

Oral presentation

Engineering a synthetic molecular oscillator based on the Lotka-Volterra dynamic

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Background

Oscillators are a fundamental building block in many engineering fields, as they provide the basis for counting, timing and synchronisation. Oscillators are found in many everyday devices such as clocks, computers or radios. Similarly, oscillations are an essential part of biological systems – providing the basis for, for example, rhythmic patterns and regulatory networks. The ability to build a stable, controllable biological oscillator would be a major step towards reliable synthetic biology based circuits. Elowitz *et al.* were part of the first ones to try to build an oscillator. Their oscillator was based on genetic network at the single cell level. However, due to the stochastic behaviour inherent at the gene expression level, the oscillations were not stable or persistent. In this paper, we present an original oscillator design produced during the iGEM-2006 competition at Imperial College. The project addressed stability and reliability issues by defining an oscillator at the population level and by applying strict engineering rules to the system development process.

Methods

This iGEM project combined genetic engineering methods with a clearly defined development cycle, copied from the

traditional engineering approach – a cycle of specification, design, modelling, testing and implementation – with biological processes to produce a synthetic biological oscillator. The overall specification aim was to build a biological oscillator in *E. coli* based on the Lotka-Volterra population dynamics. This approach was used to design a synthetic quorum sensing & quenching mechanism, made of BioBricks. In order to satisfy the requirements for the Lotka Volterra predator-prey dynamics BioBricks were constructed to show properties such as exponential growth of a prey molecule (Acyl Homoserine Lactone). In order to reduce the complexity of the overall assembly, test constructs were built to characterise the different system components. Because the design comprises BioBricks in a two population system, the parameters of the oscillations can be controlled to achieve synchronised oscillations. Furthermore, since the system uses population wide oscillations, it enables other devices to synchronise the oscillator. It is important to determine the component properties and their impact on the overall system. Hence, the modelling of the pure and modified Lotka-Volterra dynamics, along with the test constructs, were carried out. Because BioBricks were used in the design, the implementation was carried out with standard assembly. At the test-

Specifications		
		<ul style="list-style-type: none"> ● Deliver a stable biological oscillator. ● Measurable output. ● Frequency and amplitude tunable. ● Produce documentation at each step for quality control and traceability purposes.
Design		
		<ul style="list-style-type: none"> ● Based on Lotka-Volterra dynamic. ● Use of Quorum sensing-quenching BioBricks from MIT Registry. ● Population wide oscillations of AHL. ● Two cell population system defined to be able to tune frequency and amplitude. ● Definition of test constructs to break down complexity and to allow fine characterization.
Modelling		
		<ul style="list-style-type: none"> ● Test construct modelling. ● Investigation of the Lotka-Volterra dynamic. ● Full system modelling.
Implementation		
		<ul style="list-style-type: none"> ● Based on BioBricks. ● Use of the Standard Assembly. ● Quality control procedure based on sequencing.
Testing and Validation		
		<ul style="list-style-type: none"> ● Definition of testing protocols to satisfy specifications. ● Characterisation of the different test construts. ● Characterisation of the two-cell population oscillator

Figure 1
Summary of our methodology.

ing stage, the test constructs were characterised. Consequently, the properties and constraints on the full system could be extracted. (See a summary of the method in Figure 1)

Results

BioBricks composing the test constructs and the final oscillator were engineered and a number of important results were extracted from the modelling and testing stages: i) The test constructs/BioBricks representing the

different parts of the oscillator were characterised from testing in E. Coli (DH5α). ii) The model of the test constructs, which included experimental data, predicted stable oscillations as output of the full system. iii) The influence of specific system parameters on the frequency, amplitude and stability of the oscillations was investigated with the Lotka-Volterra model.

Conclusion

In the project, fundamental units of the system were fabricated and characterised. These have been added to the Registry of Standard Biological Parts. The modelling exercise was highly successful and we fully expect to complete a stable synthetic biology based oscillator to be completed in the near future. For full details visit our iGEM website <http://openwetware.org/wiki/IGEM:IMPERIAL/2006>.

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