

Diabetologia (2013) 56:350–358
 DOI 10.1007/s00125-012-2775-y

ARTICLE

Adaptive changes of human islets to an obesogenic environment in the mouse

S. Gargani · J. Thévenet · J. E. Yuan · B. Lefebvre ·
 N. Delalleau · V. Gmyr · T. Hubert · A. Duhamel ·
 F. Pattou · J. Kerr-Conte

Received: 31 May 2012 / Accepted: 24 October 2012 / Published online: 29 November 2012

© The Author(s) 2012. This article is published with open access at Springerlink.com

Abstract

Aims/hypothesis In this study, we used an immunodeficient mouse model to explore, *in vivo*, the longitudinal adaptation of human islets to an obesogenic environment.

Methods Non-diabetic *Rag2^{-/-}* mice ($n=61$) were transplanted with human islets (400 islet equivalents [IEQ]) from six pancreases: four non-diabetic and two with overt metabolic dysfunction (older, high HbA_{1c} or history of diabetes).

Electronic supplementary material The online version of this article (doi:10.1007/s00125-012-2775-y) contains peer-reviewed but unedited supplementary material, which is available to authorised users.

S. Gargani · J. Thévenet · J. E. Yuan · B. Lefebvre · N. Delalleau ·
 V. Gmyr · T. Hubert · A. Duhamel · F. Pattou · J. Kerr-Conte
 Université Lille Nord de France,
 Lille, France

S. Gargani · J. Thévenet · B. Lefebvre · N. Delalleau · V. Gmyr ·
 T. Hubert · A. Duhamel · F. Pattou · J. Kerr-Conte
 UDSL, IMPRT,
 Lille, France

S. Gargani · J. Thévenet · N. Delalleau · V. Gmyr · T. Hubert ·
 F. Pattou · J. Kerr-Conte (✉)
 Inserm U859 Biotherapies of Diabetes, Faculty of Medicine,
 1 Place de Verdun,
 F59045 Lille, France
 e-mail: jkerr-conte@univ-lille2.fr

A. Duhamel · F. Pattou · J. Kerr-Conte
 Centre Hospitalier Régional Universitaire de Lille,
 Lille, France

A. Duhamel
 CERIM, Faculty of Medicine,
 Lille, France

Present address:

B. Lefebvre
 Inserm U837, Alzheimer and Tauopathies,
 Lille, France

Animals were fed for 12 weeks with a control or high-fat diet (HFD), and followed for weight, serum triacylglycerol, fasting blood glucose and human C-peptide. After the mice were killed, human grafts and the endogenous pancreas were analysed for endocrine volume, distribution of beta and alpha cells, and proliferation.

Results Transplanted mice on an HFD gained significantly more weight ($p<0.001$) and had higher fasting glycaemia (2–12 weeks; $p=0.0002$) and consistently higher fasting human C-peptide levels (2–12 weeks; $p=0.04$) compared with those on the control diet. Histology demonstrated doubling of human islet graft volume at 12 weeks in animals on the HFD and increased beta cell volume ($p<0.001$), but no change in alpha cell volume. Human islet function (hyperbolic product HOMA2%BS) at 12 weeks was four times lower in HFD animals ($p<0.001$ vs controls) because of insufficient beta cell adaptation to decreased (70%) sensitivity (HOMA%S). Human islets obtained from donors with metabolic dysfunction failed to adapt to the HFD.

Conclusions/interpretation This longitudinal study provides direct evidence that human islets adapt both endocrine and beta cell mass, function and gene expression to obesity *in vivo*. The present model will facilitate the identification of mechanisms by which human islets adapt to obesity *in vivo* and the cell type(s) responsible, and factors predisposing human beta cells to decompensation.

Keywords Expansion · High-fat diet · Human beta cells · Human islets · Obesity · Transplantation

Abbreviations

HFD	High-fat diet
HOMA2%B	HOMA index of beta cell function
HOMA2%S	HOMA index of insulin sensitivity
IEQ	Islet equivalents

Introduction

Decoding how diabetes and obesity are interrelated is a complex and interdisciplinary research endeavour [1, 2]. Islet adaptation to obesity is well established in rodents, where islets compensate for insulin-resistant states by increasing beta cell mass and function to maintain normoglycaemia [3]. Major differences have, however, emerged in the field of beta cell biology between rodent and human cells, making work with human islets crucial [4]. Indeed, clinically, most obese individuals do not develop diabetes because islets compensate for insulin resistance [5, 6]. Yet direct evidence that human islet mass adapts longitudinally to obesity *in vivo* is lacking and, moreover, little information is available on the mechanisms and cell type(s) involved. Current evidence for increased beta cell mass in obese humans (vs lean) is based entirely on postmortem histology [7]. Beta cell mass quantification in humans by *in vivo* imaging is nascent [8] and should one day enable longitudinal studies.

The aim of this study was to explore the longitudinal adaptation of human islets to an obesogenic environment. The transplantation of human islets into immune-compromised mice is a validated technique with which to study human beta cells in an *in vivo* environment [9] and when human C-peptide levels are measured, this technique provides a quantitative assessment of human islet function [10, 11]. In this study, we transplanted human islets into an immunodeficient mouse strain sensitive to high-fat diet (HFD)-induced obesity and show direct evidence that non-diabetic human islets adapt both endocrine and beta cell mass, function and gene expression to obesity *in vivo*. However, dysfunctional human islets showed arrested adaptation. This model may foster deciphering of the mechanisms by which human islets adapt to obesity and the cell type(s) that are responsible, and possibly identify key factors predisposing human beta cells to decompensation [1].

Methods

Human islets Pancreases were harvested from human brain-deceased donors in agreement with French regulations and with our institutional ethics committee, and were processed for islet isolation as previously described [12]. Four of the pancreases were from non-diabetic donors and two were from donors with overt metabolic dysfunction (older, high HbA_{1c} or a history of diabetes; see electronic supplementary material [ESM] Tables 1 and 2). All islets showed more than 90% viability after culture, but were quantitatively insufficient for clinical islet transplantation [13].

Animals, human islet transplantation and induction of obesity Male C57BL6 *Rag2*^{-/-} immunodeficient mice of

8–9 weeks old ($n=61$; courtesy of A. Bouloumié, Inserm U858, Toulouse, France [14], and RAGN12-M, Taconic, Germantown, NY, USA) were transplanted under the kidney capsule as previously described [10] with 400 human islet equivalents (IEQ). Mouse studies were performed in accordance with the guidelines from the local animal experimentation committee. As previously described, islets from each human donor were distributed symmetrically across experimental groups to control for potential differences in islet quality between the donors [15]. Eight to ten mice were transplanted per donor per condition. Animals were fed for 12 weeks with a control diet or HFD (NJ ref D12450B; Research Diets, New Brunswick, NJ, USA; ESM Table 3).

All mice were followed for 12 weeks for weight, serum triacylglycerol, 6 h fasting blood glucose and human C-peptide. At the end of the study, mice were fasted for 6 h before receiving an oral bolus of glucose (2 g/kg body weight) diluted in saline buffer (0.9% sodium chloride). Tail blood was collected at 0, 30 and 60 min after administration to evaluate human C-peptide levels (ultrasensitive human C-peptide kit; Mercodia, Uppsala, Sweden). Blood glucose was measured up to 180 min (Accu-Chek Go glucometer; Roche, Meylan, France). HOMA, calculated using fasting human C-peptide and blood glucose levels in mice (www.dtu.ox.ac.uk/homacalculator/index.php) [16], was used as an index of insulin sensitivity (HOMA2%S), beta cell function (HOMA2%B) and the hyperbolic product (HOMA2%S×HOMA2%B/100). Results of 12 clinical-grade islet preparations (glycaemia, human C-peptide) transplanted into 20 immunodeficient mice, as previously described [10], were used to calculate theoretical HOMA2%B and HOMA2%S. Fat content was quantified by MRI using ten images/mouse before killing, as previously described [17].

Immunohistochemistry To assess proliferation, BrdU was injected 18 h before the mice were killed or administered for 7 days in the drinking water. Both the endogenous mouse pancreas and human grafts were fixed and embedded in paraffin for sectioning and analysis of endocrine mass, distribution of beta cells and proliferation. Endocrine mass of the endogenous pancreas was determined with Papanicolaou staining of sections [18]. For endocrine mass assessment in human islet grafts, the entire block was sectioned; one section every 50 μm was immunolabelled with chromogranin A primary antibody (M0869, 1/100; Dako, Glostrup, Denmark) and revealed with Envision (K4003; Dako). Similarly, beta and alpha cell volumes were determined using Alexa 488-conjugated anti-human C-peptide (green) (Monsam, Uden, the Netherlands) or anti-glucagon (G2654; Sigma, Saint-Quentin-Fallavier, France) using the Lightning-Link Conjugation Kit (Innova Biosciences, Cambridge, UK); nuclei were stained with DAPI (blue) (H-1200J; Vector Laboratories, Burlingame, CA, USA).

Endocrine or beta cell proliferation in grafts was respectively determined with three-colour fluorescence staining, BrdU–Chromogranin A–DAPI or BrdU–insulin–DAPI, or Ki67-replaced BrdU (MIB-1; M7240, 1/100; Dako). After antigen retrieval (microwave, citrate buffer pH6), tissues were incubated overnight at 4°C with primary rat anti-BrdU antibody (AbD, BU1/75, 1/100; Serotec, Paris, France) followed by goat anti-rat Alexa Fluor 594 antibody (A11007, 1/300; Invitrogen, St Aubin, France). Double labelling was performed with rabbit anti-chromogranin A (M0869, 1/100; Dako) or mouse anti-insulin/proinsulin (AbD; D6C4, 1/300; Serotec), followed by donkey anti-rabbit Alexa Fluor 488 antibody (A21206, 1/300; Invitrogen) or goat anti-mouse Alexa Fluor 488 (A11001, 1/300; Invitrogen). Double labelling for chromogranin A, the pan-endocrine marker, and cytokeratin 19, as described [19, 20] suggestive of endocrine neogenesis, was quantified as previously described (the number of CK19-positive ductal cells co-stained with chromogranin A) [7].

Morphometric analysis To quantify the total islet surface of human graft sections and islet surface in mouse pancreases, slides were scanned with a Nikon Eclipse Ti microscope, and the fractional area was digitally quantified (Nis Elements AR 3.0) (Nikon, Champigny sur Marne, France). Planimetry, used for clinical tumour volume appreciation (surface in microns²×distance in microns between paraffin sections), was used to determine endocrine, beta and alpha volumes (μm³ or mm³) of entire human islet grafts. Leica DM-R microscope and ImageJ (National Institutes of Health, Bethesda, MD, USA) were used for the quantitative analysis.

RNA tissue extraction and Q-PCR RNA was isolated from snap-frozen human islet grafts ($n=14$) immediately after killing using the RNeasy Micro Kit (Qiagen, Paris, France). RNA quantification was performed with the NanoDrop spectrophotometer (ND-1000; Thermo Scientific, Wilmington, NC, USA) and reverse transcription of 0.5 μg/μl RNA was performed with the High Capacity cDNA Reverse Transcription Kit (Applied Biosystems, Paris, France). For small human samples, pre-amplification of the gene of interest was performed with the TaqMan PreAmp Kit (Applied Biosystems), followed by quantitative PCR analysis (SYBR Green; Applied Biosystems). The *INS* primers (forward: 5'GCAACTAGACGCAGCCCGCA'3; reverse: 5'GGCTTTATTCCATCTCTCTCGGTGC'3) were designed on the basis of sequence alignments of the *INS* region of human species and unrelated to the mouse; β-actin was the reference gene. Only one amplification product for *INS* (86°C) and for actin (82°C) was observed in human islets grafts and none for genomic DNA from the mouse kidney.

Statistical analysis For each variable, the change over time (2 and 12 weeks; denoted 2–12 weeks) as well as the

difference at any given time was compared according to the two groups (control vs HFD) using analysis of variance based on a linear mixed model in order to adjust for the donor effect. The fixed effect was the group and the random effect was the donor. The same procedure was used to compare the histological data at 12 weeks according to the two groups. The analysis of repeated measurements for weight was also performed using the linear mixed model. In this analysis, the effect 'donor' was considered as a random effect and the correlations between the repeated measurements were modelled by using a first autoregressive covariance model. Our choice of this model was based on Akaike's information criterion. The validity of the linear mixed model was checked using the analysis of the residuals; the normality of residuals was tested with the Shapiro–Wilk test and the outliers were identified using box plots. The influence of each individual on the results was investigated using the Cook distance. If necessary, the stability of results after removing outliers or influential individuals was checked. Data were plotted as mean±SEM.

Statistical analyses were performed using StatView (SAS Institute, Cary, NC, USA) and SAS (SAS Institute Inc., Cary, NC, USA). p values<0.05 were considered statistically significant.

Results

HFD-induced obesity in *Rag2*^{-/-} mice transplanted with human islets A preliminary study using young (4-week-old) or adult male immunodeficient C57BL6 *Rag2*^{-/-} mice ($n=14$) confirmed that adults, as suggested [14, 21], gain weight on an HFD similar to male C57BL6 wild-type animals after 12 weeks ($p=0.110$, results not shown). All subsequent experiments used adult male *Rag2*^{-/-} mice with a human islet graft.

Even with a human islet graft, *Rag2*^{-/-} mice on a HFD gained considerable weight (Fig. 1a) as compared with mice on the control diet ($p<0.0001$). This corresponded to higher BMI, serum triacylglycerol levels (Fig. 1b) and fat indices (ESM Table 4), including abdominal fat measured with MRI (Fig. 1c: HFD 52.27±4.39% vs controls 25.7±0.604%, $p<0.05$) in adult *Rag2*^{-/-} HFD mice vs controls.

Confirmation of adaptation of endogenous pancreas Morphometric analysis of the endogenous mouse pancreas confirmed that total islet area was doubled in HFD mice. There was no difference in the number of islets (per mm² tissue surface) between the two groups (ESM Table 4), but the islets were larger and the proportion of beta cells higher in the pancreases of mice on the HFD at 12 weeks. Total pancreatic and endocrine cell proliferation was elevated in the endogenous pancreases of HFD mice.

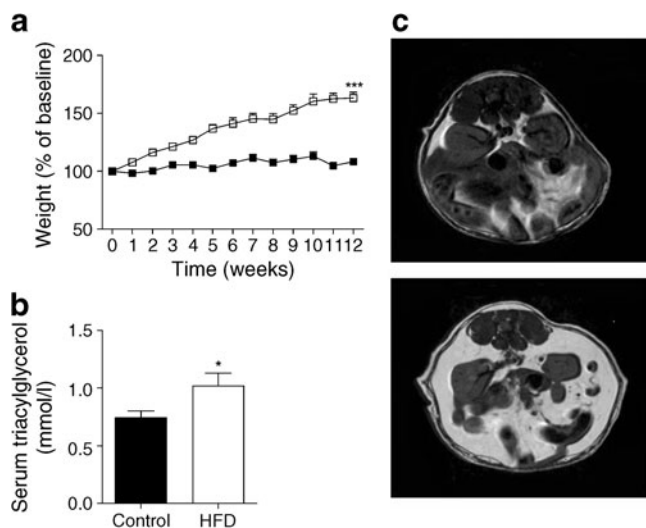


Fig. 1 (a) Weight gain (% of baseline) over 12 weeks in animals grafted with human islets from four donors. White squares, HFD grafts ($n=23$); black squares, controls ($n=22$) (linear mixed model, see Methods, $***p<0.001$). (b) Serum triacylglycerol levels at 12 weeks ($*p<0.05$ vs controls). (c) MRI was used to quantify (10 sections/mouse) abdominal fat content (expressed as a percentage \pm SEM) before the mice were killed (HFD $52.27\pm 4.39\%$ vs controls $25.7\pm 0.604\%$, $*p<0.05$, fat appears in white)

Histological assessment at 12 weeks of the human grafts in animals on control or HFD Morphometric analysis with chromogranin A (Fig. 2a, 196–210 sections/group; see Methods) confirmed the more than doubling of human endocrine graft volume (HFD 0.094 ± 0.017 vs controls 0.042 ± 0.008 mm^3 ; $p=0.0006$) (Fig. 2b). To determine if this was due to endocrine cell hypertrophy of human islets as proposed [22] in HFD animals, individual cell surfaces were measured; no difference was observed (HFD 138 ± 5.9 μm^2 vs controls 135.3 ± 4.6 μm^2 ; $p=0.7$). Indeed a 2.6-fold increase in beta cell volume (Fig. 2c; HFD 0.071 ± 0.0021 mm^3 vs controls 0.027 ± 0.001 mm^3 ; $p=0.0006$) was observed at 12 weeks (Fig. 2d), yet alpha cell volume was identical between the groups (Fig. 2e; HFD 0.011 ± 0.002 mm^3 vs controls 0.009 ± 0.002 mm^3 ; $p=0.0735$). It is worth noting that human beta cells represented $64\pm 2.0\%$ and $75\pm 1.86\%$ of the total endocrine volume at 12 weeks in control and HFD mice, respectively ($p<0.05$), whereas alpha cells made up a $18.08\pm 1.49\%$ in control animals and a statistically significantly smaller proportion in HFD animals ($12.45\pm 1.69\%$, $p<0.05$). Relative expression of human *INS* mRNA was 3.4-fold higher in human islets grafted in HFD mice (Fig. 3).

Total proliferation in human grafts, assessed at 12 weeks in all donors after overnight incubation with BrdU, was four times greater in HFD animals (BrdU-positive nuclei/total nuclei in graft: HFD $1.027\pm 0.259\%$ vs controls $0.259\pm 0.157\%$; $p=0.03$). When BrdU was administered 1 day prior to mouse killing at 12 weeks, endocrine islet cell proliferation (double chromogranin A/BrdU cells) (Fig. 4a) was low

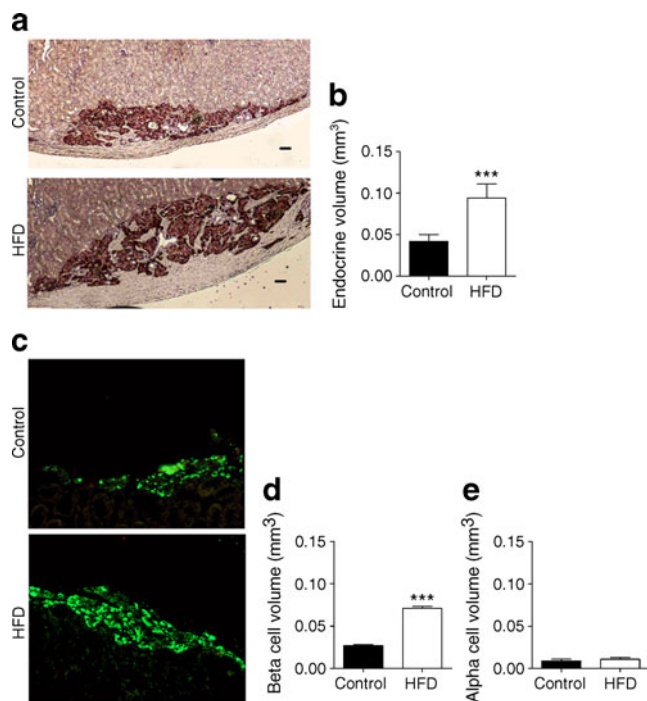


Fig. 2 (a) Representative sections of paraffin-embedded human grafts stained with the endocrine marker chromogranin A (DAB, brown) in HFD and control-diet mice (scale bar, 50 μm). (b) Morphometric analysis for total endocrine volume (chromogranin A) (HFD [$n=6$] vs control [$n=7$], >196 sections/group; linear mixed model, see Methods, $***p<0.001$). (c) Sections of paraffin-embedded human grafts stained with the beta cell marker C-peptide (green). (d) Morphometric analysis for beta cell volume (HFD [$n=6$] vs control [$n=7$], $***p<0.001$). (e) Morphometric analysis for alpha cell volume (HFD [$n=6$] vs control [$n=7$])

in all animals at 12 weeks (Fig. 4b), as was beta cell proliferation (double insulin/BrdU-positive cells; $1/1,500$ cells; $p=NS$). Anti-human anti-Ki67 antibody (Fig. 4c) showed that general proliferation in human grafts was higher in HFD mice (HFD $0.467\pm 0.082\%$ vs controls $0.214\pm 0.054\%$, $p=0.03$) and endocrine cell proliferation (double chromogranin A/Ki67) was higher, albeit insignificantly in HFD animals (HFD $0.254\pm 0.065\%$ vs controls $0.119\pm 0.034\%$; $p=0.08$) (Fig. 4d). Beta cell proliferation

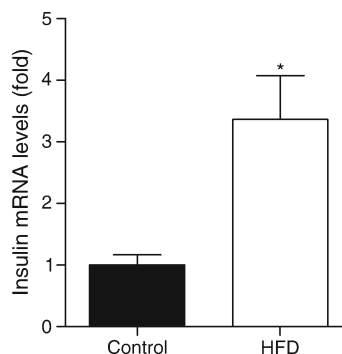
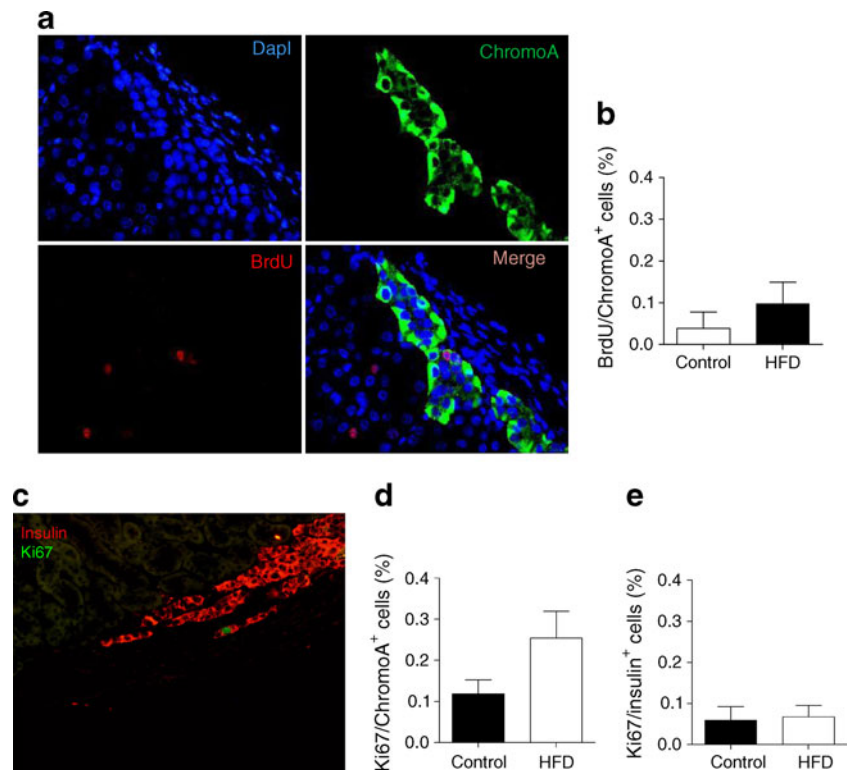


Fig. 3 Expression of the human *INS* gene in grafts (HFD [$n=7$] vs control [$n=7$]; $*p<0.05$) (see Methods)

Fig. 4 (a) Proliferation was determined on sections of human grafts at 12 weeks following an overnight incubation with BrdU (nuclei in blue, BrdU in red, chromogranin A [chromoA] in green or merge). (b) BrdU/chromogranin A endocrine-positive cell quantification in human islet grafts (HFD [$n=4$] vs control [$n=5$]). (c) Proliferation with Ki67 in beta cells (image shows merge of insulin in red, Ki67 in green). Quantification of (d) Ki67/chromogranin A and (e) Ki67/insulin-positive cells in human islets grafts (HFD [$n=4$] vs control [$n=5$]; linear mixed model, see Methods, $p=0.08$)



(double Ki67/insulin-positive) at 12 weeks showed no statistically significant difference between groups (HFD $0.067 \pm 0.033\%$ vs $0.058 \pm 0.028\%$; $p=0.1$) (Fig. 4e).

We subsequently performed a kinetic study in mice transplanted with human islets, where BrdU was administered in the drinking water for 7 days prior to killing at 4, 6, 8 or 12 weeks (ESM Fig. 1a). Cumulative 7 day chromogranin A-positive endocrine proliferation (not shown) appeared higher at 12 weeks in HFD animals vs controls in accordance with the 1 day BrdU labelling, but also at 6 weeks HFD (results not shown). We also readily detected (4%) 7 day cumulative proliferation in C-peptide-positive human beta cells at 6 and 12 weeks. Double labelling for ductal (anti-cytokeratin 19) and a pan-endocrine marker (anti-chromogranin A) was maximum at 8 weeks HFD (vs controls), suggesting the potential contribution of endocrine neogenesis (ESM Fig. 1b). Although further confirmation is required, both proliferation and neogenesis appear to potentially contribute to the expanded human islet mass in animals on an HFD.

Assessment of human islet graft function Fasting glycaemia was higher in HFD mice compared with controls at 2–12 weeks ($p=0.0002$) (Fig. 5a). It should be noted that the fasting glycaemia of *Rag2*^{-/-} mice ($n=11$) with no islet graft was 6.83 ± 0.28 mmol/l. Compared with controls, HFD animals had a tendency towards higher plasma human C-peptide levels after 12 weeks compared with 2 weeks (2 weeks: controls 172.42 ± 28.75 pmol/l vs HFD $479.54 \pm$

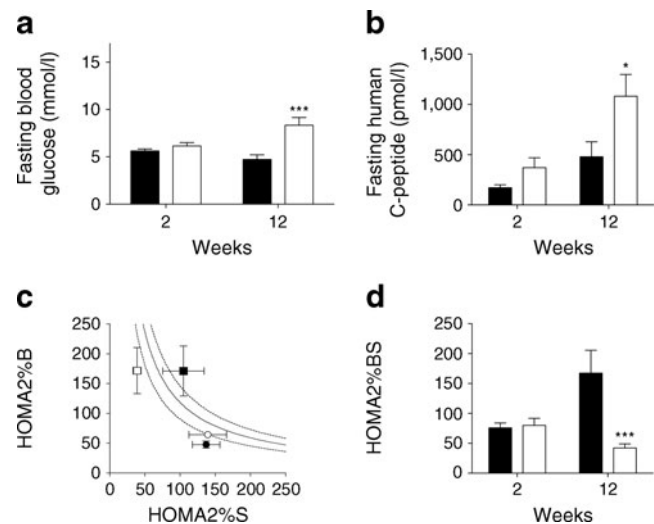


Fig. 5 (a) Fasting glycaemia (mmol/l) in animals with a human graft subsequently placed on an HFD (white bars) or control (black bars) diet (2–12 weeks, linear mixed model, see Methods, $***p<0.001$). (b) Human C-peptide in tail blood plasma over the 12 week period in HFD (white bars; $n=20$) vs control diet (black bars; $n=19$) animals (2–12 weeks, linear mixed model, see Methods, $*p<0.05$). (c) Evolution of HOMA2%B and HOMA2%S in animals at 2 weeks (circles) or 12 weeks (squares) on an HFD (white) or control diet (black) (mean \pm SEM), plotted on hyperbolic curve (see Methods; the dotted lines represent the upper and lower 95% CIs of the mean). (d) Hyperbolic product (HOMA2%BS) in mice implanted for 12 weeks with non-diabetic human islets (HFD) [white bars] vs controls [black bars] at 12 weeks ($***p<0.001$; $*p<0.05$ controls, 2 vs 12 weeks)

148.84 pmol/l; 12 weeks: controls 370.89±97.91 pmol/l vs HFD 1,081.41±216.32; 2–12 weeks, $p=0.04$) (Fig. 5b). Results for individual human islet donors are depicted in ESM Fig. 2. Human C-peptide expressed as a function of glycaemia was also higher at 12 weeks (vs controls).

We used a surrogate HOMA2 based on fasting glycaemia and human C-peptide levels to longitudinally determine changes in human islet graft function in animals at 2 and 12 weeks. Figure 5c shows HOMA2%B and HOMA2%S for animals with respect to hyperbolic product value ($\text{HOMA2}\%B \times \text{HOMA2}\%S / 100$), calculated as described in the Methods (the dotted lines represent the upper and lower 95% confidence intervals of the mean). No difference was observed at 2 weeks between the two groups. An increase in HOMA2%B was observed with time in both control and HFD animals (respectively, 2–12 weeks, $p=0.0075$ and $p=0.06$). Control animals showed no change in HOMA2%S over time ($p=0.412$); however, HFD animals showed a deterioration in HOMA2%S (>70% at 12 week vs 2 weeks, $p=0.03$) corresponding to a 1.7-fold increase in insulin resistance ($\text{HOMA-IR} = 1/\text{HOMA-S} \times 100$) over time and compared with control animals at 12 weeks. Increased peripheral insulin resistance was also confirmed in insulin tolerance tests (not shown). As a result, human islet function (hyperbolic product HOMA2%BS) showed an improvement over time (2–12 weeks, $p=0.02$) in control animals that was not observed in HFD animals at 12 weeks (Fig. 5d); HOMA2%BS at 12 weeks was four times lower in HFD animals (vs controls, $p<0.001$) due to insufficient beta cell adaptation to decreased (70%) sensitivity (HOMA%S). An OGTT ($n=18$ mice) at 12 weeks (Fig. 6a) showed slightly abnormal glucose tolerance in HFD animals (vs $\text{AUC}_{\text{glucose}}$ in control group, $p=0.001$); basal human C-peptide secretion and 60 min post glucose stimulation were higher in islets transplanted in HFD animals (vs controls, $p=0.06$; Fig. 6b), but only control islets were stimulated by glucose.

To further test the model, we evaluated the adaptation of islets isolated from two donors with overt disturbances in glucose metabolism (older donors, high HbA_{1c} or a history

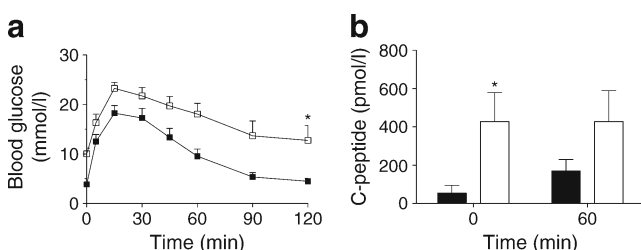


Fig. 6 (a) Representative blood glucose levels during an OGTT 12 weeks after human islet transplantation, HFD (white squares, $n=9$) vs control diet (black squares, $n=9$) (5–120 min, $*p<0.05$). (b) Human C-peptide secretion at 0 and 60 min in islets transplanted in HFD animals (white bars) compared with controls (black bars) (HFD vs controls at 0 min, $*p<0.05$, and at 60 min, $p=0.06$)

of diabetes; ESM Table 2). As described in the Methods, islets from each donor were distributed symmetrically across the experimental groups. Despite identical weight gain, higher blood glucose levels were observed earlier in HFD animals grafted with diabetic islets. No functional adaptation to HFD was observed (HOMA-B and C-peptide/glycaemia levels were lower in the HFD group vs controls), which was confirmed by low HOMA2%BS values at 12 weeks. Morphometric analysis showed no adaptation of endocrine volume to the HFD compared with control non-diabetic donors, and beta cells only accounted for approximately 10% of diabetic islets (% C-peptide surface/chromogranin A surface) compared with $64 \pm 2.0\%$ (control) to $75 \pm 1.86\%$ (HFD) in non-diabetic donor islets (ESM Fig. 3). No proliferation could be detected.

Discussion

In this study, we have explored the longitudinal adaptation of human islets to an obesogenic environment and showed direct evidence that non-diabetic human islets adapt both endocrine and beta cell mass, function and gene expression to obesity in vivo. In fact, Tyrberg et al were the first to show the proof of concept that human islets xenotransplanted into obese hyperglycaemic–hyperinsulinaemic *ob/ob* mice could adapt to murine obesity [23] but the mice required immunosuppression, which attenuates islet compensation [24], limiting graft follow-up to 2 weeks. After first testing C57BL6 male *nu/nu* mice that do not gain weight on a 12 week HFD (results not shown), adult male *Rag2^{-/-}* animals (lymphocyte B and T deficient) were selected after a pilot study, to address the question: can human islets adapt to an obesogenic environment in mice? In this particular study, we first purposely chose young donors since they are most likely to adapt. Second, we chose not to render the animals diabetic and to use a mini-human islet graft—400 IEQ vs the 2,000 IEQ classically used to cure diabetes, since this enables one to follow the adaptation to obesity in both the endogenous murine islets as well as the human cells. In fact, a mini-graft of <500 islets was chosen since higher numbers of islets (1,000, 2,000 and 3,000 IEQ) lead to an atrophy of endogenous islets (J. Kerr-Conte, unpublished observation) in accordance with previous publications [25, 26].

Concomitant with the increased weight gain, doubling of abdominal fat, increased serum triacylglycerol and reduced insulin sensitivity (HOMA-S) in 12 week HFD animals (vs controls), we report that human islet grafts (400 islets) showed functional compensation, measured as a more than doubling of fasting human C-peptide in mouse serum (vs controls), and histological adaptation of islet endocrine mass including increased beta cells. Increased human islet C-peptide secretion may be the simple result of islet

adaptation to hyperglycaemia [27] yet, in this study, increased blood glucose levels were only observed at 12 weeks; it should be noted that glycaemia in HFD animals was lower than in C57BL6 wild-type or *Rag2*^{-/-} animals on an HFD (results not shown). This has been previously attributed to the human set point (1.0 g/l) of transplanted cells compared with the mouse set point (1.5–2.0 g/l), as previously described for human embryonic stem-cell-derived islets grafted into non-diabetic immunodeficient mice [11]. In addition, increased C-peptide/glycaemia was also observed over time in mouse fed an HFD (vs controls at 12 weeks).

The doubling of human endocrine islet volume and increased percentage of beta cells (%beta/endocrine cells) paralleled the adaptation of the murine endogenous pancreas. At 12 weeks, total proliferation was higher in both the endogenous pancreas and the human graft in HFD animals. Human endocrine islet cell proliferation at 12 weeks, determined with Ki67, was higher in HFD grafts (vs controls, albeit $p=0.08$) but beta cell proliferation was extremely rare. In initial studies, BrdU was administered the day before killing and not for 4 [27] to 7 days or 1 month, as recently published [28], which might be responsible for the extremely low beta cell proliferation seen in our study. Ki67/chromogranin A-positive cells exhibited twofold greater proliferation than BrdU/chromogranin A-positive cells, but beta cell proliferation remained rare to absent.

In a subsequent kinetic study (ESM Fig. 1), BrdU was administered for 7 days in the drinking water prior to killing. Cumulative 7 day endocrine proliferation and C-peptide beta cell proliferation had a tendency to be higher at 6 and 12 weeks in HFD animals vs controls. Chromogranin A-positive ductal cells, suggestive of neogenesis, peaked at 8 weeks in HFD grafts. It should be noted that we purposely double-labelled with the pan-endocrine marker chromogranin A and cytokeratin 19, and not insulin/cytokeratin 19, in order to identify all neofomed endocrine cells as previously described [19, 20]. Further kinetic studies are required, yet it would appear that both proliferation and neogenesis can potentially contribute to the expanded islet mass. Future kinetic studies will combine permanent tagging of human beta cells (or alpha cells) prior to transplantation, as previously described [29], to bring definitive proof of the cell origin responsible for the doubling of the endocrine mass.

In this study, we used the fasting state HOMA of beta cell function (HOMA2) in the mouse as a surrogate to follow modifications in islet function in vivo. Identical at 2 weeks, marked increases were observed in HOMA2%B in both control and HFD animals with time. Only HFD led to a marked decrease in insulin sensitivity (HOMA-S). Classically, a reduction in insulin action in non-diabetic individuals is accompanied by compensatory increases in beta cell secretion

(HOMA2%B) [30], where HOMA2%BS thus remains unaltered. If islets are defective, reduced HOMA2%BS values are observed. In this study the hyperbolic product (HOMA2%BS), which was identical in the two groups at 2 weeks post-transplantation, improved significantly in control animals over time (2 vs 12 weeks), suggesting a hyperfunctional state of the animals that had both a small ectopic human graft (400 IEQ) and a persisting endogenous pancreas. HOMA2%BS did not increase in HFD animals between 2 and 12 weeks, and was four times lower than HOMA2%BS in control animals at 12 weeks. This suggests that although human islets adjust to increased insulin resistance by increasing human C-peptide secretion, normal human islets in an obese environment are defective. This was confirmed by OGTT results, which were also abnormal at 6 and 12 weeks in HFD animals (vs controls).

One limitation of our study is the use of HOMA in our combined human–murine model. Exploitation of this clinical tool in the rodent has been described [31] but remains controversial [16, 32]. Use of HOMA in this study is interesting and specific (human C-peptide), but may be the subject of debate since fasting glycaemia in this model can be regulated by both the human graft and the endogenous pancreas still in place, which also adapts as evidenced by a doubling of the murine islet mass on an HFD. Nevertheless, in our study this variable mainly allowed us to compare the different groups, all of which had a human islet graft from the same donors and endogenous pancreas.

To further validate the model, we had the opportunity to study the adaptation capacity of human islets from two dysfunctional human donors. Despite similar weight gain, animals had higher fasting glycaemia and showed no functional (human C-peptide) or histological adaptation of endocrine graft volume at 12 weeks. The hyperbolic plot, as well as HOMA2%BS, illustrates the lack of adaptation. Human islet grafts from dysfunctional donors were made up of only 10–20% beta cells compared with 75% in normal human islets. Part of the incapacity of dysfunctional islets to adapt may be linked to the age of the donors (>60 years), as has been suggested [28, 33], although other authors have incriminated BMI [27].

In conclusion, we exploited an immunodeficient mouse strain to determine the influence of an obesogenic environment on human islets, and show for the first time direct evidence that non-diabetic human islets adapt both endocrine and beta cell mass, function and gene expression to murine obesity in vivo. This novel model allows, for the first time, longitudinal studies of human islet adaptation to an obese murine environment and may be instrumental in deciphering pathways involved in human beta cell expansion, as well as in helping to identify factors predisposing human beta cells to undergo decompensation [1]. In

addition, the concomitant presence of a mini-human islet graft and the endogenous murine pancreas, both of which doubled in islet/endocrine surface, will allow scientists to study in parallel alterations in murine and human cell expansion at the same time in response to obesity. This will be key to identifying major differences between human and murine pancreatic cells.

Acknowledgements The authors would like to thank the Agence de la Biomedicine and R. Ezzouaoui and V. Raverdy (Centre Hospitalier Régional Universitaire de Lille, Lille, France) for pancreas coordination, and S. Belaich, I. Aluka, E. Moerman, B. Lukowiak and our pancreas harvesting team for their technical contribution (Inserm U859 Biotherapies of Diabetes, Faculty of Medicine, Lille, France). We also thank A. Bouloumié (Inserm U858, Toulouse, France) for mice, and M.H. Gevaert and R.M. Siminski (Laboratory of Histology, Faculty of Medicine, Lille, France) for their technical expertise.

Funding S. Gargani is the recipient of a 3 year PhD stipend from Conseil Regional Nord Pas de Calais/University Hospital of Lille. This work was supported in part by grants from Conseil Regional Nord Pas de Calais to the European Genomics Institute of Diabetes (EGID), Contrat Plan Etat Region, Inserm, European Consortium for Islet Transplantation funded by the Juvenile Diabetes Research Foundation International, Chicago Diabetes Project (J. Oberholzer), Agence de la Biomedicine and core facilities including the Biotherapies Platform provided by or funded by IFR 114 Institut de Medecine Predictive et de Recherche Therapeutique, and the University Hospital of Lille.

Duality of interest The authors declare that there is no duality of interest associated with this manuscript.

Contribution statement JKC, FP, TH, JT and SG conceived and designed the experiments. JT, JEY, SG, VG, ND and JKC acquired data. SG, JKC, JT, JEY, ND, VG and FP analysed the data. JT, SG, JKC, FP and JEY wrote the paper. AD performed statistical analysis, interpretation and writing of the Methods section. BL was involved in the conception, interpretation and drafting of the molecular biology work. JKC, JT, ND, VG, SG, TH, JKC and FP wrote the Methods, and revised the paper. All authors discussed and agreed on the results. JKC supervised the project. All authors gave final approval.

Open Access This article is distributed under the terms of the Creative Commons Attribution License which permits any use, distribution, and reproduction in any medium, provided the original author(s) and the source are credited.

References

- Eckel RH, Kahn SE, Ferrannini E et al (2011) Obesity and type 2 diabetes: what can be unified and what needs to be individualized? *J Clin Endocrinol Metab* 96:1654–1663
- Prentki M, Nolan CJ (2006) Islet beta cell failure in type 2 diabetes. *J Clin Invest* 116:1802–1812
- Sachdeva MM, Stoffers DA (2009) Minireview: meeting the demand for insulin: molecular mechanisms of adaptive postnatal beta-cell mass expansion. *Mol Endocrinol* 23:747–758
- Parnaud G, Bosco D, Berney T et al (2008) Proliferation of sorted human and rat beta cells. *Diabetologia* 51:91–100
- Kahn SE, Hull RL, Utzschneider KM (2006) Mechanisms linking obesity to insulin resistance and type 2 diabetes. *Nature* 444:840–846
- Lee YC, Nielsen JH (2009) Regulation of beta cell replication. *Mol Cell Endocrinol* 297:18–27
- Butler AE, Janson J, Bonner-Weir S, Ritzel R, Rizza RA, Butler PC (2003) Beta-cell deficit and increased beta-cell apoptosis in humans with type 2 diabetes. *Diabetes* 52:102–110
- Pattou F, Kerr-Conte J, Wild D (2010) GLP-1-receptor scanning for imaging of human beta cells transplanted in muscle. *N Engl J Med* 363:1289–1290
- Ricordi C, Scharp DW, Lacy PE (1988) Reversal of diabetes in nude mice after transplantation of fresh and 7-day-cultured (24 degrees C) human pancreatic islets. *Transplantation* 45:994–996
- Caiazzo R, Gmyr V, Kremer B et al (2008) Quantitative in vivo islet potency assay in normoglycemic nude mice correlates with primary graft function after clinical transplantation. *Transplantation* 86:360–363
- Kroon E, Martinson LA, Kadoya K et al (2008) Pancreatic endoderm derived from human embryonic stem cells generates glucose-responsive insulin-secreting cells in vivo. *Nat Biotechnol* 26:443–452
- Vantghem MC, Kerr-Conte J, Arnalsteen L et al (2009) Primary graft function, metabolic control, and graft survival after islet transplantation. *Diabetes Care* 32:1473–1478
- Kerr-Conte J, Vandewalle B, Moerman E et al (2010) Upgrading pretransplant human islet culture technology requires human serum combined with media renewal. *Transplantation* 89:1154–1160
- Duffaut C, Galitzky J, Lafontan M, Bouloumié A (2009) Unexpected trafficking of immune cells within the adipose tissue during the onset of obesity. *Biochem Biophys Res Commun* 384:482–485
- McCall M, Pawlick R, Kin T, Shapiro AM (2012) Anakinra potentiates the protective effects of etanercept in transplantation of marginal mass human islets in immunodeficient mice. *Am J Transplant* 12:322–329
- Lee S, Muniyappa R, Yan X et al (2008) Comparison between surrogate indexes of insulin sensitivity and resistance and hyperinsulinemic euglycemic clamp estimates in mice. *Am J Physiol Endocrinol Metab* 294:E261–E270
- Papademetris X, Shkarin P, Staib LH, Behar KL (2005) Regional whole body fat quantification in mice. *Inf Process Med Imaging* 19:369–380
- Lalloyer F, Vandewalle B, Percevault F et al (2006) Peroxisome proliferator-activated receptor alpha improves pancreatic adaptation to insulin resistance in obese mice and reduces lipotoxicity in human islets. *Diabetes* 55:1605–1613
- Gmyr V, Kerr-Conte J, Belaich S et al (2000) Adult human cytochrome c oxidase-positive cells reexpress insulin promoter factor 1 in vitro: further evidence for pluripotent pancreatic stem cells in humans. *Diabetes* 49:1671–1680
- Bouwens L (1998) Cytokeratins and cell differentiation in the pancreas. *J Pathol* 184:234–239
- Winer S, Chan Y, Paltser G et al (2009) Normalization of obesity-associated insulin resistance through immunotherapy. *Nat Med* 15:921–929
- Granot Z, Swisa A, Magenheim J et al (2009) LKB1 regulates pancreatic beta cell size, polarity, and function. *Cell Metab* 10:296–308
- Tyrberg B, Ustinov J, Otonkoski T, Andersson A (2001) Stimulated endocrine cell proliferation and differentiation in transplanted human pancreatic islets: effects of the ob gene and compensatory growth of the implantation organ. *Diabetes* 50:301–307
- Zahr E, Molano RD, Pileggi A et al (2007) Rapamycin impairs in vivo proliferation of islet beta-cells. *Transplantation* 84:1576–1583

25. Flatt PR, Tan KS, Bailey CJ, Powell CJ, Swanston-Flatt SK, Marks V (1986) Effects of transplantation and resection of a radiation-induced rat insulinoma on glucose homeostasis and the endocrine pancreas. *Br J Cancer* 54:685–692
26. Blume N, Skouv J, Larsson LI, Holst JJ, Madsen OD (1995) Potent inhibitory effects of transplantable rat glucagonomas and insulinomas on the respective endogenous islet cells are associated with pancreatic apoptosis. *J Clin Invest* 96:2227–2235
27. Levitt HE, Cyphert TJ, Pascoe JL et al (2011) Glucose stimulates human beta cell replication in vivo in islets transplanted into NOD-severe combined immunodeficiency (SCID) mice. *Diabetologia* 54:572–582
28. Tian L, Gao J, Weng G et al (2011) Comparison of exendin-4 on beta-cell replication in mouse and human islet grafts. *Transpl Int* 24:856–864
29. Russ HA, Bar Y, Ravassard P, Efrat S (2008) In vitro proliferation of cells derived from adult human beta-cells revealed by cell-lineage tracing. *Diabetes* 57:1575–1583
30. Bergman RN, Ader M, Huecking K, van Citters G (2002) Accurate assessment of beta-cell function: the hyperbolic correction. *Diabetes* 51(Suppl 1):S212–S220
31. Ng SF, Lin RC, Laybutt DR, Barres R, Owens JA, Morris MJ (2010) Chronic high-fat diet in fathers programs beta-cell dysfunction in female rat offspring. *Nature* 467:963–966
32. Wallace TM, Levy JC, Matthews DR (2004) Use and abuse of HOMA modeling. *Diabetes Care* 27:1487–1495
33. Reers C, Erbel S, Esposito I et al (2009) Impaired islet turnover in human donor pancreata with aging. *Eur J Endocrinol* 160:185–191