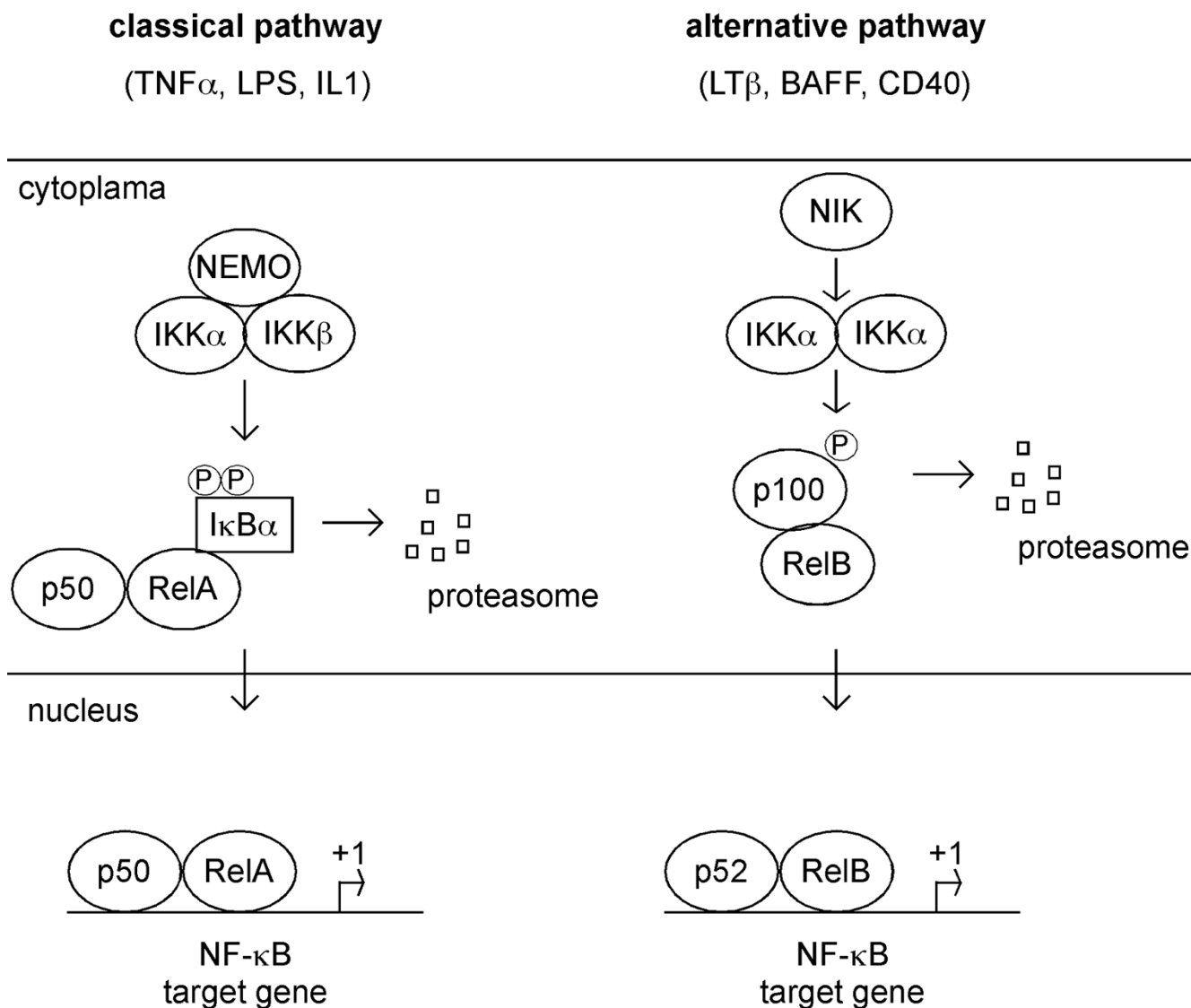
In addition, Survivin, the smallest member (16,5 kD) of the IAP protein family contributes to anti-apoptotic signaling in PDAC cells. Survivin is selectively expressed in most human neoplasms including pancreatic cancer, but not in normal adult tissue [27,61,62]. Survivin has been implicated in control of cell division and inhibition of apoptosis, by molecular mechanisms recently summarized in an excellent review by Altieri [63]. The control of cell division might explain the induction of apoptosis observed after downregulation of Survivin expression in PDAC cells, which was not overt after downregulation of cIAP2 and XIAP [64-66]. In addition, downregulation of Survivin by RNA interference sensitizes PDAC cells towards TRAIL- and γ -irradiation-induced apoptosis [65,66]. Interestingly, Survivin is expressed in a cell-cycle-dependent manner in PDAC cells. Low expression was observed at the RB-dependent restriction point in the late G1-phase and this particular cell cycle phase was TRAIL-sensitive [66]. Since cell cycle inhibitors are investigated in clinical trials a combination therapy of G1-phase cell cycle inhibition and death receptor activation might be a feasible approach for the treatment of PDAC.

NF- κ B – a key anti-apoptotic transcription factor in PDAC

NF- κ B is a nuclear transcription factor, which is activated by a variety of different stimuli. NF- κ B controls different biological processes like inflammation, cell cycle, apoptosis or epithelial to mesenchymal transition [67]. This transcription factor binds DNA as a dimer, composed of the

NF- κ B factors RelA (p65), RelB, c-Rel, NF- κ B1 (p50/p105) and NF- κ B2 (p52/p100). Classical NF- κ B (p50/p65 dimer) is usually kept in an inactive form in the cytoplasm by stable association with inhibitor proteins such as I κ B α or I κ B β . The I κ Bs are phosphorylated by a multi-protein kinase complex, the Inhibitor of I κ B kinase (IKK). The IKK complex contains the two catalytical kinases IKK α and IKK β and the structural component IKK γ /NEMO [68,69]. Activation of the IKK complex causes phosphorylation of the I κ Bs, subsequent ubiquitination and proteasomal degradation [68] (figure 2). This leads to the liberation and nuclear translocation of classical NF- κ B and target gene activation. In addition to the canonical NF- κ B activation pathway, an alternative pathway involves IKK α -dependent processing of NF- κ B2 and nuclear translocation of a p52/RelB dimer (figure 2). The contribution of NF- κ B to the tumorigenesis of various cancers is well documented [70,71]. Also in PDAC, NF- κ B plays an important role [72-75]. In almost all PDAC cell lines the IKK complex is activated, leading to the nuclear translocation of NF- κ B [76-78]. Interference with IKK activity in PDAC cell lines leads to the induction of apoptosis and/or sensitization towards cytotoxic therapies [77,78]. Although IKK α was shown to be overexpressed in PDAC, downregulation of IKK α by RNA interference was not accompanied by significant apoptosis [79,80]. Instead, IKK α was shown to promote cell cycle progression of PDAC cells by inducing transcription of the onco-gene S-phase kinase associated protein 2 (SKP2) in a p52/RelB-dependent fashion [80]. Therefore, anti-apoptotic function of NF- κ B signaling seems to be mediated by the classical NF- κ B pathway, but the contribution of canonical and non-canonical signaling to apoptosis resistance has to be investigated in more detail. In line with activated IKK, the NF- κ B subunit RelA/p65 is nuclear translocated in around 45 to 70% of pancreatic cancers [76,77,81]. Furthermore, nuclear RelA/p65 is a predictor for poor patients survival and was linked to a higher expression of NF- κ B target genes, like BCL-X_L [81]. Based on the broad impact of NF- κ B signaling towards the apoptotic machinery, it is not surprising that inhibition of NF- κ B activity sensitizes pancreatic cancer cells to chemotherapeutic agents and death-receptor mediated apoptosis [82-86]. Target genes of NF- κ B in PDAC explaining the sensitizer effect of NF- κ B inhibition are BCL-X_L and c-Flip [82,84-86].

Although NF- κ B is activated in PDAC, the signaling pathways leading to IKK activation in PDAC cells are not entirely clear. Here the EGF-receptor-, PI3K-AKT- and K-Ras-pathways were shown to contribute to NF- κ B activity [76,77]. Furthermore, NF- κ B is modulated by several other important signaling pathways in PDAC. For example, Sarkar's group demonstrated that depleting Notch-1 expression in PDAC cells induces apoptosis and was cor-

**Figure 2**

Representation of the two NF- κ B signaling pathways. Classical signaling (depicted left) is activated by TNF α , IL1 β or LPS. Activation of canonical signaling depends on the IKK holocomplex (IKK α , IKK β , IKK γ /NEMO). Activated IKK phosphorylates I κ Bs, leading to the proteasomal degradation and subsequent nuclear translocation of classical NF- κ B. An alternative signaling pathway (depicted right) is activated by lymphotoxin, CD40L or BAFF. Signaling depends on IKK α and leads to the nuclear translocation of a p52/RelB dimer.

related with reduced NF- κ B activity and impaired expression of the NF- κ B target gene BCL-X_L [87,88]. Moreover, Notch-1/NF- κ B signaling contributes to invasiveness of PDAC cells [87]. Furthermore, casein kinase 2 (CK2)

maintains NF- κ B activity and anti-apoptotic signaling in PDAC cells [89]. A further modulator of NF- κ B/RelA transcriptional activity is the early response gene IEX-1, which is involved in the regulation of apoptosis. In a recent

study, Arlt et al. observed that IEX-1 interacts directly with the C-terminal activation domain of RelA and is involved in inhibition of RelA-dependent transcriptional activity and therefore negatively regulates NF- κ B target genes, like Bcl-X_L, cIAP1 and cIAP2 [90]. This molecular observation might explain the recent demonstration that IEX-1 expression is a significantly favorable factor for survival of patients with PDAC [91].

The PI3K/AKT survival pathway

The phosphatidylinositol-3' kinase (PI3K)/AKT-pathway is a pro-survival pathway in PDAC cells. This pathway is activated by survival signals like growth factors, cytokines, hormones and oncogenic Ras [92]. Activation of PI3K leads to the phosphorylation of AKT/PKB. Besides the growth promoting potential, activated AKT/PKB favors survival via the direct regulation of apoptotic proteins, like the BCL-2 members BAD and BCL-X_L or caspase 9 [93-96]. In addition, AKT modulates the activity of transcription factors, like FOXO, NF- κ B or c-myc. Schlieman et al. has shown that the PI3K/AKT pathway is activated in about 60% of pancreatic adenocarcinoma tissues and also most pancreatic cancer cell lines display elevated levels of activated AKT [97,98]. Although it was shown that PI3K/AKT signaling modulates NF- κ B and c-myc activity in PDAC cells, the molecular activation of this pathway in PDAC is not entirely clear [77,99,100]. Here, the contribution of the tumor suppressor phosphatase and tensin homolog deleted in chromosome ten (PTEN) has been demonstrated [100]. Low expression of this natural antagonist of PI3K signaling was demonstrated in 75% of PDACs [100]. In addition, a receptor tyrosin-kinase is involved in PI3K activation, since the insulin receptor substrate 1 (IRS-1) was shown to be an essential mediator of PI3K activation in quiescent PDAC cells [101]. Furthermore, an unexpected pathway mediates the activation of PI3K-AKT signaling in pancreatic cancer cells. This pathway is regulated by the architectural transcription factor HMGA1. HMGA1 is highly overexpressed in PDAC and known to account for unrestrained proliferation of pancreatic cancer cells [102,103]. Recently, it was shown that HMGA1 activates PI3K-AKT signaling in PDAC cells [104,105]. By forming a complex with the transcription factor C/EBP β , HMGA1 activates the insulin receptor gene and the insulin receptor contributes to the activation of PI3K-AKT signaling [106]. This pathway seems to mediate therapeutic resistance, since HMGA1-mediated gemcitabine resistance depends onto intact AKT signaling [107].

Although we observed no overt apoptosis in PDAC cells treated either with the PI3K inhibitor Ly294002 or by interference with AKT1 expression using siRNAs (unpublished data), inhibition of the PI3K/AKT signaling sensitizes PDAC cells towards therapeutic induction of

apoptosis [97,108-111]. Since very high concentrations of the PI3K inhibitor Ly294002 (up to 100 μ M) were used in many studies and contribution of the PI3K/AKT signaling towards gemcitabine resistance were not detected in BxPc-3, Capan-1 and PancTu-1 cells, the impact of PI3K/AKT signaling for therapeutic resistance of PDAC cells has to be clarified in more detail using functional genomics [112].

Conclusion

Deregulation of the apoptosis machinery and evasion of apoptosis is a general mechanism in PDAC. Since nearly every step during carcinogenesis activates apoptosis, the development of anti-apoptotic strategies must be an early and essential event. Contribution of molecules like cIAP2 or Survivin to early steps in the carcinogenesis of PDAC should be investigated in relevant mouse models of PDAC in the future. Also therapeutic resistance is mediated by the profound deregulation of the apoptotic machinery in PDAC cells. Detailed molecular knowledge of the anti-apoptotic mechanisms of PDAC cells is essential for the improvement of conventional chemotherapies ("sensitizer strategies") and the development of new potent targeted therapies. Hopefully, this will lead to an improvement of the prognosis of this dismal disease.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

RH: drafted and wrote the manuscript. RMS: drafted and revised the manuscript critically for important intellectual content. DS: drafted and revised the manuscript critically for important intellectual content. GS: drafted and wrote the manuscript, revised the manuscript critically for important intellectual content, given final approval of the version to be published. All authors read and approved the final manuscript.

Acknowledgements

The authors are supported by Deutsche Forschungsgemeinschaft, SFB456, Else Kröner-Fresenius-Stiftung, Fritz Thyssen-Stiftung and Kommission für klinische Forschung. We would like to apologize for not citing any relevant reports due to lack of space, the need to selectively choose examples, or an oversight on our part.

References

1. Jemal A, Siegel R, Ward E, Murray T, Xu J, Thun MJ: **Cancer statistics, 2007**. *CA Cancer J Clin* 2007, **57**:43-66.
2. Schneider G, Siveke JT, Eckel F, Schmid RM: **Pancreatic cancer: basic and clinical aspects**. *Gastroenterology* 2005, **128**:1606-1625.
3. Schneider G, Schmid RM: **Genetic alterations in pancreatic carcinoma**. *Mol Cancer* 2003, **2**:15.
4. Schneider G, Saur D, Schmid RM: **Pancreatic cancer – Molecular alterations**. *The Chinese-German Journal of Clinical Oncology* 2007, **6**:102-106.
5. Hezel AF, Kimmelman AC, Stanger BZ, Bardeesy N, Depinho RA: **Genetics and biology of pancreatic ductal adenocarcinoma**. *Genes Dev* 2006, **20**:1218-1249.
6. Maitra A, Hruban RH: **Pancreatic Cancer**. *Annu Rev Pathol* 2008, **3**:157-188.

7. Welsch T, Kleeff J, Friess H: **Molecular pathogenesis of pancreatic cancer: advances and challenges.** *Curr Mol Med* 2007, **7**:504-521.
8. Magistrelli P, Coppola R, Tonini G, Vincenzi B, Santini D, Borzomati D, Vecchio F, Valeri S, Castri F, Antinori A, Nuzzo G, Caraglia M, Picciocchi A: **Apoptotic index or a combination of Bax/Bcl-2 expression correlate with survival after resection of pancreatic adenocarcinoma.** *J Cell Biochem* 2006, **97**:98-108.
9. Schulze-Bergkamen H, Krammer PH: **Apoptosis in cancer – implications for therapy.** *Semin Oncol* 2004, **31**:90-119.
10. Hager JH, Hanahan D: **Tumor cells utilize multiple pathways to down-modulate apoptosis. Lessons from a mouse model of islet cell carcinogenesis.** *Ann N Y Acad Sci* 1999, **887**:150-163.
11. Igney FH, Krammer PH: **Death and anti-death: tumour resistance to apoptosis.** *Nat Rev Cancer* 2002, **2**:277-288.
12. Okada H, Mak TW: **Pathways of apoptotic and non-apoptotic death in tumour cells.** *Nat Rev Cancer* 2004, **4**:592-603.
13. Degtarev A, Boyce M, Yuan J: **A decade of caspases.** *Oncogene* 2003, **22**:8543-8567.
14. Adams JM, Cory S: **Bcl-2-regulated apoptosis: mechanism and therapeutic potential.** *Curr Opin Immunol* 2007, **19**:488-496.
15. Saelens X, Festjens N, Walle L Vande, van Gurp M, van Loo G, Vandenabeele P: **Toxic proteins released from mitochondria in cell death.** *Oncogene* 2004, **23**:2861-2874.
16. Fulda S: **Inhibitor of apoptosis proteins as targets for anticancer therapy.** *Expert Rev Anticancer Ther* 2007, **7**:1255-1264.
17. Debatin KM, Krammer PH: **Death receptors in chemotherapy and cancer.** *Oncogene* 2004, **23**:2950-2966.
18. Fas SC, Fritzsching B, Suri-Payer E, Krammer PH: **Death receptor signaling and its function in the immune system.** *Curr Dir Autoimmun* 2006, **9**:1-17.
19. Wajant H: **CD95/FasL and TRAIL in tumour surveillance and cancer therapy.** *Cancer Treat Res* 2006, **130**:141-165.
20. Walczak H, Krammer PH: **The CD95 (APO-1/Fas) and the TRAIL (APO-2L) apoptosis systems.** *Exp Cell Res* 2000, **256**:58-66.
21. Trauzold A, Wermann H, Arlt A, Schutze S, Schafer H, Oestern S, Roder C, Ungefroren H, Lampe E, Heinrich M, Walczak H, Kalthoff H: **CD95 and TRAIL receptor-mediated activation of protein kinase C and NF-kappaB contributes to apoptosis resistance in ductal pancreatic adenocarcinoma cells.** *Oncogene* 2001, **20**:4258-4269.
22. Hinz S, Trauzold A, Boenicke L, Sandberg C, Beckmann S, Bayer E, Walczak H, Kalthoff H, Ungefroren H: **Bcl-XL protects pancreatic adenocarcinoma cells against CD95- and TRAIL-receptor-mediated apoptosis.** *Oncogene* 2000, **19**:5477-5486.
23. Sprick MR, Walczak H: **The interplay between the Bcl-2 family and death receptor-mediated apoptosis.** *Biochim Biophys Acta* 2004, **1644**:125-132.
24. Lowe SV, Cepero E, Evan G: **Intrinsic tumour suppression.** *Nature* 2004, **432**:307-315.
25. Luttes J, Neumann S, Jesnowski R, Borries V, Lohr M, Kloppel G: **Lack of apoptosis in PanIN-1 and PanIN-2 lesions associated with pancreatic ductal adenocarcinoma is not dependent on K-ras status.** *Pancreas* 2003, **27**:e57-62.
26. Esposito I, Kleeff J, Abiatari I, Shi X, Giese N, Bergmann F, Roth W, Friess H, Schirmacher P: **Overexpression of cellular inhibitor of apoptosis protein 2 is an early event in the progression of pancreatic cancer.** *J Clin Pathol* 2007, **60**:885-895.
27. Bhanot U, Heydrich R, Moller P, Hasel C: **Survivin expression in pancreatic intraepithelial neoplasia (PanIN): steady increase along the developmental stages of pancreatic ductal adenocarcinoma.** *Am J Surg Pathol* 2006, **30**:754-759.
28. Ungefroren H, Voss M, Jansen M, Roeder C, Henne-Bruns D, Kremer B, Kalthoff H: **Human pancreatic adenocarcinomas express Fas and Fas ligand yet are resistant to Fas-mediated apoptosis.** *Cancer Res* 1998, **58**:1741-1749.
29. Gerhard MC, Schmid RM, Hacker G: **Analysis of the cytochrome c-dependent apoptosis apparatus in cells from human pancreatic carcinoma.** *Br J Cancer* 2002, **86**:893-898.
30. Kornmann M, Ishiwata T, Kleeff J, Beger HG, Korc M: **Fas and Fas-ligand expression in human pancreatic cancer.** *Ann Surg* 2000, **231**:368-379.
31. Satoh K, Kaneko K, Hirota M, Masamune A, Satoh A, Shimosegawa T: **Tumor necrosis factor-related apoptosis-inducing ligand and its receptor expression and the pathway of apoptosis in human pancreatic cancer.** *Pancreas* 2001, **23**:251-258.
32. Trauzold A, Schmiedel S, Roder C, Tams C, Christgen M, Oestern S, Arlt A, Westphal S, Kapischke M, Ungefroren H, Kalthoff H: **Multiple and synergistic deregulations of apoptosis-controlling genes in pancreatic carcinoma cells.** *Br J Cancer* 2003, **89**:1714-1721.
33. Westphal S, Kalthoff H: **Apoptosis: targets in pancreatic cancer.** *Mol Cancer* 2003, **2**:6.
34. Ungefroren H, Kruse ML, Trauzold A, Roeschmann S, Roeder C, Arlt A, Henne-Bruns D, Kalthoff H: **FAP-1 in pancreatic cancer cells: functional and mechanistic studies on its inhibitory role in CD95-mediated apoptosis.** *J Cell Sci* 2001, **114**:2735-2746.
35. Elnemr A, Ohta T, Yachie A, Kayahara M, Kitagawa H, Fujimura T, Ninomiya I, Fushida S, Nishimura GI, Shimizu K, Miwa K: **Human pancreatic cancer cells disable function of Fas receptors at several levels in Fas signal transduction pathway.** *Int J Oncol* 2001, **18**:311-316.
36. Mori T, Doi R, Kida A, Nagai K, Kami K, Ito D, Toyoda E, Kawaguchi Y, Uemoto S: **Effect of the XIAP inhibitor Embelin on TRAIL-induced apoptosis of pancreatic cancer cells.** *J Surg Res* 2007, **142**:281-286.
37. Trauzold A, Roder C, Sipos B, Karsten K, Arlt A, Jiang P, Martin-Subero JI, Siegmund D, Muerkoster S, Pagerols-Raluy L, Siebert R, Wajant H, Kalthoff H: **CD95 and TRAF2 promote invasiveness of pancreatic cancer cells.** *Faseb J* 2005, **19**:620-622.
38. Vogler M, Durr K, Jovanovic M, Debatin KM, Fulda S: **Regulation of TRAIL-induced apoptosis by XIAP in pancreatic carcinoma cells.** *Oncogene* 2007, **26**:248-257.
39. Evans JD, Cornford PA, Dodson A, Greenhalf W, Foster CS, Neoptolemos JP: **Detailed tissue expression of bcl-2, bax, bak and bcl-x in the normal human pancreas and in chronic pancreatitis, ampullary and pancreatic ductal adenocarcinomas.** *Pancreatol* 2001, **1**:254-262.
40. Campani D, Esposito I, Boggi U, Cecchetti D, Menicagli M, De Negri F, Colizzi L, Del Chiaro M, Mosca F, Fornaciari G, Bevilacqua G: **Bcl-2 expression in pancreas development and pancreatic cancer progression.** *J Pathol* 2001, **194**:444-450.
41. Fulda S, Debatin KM: **Extrinsic versus intrinsic apoptosis pathways in anticancer chemotherapy.** *Oncogene* 2006, **25**:4798-4811.
42. Gutierrez-Puente Y, Zapata-Benavides P, Tari AM, Lopez-Berestein G: **Bcl-2-related antisense therapy.** *Semin Oncol* 2002, **29**:71-76.
43. Boucher MJ, Morisset J, Vachon PH, Reed JC, Laine J, Rivard N: **MEK/ERK signaling pathway regulates the expression of Bcl-2, Bcl-X(L), and Mcl-1 and promotes survival of human pancreatic cancer cells.** *J Cell Biochem* 2000, **79**:355-369.
44. Miyamoto Y, Hosotani R, Wada M, Lee JU, Koshiba T, Fujimoto K, Tsuji S, Nakajima S, Doi R, Kato M, Shimada Y, Imamura M: **Immunohistochemical analysis of Bcl-2, Bax, Bcl-X, and Mcl-1 expression in pancreatic cancers.** *Oncology* 1999, **56**:73-82.
45. Greten FR, Weber CK, Greten TF, Schneider G, Wagner M, Adler G, Schmid RM: **Stat3 and NF-kappaB activation prevents apoptosis in pancreatic carcinogenesis.** *Gastroenterology* 2002, **123**:2052-2063.
46. Fritsch RM, Schneider G, Saur D, Scheibel M, Schmid RM: **Translational repression of MCL-1 couples stress-induced eIF2alpha phosphorylation to mitochondrial apoptosis initiation.** *J Biol Chem* 2007, **282**:22551-22562.
47. Labi V, Grespi F, Baumgartner F, Villunger A: **Targeting the Bcl-2-regulated apoptosis pathway by BH3 mimetics: a breakthrough in anticancer therapy?** *Cell Death Differ* 2008, **15**:977-987.
48. van Delft MF, Wei AH, Mason KD, Vandenberg CJ, Chen L, Czabotar PE, Willis SN, Scott CL, Day CL, Cory S, Adams JM, Roberts AW, Huang DC: **The BH3 mimetic ABT-737 targets selective Bcl-2 proteins and efficiently induces apoptosis via Bak/Bax if Mcl-1 is neutralized.** *Cancer Cell* 2006, **10**:389-399.
49. Huang S, Sinicrope FA: **BH3 mimetic ABT-737 potentiates TRAIL-mediated apoptotic signaling by unsequestering Bim and Bak in human pancreatic cancer cells.** *Cancer Res* 2008, **68**:2944-2951.
50. Gukovskaya AS, Pandolfi SJ: **Cell death pathways in pancreatitis and pancreatic cancer.** *Pancreatol* 2004, **4**:567-586.
51. Vaux DL, Silke J: **IAPs, RINGs and ubiquitylation.** *Nat Rev Mol Cell Biol* 2005, **6**:287-297.

52. Satoh K, Kaneko K, Hirota M, Masamune A, Satoh A, Shimosegawa T: **Expression of survivin is correlated with cancer cell apoptosis and is involved in the development of human pancreatic duct cell tumors.** *Cancer* 2001, **92**:271-278.
53. Sarela AI, Verbeke CS, Ramsdale J, Davies CL, Markham AF, Guillou PJ: **Expression of survivin, a novel inhibitor of apoptosis and cell cycle regulatory protein, in pancreatic adenocarcinoma.** *Br J Cancer* 2002, **86**:886-892.
54. Trauzold A, Schmiedel S, Oestern S, Christgen M, Westphal S, Roederer C, Kalthoff H: **Concerted deregulations of multiple apoptosis-controlling genes in pancreatic carcinoma cells.** *Ann N Y Acad Sci* 2003, **1010**:510-513.
55. Lopes RB, Gangeswaran R, McNeish IA, Wang Y, Lemoine NR: **Expression of the IAP protein family is dysregulated in pancreatic cancer cells and is important for resistance to chemotherapy.** *Int J Cancer* 2007, **120**:2344-2352.
56. Li Y, Jian Z, Xia K, Li X, Lv X, Pei H, Chen Z, Li J: **XIAP is related to the chemoresistance and inhibited its expression by RNA interference sensitize pancreatic carcinoma cells to chemotherapeutics.** *Pancreas* 2006, **32**:288-296.
57. Shrikhande SV, Kleeff J, Kaye H, Keleg S, Reiser C, Giese T, Buchler MW, Esposito I, Friess H: **Silencing of X-linked inhibitor of apoptosis (XIAP) decreases gemcitabine resistance of pancreatic cancer cells.** *Anticancer Res* 2006, **26**:3265-3273.
58. Giagkousiklidis S, Vellanki SH, Debatin KM, Fulda S: **Sensitization of pancreatic carcinoma cells for gamma-irradiation-induced apoptosis by XIAP inhibition.** *Oncogene* 2007, **26**:7006-7016.
59. Karikari CA, Roy I, Tryggestad E, Feldmann G, Pinilla C, Welsh K, Reed JC, Armour EP, Wong J, Herman J, Rakheja D, Maitra A: **Targeting the apoptotic machinery in pancreatic cancers using small-molecule antagonists of the X-linked inhibitor of apoptosis protein.** *Mol Cancer Ther* 2007, **6**:957-966.
60. Oost TK, Sun C, Armstrong RC, Al-Assaad AS, Betz SF, Deckwerth TL, Ding H, Elmore SW, Meadows RP, Olejniczak ET, Oleksijew A, Oltersdorf T, Rosenberg SH, Shoemaker AR, Tomaselli KJ, Zou H, Fesik SW: **Discovery of potent antagonists of the antiapoptotic protein XIAP for the treatment of cancer.** *J Med Chem* 2004, **47**:4417-4426.
61. Kami K, Doi R, Koizumi M, Toyoda E, Mori T, Ito D, Fujimoto K, Wada M, Miyatake S, Imamura M: **Survivin expression is a prognostic marker in pancreatic cancer patients.** *Surgery* 2004, **136**:443-448.
62. Qiao JG, Zhang YQ, Yin YC, Tan Z: **Expression of Survivin in pancreatic cancer and its correlation to expression of Bcl-2.** *World J Gastroenterol* 2004, **10**:2759-2761.
63. Altieri DC: **Survivin, cancer networks and pathway-directed drug discovery.** *Nat Rev Cancer* 2008, **8**:61-70.
64. Tsuji N, Asanuma K, Kobayashi D, Yagihashi A, Watanabe N: **Introduction of a survivin gene-specific small inhibitory RNA inhibits growth of pancreatic cancer cells.** *Anticancer Res* 2005, **25**:3967-3972.
65. Guan HT, Xue XH, Dai ZJ, Wang XJ, Li A, Qin ZY: **Down-regulation of survivin expression by small interfering RNA induces pancreatic cancer cell apoptosis and enhances its radiosensitivity.** *World J Gastroenterol* 2006, **12**:2901-2907.
66. Retzer-Lidl M, Schmid RM, Schneider G: **Inhibition of CDK4 impairs proliferation of pancreatic cancer cells and sensitizes towards TRAIL-induced apoptosis via downregulation of survivin.** *Int J Cancer* 2007, **121**:66-75.
67. Hayden MS, Ghosh S: **Signaling to NF-kappaB.** *Genes Dev* 2004, **18**:2195-2224.
68. Scheidereit C: **IkappaB kinase complexes: gateways to NF-kappaB activation and transcription.** *Oncogene* 2006, **25**:6685-6705.
69. Perkins ND: **Integrating cell-signalling pathways with NF-kappaB and IKK function.** *Nat Rev Mol Cell Biol* 2007, **8**:49-62.
70. Karin M, Cao Y, Greten FR, Li ZW: **NF-kappaB in cancer: from innocent bystander to major culprit.** *Nat Rev Cancer* 2002, **2**:301-310.
71. Basseres DS, Baldwin AS: **Nuclear factor-kappaB and inhibitor of kappaB kinase pathways in oncogenic initiation and progression.** *Oncogene* 2006, **25**:6817-6830.
72. Algul H, Adler G, Schmid RM: **NF-kappaB/Rel transcriptional pathway: implications in pancreatic cancer.** *Int J Gastrointest Cancer* 2002, **31**:71-78.
73. Scwabas GM, Fujioka S, Schmidt C, Evans DB, Chiao PJ: **NF-kappaB in pancreatic cancer.** *Int J Gastrointest Cancer* 2003, **33**:15-26.
74. Holcomb B, Yip-Schneider M, Schmidt CM: **The role of nuclear factor kappaB in pancreatic cancer and the clinical applications of targeted therapy.** *Pancreas* 2008, **36**:225-235.
75. Sebens S, Arlt A, Schafer H: **NF-kappaB as a molecular target in the therapy of pancreatic carcinoma.** *Recent Results Cancer Res* 2008, **177**:151-164.
76. Wang W, Abbruzzese JL, Evans DB, Larry L, Cleary KR, Chiao PJ: **The nuclear factor-kappa B RelA transcription factor is constitutively activated in human pancreatic adenocarcinoma cells.** *Clin Cancer Res* 1999, **5**:119-127.
77. Liptay S, Weber CK, Ludwig L, Wagner M, Adler G, Schmid RM: **Mitogenic and antiapoptotic role of constitutive NF-kappaB/Rel activity in pancreatic cancer.** *Int J Cancer* 2003, **105**:735-746.
78. Li L, Aggarwal BB, Shishodia S, Abbruzzese J, Kurzrock R: **Nuclear factor-kappaB and IkappaB kinase are constitutively active in human pancreatic cells, and their down-regulation by curcumin (diferuloylmethane) is associated with the suppression of proliferation and the induction of apoptosis.** *Cancer* 2004, **101**:2351-2362.
79. Farrow B, Sugiyama Y, Chen A, Uffort E, Nealon W, Mark Evers B: **Inflammatory mechanisms contributing to pancreatic cancer development.** *Ann Surg* 2004, **239**:763-769. discussion 769-771.
80. Schneider G, Saur D, Sivek JT, Fritsch R, Greten FR, Schmid RM: **IKKalpha controls p52/RelB at the skp2 gene promoter to regulate G1- to S-phase progression.** *Embo J* 2006, **25**:3801-3812.
81. Weichert W, Boehm M, Gekeler V, Bahra M, Langrehr J, Neuhaus P, Denkert C, Imre G, Weller C, Hofmann HP, Niesporek S, Jacob J, Dietel M, Scheidereit C, Kristiansen G: **High expression of RelA/p65 is associated with activation of nuclear factor-kappaB-dependent signaling in pancreatic cancer and marks a patient population with poor prognosis.** *Br J Cancer* 2007, **97**:523-530.
82. Arlt A, Vorndamm J, Breitenbroich M, Folsch UR, Kalthoff H, Schmidt WE, Schafer H: **Inhibition of NF-kappaB sensitizes human pancreatic carcinoma cells to apoptosis induced by etoposide (VP16) or doxorubicin.** *Oncogene* 2001, **20**:859-868.
83. Fujioka S, Scwabas GM, Schmidt C, Niu J, Frederick WA, Dong QG, Abbruzzese JL, Evans DB, Baker C, Chiao PJ: **Inhibition of constitutive NF-kappa B activity by I kappa B alpha M suppresses tumorigenesis.** *Oncogene* 2003, **22**:1365-1370.
84. Muerkoster S, Arlt A, Witt M, Gehrz A, Haye S, March C, Grohmann F, Wegehenkel K, Kalthoff H, Folsch UR, Schafer H: **Usage of the NF-kappaB inhibitor sulfasalazine as sensitizing agent in combined chemotherapy of pancreatic cancer.** *Int J Cancer* 2003, **104**:469-476.
85. Sato T, Odagiri H, Ikenaga SK, Maruyama M, Sasaki M: **Chemosen-sitivity of human pancreatic carcinoma cells is enhanced by IkappaBalpha super-repressor.** *Cancer Sci* 2003, **94**:467-472.
86. Thomas RP, Farrow BJ, Kim S, May MJ, Hellmich MR, Evers BM: **Selective targeting of the nuclear factor-kappaB pathway enhances tumor necrosis factor-related apoptosis-inducing ligand-mediated pancreatic cancer cell death.** *Surgery* 2002, **132**:127-134.
87. Wang Z, Banerjee S, Li Y, Rahman KM, Zhang Y, Sarkar FH: **Down-regulation of notch-1 inhibits invasion by inactivation of nuclear factor-kappaB, vascular endothelial growth factor, and matrix metalloproteinase-9 in pancreatic cancer cells.** *Cancer Res* 2006, **66**:2778-2784.
88. Wang Z, Zhang Y, Li Y, Banerjee S, Liao J, Sarkar FH: **Down-regulation of Notch-1 contributes to cell growth inhibition and apoptosis in pancreatic cancer cells.** *Mol Cancer Ther* 2006, **5**:483-493.
89. Hamacher R, Saur D, Fritsch R, Reichert M, Schmid RM, Schneider G: **Casein kinase II inhibition induces apoptosis in pancreatic cancer cells.** *Oncol Rep* 2007, **18**:695-701.
90. Arlt A, Rosenstiel P, Kruse ML, Grohmann F, Minkenberg J, Perkins ND, Folsch UR, Schreiber S, Schafer H: **IEX-1 directly interferes with RelA/p65 dependent transactivation and regulation of apoptosis.** *Biochim Biophys Acta* 2007.
91. Sasada T, Azuma K, Hirai T, Hashida H, Kanai M, Yanagawa T, Takabayashi A: **Prognostic significance of the immediate early**

- response gene X-1 (IEX-1) expression in pancreatic cancer.** *Ann Surg Oncol* 2008, **15**:609-617.
92. Bader AG, Kang S, Zhao L, Vogt PK: **Oncogenic PI3K deregulates transcription and translation.** *Nat Rev Cancer* 2005, **5**:921-929.
 93. Cardone MH, Roy N, Stennicke HR, Salvesen GS, Franke TF, Stanbridge E, Frisch S, Reed JC: **Regulation of cell death protease caspase-9 by phosphorylation.** *Science* 1998, **282**:1318-1321.
 94. Datta SR, Dudek H, Tao X, Masters S, Fu H, Gotoh Y, Greenberg ME: **Akt phosphorylation of BAD couples survival signals to the cell-intrinsic death machinery.** *Cell* 1997, **91**:231-241.
 95. del Peso L, Gonzalez-Garcia M, Page C, Herrera R, Nunez G: **Interleukin-3-induced phosphorylation of BAD through the protein kinase Akt.** *Science* 1997, **278**:687-689.
 96. Jones RG, Parsons M, Bonnard M, Chan VS, Yeh WC, Woodgett JR, Ohashi PS: **Protein kinase B regulates T lymphocyte survival, nuclear factor kappaB activation, and Bcl-X(L) levels in vivo.** *J Exp Med* 2000, **191**:1721-1734.
 97. Bondar VM, Sweeney-Gotsch B, Andreeff M, Mills GB, McConkey DJ: **Inhibition of the phosphatidylinositol 3'-kinase-AKT pathway induces apoptosis in pancreatic carcinoma cells in vitro and in vivo.** *Mol Cancer Ther* 2002, **1**:989-997.
 98. Schlieman MG, Fahy BN, Ramsamooj R, Beckett L, Bold RJ: **Incidence, mechanism and prognostic value of activated AKT in pancreas cancer.** *Br J Cancer* 2003, **89**:2110-2115.
 99. Shah SA, Potter MW, Hedeshian MH, Kim RD, Chari RS, Callery MP: **PI-3' kinase and NF-kappaB cross-signaling in human pancreatic cancer cells.** *J Gastrointest Surg* 2001, **5**:603-612. discussion 612-603.
 100. Asano T, Yao Y, Zhu J, Li D, Abbruzzese JL, Reddy SA: **The PI 3-kinase/Akt signaling pathway is activated due to aberrant Pten expression and targets transcription factors NF-kappaB and c-Myc in pancreatic cancer cells.** *Oncogene* 2004, **23**:8571-8580.
 101. Asano T, Yao Y, Shin S, McCubrey J, Abbruzzese JL, Reddy SA: **Insulin receptor substrate is a mediator of phosphoinositide 3-kinase activation in quiescent pancreatic cancer cells.** *Cancer Res* 2005, **65**:9164-9168.
 102. Abe N, Watanabe T, Masaki T, Mori T, Sugiyama M, Uchimura H, Fujioka Y, Chiappetta G, Fusco A, Atomi Y: **Pancreatic duct cell carcinomas express high levels of high mobility group I(Y) proteins.** *Cancer Res* 2000, **60**:3117-3122.
 103. Trapasso F, Sarti M, Cesari R, Yendamuri S, Dumon KR, Aqeilan RI, Pentimalli F, Infante L, Alder H, Abe N, et al.: **Therapy of human pancreatic carcinoma based on suppression of HMGA1 protein synthesis in preclinical models.** *Cancer Gene Ther* 2004, **11**:633-641.
 104. Liau SS, Jazag A, Ito K, Whang EE: **Overexpression of HMGA1 promotes anoikis resistance and constitutive Akt activation in pancreatic adenocarcinoma cells.** *Br J Cancer* 2007, **96**:993-1000.
 105. Liau SS, Jazag A, Whang EE: **HMGA1 is a determinant of cellular invasiveness and in vivo metastatic potential in pancreatic adenocarcinoma.** *Cancer Res* 2006, **66**:11613-11622.
 106. Kolb S, Fritsch R, Saur D, Reichert M, Schmid RM, Schneider G: **HMGA1 controls transcription of insulin receptor to regulate cyclin D1 translation in pancreatic cancer cells.** *Cancer Res* 2007, **67**:4679-4686.
 107. Liau SS, Whang EE: **HMGA1 is a molecular determinant of chemoresistance to gemcitabine in pancreatic adenocarcinoma.** *Clin Cancer Res* 2008, **14**:1470-1477.
 108. Ng SSV, Tsao MS, Chow S, Hedley DW: **Inhibition of phosphatidylinositide 3-kinase enhances gemcitabine-induced apoptosis in human pancreatic cancer cells.** *Cancer Res* 2000, **60**:5451-5455.
 109. Ng SS, Tsao MS, Nicklee T, Hedley DW: **Wortmannin inhibits pkb/akt phosphorylation and promotes gemcitabine antitumor activity in orthotopic human pancreatic cancer xenografts in immunodeficient mice.** *Clin Cancer Res* 2001, **7**:3269-3275.
 110. Fahy BN, Schlieman M, Virudachalam S, Bold RJ: **AKT inhibition is associated with chemosensitisation in the pancreatic cancer cell line MIA-PaCa-2.** *Br J Cancer* 2003, **89**:391-397.
 111. Fahy BN, Schlieman MG, Virudachalam S, Bold RJ: **Inhibition of AKT abrogates chemotherapy-induced NF-kappaB survival mechanisms: implications for therapy in pancreatic cancer.** *J Am Coll Surg* 2004, **198**:591-599.
 112. Arlt A, Gehrz A, Muerkoster S, Vorndamm J, Kruse ML, Folsch UR, Schafer H: **Role of NF-kappaB and Akt/PI3K in the resistance of pancreatic carcinoma cell lines against gemcitabine-induced cell death.** *Oncogene* 2003, **22**:3243-3251.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

