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Females tend to prefer genetically similar mates in an island population of house sparrows

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Abstract

Background: It is often proposed that females should select genetically dissimilar mates to maximize offspring genetic diversity and avoid inbreeding. Several recent studies have provided mixed evidence, however, and in some instances females seem to prefer genetically similar males. A preference for genetically similar mates can be adaptive if outbreeding depression is more harmful than inbreeding depression or if females gain inclusive fitness benefits by mating with close kin. Here, we investigated genetic compatibility and mating patterns in an insular population of house sparrow (*Passer domesticus*), over a three-year period, using 12 microsatellite markers and one major histocompatibility complex (MHC) class I gene. Given the small population size and the distance from the mainland, we expected a reduced gene flow in this insular population and we predicted that females would show mating preferences for genetically dissimilar mates.

Results: Contrary to our expectation, we found that offspring were less genetically diverse (multi-locus heterozygosity) than expected under a random mating, suggesting that females tended to mate with genetically similar males. We found high levels of extra-pair paternity, and offspring sired by extra-pair males had a better fledging success than those sired by the social male. Again, unexpectedly, females tended to be more closely related to extra-pair mates than to their social mates. Our results did not depend on the type of genetic marker used, since microsatellites and MHC genes provided similar results, and we found only little evidence for MHC-dependent mating patterns.

Conclusions: These results are in agreement with the idea that mating with genetically similar mates can either avoid the disruption of co-adapted genes or confer a benefit in terms of kin selection.

Keywords: Sexual selection, Mate choice, *Passer domesticus*, Major Histocompatibility Complex (MHC), Microsatellites, Extra-pair paternity

Background

The adaptive function of mate choice remains unclear and one of the most challenging problems in behavioural ecology [1-3]. Choosy females can potentially obtain direct or indirect genetic benefits for their progeny (genetic quality or compatibility) [3-5]. Females that avoid mating with related or genetically similar mates avoid the costs of inbreeding depression [6-10]. Inbreeding avoidance may also increase the genetic diversity of progeny, which might confer fitness benefits in temporally and spatially heterogeneous environments. Yet, several recent studies

have surprisingly reported evidence for mate choice for *genetically related* reproductive partners [11-13]. It is unclear whether or how inbreeding *per se* could be beneficial, though such findings may be due to outbreeding avoidance. If local environmental conditions selectively favour co-adapted ensembles of genes, mating with genetically distant partners could disrupt these assemblages and result in a loss of fitness (outbreeding depression) [14-17]. Bateson [18,19] suggested that maximal reproductive success may be achieved by pairs with intermediate genetic relatedness, and there is support for his 'optimal outbreeding' hypothesis [20-22]. Mating preferences for kin could also allow females to increase their inclusive fitness [23-25] in the absence of significant inbreeding depression [18,26,27]. Similarly, selecting kin as social

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mates may improve cooperation between the sexes and reduce sexual conflict over parental investment [28].

The rapid development of molecular genetic tools in recent years has considerably aided mate choice research in several ways. First, although the vast majority of bird species are socially monogamous, many species also engage in extra-pair copulations so that broods are usually composed of chicks sired by different fathers [29-32]. Genetic paternity analyses allow the detection of extra-pair (EP) paternity and identification of EP males and EP offspring. Moreover, extra-pair mating presents an opportunity to examine the processes governing mate choice in the absence of any potential direct benefit since the extra-pair male does not contribute to parental care. According to the inbreeding avoidance hypothesis, for example, females mated with closely related males should engage in extra-pair copulations with genetically dissimilar mates [3], whereas the outbreeding avoidance and the kin selection hypotheses predict that females should engage in extra-pair copulations with genetically similar males [8,33]. Second, molecular genetic tools make it possible to infer individuals' genome-wide diversity and relatedness between partners using both neutral loci (e.g. microsatellites) and functional selected genes [1-5]. Third, molecular tools have helped to test the hypothesis that genetic benefits from mate choice include increasing offspring heterozygosity at the major histocompatibility complex (MHC) loci [34-39]. MHC genes encode cell-surface glycoproteins that control antigen presentation, and MHC heterozygotes are supposed to better face infectious diseases [40,41]. Yet, just as different species show inbreeding or outbreeding preferences, recent work indicates the MHC-dependent mating preferences can be disassortative or assortative for alleles, disassortative for allelic diversity, or for specific alleles. It has been suggested that 'if there is an optimal level of MHC heterozygosity for combating infections, then females should prefer to mate with males that have intermediate levels of MHC dissimilarity' [35], and subsequent work has provide evidence that female preferences depend on males' individual allelic diversity [39,42,43].

We studied mate choice both for social and extra-pair mates in a small insular population of the house sparrow (*Passer domesticus*), using microsatellite loci and MHC

class I genes, over three consecutive years. Mating patterns were assessed by examining offspring genetic diversity. This indirect assessment only allowed us to infer realized mate choice, whereas assessing female preference would have required letting females choose a partner in the absence of constraints [44]. Given the small population size and the isolated nature of the studied population, we expected reduced gene flow. This led us to predict that females should preferentially mate with diverse and genetically dissimilar males to i) reduce the risk of inbreeding; ii) enhance genetic diversity of their progeny.

Methods

The study population

The house sparrow population studied here is located at Hoëdic, a small (2.08 km²) island off the French coast of Brittany (47°20'24.40"N-2°52'43.09"W). Adult house sparrows were captured using mist nets and banded with a metal ring and a unique combination of coloured rings which allowed individual recognition. At the first capture, we obtained a small amount of blood (20 µl) by brachial vein puncture and stored it in 500 µl of Queen's Lysis Buffer (QLB) [45]. We monitored pairs breeding in nest boxes that were set up in the village, from 2009 to 2011. For logistic reasons, we were only able to monitor the first two broods during each year, even though house sparrows can lay up to 3-4 clutches per breeding season. Between late April and the end of June, we visited nest boxes at least twice per week and recorded clutch size and the number of hatched and fledged chicks. When chicks were 8 days old, they were banded with a metal ring and a drop of blood was collected and stored as for adults. The identities of social parents were assessed during focal observations when adults were brooding or feeding the chicks. Sample sizes are summarized in Table 1. Ringing licence and permit to take blood samples were given by the Muséum National d'Histoire Naturelle (Paris) and the Préfecture du Morbihan.

We estimated population size using the POPAN module of the software MARK [46]. POPAN gives an estimate of the population size while taking into account the probabilities of recapture (p) and survival (ϕ), as well as the probability of new individuals entering the population

Table 1 Total number of individuals, breeding pairs, and individuals genotyped

| Year | Number of sampled individuals | Number of breeding pairs with known identity | Number of microsatellite genotyped individuals (number of chicks) | Number of MHC genotyped individuals (number of chicks) |
|--------|-------------------------------|--|---|--|
| 2009 | 225 | 15 | 222 (51) | 190 (51) |
| 2010 | 341 | 40 | 335 (82) | 294 (82) |
| 2011 | 316 | 49 | 311 (89) | 275 (89) |
| Total* | 574 | 96 | 565 (222) | 494 (222) |

*Total of unique individuals.

(p_{ent}). We ran models where each of the three parameters was either constant or varied as a function of capture session (time) (three sessions per year, three years = nine capture sessions). The best model was selected based on the AIC criterion and was the model where the three parameters varied with time. We also used the U-CARE module to check any violation of the assumptions underlying the use of capture-mark-recapture models and did not find any departure from these assumptions [47].

Microsatellite genotyping

DNA was extracted using the Wizard® SV 96 Genomic DNA Purification kit (Promega) according to the manufacturer's instructions. All individuals were genotyped using the following twelve microsatellite loci: PdomD09, PdomA08, PdomB01, PdomH05 [48], Mjg1 [49], Ase18 [50], Pdo3, Pdo5 [51], Pdo1 [52], Pdo10 [53], Pdo16 and Pdo27 [54]. Polymerase chain reactions (PCRs) were performed in a final volume of 10 μ l including 10 to 50 ng of DNA, 2 μ l of 5X buffer, 1.5 to 2 mM of MgCl_2 , 400 μ l of dNTPs, 1 μ M of each primers and 0,2 U of *Taq* DNA polymerase (Promega). The PCR program comprised: 94°C 3 to 4 min, 30 to 35 cycles of 94°C 20s, 20s for annealing (48°C to 62°C according to the different loci), and 72°C 30 to 40s, followed by a final extension of 72°C 5 to 7 min. Samples were then run in an ABI3730 automated sequencer. Allele sizes were determined using GeneMapper v4.0.

MHC class I genotyping

We amplified the MHC class I exon 3, which corresponds to the highly variable peptide-binding region (PBR) of the protein [55]. Passerines have been shown to have several loci at the MHC class I exon 3 due to gene duplication and fragmentation, which makes it impossible to determine the number of amplified loci or estimate heterozygosity at each locus [42,55-58].

PCR amplifications were performed using a fluorescent (6'FAM) labelled primer (A23M – GCG CTC CAG CTC CTT CTG CCC ATA) and an unlabeled primer (A21M – GTA CAG CGC CTT GTT GGC TGT GA). PCRs were performed in a final volume of 10 μ l, including 50 to 100 ng of genomic DNA, 0,6 μ M of each primer and 5 μ l of Multiplex PCR reagent (QIAGEN GmbH) containing hot-start DNA polymerase, buffer and dNTPs. The PCR program began with 5min initial denaturation at 95°C, followed by 35 cycles of 30s denaturation at 94°C, 90s annealing at 56°C and 90s extension at 72°C. A final elongation step was run for 10 min at 72°C. To control for PCR artefacts, we used 2 negative controls, for PCR and for sequencer, by adding purified water instead of DNA or PCR products. MHC diversity was screened using capillary electrophoresis single conformation polymorphism (CE-SSCP) [43]. PCR samples were prepared

for electrophoresis by combining 1 μ l PCR product, with 8.75 μ l Hi-Di formamide and 0.25 μ l of in-house prepared ROX size standard [59]. This mix was heated for 5min at 95°C to separate the complementary DNA strands. Electrophoresis was conducted in an automated DNA sequencer (ABI PRISM 3130 xl automated DNA Sequencer, Applied Biosystems). The retention time of allelic variants was assessed relative to the ROX size standard.

Statistical analyses

Estimations of within-individual genetic diversity and between-individual dissimilarity

Genome-wide inbreeding was assessed with individual Internal Relatedness (IR, [60]). IR corresponds to the number of homozygous loci, divided by the number of genotyped loci, weighted by the allele frequencies. To measure the genetic similarity between paired males and females, we also computed unbiased pairwise relatedness (r , [61]), where each locus is weighted using the method described in [62,63]. IR and r were assessed from multi-locus microsatellite genotypes.

Allele-sharing was calculated to estimate MHC similarity between males and females forming a pair-bond. Allele-sharing is twice the number of shared alleles divided by the number of different alleles of each individual [$D = 2F_{\text{ab}}/(F_{\text{a}} + F_{\text{b}})$] [42,64].

Paternity analysis

In each nest, social parents were identified during the brooding and chick feeding period. To assess extra-pair paternity, we used the likelihood-based approach implemented in the software CERVUS 3.0 [65]. The software allows excluding and assigning putative fathers based on their multi locus genotypes. The probability of exclusion and assignment was fixed to 95%. We also tested if there was any mismatch between the maternal identity based on the field observations and the one based on microsatellites. Maternal mismatches would indicate that brood parasitism had occurred. Over the entire study period, in only two instances we found a mismatch between the maternal identity based on the field observations and the genetic markers. However, these mismatches involved complete clutches which likely reflect errors in the reading of the color bands rather than brood parasitism. Accordingly these two records were excluded from the statistical analyses. We considered a chick as being extra-pair if it was sired by a male other than the social male that was identified by field observations. We also tested whether there was any evidence suggesting a departure from Hardy-Weinberg equilibrium of the microsatellite loci using CERVUS 3.0 [65]. We did not find any departure from Hardy-Weinberg equilibrium (all $p < 0.05$; Table 2).

Table 2 Number of alleles (k), number of genotyped adults (N), observed heterozygosity (Hobs), expected heterozygosity (Hexp) and deviation from Hardy-Weinberg equilibrium (HW) for each microsatellite locus over the three study years (NS = non significant)

| Locus | k | N | Hobs | Hexp | HW |
|---------|----|-----|------|------|----|
| PdomD09 | 8 | 343 | 0.73 | 0.76 | NS |
| PdomA08 | 12 | 335 | 0.72 | 0.80 | NS |
| PdomB01 | 9 | 343 | 0.62 | 0.61 | NS |
| PdomH05 | 16 | 343 | 0.70 | 0.71 | NS |
| Mjg1 | 21 | 343 | 0.90 | 0.92 | NS |
| Ase18 | 15 | 343 | 0.86 | 0.88 | NS |
| Pdo3 | 16 | 343 | 0.91 | 0.89 | NS |
| Pdo5 | 18 | 343 | 0.83 | 0.84 | NS |
| Pdo1 | 15 | 334 | 0.84 | 0.86 | NS |
| Pdo10 | 13 | 337 | 0.87 | 0.88 | NS |
| Pdo16 | 13 | 343 | 0.81 | 0.82 | NS |
| Pdo27 | 14 | 322 | 0.84 | 0.84 | NS |

Mate choice

The null hypothesis of random mating with respect to parental relatedness was tested by comparing observed chick IR values to expected distributions under this null hypothesis. We computed observed IRs for i) the overall sample of chicks produced over the three-year study period ($n = 222$), ii) chicks produced by social males ($n = 111$), and iii) chicks produced by extra-pair males ($n = 65$). Over the entire sample of 222 chicks, paternal identity could not be assigned for 46 chicks. We also compared the relatedness (r) between pair members (between the female and her social mate, and her genetic mate when she engaged in EP copulations) to the expected distribution of values under random mating. Expected values were generated using the software STORM [66] by randomly sampling (1000 iterations for each year) reproductive males and females that were observed in a given year in order to generate the same number of chicks and mating pairs to the observed ones in the same year.

MHC allele-sharing (D , between the female and her social mate, and her genetic mate when she engaged in EP copulations) was compared with the distribution of expected D values obtained by randomly generating mating pairs (1000 bootstraps for each year, R version 2.15.0, R Development Core Team 2011). As for IR and r , D was computed using social and extra-pair partners.

Hypothesis testing

We used General Linear Mixed Models (GLMMs) to test the hypothesis that females mated with more closely related social males would also engage in extra-pair fertilization. Brood type was modeled as a binary response variable (with or without extra-pair chicks). The

explanatory (fixed) variables were social male IR, relatedness to the female (r), band-sharing within the male-female pair, and year. Since some females laid several clutches during the three-year period covered by the study, female identity nested within year was set as a random factor.

We also compared the hatching and fledging success of broods with no extra-pair chicks to the hatching and fledging success of broods containing at least one extra-pair young. Here, fledging success was entered as a binomial response variable (fledged or not), year and brood type (broods with extra-pair or no extra-pair young) were also included as fixed factors and female identity was nested within year as a random factor.

Internal Relatedness (IR) of chicks produced by social males was compared to IR of chicks produced by extra-pair males using a GLMM with a binomial distribution of errors. Year, chick type (sired by the social or the extra-pair male), and their interactions were added as fixed factors. Female identity was nested within year and entered as a random factor.

We also compared IR, r and D between social and extra-pair males, for the restricted sample of females that engaged in extra-pair copulations, with the aim of investigating if male genetic characteristics and the relatedness with the female affected the likelihood of being a social or an extra-pair male. We constructed a GLMM where male status (social or extra-pair) was entered as a binomial response variable. Male IR, D , r and year were included as fixed factors. Breeding event nested within female and within year was also declared as a random factor. When a female mated with several extra-pair mates during a single reproductive event, we computed the mean IR, r and D and used these values in the statistical models.

We used a similar GLMM to test if within-brood chicks sired by the social or the extra-pair male(s) differed in their fledging success. Fledging success corresponds to the number of fledged chicks sired by a given male, divided by the total number of eggs laid and this was modeled as a binomial response variable. Male type (social or extra-pair), year and their interaction were added as fixed factors. Breeding event, nested within female identity and year was added as a random factor.

We used the package lme4 [67], implemented in R 2.15.0 to run all GLMMs. We used the information-theoretic (IT) approach to perform model selection [68]. Model support was assessed using the corrected version of Akaike Information Criterion (AICc) for small sample sizes, and Δ AIC was used to infer support for models in the candidate set [69]. Δ AIC corresponds to the difference in AICc of the focus model minus the AICc of the best model (the model with the lowest AIC) [68]. We calculated the Akaike weights (ω) for each model, which

is the probability that a model is selected as the best in a model set [68]. Using the package MuMIn [70], we also calculated the summed AIC weight ($\Sigma AIC\omega$) for each variable. This corresponds to the sum of the weights of the models in which the variable is present and can be interpreted as the probability that a given variable is retained in the selected model [68,71]. Following Burnham and Anderson [68], we considered that a model had substantial empirical support if its ΔAIC was lower than 2.

Results and discussion

The number of alleles for each microsatellite locus varied between 8 and 21, whereas observed heterozygosity varied between 0.70 and 0.91 (Table 2). Individuals had a mean of 21.50 (± 1.05 SE) microsatellite alleles over 12 loci, and a mean of 2.31 (± 0.05 SE) MHC class I alleles. The individual MHC allele number varied between 1 and 6 (from as many as 3 class I loci amplified) and a total of 37 MHC alleles were found in the entire population.

Over the three study years the average population size of adult birds was 204 individuals (± 21 SE). Therefore, we sampled a substantial fraction of the total estimated breeding population each year.

In 2009, 2010 and 2011, respectively 35.3, 40.2 and 48.3% of chicks were from extra-pair matings. Similarly, 64.3, 57.7 and 69.0% of broods contained at least one extra-pair young, respectively.

The observed IR computed over the entire sample of chicks (IRobs) was significantly higher than expected under random mate choice (mean = 0.037 \pm 0.001 SE; $n = 222$; $p = 0.020$; Figure 1A). The results were similar when IR was computed on within-pair only (IRobs_{within-pair} = 0.032 \pm 0.014 SE; $p = 0.046$; $n = 112$), or extra-pair only chicks (IRobs_{extra-pair} = 0.044 \pm 0.020 SE; $p = 0.009$; $n = 53$) (Figure 1A).

Mean pairwise relatedness (robs_{social}) between females and their social mates was -0.008 (± 0.016 SE; $n = 96$) and did not differ from expected values under a random mate choice (Figure 1B). Mean pairwise relatedness (robs_{genetic}) between females and their genetic mates was 0.011 (± 0.018 SE; $n = 79$) and did not differ from expected values under random mate choice (Figure 1B).

MHC allele-sharing (Dobs_{social}) between females and their social mates and allele-sharing (Dobs_{genetic}) between females and their extra-pair mates were respectively 0.263 (± 0.024 SE; $n = 56$) and 0.333 (± 0.045 SE; $n = 75$), and did not differ from expected values under random mate choice (Figure 1C).

None of the measures of relatedness between the female and her social mate (r), and inbreeding level of the male (IR) affected the female likelihood to engage in extra-pair fertilizations, since the model with the lowest AIC value was the null model (Table 3, $n = 70$). Similarly, none of the variables improved the fit of the model

exploring the variation in hatching success (the null model had the lowest AIC and the highest ω (Table 4, $n = 61$). However, fledging success did vary among years (0.89 in 2009, 0.75 in 2010, and 0.59 in 2011) and the model including year had the lowest AIC and the highest ω values (Table 4, $n = 53$). IR of chicks was not influenced by mating type (social or extra-pair) but tended to vary across years. However, we should note that the null model is very close suggesting that the effect of year is rather weak (Table 5, $n = 222$).

We then focused on broods that contained both within and extra-pair chicks to compare the genetic characteristics (IR, r and D) between social and extra-pair mates. The best model was the one that included pairwise relatedness (r , Table 6, $n = 60$). The $\Delta AICc$ and $\Sigma AIC\omega$ of the other competitive models also suggested a possible role for MHC band sharing and pairwise relatedness (Table 6). The mean pairwise relatedness was 0.074 (± 0.029 SE, $n = 30$) between females and extra-pair mates whereas it was 0.008 (± 0.029 SE, $n = 30$) between females and social mates. MHC allele-sharing was 0.34 (± 0.050 SE, $n = 30$) between females and extra-pair mates, and 0.24 (± 0.047 SE, $n = 30$) between females and social mates. We should however mention that the null model is also competitive suggesting that the contribution of these variables is weak.

Fledging success of chicks sired by extra-pair males (0.41 \pm 0.047 SE, $n = 30$) was higher than for chicks sired by the social mate (0.20 \pm 0.044 SE, $n = 30$). The model including sire type (social vs. extra-pair) had the lowest AIC (Table 7, $n = 60$), and sire type had a very high $\Sigma AIC\omega$ (0.99).

We expected that the particular demographic and ecological characteristics of the studied insular population of house sparrows would have promoted the evolution of mating preference for dissimilar mates to avoid the depletion of genetic diversity and inbreeding [6-13,72]; however, our results do not provide support for the "inbreeding avoidance" hypothesis. On the contrary, we found evidence suggesting a preference for genetically similar mates. Over the three years covered by the study, we found that offspring were less heterozygous than expected under random mate choice (based on 12 microsatellite loci), though there was little support that MHC class I genes influenced pair formation. This pattern was consistent even when taking into account the relatively high proportion of extra-pair fertilizations. When focusing on broods containing both within and extra-pair chicks, we also found that extra-pair mates tended to be genetically more similar to the females than social mates both for microsatellite markers and MHC genes, even though the statistical support for these findings was less clear-cut than for offspring heterozygosity. Interestingly, in broods containing both within- and extra-pair young, fledging

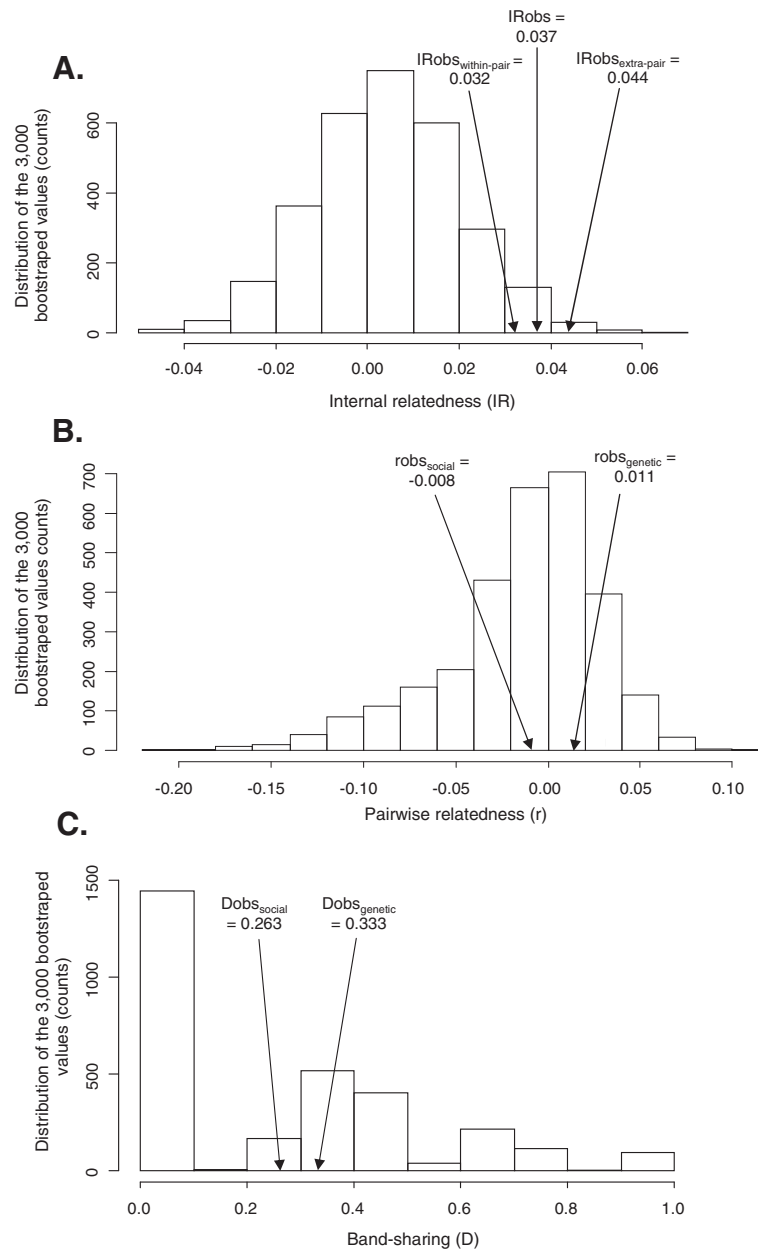


Figure 1 Distribution of the 1,000 bootstrapped Internal Relatedness IR (A), pairwise-relatedness r (B) and MHC allele-sharing (C). I_{Robs} , $I_{\text{Robs_within-pair}}$ and $I_{\text{Robs_extra-pair}}$ correspond to the observed IR computed on the entire sample of chicks, chicks produced by social males and chicks produced by extra-pair males, respectively. $r_{\text{obs_social}}$, $D_{\text{obs_social}}$ correspond to the observed r and allele-sharing between the female and her social mate. $r_{\text{obs_genetic}}$, $D_{\text{obs_genetic}}$ corresponds to the observed r and allele-sharing between the female and her extra-pair mate.

success of chicks sired by extra-pair males was higher than for chicks sired by social mates.

We could only assess realized mate choice (actual mating patterns) and not female preference (the preference that females might express in the absence of constraints). A number of environmental constraints might prevent females from expressing their actual preferences. For example, imperfect sampling of potential mates or the

cost of mate searching can indeed affect the pattern of realized mate choice [44].

Our prediction that females would tend to mate with genetically dissimilar mates was based on the assumption that this insular population has low genetic diversity and that it is therefore vulnerable to inbreeding depression. However, as a part of a larger study on the population genetics of the house sparrow, we found that the

Table 3 A: GLMM exploring the effects of Internal Relatedness (IR), pairwise relatedness (r), and MHC allele-sharing (D) of the social male on the female likelihood to engage in extra-pair fertilizations, B: Relative variable importance given by Akaike weights ($\Sigma AIC\omega$)

| A. Variables | Model | K | AICc | $\Delta AICc$ |
|---|---------------|------|------|---------------|
| Brood type (broods with extra-pair chicks vs. broods with no extra-pair chicks) (n = 66) | Null | 1 | 90.6 | 0.00 |
| | D | 2 | 91.4 | 0.78 |
| | IR | 2 | 92.0 | 1.37 |
| | r | 2 | 92.0 | 1.39 |
| | D + r | 3 | 92.5 | 1.88 |
| | D + IR | 3 | 93.1 | 2.53 |
| | IR + r | 3 | 93.4 | 2.86 |
| | Year | 3 | 94.3 | 3.74 |
| | D + IR + r | 4 | 94.4 | 3.8 |
| | D + year | 4 | 95.5 | 4.89 |
| | IR + year | 4 | 95.6 | 5.02 |
| | r + year | 4 | 95.8 | 5.18 |
| | D + r + year | 5 | 96.7 | 6.08 |
| | IR + r + year | 5 | 97.2 | 6.61 |
| D + IR + year | 5 | 97.2 | 6.62 | |
| D + IR + r + year | 6 | 98.6 | 8.03 | |

| B. Variables | Source of variation | $\Sigma AIC\omega$ |
|---|---------------------|--------------------|
| Brood type (broods with extra-pair chicks vs. broods with no extra-pair chicks) | D | 0.39 |
| | r | 0.34 |
| | IR | 0.31 |
| | Year | 0.13 |

K = number of parameters.

within-population diversity of both microsatellites and MHC was similar between this (Hoëdic) and six mainland populations located within a radius of 200 km (Bichet et al., unpublished observations). The Hoëdic population was nevertheless genetically differentiated (based on F_{st} values) from the other populations used in this study. Therefore, while genetic variation has been maintained in this insular population, isolation and reduced gene flow have still produced genetic divergence from the mainland populations (Bichet et al., unpublished observations).

Insofar as the elevated IR of both intra- and extra-pair offspring reflects a female mating preference for genetically similar males (but see findings on pairwise relatedness between mates, Figure 1B), there are several possible mechanisms that could explain such a preference. First, spatially variable environmental conditions (such as variable risks to contract infectious diseases) might have promoted the evolution of co-adapted genes conferring a

Table 4 A: GLMM exploring the effects of brood type (broods with extra-pair chicks vs. broods with no extra-pair chicks) on hatching and fledging success, B: Relative variable importance given by Akaike weights ($\Sigma AIC\omega$)

| A. Variables | Model | K | AICc | $\Delta AICc$ | ω |
|------------------------------|---------------------------|---|-------|---------------|----------|
| Hatching success (n = 65) | Null | 1 | 74.1 | 0.00 | 0.523 |
| | Brood type | 2 | 76.1 | 1.96 | 0.196 |
| | Year | 3 | 76.6 | 2.46 | 0.153 |
| | Year + brood type + year* | 5 | 78.1 | 3.97 | 0.072 |
| | Year + brood type | 4 | 78.6 | 4.47 | 0.056 |
| Fledging success (n = 55) | Year | 3 | 96.6 | 0.00 | 0.722 |
| | Year + brood type | 4 | 99.0 | 2.42 | 0.215 |
| | Null | 1 | 102.7 | 6.07 | 0.035 |
| | Year + brood type + year* | 4 | 104.1 | 7.54 | 0.017 |
| | Brood type | 2 | 104.9 | 8.25 | 0.012 |

| B. Variables | Source of variation | $\Sigma AIC\omega$ |
|------------------|---------------------|--------------------|
| Hatching success | Brood type | 0.32 |
| | Year | 0.28 |
| | Year*brood type | 0.07 |
| Fledging success | Year | 0.95 |
| | Brood type | 0.24 |
| | Year*brood type | 0.02 |

K = number of parameters. Asterisk means the interaction.

benefit under the locally prevailing conditions. Second, preference for genetically similar mates might also evolve through kin selection where females seek to increase their own inclusive fitness [23,25]. Our finding that females engaging in extra-pair matings showed a preference for

Table 5 A: GLMM exploring the effects of chick type (sired by the social or extra-pair mate) on chick multi-locus internal relatedness (IR), B: Relative variable importance given by Akaike weights ($\Sigma AIC\omega$)

| A. Variables | Model | K | AICc | $\Delta AICc$ | ω |
|-----------------|-------------------------------------|---|--------|---------------|----------|
| IR (n = 222) | Year | 3 | -231.1 | 0.00 | 0.308 |
| | Null | 1 | -231.0 | 0.05 | 0.300 |
| | Year + chick type + year*chick type | 5 | -229.5 | 1.55 | 0.142 |
| | Year + chick type | 4 | -229.4 | 1.70 | 0.132 |
| | Chick type | 2 | -229.2 | 1.90 | 0.119 |

| B. Variables | Source of variation | $\Sigma AIC\omega$ |
|--------------|---------------------|--------------------|
| IR | Year | 0.58 |
| | Chick type | 0.39 |
| | Year*chick type | 0.14 |

K = number of parameters. Asterisk means the interaction.

Table 6 A: GLMM exploring the effects of Internal Relatedness (IR), pairwise relatedness to the female (r), and MHC allele-sharing (D) on the likelihood of being a social or an extra-pair male, B: Relative variable importance given by Akaike weights ($\Sigma AIC\omega$)

| A. Variables | Model | K | AICc | $\Delta AICc$ | ω |
|--|---------------|-------|-------|---------------|----------|
| Male type (social vs. extra-pair) (n = 60) | r | 2 | 98.1 | 0.00 | 0.183 |
| | D | 2 | 98.3 | 0.21 | 0.164 |
| | Null | 1 | 98.5 | 0.32 | 0.156 |
| | D + r | 3 | 99.1 | 0.93 | 0.115 |
| | IR | 2 | 99.5 | 1.39 | 0.091 |
| | IR + r | 3 | 99.8 | 1.64 | 0.080 |
| | IR + D | 3 | 100.0 | 1.83 | 0.073 |
| | IR + D + r | 4 | 101.0 | 2.87 | 0.044 |
| | r + year | 4 | 102.6 | 4.51 | 0.019 |
| | D + year | 4 | 102.8 | 4.65 | 0.018 |
| | Year | 3 | 102.9 | 4.77 | 0.017 |
| | D + r + year | 5 | 103.6 | 5.43 | 0.012 |
| | IR + year | 4 | 104.1 | 5.97 | 0.009 |
| | IR + r + year | 5 | 104.4 | 6.26 | 0.008 |
| IR + D + year | 5 | 104.6 | 6.43 | 0.007 | |
| IR + D + r + year | 6 | 105.6 | 7.51 | 0.004 | |

| B. Variables | Source of variation | $\Sigma AIC\omega$ |
|--------------------------------------|---------------------|--------------------|
| Male type (social vs. extra-pair) | r | 0.46 |
| | D | 0.44 |
| | IR | 0.32 |
| | Year | 0.09 |

K = number of parameters.

Table 7 A: GLMM exploring the effects of male type (social vs. extra-pair) on fledging success, Fledging success corresponds to the number of chicks sired by a male, divided by the total number of eggs laid in the clutch, B: Relative variable importance given by Akaike weights ($\Sigma AIC\omega$)

| A. Variables | Model | K | AICc | $\Delta AICc$ | ω |
|------------------------------|-----------------------------------|---|-------|---------------|----------|
| Fledging success (n = 74) | Male type | 2 | 94.1 | 0.00 | 0.891 |
| | Male type + year | 4 | 98.7 | 4.61 | 0.089 |
| | Male type + year + year*male type | 5 | 101.8 | 7.75 | 0.019 |
| | Null | 1 | 106.9 | 12.80 | 0.001 |
| | Year | 2 | 111.3 | 17.25 | 0.000 |

| B. Variables | Source of variation | $\Sigma AIC\omega$ |
|------------------|---------------------|--------------------|
| Fledging success | Male type | 1.00 |
| | Year | 0.11 |
| | Year*male type | 0.02 |

K = number of parameters. Asterisk means the interaction.

more genetically similar males is also in agreement with the results of two recent avian studies on ground tits (*Parus humilis*) [13] and barn swallows (*Hirundo rustica*) [33]. Wang and Lu [13] showed that even though the propensity of females to engage in extra-pair matings did not depend on the relatedness with the social mates, females nevertheless sought extra-pair copulations with males with whom they were more related to than their social mates. Since there was no cost due to mating with relatives, Wang and Lu [13] suggested that these results support the hypothesis that females gain inclusive fitness by mating with related males.

Findings based on offspring heterozygosity (IR) and based on relatedness between pair members (r) provided a quite different picture, which might appear puzzling. One possible explanation involves the residual variation in IR that is not accounted for by r. Alternatively, we would also like to remind that the two indices refer to two different steps in the process of mate choice, r refers to the similarity between mates, IR refers to the product of the mate choice where recombination, early embryo failure, etc. might contribute to generate the observed discrepancy.

Population structure or 'environmental constraints' might also affect pair formation potentially interfering with mating preferences [44]. For instance, related individuals might tend to cluster in the same flock, as reported for instance in lekking peacocks (*Pavo cristatus*) [73] and to occupy nearby nest boxes, which might increase the likelihood of mating with genetically similar mates. However, since the relatedness between social mates was not higher than for random pairs, this type of clustering seems unlikely in the present case.

A large proportion of chicks were sired by an extra-pair mate. The proportion of extra-pair chicks was higher than those previously reported for other house sparrow populations [51,74-77]. One possible explanation is a bias in estimates of extra-pair chicks due to mistaken identification of birds (coloured leg bands), however, this is unlikely as we found a mismatch in maternal identity in only 3% (2/69) of broods (and in both cases the entire brood was assigned to a single mother). In five broods, none of the chicks was assigned to the social father. However, in all of these five cases, chicks were assigned to at least two different males, strongly suggesting that the high proportion of extra-pair chicks was not the result of mistakes in the identification of the social father. Most of the previous work found that extra-pair paternity is low in insular populations [12,51,78], although some exceptions have been reported [79-81], with as much as 55% of extra-pair chicks reported for an insular population of tree swallows (*Tachycineta bicolor*) [75]. The ecological factors that might explain why insular populations have lower or higher levels of extra-pair paternity compared to mainland populations

are unclear and contentious. One might speculate that insular populations with high breeding density and synchrony might be more prone to extra-pair copulations because of the increased availability of extra-pair mates, but this would require additional work.

There have been extensive reports of MHC-dependent mating preferences [9,39,82-88]; however, we found no support for a potential role of MHC genes in mate choice, other than the general assessment of relatedness. Insular populations might be less exposed to parasites and pathogens [89-92], which could weaken the selection for specific MHC alleles and diversity (but see [9]).

Conclusions

These results tend to support the idea that mating with genetically similar mates can either avoid the disruption of co-adapted genes or confer a benefit in terms of kin selection. Definitely, more work should be devoted to the role of MHC-based mate choice in mainland and insular populations that differ in their exposure to infectious diseases.

Abbreviations

MHC: Major Histocompatibility Complex; EP: Extra-pair; PBR: Peptide-binding region; IR: Internal relatedness; IRobS: Observed internal relatedness; IRobS_{extra-pair}: Observed internal relatedness for extra-pair chicks; IRobS_{within-pair}: Observed internal relatedness for within-pair chick; MLH: Multi-locus heterozygosity; r: Pairwise relatedness; robs_{social}: Observed pairwise relatedness for social pairs, between the female and her social male; robs_{genetic}: Observed pairwise relatedness for genetic pairs, between the female and the male who sired the chicks; D: Number of shared MHC alleles; Dobs_{social}: Observed number of shared MHC alleles for social pairs; Dobs_{genetic}: Observed number of shared MHC alleles for genetic pairs; GLMM: General Linear Mixed Model; AIC: Akaike Information Criterion; ω : AIC weights; $\Sigma AIC\omega$: Summed AIC weights.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

Study design: DJP, YM, GS. Data collection in the field: CB, EC-H, MB, SG, GS. Molecular genetic analyses: CB, MB. Statistical analyses: CB, LD. Manuscript writing: CB, DJP, YM, LD, SG, GS. All authors read and approved the final manuscript.

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