



## RESEARCH

## Open Access

# Predictive value of the SLC22A18 protein expression in glioblastoma patients receiving temozolomide therapy

Sheng-Hua Chu<sup>1\*</sup>, Yan-Bin Ma<sup>1</sup>, Dong-Fu Feng<sup>1</sup>, Zhi-Qiang Li<sup>2</sup> and Pu-Cha Jiang<sup>2</sup>

## Abstract

**Background:** Our previous study showed that SLC22A18 downregulation and promoter methylation were associated with the development and progression of glioma and the elevated expression of SLC22A18 was found to increase the sensitivity of glioma U251 cells to the anticancer drug 1,3-bis(2-chloroethyl)-1-nitrosourea (BCNU). In this study, we investigated the predictive value of SLC22A18 promoter methylation and protein expression in glioblastoma multiforme (GBM) patients receiving temozolomide (TMZ) therapy.

**Patients and methods:** SLC22A18 promoter methylation and protein expression were examined by methylation-specific polymerase chain reaction (MSP) and Western blotting respectively, then we compared SLC22A18 promoter methylation and protein expression in tumor cell explants in regard to prediction of TMZ response and survival time of 86 GBM patients.

**Results:** SLC22A18 promoter methylation was detected in 61 of 86 (71%) samples, whereas 36 of 86 (42%) cases were scored positive for SLC22A18 protein expression. Overall SLC22A18 promoter methylation was significantly related to SLC22A18 protein expression, but a subgroup of cases did not follow this association. Multivariate Cox regression analysis indicated that SLC22A18 protein expression, but not promoter methylation, was significantly correlated with TMZ therapy. SLC22A18 protein expression predicted a significantly shorter overall survival in 51 patients receiving TMZ therapy, whereas no differences in overall survival were observed in 35 patients without TMZ therapy.

**Conclusions:** These results show that lack of SLC22A18 protein expression is superior to promoter methylation as a predictive tumor biomarker in GBM patients receiving temozolomide therapy.

## Background

Glioblastoma multiforme (GBM) is the most common and lethal glial tumor of the adult brain, accounting for about fifty percent of all gliomas. It is characterized by an aggressive growth pattern, a marked degree of the invasiveness and very poor prognosis. The standard treatment for malignant glioma patients was resection followed by radiotherapy in the past many years. Lately a great lot of researches revealed a statistically significant survival benefit for GBM patients treated with radiotherapy plus temozolomide (TMZ) [1,2]. Consequently, radiotherapy

plus concurrent TMZ therapy currently represents the standard of care for newly diagnosed GBM patients [3].

Solute carrier family 22 (organic cation transporter) member 18 (SLC22A18), also known as IMPT1/BWR1A/TSSC5, is located within the human 11p15.5 cluster [4,5]. Blast homology analysis suggests that SLC22A18 is a member of the family of polyspecific transporters and multidrug resistance genes [5]. More recently, SLC22A18 has been shown to be a tumor suppressor candidate and a substrate for RING105 [6]. Structural mutations in SLC22A18 are rare, with isolated reports of point mutations in a breast cancer cell line [7], a rhabdomyosarcoma cell line [5], and Wilms' tumors and lung tumors [8]. Exonic deletions in Wilms' tumors and loss of heterozygosity in hepatoblastomas have also been reported [9], indicating that SLC22A18 may play a role in tumorigenesis. We have

\* Correspondence: [shenghuachu@126.com](mailto:shenghuachu@126.com)

<sup>1</sup>Department of Neurosurgery, Shanghai 3rd People's Hospital, School of Medicine, Shanghai Jiao Tong University, 280 Mohe Road, Baoshan District, Shanghai 201900, China

Full list of author information is available at the end of the article

previously found that SLC22A18 downregulation and promoter methylation were associated with the development and progression of glioma and SLC22A18 represented a candidate biomarker for long-term survival in this disease, suggesting that SLC22A18 is an important tumor suppressor in glioma [10,11]. We have also found that the elevated expression of SLC22A18 was found to increase the sensitivity of glioma U251 cells to the anticancer drug 1,3-bis(2-chloroethyl)-1-nitrosourea (BCNU) [12].

In this study, for the first time we used primary tumor cell explants obtained from GBM surgical specimens instead of tissue samples to investigate SLC22A18 methylation promoter and protein expression of tumor cell. Our data presented show that expression of SLC22A18 protein has a very strong predictive value for TMZ response and survival time in GBM patients.

## Materials and methods

### Study patients

We collected 86 cases of surgically resected GBMs in the period ranging from 2007–2010 at the Department of Neurosurgery, NO.3 People's Hospital Affiliated to Shanghai Jiao Tong University School of Medicine and Zhongnan Hospital of Wuhan University. Informed patient consent and prior approval from the NO.3 People's Hospital Affiliated to Shanghai Jiao Tong University School of Medicine and Zhongnan Hospital of Wuhan University

Ethics Committees (Ethical approval ZNHWHU0389, NTPH-SHJTUSM046) was obtained before the clinical materials were used for research purposes. All experiments on humans in the present study were performed in compliance with the Helsinki Declaration. Gadolinium-enhanced MRI performed within 1 week after surgery was used to categorize the surgical results according to the removed tumor proportion, i.e., biopsy, ≤ 50%; partial removal, 50-95%; subtotal removal, 96-99%; total removal, >99%. There were 46 men and 40 women with a mean age of 62.5 years (range from 14 to 78 years). None of the patients had received chemical therapy or radiotherapy prior to surgery. 50 patients received concurrent radiotherapy and chemotherapy (60 Gy and daily TMZ at 75 mg/m<sup>2</sup>; 7 days per week over a 42-day period) after surgery. 30 patients received the adjuvant TMZ after concurrent radio-temozolomide therapy. 36 patients did not receive chemotherapy, out of which 18 had only radiotherapy and 18 did not have further therapy. Characteristics of the patients according to SLC22A18 promoter methylation and protein expression status were shown in Table 1.

### Primary tumor cell cultures

After removing visible blood vessel and necrotic tissues, surgery specimens were processed for primary cell cultures as described previously [13-15], and analyzed between passages 2 and 6. During resection patches from non-necrotic

**Table 1 Characteristics of the patients according to SLC22A18 promoter methylation and protein expression status**

Variables	N = 86 No. (%)	SLC22A18 promoter methylation status, No. (%)		P-value	SLC22A18 protein expression, No. (%)		P-value
		Methylated N = 61(71)	Unmethylated N = 25(29)		Positive N = 36(42)	Negative N = 50(58)	
Gender							
Male	46(53)	36(59)	10(40)	0.11	21(58)	25(50)	0.45
Female	40(47)	25(41)	15(60)		15(42)	25(50)	
Age (years)							
<45	13(15)	8(13)	5(20)	0.42	4(11)	9(18)	0.38
≥45	73(85)	53(87)	20(80)		32(89)	41(82)	
Performance status							
<90	39(45)	27(44)	12(48)	0.75	16(44)	23(46)	0.89
≥90	47(55)	34(56)	13(52)		20(56)	27(54)	
Extent of surgery							
Total	45(52)	34(56)	11(44)	0.32	19(67)	26(42)	0.94
Not total	41(48)	27(44)	14(56)		17(33)	24(58)	
MGMT promoter							
Unmethylation	29(34)	18(30)	11(44)	0.20	12(33)	17(34)	0.95
Methylation	57(66)	43(70)	14(56)		24(67)	33(66)	
TMZ therapy							
Yes	50(58)	37(61)	13(52)	0.46	23(64)	27(54)	0.36
No	36(42)	24(39)	12(48)		13(36)	23(46)	

parts of the tumors were gained, disaggregated mechanically and transferred into culture flasks, containing growth medium (RPMI-1640, 20% fetal bovine serum, 1% penicillin/streptomycin, 1% glutamine; Gibco Life Technologies, Paisley, Scotland, UK). Three to five culture flasks were set up from each surgery specimen. Cells were then cultured in RPMI-1640 supplemented with 10% fetal bovine serum and 1% glutamine and without antibiotics. The presence of predominantly malignant cells in primary cell cultures was proved by comparative genomic hybridization and/or fluorescent *in situ* hybridization analyses [13]. All cell cultures were periodically checked for Mycoplasma contamination using a Mycoplasma Stain Kit (Sigma-Aldrich, St. Louis, Missouri, USA) [16].

#### DNA extraction and methylation-specific polymerase chain reaction

As we've previously described [10,17], genomic DNA was extracted from tumor cell cultures or frozen surgery tissues by the digestion with proteinase K (Fisher Scientific, Fairlawn, NJ, USA) using a genomic DNA purification kit (PureGene kit, Gentra Systems, Inc., Minneapolis, MN, USA), and 1 µg genomic DNA was treated with a CpG WIZ DNA Modification Kit (EMD Millipore Corporation, Billerica, MA, USA) to convert unmethylated cytosines to uracil, leaving methylated cytosines unchanged. The modified DNA was diluted in TE buffer (10 mM Tris-Cl, pH 7.5, 1 mM EDTA). SLC22A18 and MGMT promoter methylation analysis was performed by PCR, using bisulfite-treated DNA as template, with specific primers for the methylated (unmodified by bisulfite treatment) and unmethylated (bisulfite modified) gene sequences using the methylation-specific polymerase chain reaction (MSP) method [10,17]. Expression levels were quantified by Quantity One Quantitation software (BioRad) and calculated relative to the controls. The results were confirmed by repeating the bisulfite treatment and MSP assays for all samples.

#### Western blot analysis

As we've previously described [18-20], tumor cell cultures were washed in ice-cold PBS and lysed in buffer using standard methods. Tumor tissues were homogenized in a RIPA lysis buffer. Lysates were cleared by centrifugation (14,000 rpm) at 4°C for 30 minutes. Protein samples (approximately 40 µg) were separated by SDS-PAGE (15% gel), transferred to PVDF membrane and non-specific binding sites blocked by incubation in 5% non-fat milk for 60 minutes. Membranes were incubated overnight at 4°C with polyclonal anti-SLC22A18 antibody (1:1,000 dilution; Santa Cruz, CA). The membrane was then washed three times with TBST for 10 minutes and probed with HRP-conjugated secondary antibody (at 1:2,000 dilution; Dako, Glostrup, Denmark) for 30 minutes at room temperature. Visualization and quantification

were done using the ChemiDoc System (BioRad). Data gained by the densitometric analysis of Western blotting for SLC22A18 were expressed relative (arbitrary units) to the SLC22A18 expressing human neurons (ScienCell Research Laboratories, San Diego, CA, USA) that is included as a positive control in each blot and set arbitrarily as 1. Each assay was performed in triplicate.

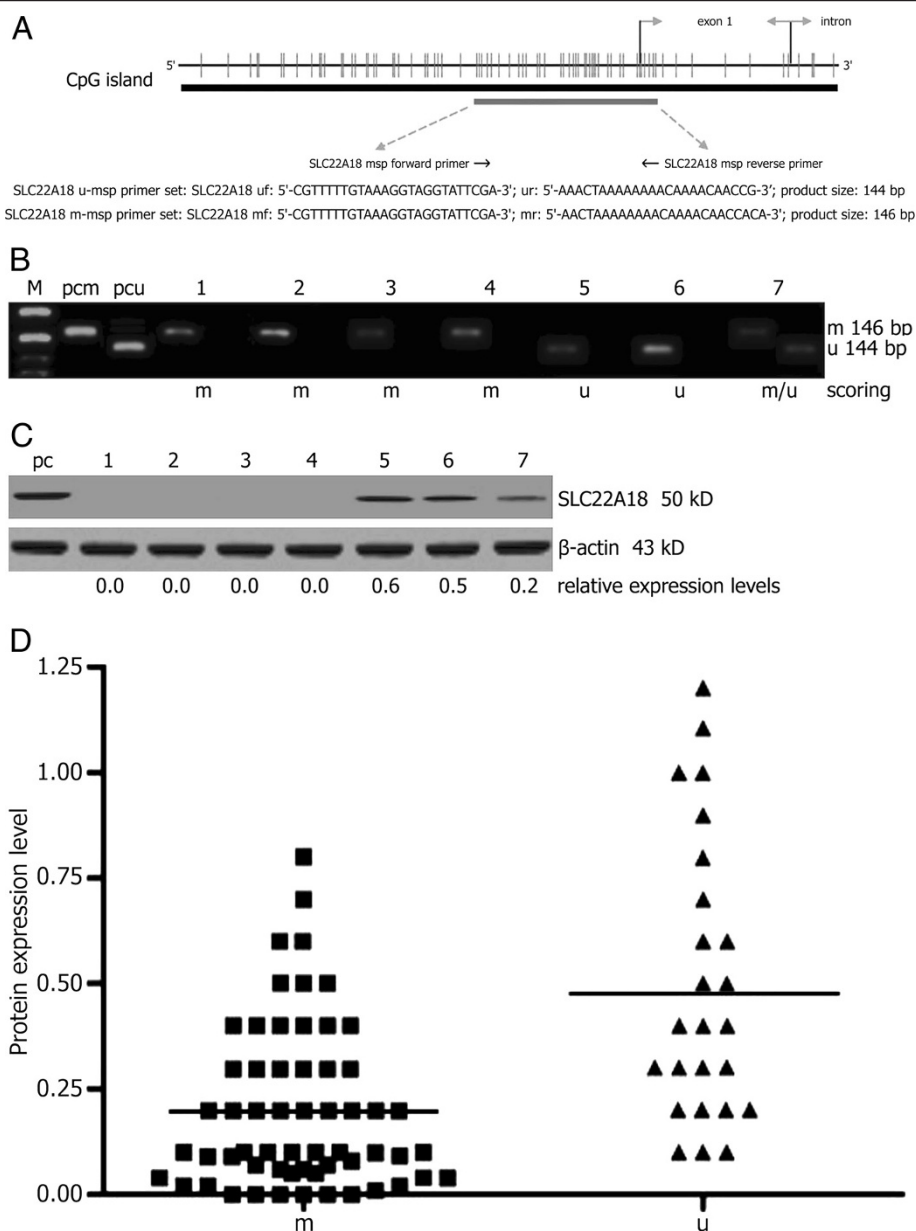
#### Statistical analyses

Statistical analyses and graphs were performed using the Statistical Package for the Social Sciences (version 16.0, for Windows) (SPSS, Chicago, IL, USA). Quantitative values were expressed as mean ± SD. Statistical differences between groups were examined using the Fisher's exact test. *P*-values less than 0.05 were considered statistically significant.

#### Results

##### SLC22A18 promoter methylation and protein expression

Primary tumor cell culture was successful *in vitro* in all surgery specimens, allowing SLC22A18 promoter methylation and protein expression analyses. In 61 of 86 (71%) GBM cases methylated SLC22A18 gene promoter sequences were detected. Out of the 61 samples with methylated sequences, 24 samples contained a mixed profile with varying proportions of unmethylated and methylated sequences, whereas the remaining 37 cases (61%) lacked any unmethylated DNA (Figure 1A-B). SLC22A18 promoter was completely unmethylated in 25 of 86 (29%) GBM cases. To investigate the potential influence of tumor cell culture, SLC22A18 promoter methylation was determined in both surgical tissues and the respective tumor cell cultures in 86 cases. SLC22A18 methylation status agreed well in all patients (see Additional file 1: Table S1), and semi-quantification of MSP products for methylated DNA sequences showed a significant correlation (linear regression analysis,  $P < 0.001$ ). In all cell cultures SLC22A18 protein expressions were successfully analyzed by Western blotting (Figure 1C). 36 of the 86 cases (42%) were scored positive for SLC22A18 protein expression based on a visible band and corresponding to a relative expression level > 0.1. Neither SLC22A18 promoter methylation nor protein expression was significantly correlated with any of the clinical features listed in Table 1. The mean SLC22A18 expression levels were significantly lower in tumor cell cultures harboring methylated SLC22A18 promoter sequences when compared with only unmethylated subgroup ( $P < 0.001$ ) (Figure 1D). Therefore, SLC22A18 protein expression was significantly associated with promoter methylation status when analyzed by 2-sided  $\chi^2$  test ( $P < 0.001$ ). SLC22A18 protein expression levels in the subgroups were strongly overlapping according to promoter methylation despite this association (Figure 1D). In Figure 2 data of 4 representative samples that did not follow the correlation between SLC22A18 protein expression and

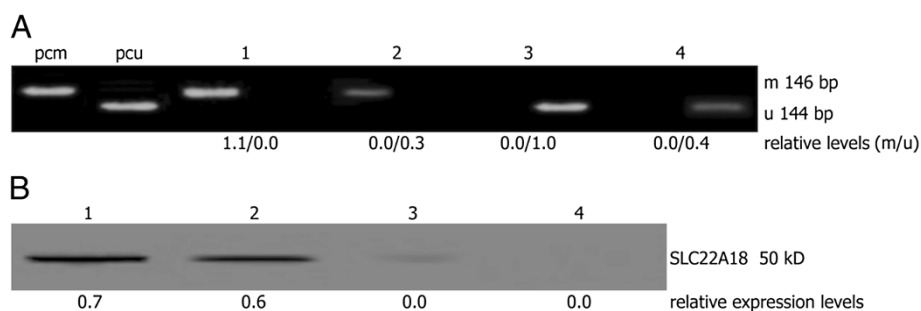


**Figure 1 Analysis of SLC22A18 promoter methylation and protein expression in GBM cell cultures explanted from surgical specimens.**

(A) CpG island of SLC22A18 gene with location and sequence of methylation-specific polymerase chain reaction (MSP) primers (uf, forward U-MSP primer; ur, reverse U-MSP primer; mf, forward M-MSP primer; mr, reverse M-MSP primer). (B) Representative images of MSP analysis of SLC22A18 promoter methylation. Amplification products with specific primers for unmethylated (u) and methylated (m) DNA sequences are indicated, and scoring as unmethylated (u), methylated (m), and mixed (u/m) is shown. Positive controls for unmethylated (pcu) and methylated (pcm) sequences were included. M, size marker. (C) Representative images of western blotting analysis of SLC22A18 protein expression, and levels of relative expression were gained by the densitometric analysis of Western blotting compared with the SLC22A18 expressing human neurons included as positive control (pc) and set arbitrarily as 1. β-actin was used as loading control. (D) Scatter gram analysis of SLC22A18 protein expression in two subgroups with methylated and unmethylated SLC22A18 promoter sequences.

promoter methylation status were shown. These findings included cases with exclusively unmethylated promoter sequences but lack of SLC22A18 protein expression as well as high expression of SLC22A18 protein, despite completely methylated promoter.

**Relationship between SLC22A18 status and overall survival of GBM patients with/without TMZ therapy**  
 58 (67%) of 86 cases had died (34 of 35 cases without TMZ treatment and 24 of 51 cases with TMZ treatment) at a median follow-up time of 27.1 months (95% CI



**Figure 2 Discrepancy between SLC22A18 promoter methylation and protein expression.** Representative samples for SLC22A18 promoter methylation and protein expression (1, 2) as well as lack of promoter methylation and protein expression (3, 4) are shown. **(A)** Polyacrylamide gel showing amplification products of unmethylated and methylated DNA sequences by MSP. Positive controls for unmethylated (pcu) and methylated (pcm) sequences were shown. **(B)** Western blot analysis for the corresponding cases.

25.8-30.2). By univariate analyses the overall survival of GBM patients was compared to SLC22A18 protein expression or promoter methylation status (Table 2). In the univariate analysis lack of SLC22A18 protein expression (hazard ratio [HR] of death 2.14, 95% CI 1.24-3.72;  $P = 0.01$ ), but not promoter methylation status (HR of death 0.64, 95% CI 0.32-1.15;  $P = 0.13$ ), was significantly connected with shorter overall survival in addition to well-known prognostic factors such as age, performance status, extent of surgery, MGMT promoter methylation and TMZ therapy. Besides age, performance status, extent of surgery, MGMT promoter methylation and TMZ therapy, only lack of SLC22A18 protein expression, but not SLC22A18 promoter methylation status, remained as an independent prognostic factor (HR of death 2.11, 95% CI 1.09-3.77;  $P = 0.02$ ) in multivariate Cox regression analysis (Table 3). An interaction term, the product of SLC22A18 parameter and TMZ therapy, was incorporated into the multivariate Cox regression models to evaluate a possible interaction between the SLC22A18 parameters and TMZ treatment. These findings showed a statistically significant association between SLC22A18 protein expression and TMZ treatment ( $P = 0.004$ ), whereas in the case of SLC22A18 promoter methylation this interaction was not significant. Because the interaction term of SLC22A18 protein expression and TMZ therapy was

significant, we studied the relationship between SLC22A18 protein expression and overall survival time in TMZ therapy and no TMZ therapy subgroups (Table 4, Figure 3A and B). In patients with TMZ therapy, SLC22A18 protein expression was significantly associated with longer overall survival time (HR of death 5.65, 95% CI 1.79-17.45;  $P = 0.002$ ). Conversely, no differences in overall survival time according to SLC22A18 protein expression (HR of death 1.12, 95% CI 0.56-2.28;  $P = 0.62$ ) were shown in patients without TMZ therapy. A similar study using the subgroups according to promoter methylation status showed no significant differences with overall survival time in the TMZ therapy subgroup (Figure 3C and D).

## Discussion

We have previously found that SLC22A18 protein expression was significantly decreased in human gliomas compared to the adjacent normal brain tissues. SLC22A18 protein expression was significantly lower in gliomas which recurred within six months after surgery than gliomas which did not recur within six months. Multivariate Cox regression analysis indicated that SLC22A18 expression level was an independent survival prognostic factor for patients with glioma. SLC22A18 promoter methylation was detected in 50% of the gliomas, but not in the adjacent normal tissues of any patient. SLC22A18 expression was significantly decreased in gliomas with SLC22A18 promoter methylation, compared to gliomas without methylation. The SLC22A18 promoter was methylated in U251 cells and treatment with the demethylating agent 5-aza-2-deoxycytidine increased SLC22A18 expression and reduced cell proliferation. Stable overexpression of SLC22A18 inhibited growth and adherence, induced apoptosis *in vitro* and reduced *in vivo* tumor growth of U251 cells [10,11,21]. We have also found that elevated expression of SLC22A18 increased the sensitivity of U251 glioma cells to BCNU [12]. Recently we have found that by increasing the expression of SLC22A18 hydroxyapatite nanoparticles could

**Table 2 Univariate survival analysis**

Variable	HR	95% CI	P value
Gender	0.75	0.38-1.32	0.24
Age	1.12	1.02-1.14	< 0.001
Performance status	0.98	0.92-1.02	< 0.001
Extent of surgery	0.14	0.10-0.28	< 0.001
MGMT methylation	1.25	1.12-1.32	0.006
TMZ therapy	0.24	0.12-0.41	< 0.001
SLC22A18 promoter methylation	0.64	0.32-1.15	0.13
SLC22A18 protein expression	2.14	1.24-3.72	0.01

**Table 3 Multivariate survival analysis**

	Promoter methylation			Protein expression		
	HR	95% CI	P value	HR	95% CI	P value
Variable without interaction term						
Gender	0.83	0.41-1.52	0.44	0.95	0.46-1.76	0.86
Age	1.10	1.01-1.20	0.002	1.10	1.01-1.12	0.002
Performance status	0.98	0.92-1.02	0.001	0.98	0.92-1.02	0.001
Extent of surgery	0.14	0.08-0.22	< 0.001	0.14	0.08-0.24	< 0.001
MGMT promoter	1.32	1.11-1.56	0.012	1.32	1.11-1.55	0.012
TMZ therapy	0.29	0.14-0.50	< 0.001	0.29	0.14-0.49	< 0.001
Promoter methylation	0.62	0.31-1.19	0.15			
Protein expression				2.11	1.09-3.77	0.02
Variable with interaction terms						
Gender	0.83	0.41-1.55	0.46	1.02	0.50-1.87	0.95
Age	1.10	1.01-1.20	0.002	1.10	1.01-1.22	0.001
Performance status	0.98	0.92-1.03	0.002	0.98	0.92-1.02	0.001
Extent of surgery	0.14	0.08-0.22	< 0.001	0.14	0.08-0.34	< 0.001
MGMT promoter	1.32	1.11-1.57	0.018	1.32	1.11-1.55	0.012
TMZ therapy	0.32	0.11-0.83	0.02	0.11	0.03-0.25	< 0.001
Promoter methylation	0.67	0.59-2.91	0.33			
Protein expression				1.06	0.45-2.11	0.45
Promoter methylation × TMZ therapy			0.83			
Protein expression × TMZ therapy						0.004

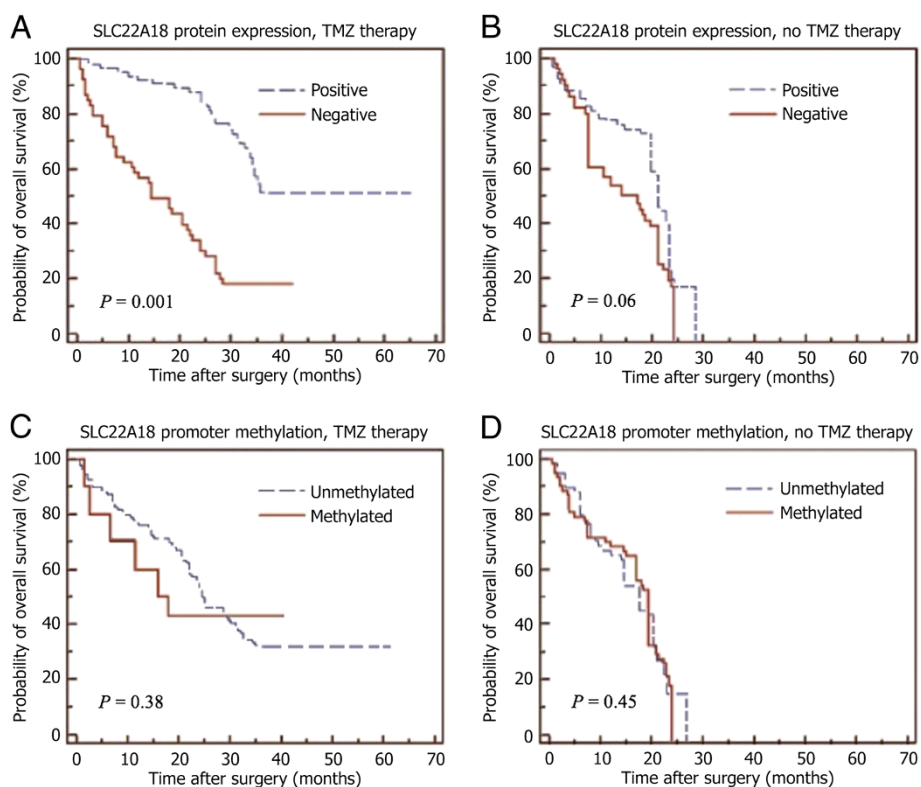
inhibit the growth of human glioma cells *in vitro* and *in vivo* [18].

In the present study, we have studied the relationship between SLC22A18 promoter methylation and protein expression and the association of these two parameters with TMZ treatment as well as overall survival time of GBM patients. We determined SLC22A18 protein expression by Western blot analysis in primary tumor cells derived from the surgical specimens instead of tissue sections or tumor extracts in order to focus completely on the malignant cell compartment. To avoid problems arising from the tissue fixation and paraffin embedding, we used cell culture-derived DNA for detection of SLC22A18 promoter methylation status; and MSP was performed comparably from

tumor cells and snap-frozen tissue in a subgroup of tumors, confirming the presence of methylated SLC22A18 promoter sequences in all cases. During short-term *in vitro* cultivation of tumor cells, the observations also exclude possible changes in the SLC22A18 promoter methylation status. Using this new method, we demonstrate that SLC22A18 protein expression is a strong important predictive marker for the response of TMZ therapy and the related survival benefit. Our results showed SLC22A18 protein expression was associated with longer overall survival time. Especially in patients with TMZ therapy, SLC22A18 protein expression was significantly associated with longer overall survival time, and in patients without TMZ therapy no significant differences in overall survival time according to

**Table 4 Multivariate survival analysis in subgroups of cases regarding TMZ therapy**

Variable	TMZ therapy			No TMZ therapy		
	HR	95% CI	P value	HR	95% CI	P value
Gender	0.45	0.12-1.25	0.08	1.82	0.76-4.06	0.12
Age	1.12	1.01-1.16	0.005	1.10	0.98-1.14	0.03
Performance status	0.97	0.90-1.02	0.006	0.99	0.92-1.04	0.01
Extent of surgery	1.05	0.98-1.12	0.003	1.02	0.96-1.18	0.01
MGMT promoter	5.85	1.86-18.02	0.001	1.25	0.62-2.34	0.74
SLC22A18 Protein expression	5.65	1.79-17.45	0.002	1.12	0.56-2.28	0.62



**Figure 3** SLC22A18 promoter methylation status and protein expression when compared with overall survival time of cases. Results of Kaplan-Meier analyses of overall survival time according to SLC22A18 protein expression (A and B) or promoter methylation (C and D) of GBM patients with TMZ therapy (A and C) and without TMZ therapy (B and D) are shown.

SLC22A18 protein expression were shown. The reasons for the discrepancy of SLC22A18 expression in different tumor patients and a patient subgroup are unclear so far. Thus, future studies with larger sample sizes should be done to confirm this trend. SLC22A18 protein expression was of superior predictive value in GBM patients in our study despite a strong association between SLC22A18 promoter methylation and protein expression. These findings show that reliable SLC22A18 protein detection approach has to be developed to identify GBM patients likely to benefit from TMZ treatment. Consistent with this idea [10], the SLC22A18 parameters showed significant correlation in this study, confirming that SLC22A18 promoter methylation is a major factor regulating SLC22A18 protein expression. However, as obvious from the scatter grams (Figure 1D) and depicted by selected patients (Figure 2), there are some tumor cell samples not following this association. This shows that SLC22A18 protein is still expressed in a subgroup of tumors with SLC22A18 promoter methylation and vice versa. The reasons for the discrepancy between SLC22A18 expression and promoter methylation in a patient subgroup are unclear so far. Our results show that epigenetic silencing via promoter methylation is an important but obviously not always decisive factor regulating

SLC22A18 expression in GBM cells. With respect to therapy response and overall survival, SLC22A18 protein expression proved to be of significantly higher predictive power when compared with promoter methylation status in our samples. Our attempts to analyze SLC22A18 expression by immunohistochemistry in selected patient samples used in this study were inconclusive, and immunostaining neither correlated with SLC22A18 expression nor promoter methylation detected by Western blot, even when using identical antibodies (data not shown). Our data based on tumor cell explants and Western blot analysis turn out the high quality of SLC22A18 expression as a predictive marker for TMZ response in patients with GBM.

## Conclusions

Collectively, we analyzed the influence of SLC22A18 status on the response of TMZ treatment and survival time of GBM patients by using a completely novel approach. Evaluation of SLC22A18 protein levels in GBM cell cultures derived from surgical specimens was proved to be a strongly predictive indicator for TMZ therapy-mediated survival time benefit with superior information power as compared to the MSP analysis of SLC22A18 promoter methylation. Accordingly, reproducible and reliable SLC22A18 protein

detection methods have to be developed and tested with respect to their predictive value for the response of TMZ treatment in prospective studies involving more GBM patients.

## Additional file

**Additional file 1: Table S1.** SLC22A18 promoter methylation of DNA extracted from primary GBM cell cultures and surgical tissues.

## Abbreviations

SLC22A18: Solute carrier family 22 (organic cation transporter) member 18; RPMI: Roswell park memorial institute; PBS: Phosphate-buffered saline; SDS: Sodium dodecyl sulfate; MSP: Methylation-specific polymerase chain reaction.

## Competing interests

The authors declare that they have no competing interests.

## Authors' contributions

SHC, ZQL and PCJ carried out the laboratory analysis. SHC, YBM and DFF participated in the design of the study and drafted the manuscript. SHC, ZQL and PCJ conceived of the study, and participated in its design and coordination and helped to draft the manuscript. All authors read and approved the final manuscript.

## Acknowledgements

This work was supported by grants from the Natural Science Foundation of China (30901535), Innovation Program of Shanghai Municipal Education Commission (12YZ046), Research Fund of Xinhua Hospital Affiliated to Shanghai Jiao Tong University School of Medicine (10XJ22010) and the New One Hundred Person Project of Shanghai Jiao Tong University of School of Medicine (10XBR01). The authors would like to thank Department of Epidemiology and Health Statistics, Wuhan University for providing help.

## Author details

<sup>1</sup>Department of Neurosurgery, Shanghai 3rd People's Hospital, School of Medicine, Shanghai Jiao Tong University, 280 Mohe Road, Baoshan District, Shanghai 201900, China. <sup>2</sup>Department of Neurosurgery, Zhongnan Hospital of Wuhan University, Wuhan 430071, China.

Received: 1 October 2012 Accepted: 13 March 2013

Published: 20 March 2013

## References

1. Stupp R, Mason WP, van den Bent MJ, Weller M, Fisher B, Taphoorn MJ, Belanger K, Brandes AA, Marosi C, Bogdahn U, Curschmann J, Janzer RC, Ludwin SK, Gorlia T, Allgeier A, Lacombe D, Cairncross JG, Eisenhauer E, Mirimanoff RO: European organisation for research and treatment of cancer brain tumor and radiotherapy groups; national cancer institute of Canada clinical trials group: radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *N Engl J Med* 2005, **352**:987–996.
2. Stupp R, Hegi ME, Gilbert MR, Chakravarti A: Chemoradiotherapy in malignant glioma: standard of care and future directions. *J Clin Oncol* 2007, **25**:4127–4136.
3. Grossman SA, Ye X, Piantadosi S, Desideri S, Nabors LB, Rosenfeld M, Fisher J: NABTT CNS consortium: survival of patients with newly diagnosed glioblastoma treated with radiation and temozolomide in research studies in the United States. *Clin Cancer Res* 2010, **16**:2443–2449.
4. Lee MP, Reeves C, Schmitt A, Su K, Connors TD, Hu RJ, Brandenburg S, Lee MJ, Miller G, Feinberg AP: Somatic mutation of TSSCS, a novel imprinted gene from human chromosome 11p15.5. *Cancer Res* 1998, **58**:4155–4159.
5. Schwienbacher C, Sabbioni S, Campi M, Veronese A, Bernardi G, Menegatti A, Hatada I, Mukai T, Ohashi H, Barbanti-Brodano G, Croce CM, Negrini M: Transcriptional map of 170-kb region at chromosome 11p15.5: identification and mutational analysis of the BWR1A gene reveals the presence of mutations in tumor samples. *Proc Natl Acad Sci USA* 1998, **95**:3873–3878.
6. Yamada HY, Gorbysky GJ: Tumor suppressor candidate TSSC5 is regulated by Ubch6 and a novel ubiquitin ligase RING105. *Oncogene* 2006, **25**:1330–1339.
7. Gallagher E, Mc Goldrick A, Chung WY, Mc Cormack O, Harrison M, Kerin M, Dervan PA, Mc Cann A: Gain of imprinting of SLC22A18 sense and antisense transcripts in human breast cancer. *Genomics* 2006, **88**:12–17.
8. Schwienbacher C, Angioni A, Scelfo R, Veronese A, Calin GA, Massazza G, Hatada I, Barbanti-Brodano G, Negrini M: Abnormal RNA expression of 11p15 imprinted genes and kidney developmental genes in Wilms' tumor. *Cancer Res* 2000, **60**:1521–1525.
9. Albrecht S, Hartmann W, Houshdaran F, Koch A, Gärtner B, Prawitt D, Zabel BU, Russo P, Von Schweinitz D, Pietsch T: Allelic loss but absence of mutations in the polyspecific transporter gene BWR1A on 11p15.5 in hepatoblastoma. *Int J Cancer* 2004, **111**:627–632.
10. Chu SH, Feng DF, Ma YB, Zhang H, Zhu ZA, Li ZQ, Jiang PC: Promoter methylation and downregulation of SLC22A18 are associated with the development and progression of human glioma. *J Transl Med* 2011, **9**:156.
11. Chu SH, Ma YB, Feng DF, Zhang H, Zhu ZA, Li ZQ, Jiang PC: Correlation of low SLC22A18 expression with poor prognosis in patients with glioma. *J Clin Neurosci* 2012, **19**:95–98.
12. Chu SH, Ma YB, Feng DF, Zhang H, Qiu JH, Zhu ZA: Elevated expression of solute carrier family 22 member 18 increases the sensitivity of U251 glioma cells to BCNU. *Oncol Lett* 2011, **2**:1139–1142.
13. Spiegl-Kreinecker S, Pirker C, Filipits M, Lötsch D, Buchroithner J, Pichler J, Silye R, Weis S, Micksche M, Fischer J, Berger W: O6-Methylguanine DNA methyltransferase protein expression in tumor cells predicts outcome of temozolomide therapy in glioblastoma patients. *Neuro Oncol* 2010, **12**:28–36.
14. Spiegl-Kreinecker S, Pirker C, Marosi C, Buchroithner J, Pichler J, Silye R, Fischer J, Micksche M, Berger W: Dynamics of chemosensitivity and chromosomal instability in recurrent glioblastoma. *Br J Cancer* 2007, **96**:960–969.
15. Peigñan L, Garrido W, Segura R, Melo R, Rojas D, Cárcamo JG, San Martín R, Quezada C: Combined use of anticancer drugs and an inhibitor of multiple drug resistance-associated protein-1 increases sensitivity and decreases survival of glioblastoma multiforme cells *in vitro*. *Neurochem Res* 2011, **36**:1397–1406.
16. Lu DY, Yeh WL, Huang SM, Tang CH, Lin HY, Chou SJ: Osteopontin increases heme oxygenase-1 expression and subsequently induces cell migration and invasion in glioma cells. *Neuro Oncol* 2012, **14**:1367–1378.
17. Chu SH, Ma YB, Feng DF, Zhang H, Zhu ZA, Li ZQ, Jiang PC: Upregulation of SATB1 is associated with the development and progression of glioma. *J Transl Med* 2012, **10**:149.
18. Chu SH, Feng DF, Ma YB, Li ZQ: Hydroxyapatite nanoparticles inhibit the growth of human glioma cells *in vitro* and *in vivo*. *Int J Nanomed* 2012, **7**:3659–3666.
19. Chu SH, Feng DF, Zhang H, Chen ET, Duan ZX, Li XY, Li J, Ma YB, Zhu ZA, Qiu JH: c-Met-targeted RNA interference inhibits growth and metastasis of glioma U251 cells *in vitro*. *J Neurooncol* 2009, **93**:183–189.
20. Chu SH, Ma YB, Feng DF, Zhang H, Qiu JH, Zhu ZA: c-Met antisense oligodeoxynucleotides increase sensitivity of human glioma cells to paclitaxel. *Oncol Rep* 2010, **24**:189–194.
21. Chu SH, Ma YB, Feng DF, Zhang H, Qiu JH, Zhu ZA: Effect of 5-Aza-2'-deoxycytidine on SLC22A18 in glioma U251 cells. *Mol Med Report* 2012, **5**:138–141.

doi:10.1186/1479-5876-11-69

**Cite this article as:** Chu et al.: Predictive value of the SLC22A18 protein expression in glioblastoma patients receiving temozolomide therapy. *Journal of Translational Medicine* 2013 **11**:69.