

Characterization of a hydroxyproline-rich glycoprotein in pearl millet and its differential expression in response to the downy mildew pathogen *Sclerospora graminicola*

Neerakkal Sujeeth · Ramachandra K. Kini · Sekhar Shailasree · Eelco Wallaart · Shekar H. Shetty · Jacques Hille

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Abstract A monoclonal antibody, JIM 20, derived against an extensin type of hydroxyproline-rich glycoprotein (HRGP) from pea, showed high affinity for HRGP in pearl millet [*Pennisetum glaucum* (L.) R. Br.]. Electrophoretic separation of Tris–SDS extracted proteins from suspension cells of pearl millet revealed a range of PM-HRGP polypeptides having a glycan epitope, which reacted with JIM 20. A high molecular mass band, probably an HRGP aggregate or polymer, and a few low molecular mass polypeptides were recognized by JIM 20 during Western blot analysis. Treatment of pearl millet suspension cells with hydrogen peroxide in the presence of an endogenous peroxidase resulted in insolubilization of HRGP polypeptides with molecular weights between 45 and 33 kDa. To investigate the gene coding for an extensin type of HRGP, a fosmid-based genomic library of pearl millet having a fourfold genome coverage was constructed. A partial sequence of 378 bp of an HRGP gene was obtained by PCR amplification of pearl millet DNA with a primer pair designed from the conserved regions of

monocotyledon extensin type of HRGPs. Screening the genomic library using the homologous probe developed from the 378-bp PCR product resulted in the isolation of five fosmid clones. Restriction mapping of these fasmids resulted in an 11.8-kb region around an HRGP gene in pearl millet. The newly characterized gene, *PM-HRGP*, had all the characteristic features of a monocotyledon extensin type of HRGP. An intron at the 3' untranslated region of the gene was identified by cDNA cloning. Differential expression of the *PM-HRGP* gene was observed during compatible and incompatible interactions of pearl millet with the downy mildew pathogen *Sclerospora graminicola* (Sacc) Schroet. Induced expression of the gene was observed only in case of an incompatible interaction.

Keywords Extensins · PM-HRGP · Fosmid pearl millet library · Downy mildew · *Sclerospora graminicola*

Abbreviations

HRGP	Hydroxyproline-rich glycoprotein
PRP	Proline-rich proteins
SDS-PAGE	Sodium dodecyl sulfate-polyacrylamide gel electrophoresis
qRT-PCR	Quantitative real-time PCR

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N. Sujeeth (✉) · J. Hille
Molecular Biology of Plants, Groningen Biomolecular Sciences and Biotechnology Institute, Centre for Life Sciences, University of Groningen, Nijenborgh 7, 9747 AG Groningen, The Netherlands
e-mail: s.neerakkal@rug.nl

N. Sujeeth · R. K. Kini · S. Shailasree · S. H. Shetty
Department of Studies in Biotechnology, Manasagangotri, University of Mysore, Mysore 570006, Karnataka, India

E. Wallaart
Sylphium Life Sciences, Duurswoldlaan 22, 9727 DJ Groningen, The Netherlands

Introduction

The mature plant cell wall contains proteins forming a positively charged network or frame to which negatively charged carbohydrates are attached (Cannon et al. 2008). Among the glycoprotein components of the plant cell wall, hydroxyproline-rich glycoproteins (HRGPs) are one of the important structural components with a wide range of roles

including defense against plant pathogens. This superfamily of glycoproteins includes extensins, proline-rich proteins (PRPs) and arabinogalactan proteins (AGPs) (Sommer-Knudsen et al. 1998; Jose-Estanyol and Puigdomenech 2000; Showalter et al. 2010). The extensins and PRPs are known to be insoluble proteins, whereas AGPs are soluble proteins.

The dicotyledonous extensins constitute a large family of glycoproteins that function in diverse aspects of plant growth, development and in some cases in plant defense (Showalter 1993). Accumulation and cross-linking of extensins provide protective functions during pathogen attack and wounding in many plants (Templeton et al. 1990; Bowles 1990; Jackson et al. 2001). The expression of extensin transcripts can be induced by various treatments, such as wounding, pathogen infection and mechanical signals (Merkouropoulos et al. 1999; Yoshida et al. 2001). However, oxidative cross-linking of these cell wall proteins is elicited very rapidly upon pathogen infection, before deployment of transcription-dependent defenses (Brisson et al. 1994; El-Gendy et al. 2001).

Extensin genes are well characterized in dicots, and the existence of a multigene family for extensin has been described in carrot (Chen and Varner 1985) and oil seed rape (Evans et al. 1990). The *Arabidopsis* genome encodes 20 closely homologous extensin polypeptides with distinct patterns of expression at the organ/tissue level (Zimmermann et al. 2004). The search for extensin in monocots resulted in the characterization of a much simpler system, probably encoded by a single gene as in the case of maize, sorghum and rice. The first monocot *HRGP* gene with a single Ser-(Hyp)₄ motif near the C terminus, a variant of the Ser-(Hyp)₄-rich extensin family in dicots, was identified in maize (Stiefel et al. 1988, 1990). Similar structures were reported in sorghum (Raz et al. 1991) and also in rice with the exception that the C-terminus Ser-Hyp₄ motif is absent (Caelles et al. 1992). All these monocot variants of extensin were classified as a separate group called proline/hydroxyproline-rich glycoproteins (P/HRGPs) along with other PRPs by Sommer-Knudsen et al. (1998). They also indicated that P/HRGPs may have been evolved from extensin by deletion of Ser(Pro)₄ repeats. Jose-Estanyol and Puigdomenech (2000) reviewed and classified them under the extensin class based on the presence of a single Ser (Pro)₄ motif and having similar functional properties compared to other extensins characterized.

The cereal pearl millet [*Pennisetum glaucum* (L.) R. Br.] is a staple food grain of people living in the semi-arid tropics including parts of India and Africa. It is the seventh most important cereal crop worldwide, grown on about 15 million ha in Africa and around 12 million ha in Asia (Léder 2004). Due to its adaptation to drought and heat, pearl millet is an important crop that helps to attain food security where other cereals fail. The major constraint for its production is the downy mildew disease caused by the

obligate, biotrophic, oomycetous pathogen *Sclerospora graminicola* (Sacc.) Schroet. It causes systemic infection in pearl millet that manifests itself as foliar symptoms and malformation of the panicles, resulting in severe grain loss. Even though several resistant cultivars have been developed by classical breeding programs, the resistance breaks down over a period of time due to the continuous evolution of the pathogen. A better understanding of the basis of resistance in pearl millet to the pathogen is essential for developing durable resistance strategies.

Previously, we showed that cell wall HRGPs in pearl millet accumulate in response to pathogen infection and elicitor treatments (Shailasree et al. 2004; Sujeeth et al. 2010). Here, we report the presence of an extensin type of PM-HRGP in pearl millet. A putative role of the protein in plant defense, as suggested by cross-linking observed in vitro, is discussed. Furthermore, a gene encoding the extensin type of *PM-HRGP* in pearl millet was characterized. The accumulation of transcripts of this gene during interaction of pearl millet with *S. graminicola*, the downy mildew disease pathogen, has been studied.

Materials and methods

Analysis of hydroxyproline-rich glycoproteins

Pearl millet suspension cell culture

The pearl millet cell culture was raised from IP18296, a highly resistant cultivar by following the method of Vasil and Vasil (1981). The well-established suspension cells were regularly sub-cultured onto fresh medium at 1:5 dilution rates, at 10-day intervals and after ten sub-cultures the cells were used for the study. A cell culture (10^8 cells ml⁻¹) at the midpoint of the log phase of growth (16-day-old) was used for the experiments.

Protein isolation and deglycosylation

Total protein was extracted from the suspension cells by resuspending in 1 ml of 100 mM Tris-HCl (pH 7.4) containing 4% SDS in a handheld homogenizer. The homogenates were centrifuged for 5 min at 10,000×g and the supernatant was collected. A part of the extracted total protein was dialyzed against deionized H₂O, lyophilized and deglycosylated using HF in pyridine (Aldrich) following Vanholst and Varner (1984).

Electrophoresis and Western blot analysis

The supernatants collected from the crude and deglycosylated samples were assayed for protein (Lowry et al. 1951)

and 60 µg of protein was subjected to 12% SDS-PAGE. The separated proteins were stained with Coomassie blue. Glycoproteins in the gels were identified by Periodic acid Schiff (PAS) stain (Shailasree et al. 2004). For Western blot analysis, the separated proteins on SDS-PAGE were immediately blotted onto polyvinylidene difluoride (PVDF) membranes using the Multiphor II (Pharmacia, Freiburg, Germany) electrophoretic transfer apparatus according to the manufacturer's protocol. The blots were blocked in 2% fat-free milk in Tris-buffered saline (TBS: 10 mM Tris-HCl, pH 8.0 containing 150 mM NaCl). All blots developed were probed separately with the monoclonal antibodies JIM20 and JIM12, specific to dicot extensins, and LM1, specific toward P/HRGPs from maize (Plant Probes, <http://www.plantprobes.net/>).

Cross-linking analysis

Aliquots of 1-ml cell culture were placed in microfuge tubes. Peroxidase-mediated cross-linking analysis was carried out by addition of 0.03% H₂O₂ and 0.03% H₂O₂ + 0.05% sodium azide in two aliquots for 10 min. Untreated cells were kept as a control. The cells were separated from the medium by centrifugation at 5,000×g for 5 min at room temperature and used for protein extraction and Western blot analysis using JIM 20.

PCR amplification of a conserved region of the *HRGP* gene

Pearl millet var.7042S, an inbred line (Thakur et al. 2001), was used as the standard pearl millet genotype. Genomic DNA isolation from pearl millet seedlings was carried out according to the protocol of Fulton et al. (1995). For PCR amplification of the *HRGP* gene, primers were designed from the conserved regions of *HRGP* gene sequences of maize (AJ131535), sorghum (X56010) and rice (X61280).

The primer sequences were PrRuG 2796 and PrRuG 2797 (Table 1). PCR of pearl millet genomic DNA was carried out using a high GC-rich PCR system (Roche, Mannheim, Germany). The GC-rich solution at a concentration of 0.5 M was used for specific amplification of the gene product. The amplification was carried out by the PCR program: initial denaturation for 5 min at 95°C followed by 30 cycles of 30 s at 95°C, 1 min at 65°C, 1 min at 72°C, and a final extension step of 7 min at 72°C. To the blunt end PCR product obtained, a T overhang was added using Taq DNA polymerase (Fermentas). It was further cloned in the pGEM-T Easy vector (Promega, Madison, USA) and sequenced. This clone was used as an HRGP homologous probe for further studies.

Fosmid library construction

High molecular weight DNA (HMW DNA) was isolated from roots of 3-day-old pearl millet seedlings using the G NOME DNA kit (MP Biomedicals, Illkirch, France) according to the manufacturer's protocol. To bring 80% of HMW DNA to 40-kb size, it was subjected to random physical shearing by several passages through a sterile tip of a micropipette. It was further run on a 1% agarose gel in a CHEF-DR II pulsed field electrophoresis system (BIO-RAD) under the following conditions: 16-h run time, 14°C, 1-80 s pulses, 120° angle in 0.5× TAE at 6 V/cm. The gel eluted 40-kb DNA was end-repaired to blunt end and cloned into the pCC1FOS fosmid vector using the Copycontrol Fosmid Library Production kit (Epicentre Biotechnologies, Madison, USA), according to the manufacturer's instructions. The ligation mixture was then packed into phage particles using MaxPlax lambda packaging extracts (Epicentre Biotechnologies, Madison, USA). The packaged fosmid clones were titrated by serial dilutions in phage dilution buffer (PDB) and infected to 100 µl of Phage T1-resistant *E. coli* (EP1300™ plating strain) cells

Table 1 PCR primers used in this work

Primer	Description	Sequence (5'–3')
PrRuG 2796	Forward for 378-bp PCR product from pearl millet DNA	TCCAGGCCGACGCCGGCTACGGCTAC
PrRuG 2797	Reverse for 378-bp PCR product from pearl millet DNA	TACGTCGGCGGAGTCGGCTTGG
PrRuG 2801	Forward for <i>PM-HRGP</i> intron search	GAGGATGGGCGCCGGAAG
PrRuG 2812	Reverse for <i>PM-HRGP</i> intron search	CTTGCCGGCTGGCTAGCACAC
PrRuG 2817	Forward for <i>GAPDH</i> gene characterization in pearl millet	ATGGGCAAGATTAAGATCGGAATCAACG
PrRuG 2818	Reverse for <i>GAPDH</i> gene characterization in pearl millet	CATGTGGCCGATCAGGTCGAC
PrRuG 2892	Forward for <i>GAPDH</i> gene qRT-PCR	CCAAGGCTGTCGGTAAGGTTC
PrRuG 2893	Reverse for <i>GAPDH</i> gene qRT-PCR	TGCTCGTTCAGAGCGATTCC
PrRuG 3293	Forward for <i>PM-HRGP</i> gene qRT-PCR	GCCTAAGCCGAAGCCACCAA
PrRuG 3294	Reverse for <i>PM-HRGP</i> gene qRT-PCR	GCGTGTAGGTCGGAGGAGTT

for 20 min at 37°C. The phage-infected *E. coli* cells were plated onto LB agar plates containing chloramphenicol (12.5 µg/ml).

Pearl millet has a haploid genome size of 2,352 Mb (Bennett et al. 2000). The required number of fosmid clones for a complete library of pearl millet was calculated as N using the formula, $N = 1n(1 - P)/1n(1 - f)$ (Sambrook et al. 1989). A total of at least 2.7×10^5 clones were developed to get a complete library of pearl millet with a fourfold coverage of its haploid genome. For storage of the pearl millet library, 2.7×10^5 clones from the library plates were suspended in LB broth, pooled, aliquoted into 1 ml sterile vials containing 20% glycerol and stored at -80°C .

Screening of the pearl millet DNA library

One aliquot of the pearl millet DNA library from -80°C stocks was serially diluted to obtain about 20,000 colonies/LB agar plate (14-cm diameter \times 2.3-cm thickness) containing chloramphenicol. A total number of 18 of such plates were developed to obtain the complete library of pearl millet with fivefold genome equivalence (3.6×10^5 clones). A colony blot hybridization (Sambrook et al. 1989) with the homologous HRGP probe labeled with [^{32}P]-dCTP was carried out. Positive clones isolated for the HRGP gene were sub-cultured overnight in LB broth and stored.

Restriction analysis

The fosmids were isolated from positive clones by the alkaline lysis method of Sambrook et al. (1989). The presence of a 40-kb pearl millet DNA insert in the isolated fosmids was confirmed by *Not* I digestion followed by electrophoresis on the CHEF-DR II pulsed field electrophoresis system with the same conditions as mentioned in the library preparation protocol. Positive fosmids were digested with restriction enzymes, *Ava* I, *Bam* HI, *Hind* III and *Hpa* I, in single and double combinations, separated on an 0.8% agarose gel and transferred by Southern blotting to Hybond N⁺ nylon membrane (Amersham, UK) following the protocol of Sambrook et al. (1989). The labeling and analysis of blots was carried out using the homologous HRGP probe labeled with [^{32}P]-dCTP.

HRGP sequence annotation

A 3.5-kb *Bam* HI fragment containing the *HRGP* full-length gene sequence identified from restriction analysis was ligated into a dephosphorylated pUC18/19 vector linearized at the *Bam* HI site. The ligated product was cloned into the MM294 competent *E. coli* strain (Sylphium Life Sciences, Groningen, The Netherlands) and sequenced.

The sequence of the 3.5-kb *Bam* HI fragment with the *HRGP* full-length gene was deposited in the database [GenBank: GQ223398].

Cloning of the *HRGP* gene from cDNA of pearl millet

For cDNA synthesis, total RNA was extracted from seedlings of pearl millet using Spectrum Plant Total RNA kit (Sigma, St. Louis, USA). The DNA-free RNA was converted to cDNA by reverse transcription using the oligo-5'-GCAGCGATC(T)₂₈V(A/C/G)N(A/C/G/T)-3'. The forward primer spanning the start codon of the *PM-HRGP* gene PrRuG 2801 and a reverse primer at a region 181-bp downstream from the stop codon, PrRuG 2812 (Table 1), was used for amplification of the *HRGP* gene area simultaneously from cDNA and genomic DNA of pearl millet. The resulting products from both reactions were further ligated into the pGEM-T easy vector, cloned and sequenced.

Differential gene expression study

For the analysis, an endogenous control gene glyceraldehyde-3-phosphate dehydrogenase (*GAPDH*) was newly isolated and characterized from pearl millet [GenBank: GQ398107]. The primer pair used for amplification was forward primer-PrRuG 2817 and reverse primer-PrRuG 2818 (Table 1).

Plant material

Pearl millet cultivars 7042 S (highly susceptible, HS) with >25% downy mildew disease incidence (DMDI) and IP18296 (highly resistant, HR) with 0% DMDI after inoculation with *S. graminicola* under field conditions were used. Two-day-old seedlings were root dip inoculated with zoospores of *S. graminicola* (Singh and Gopinath 1985). Seedlings dipped in sterile distilled water served as uninoculated control. The seedlings were harvested at different time intervals for total RNA isolations using Spectrum Plant Total RNA kit (Sigma, St. Louis, USA).

Quantification of *HRGP* expression

Reverse transcription of 1 µg of DNA-free total RNA was performed using oligo (dT) primers for analysis. The relative expression of *PM-HRGP* in relation to the endogenous control gene *GAPDH* was assessed by quantitative real-time PCR (qRT-PCR) using a 7500 Fast Real-Time PCR system (Applied Biosystems). The primers for qRT-PCR were designed from the coding regions of *PM-HRGP* (PrRuG 3293 and PrRuG 3294) and the *GAPDH* gene (PrRuG 2892 and PrRuG 2893) (Table 1). Twenty-microliter PCR reactions contained diluted cDNA, $2 \times$ Power

SYBR Green Master mix (Applied Biosystems, Foster City, CA) and 250 nM of forward and reverse primers (both for *HRGP* and *GAPDH*). Thermal cycling conditions were 95°C for 10 min followed by 40 cycles of 95°C for 15 s and 60°C for 1 min. Following amplification, a dissociation stage was performed to check for nonspecific PCR products. Specificity of the amplicons was checked on an agarose gel of 1.2%. Induction of genes (fold increase) at different time points of inoculation was compared with RNAs from the 0-h sample and calculated using the comparative threshold ($2^{-\Delta\Delta C_T}$) method of Livak and Schmittgen (2001). The *PM-HRGP* gene expression was normalized to that of the control *GAPDH* gene during quantitative measurement.

Results

Monoclonal antibody JIM20 binds to PM-HRGP in pearl millet

Coomassie blue staining of SDS-PAGE separated total proteins from suspension cells of pearl millet showed several bands ranging from lower to higher molecular mass (Lane 1, Fig. 1). The deglycosylated sample showed two proteins with molecular weights 37 and 30 kDa (Lane 2, Fig. 1). The PAS staining for glycoproteins staining revealed the presence of a high molecular mass range fuzzy band, a glycan-rich aggregate or polymer in the total protein extract (Lane 3, Fig. 1), which disappeared upon deglycosylation of the sample (Lane 4, Fig. 1). The high

molecular mass band observed during the PAS staining gave an intense reaction with JIM 20 antibodies during Western blot analysis. Upon deglycosylation, the reactivity with the antibody was lost due to the specificity of the JIM 20 toward the carbohydrate epitope of the isolated protein. Furthermore, a lower molecular mass polypeptide of 56 kDa and three other polypeptides falling at a low molecular mass between 45 and 33 kDa also showed a similar pattern of cross-reactivity (lanes 5 and 6, Fig. 1). These HRGPs could not be detected when probed with the antibodies LM1 and JIM12 during Western blot analysis (results not shown).

Cross-linking analysis of PM-HRGP

Cross-linking analysis revealed that three polypeptides with molecular size between 45 and 33 kDa were reduced in the protein extracts from H₂O₂-treated cell suspensions, probably due to the insolubilization process of PM-HRGP (Lane 8, Fig. 1). When sodium azide, a well-known inhibitor of peroxidase, was added to the growth medium prior to the addition of H₂O₂, the levels of the polypeptides were partially restored to the previous levels indicating that they were not insolubilized (lane 9, Fig. 1).

PCR amplification of a conserved region of the *HRGP* gene from pearl millet

A 378-bp DNA fragment was obtained by PCR amplification of pearl millet DNA using an *HRGP* gene-specific primer pair. The sequence of the fragment showed 77%

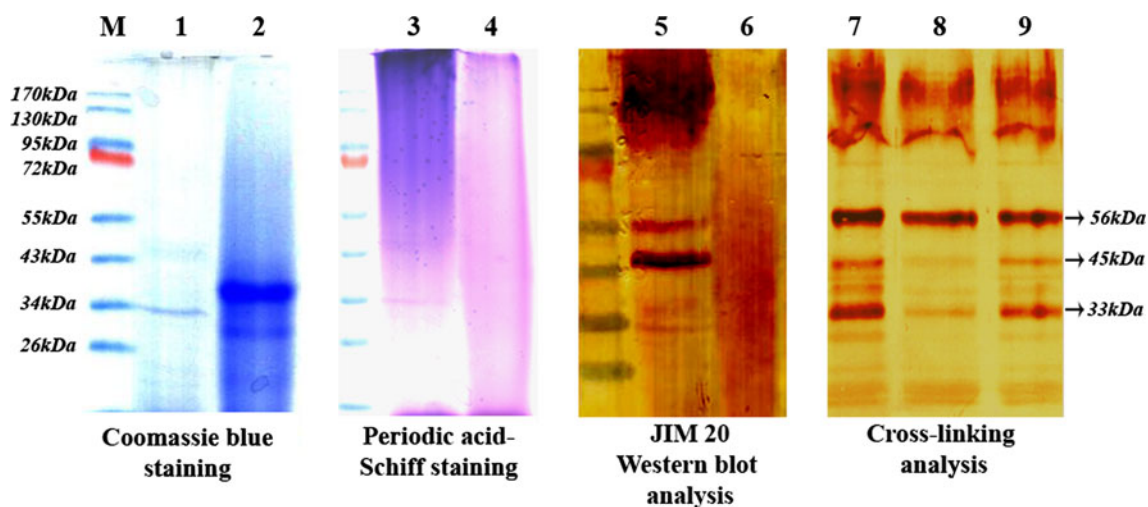


Fig. 1 Analysis of Tris-HCl and SDS extracted proteins from pearl millet suspension cells. *Lanes 1* and *2* Coomassie blue staining of crude and deglycosylated protein samples, respectively; *lanes 3* and *4* periodic acid Schiff staining of crude and deglycosylated protein samples, respectively; *lanes 5* and *6* Western blot analysis using JIM 20 monoclonal antibody on crude and deglycosylated protein

samples, respectively. For cross-linking analysis, 0.03% H₂O₂ was added to 1 ml of aliquots of pearl millet suspension cultured cells in the presence and absence of 0.05% sodium azide. Cells and culture medium were separated by centrifugation, subjected to SDS-PAGE, transferred to PVDF membrane and developed with JIM20. *Lane 7* Untreated; *8* 0.03% H₂O₂; *9* 0.03% H₂O₂ and 0.05% sodium azide

similarity to the *Sorghum vulgare* extensin gene (GenBank: X56010.1). The PCR product thus obtained was cloned and used as a homologous probe for obtaining the full-length *HRGP* gene from pearl millet.

Genomic library preparation and screening for a full-length *HRGP* gene in pearl millet

A fosmid genome library was prepared from physically sheared HMW DNA from pearl millet. The library was constructed using two ligation and packaging reactions to include the large-sized pearl millet genome (2,352 Mb). Furthermore, screening of 3.6×10^5 clones from the library using the *HRGP* homologous probe identified five positive clones for the *HRGP* gene, each of which had the gene in a 40-kb DNA fragment of pearl millet.

Restriction mapping of positive clones

An identical hybridization reaction/pattern in the gene area for all five positive fosmids was recorded during restriction analysis. This result confirms that all five positive fosmid clones isolated from the library contain the same copy of the *HRGP* gene. This was also confirmed by PCR of these clones with the primers which had earlier amplified the 378-bp partial sequence of the pearl millet *HRGP* and sequencing of the products. Furthermore, by carrying out single and double digestions with the four enzymes

selected for restriction analysis, restriction sites present in an 11.8-kb region around the *HRGP* gene in each fosmid were mapped. The gene was thus predicted to be present in a region between two *Bam* HI sites situated 3.5-kb apart in the 11.8-kb region (Fig. 2). The 3.5-kb region was further sub-cloned and sequenced.

Identification of the full-length *HRGP* gene in pearl millet

The sequence of the 3.5-kb *Bam* HI fragment showed the presence of a proline-rich gene sequence with a single long open reading frame of 939 bp. BLAST search of this 939-bp sequence showed close similarities of the proline-rich sequence to sorghum bicolor extensin (UniProtKB Accession Number: P24152), *Zea mays* extensin (P14918), *Nicotiana tabacum* extensin (P13983), *Arabidopsis thaliana* extensin protein (Q9M1G9), also with a vegetative cell wall protein of *Chlamydomonas reinhardtii* (Q9FPQ6), and a basic proline-rich protein of *Sus scrofa* (Q95JC9).

Characterization of the *PM-HRGP* gene

In silico translational analysis revealed that the 939-bp *HRGP* sequence encoded a protein of 312 amino acids (Fig. 3). The predicted protein of the *HRGP* gene from pearl millet was rich in amino acids like proline, threonine, lysine and tyrosine, showing similarity to the other

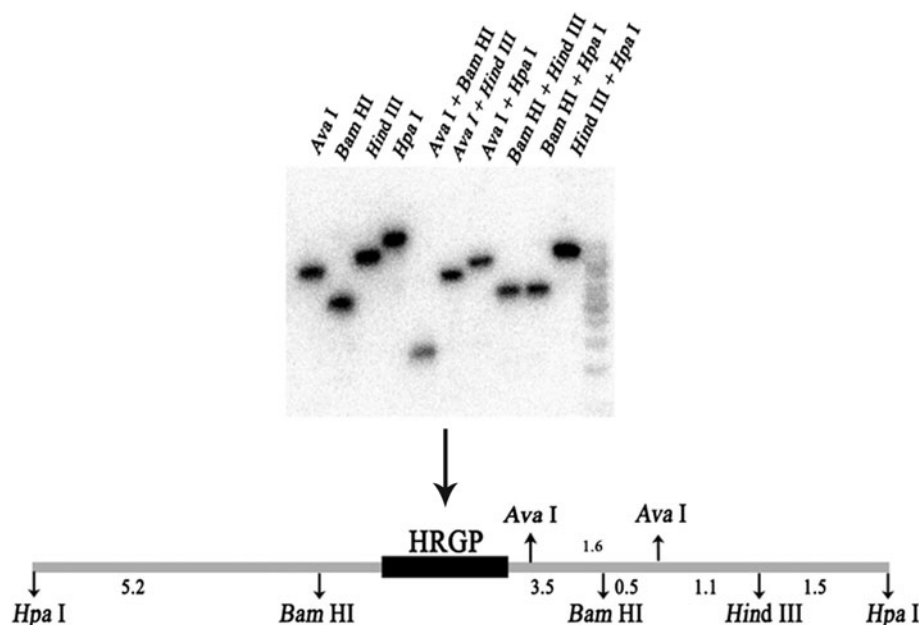


Fig. 2 Restriction analysis of the positive clones identified for the *HRGP* gene from a genomic fosmid library of pearl millet. Restriction digestion using *Bam* HI, *Ava* I, *Hind* III and *Hpa* I on each isolated fosmid was carried out and blotted. After *HRGP* probe radiolabeling of the restriction blots, the 11.8-kb region around the *HRGP* gene in all five positive fosmid clones were mapped. All five clones were

confirmed to have the same type of *HRGP* gene from the restriction pattern obtained. The figure represents the restriction mapping of one such positive clone from the library. The *PM-HRGP* gene was predicted to be situated between two *Bam*HI sites of 3.5-kb apart on the 11.8-kb region from the restriction pattern obtained

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-240 GACCACTGCAGTGCACGGCGAACCACAAAGGGACCAACGGGGAAAGCCTCCGGGCTCCTCCTCC
-180 CCCCCACATGCCTATCCTCTCGATCGATCAGGAGCGGGGCATCCGAGGCCCCACCCGG
-120 CCGCTTGCTCCGCGTGTATATRRAGAGGTGGCAGGGTGAGCTCTTCTCCTCAGGCCAGC
-60 CACACACTGATCAGAGCTTG TAGACTGAAGT GAGTGGCGAGAGTGAGAGCCAGTGAGAGG

1 ATG GGC GCC GGC AAG GCC GCT CTG CTG CTG GCC CTG GTG GCC GTG CGC CTG GCC GTC GAG
  M G A G K A A L L L A L V A V R L A V E 20
61 ATC CAG GCC GAC GCC GGC TAC GGC TAC GGC GGC GGG TAC GCG CCC ACG CCG ACC ACC CCG
  I Q A D A G Y G Y G G Y A P T P T T P 40
121 ACG CCC ACT CCG GAG AAG CCA CCC AAG GGC GGC AAG CCG CCC AAG GAG CAC GGT CCC AAG
  T P T P E K P P K G G K P P K E H G P K 60
181 CCT GAG AAG CCA CCC AAG GGC GAC AAG CCG CCC AAG GAG CAC GGG CAC AAG CCG GAG AAG
  P E K P P K G D K P P K E H G H K P E K 80
241 CCG CCC AAG GGG AAC AAG CCT CCC ACC TAC ACG CCG TCC CCG AAG CCG CCG ACG CCG CCG
  P P K G N K P P T Y T P S P K P P T P P 100
300 ACG TAC ACT CCC ACA CCG AAG CCC ACT CCG CCG GCG CCG AAG CCG AAG CCG ACG CCG CCC
  T Y T P T P K P T P P A P K P K P T P P 120
361 ACG TAC ACT CCC TCC CCG AAG CCG ACT CCG CCG ACG CCC AAG CCG ACT CCT CCG ACG CCA
  T Y T P S P K P T P P T P K P T P P T P 140
421 AAG CCC ACC CCG CCG ACG TAC ACT CCG TCG CCG AAG CCT ACC CCG CCG ACA TAC ACT CCC
  K P T P P T Y T P S P K P T P P T Y T P 160
481 ACG CCA AAA CCC ACT CCG CCG ACG GAC ACC CCG GCT CCC AAG CCA CCG GCG ACC AAG CCG
  T P K P T P P T D T P A P K P P A T K P 180
541 CCG ACC TAC ACC CCA ACT CCC AAG CCC ACC CCG CCA TCC TAC ACC CCG TCG CCT AAG CCG
  P T Y T P T P K P T P P S Y T P S P K P 200
601 AAG CCA CCA ACC AAG CCG CCA ACC TAC ACC CCC ACG CCC AAG CCA ACC CCG CCA ACC TAC
  K P P T K P P T Y T P T P K P T P P T Y 220
661 CCG CCG ACG CCC AAG CCC ACT CCA CCG GCG CCG AAG CCG AAG CCC ACT CCC CCG ACC TAC
  P P T P K P T P P A P K P K P T P P T Y 240
721 ACC CCT TCG CCG ACG AAG CCG AAG CCC ACT CCT CCC CCC ACT CCC AAG CCA ACT CCT CCG
  T P S P T K P K P T P P P T P K P T P P 260
781 ACC TAC ACG CCG AAC CCC AAG CCA ACT CCC ACT ACT CCC AAC CCC GAA CCC ACG CCG CCG
  T Y T P N P K P T P T T P N P E P T P P 280
841 ACC TAC AAG CCT GCT CCC AAG CCC ACA CCG CCG TCG TAC AAC AAG CCG CCG GCG GCC TCC
  T Y K P A P K P T P P S Y N K P P A A S 300
901 TAC ACC CCC AGC CCA CCG CCG CCG CAC TAC TAC AAC TGA GATCCGCCGTCTCAAGCTCGGAGCATAG
  Y T P S P P P P H Y Y N - 312

9 6 2 GGTACGTACACAGCACGTCGCAAAACATTTTCTAACCTCTCTGTGTCAGACGCATAT
1082 TCTGTTCACCTTATTATTATCTGAAAAAATGTTTTGCTTAACAAAATGC TGCTCTTTTGT
1142 TTGTTTGTGTTGCAGAAAGGT GATGGAAGACACAGTATAATGTGTGCTAGCCAGCCGGCCA
1202 AGATGAGCTGATGAAGAGAGGCCCTCGAGATCCTCCTCGTGTGTTTTCTGCCGTCCATCT

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Fig. 3 The nucleotide sequence and derived amino acid sequence of the *PM-HRGP* gene. Nucleotides are numbered from the A of the start codon (+1) on the left and amino acids on the right. A putative TATA box is shown in red and underlined. The intron found in the 3'-untranslated region is in italics. The putative site of processing of the

mature protein between Ala 23 and Asp 24 is indicated by a vertical arrow. The C-terminal single Ser (Hyp)₄ motif is highlighted with a box. Repeating tyrosine containing peptide motifs that may play a role in the cross-linking process are underlined (color figure online)

monocotyledon extensins found in sorghum, maize and rice. A single SPPPP sequence characteristic of most HRGPs of monocots except in rice (Caelles et al. 1992) was present in the motif at the C-terminal region of the

protein (Fig. 3). The new gene identified in pearl millet was named as *PM-HRGP*.

Cloning and sequencing of the *PM-HRGP* gene region from genomic DNA and cDNA of pearl millet revealed the

presence of an intron in the 3' UTR of the *PM-HRGP* gene with a size of 127 bp and a splice site starting at 28-bp downstream of the stop codon (Fig. 3). An unrooted phylogenetic tree was constructed based on the comparison of *PM-HRGP* sequences with other *HRGPs* from monocot and dicot species. In the phylogenetic tree, *PM-HRGP* was positioned in a single clade with other monocot extensins and an extensin from *Arabidopsis* in close vicinity (Fig. 4).

Quantitative analysis of *PM-HRGP* gene expression

Quantification of expression of the *PM-HRGP* gene in resistant and susceptible pearl millet samples indicated significant differences in the levels of *PM-HRGP* gene expression. High levels of expression of the gene were observed in resistant inoculated samples from 4 to 8 h after

inoculation with the pathogen. A fourfold increase in the *PM-HRGP* mRNA was observed in the 6 h upon inoculation with the pathogen. In case of susceptible pearl millet cultivar, no significant increase was observed both in control and inoculated seedlings at different time points (Fig. 5).

Discussion

A range of extensin class of *PM-HRGP* polypeptides from suspension cells of pearl millet showed affinity for the JIM 20 monoclonal antibody

The *HRGPs* are a superfamily of glycoproteins in higher plants. The subgroups extensins and proline-rich proteins,

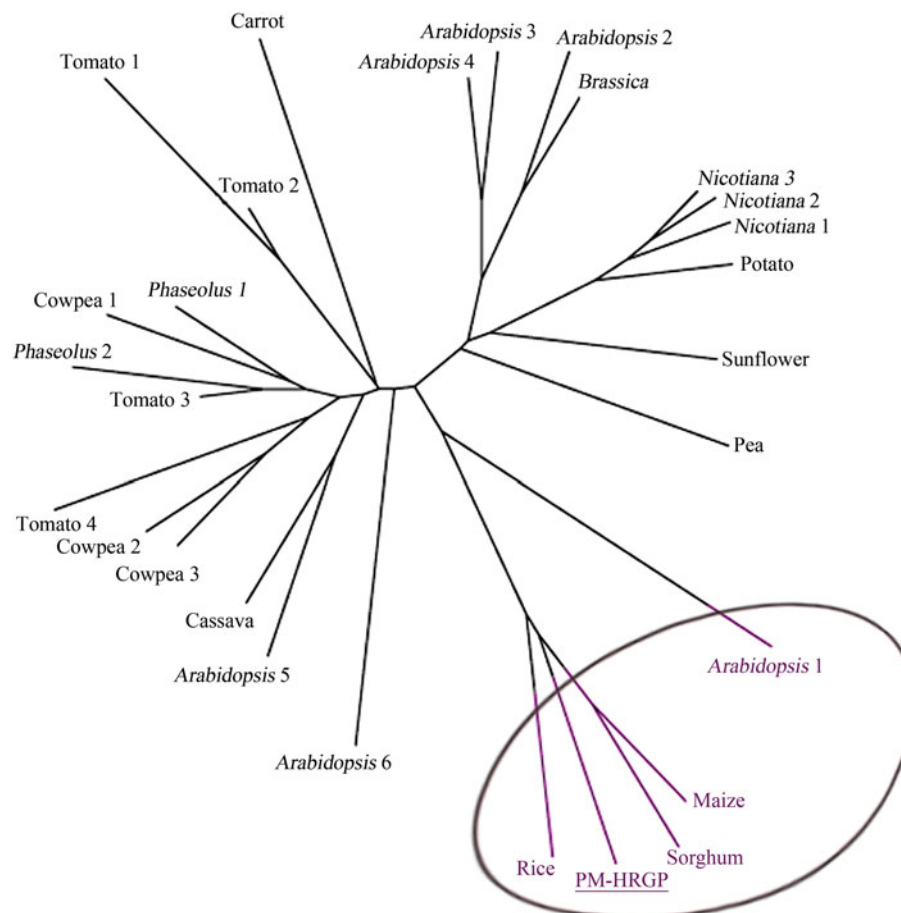


Fig. 4 Phylogenetic status of the *PM-HRGP* gene of pearl millet among other monocot variants of extensin (*HRGP*) and dicot extensins. The *PM-HRGP* was positioned in a single clade with monocot extensin genes and an extensin gene from *Arabidopsis* in the tree. The sequences of 27 plant extensins were obtained from the GenBank database: sorghum (X56010) maize (AJ131535) rice (X61280), cassava (AF239615) *Phaseolus* (M18093) (M18095) carrot (X02873) potato (Z21937) *Brassica* (A18812) Cowpea 1, 2 and 3 (X86029) (X86028) (X91836) *Arabidopsis* 1, 2, 3, 4, 5 and 6

(NM_128298, NM_101983, NM_179564, NM_129877, U43627, NM_122910) sun flower (M76546) *Nicotiana* 1, 2 and 3 (M34371, D13951, X71602) tomato 1,2,3 and 4 (M76671, X55681, X55685, M76670), pea (AF397029). The tree shown is an unrooted, neighbor-joined tree demonstrating the relationships between various members of the *HRGP* extensin family from monocots and dicots. The sequences present in this tree are by no means exhaustive; rather they constitute a representative sampling of sequences available in various databases

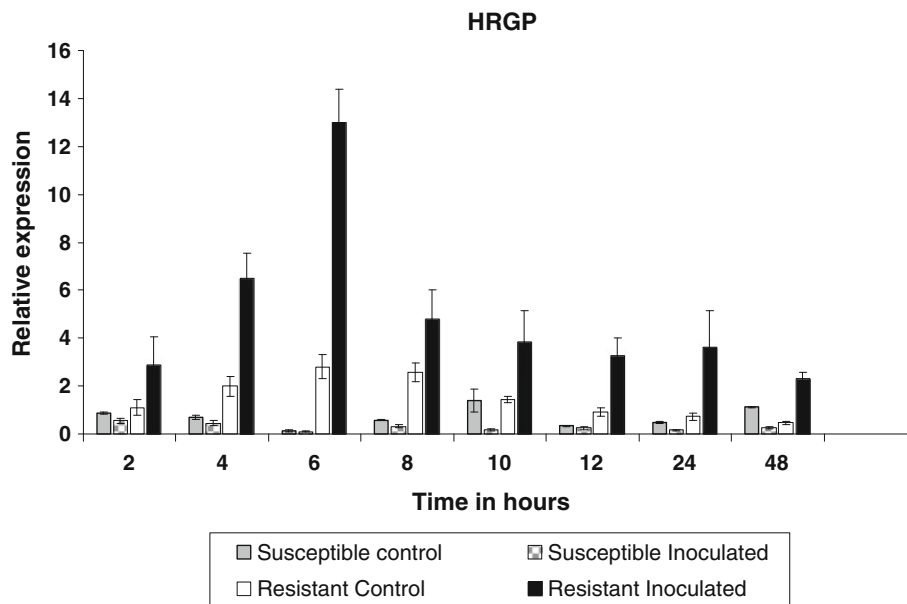


Fig. 5 Relative expression of the *PM-HRGP* gene in susceptible and resistant cultivars of pearl millet before and after challenge inoculation using *S. graminicola*. DNA-free total RNA was isolated from 2-day-old seedlings after pathogen inoculation. Reverse transcription was carried out using oligo(dT) primer. The resulting cDNAs were amplified using gene-specific qRT-PCR primers and fold changes

measured using real-time PCR. Bars represent the expression level of *PM-HRGP* relative to zero-time controls. The values shown are means of three repeats \pm SE (indicated by error bars). *GAPDH* was chosen as an endogenous control during the experiments. Experiments were repeated twice and showed similar results

of the HRGP superfamily may have a role to play in plant cell wall defense (Kieliszewski and Lamport 1994; Jose-Estanyol and Puigdomenech 2000; Seifert and Roberts 2007; Showalter et al. 2010). Our previous research work suggested the involvement of functional HRGPs in successful defense against the phytopathogenic oomycete *S. graminicola* (Shailasree et al. 2004; Sujeeth et al. 2010). We reported a salt-inextractable, proline-rich class of HRGP reacting with the MAC265 monoclonal antibody from pearl millet seedlings (Shailasree et al. 2004; Deepak et al. 2007). MAC 265 was originally isolated by Vandebosch et al. (1989) based on its recognition of legume-specific glycoproteins during pea–rhizobium symbiosis. In the current study, we report the presence of a Tris–SDS extractable extensin class of HRGP protein from suspension cells of pearl millet.

To investigate the presence of an extensin type of HRGP in pearl millet, we selected a panel of extensin-specific monoclonal antibodies. Western blot analysis on the Tris–SDS extracted total protein from pearl millet suspension cells showed an intense reaction with the JIM 20 monoclonal antibody. JIM 20 is a monoclonal antibody, which recognizes the glycan epitope found in pea extensins (Smallwood et al. 1994; Knox et al. 1995). The monoclonal antibody JIM 12 (specific for the glycan/protein epitope) of carrot extensins (Smallwood et al. 1994) did not give any cross-reactivity with any of the polypeptides extracted from pearl millet. Interestingly, the LM1 monoclonal

antibody, specific for the glycan epitope of extensin type of HRGP from rice (Smallwood et al. 1995), was also negative toward pearl millet extensins. The cross-reactivity of pearl millet extensin polypeptides (PM-HRGP) with JIM 20 in the current study and pearl millet PRPs with MAC 265 (Shailasree et al. 2004) suggests that the glycosylation pattern of the HRGP superfamily in pearl millet is closer to the HRGP superfamily of glycoproteins found in pea. A high molecular mass band, probably an extensin protein aggregate or polymer, and a range of lower molecular mass extensin polypeptides were identified using JIM 20. Furthermore, these extensin type of HRGP identified, upon deglycosylation with HF-pyridine, did not give any cross-reaction with JIM 20, thus confirming the specificity of the antibody toward glycan epitopes in PM-HRGP.

Low molecular mass PM-HRGP polypeptides insolubilized rapidly upon treatment of pearl millet cell suspensions with hydrogen peroxide

Our previous studies showed intense accumulation of H_2O_2 within cells close to the parasite haustoria undergoing a hypersensitive-like reaction in resistant pearl millet seedlings. Furthermore, basic isoforms of a peroxidase enzyme were found more in pearl millet upon pathogen attack and elicitor treatments (Sujeeth et al. 2010). In the current study, immunoblot cross-linking analysis using JIM 20 revealed the insolubilization of three low molecular mass

PM-HRGP polypeptides, ranging between 45 and 33 kDa upon 10-min treatment of suspension cells with H₂O₂. Sodium azide is a well-known peroxidase inhibitor. The sodium azide sensitivity for PM-HRGP insolubilization in our analysis suggests that this process is mediated by peroxidases in the cell suspensions of pearl millet. This rapid cross-linking process is hypothesized to slow the spread of the pathogen in plants prior to the activation of transcription-dependent defenses (Lamb and Dixon 1997).

Construction of a fosmid library in pearl millet and isolation of a full-length *PM-HRGP* gene

To investigate the functions of extensin type of HRGP and its responses to the downy mildew pathogen *S. graminicola*, our aim was to isolate the full-length extensin gene in pearl millet. For this, a fosmid genomic library of pearl millet of fourfold haploid equivalents was constructed and screened for the presence of the extensin gene. Screening of the complete genomic library resulted in the isolation of only a single type of *HRGP* gene. The presence of a single gene coding for an extensin type of HRGP has been reported in other monocots like maize (Stiefel et al. 1988), sorghum (Raz et al. 1991) and rice (Caelles et al. 1992). The newly characterized *HRGP* gene from pearl millet was termed as *PM-HRGP*.

The *PM-HRGP* gene in pearl millet has all the characteristic features of a monocotyledon extensin type of *HRGP*

A potential TATA box was present at position –104 bp from the start site in the upstream region of the *PM-HRGP* gene. The 500-bp region upstream to the transcription start site of *PM-HRGP* showed 60% homology with the upstream element of the maize extensin gene. The upstream regulatory sequences of the *HRGP* genes responsible for tissue specificity, wound inducibility and activation during infection have been extensively studied in different species such as maize, carrot, tobacco and oilseed rape (Jose and Puigdomenech 1993). Further studies directed toward the identification of the regulatory elements present in the *PM-HRGP* gene could lead to a better understanding of its role during stress, pathogen attack and elicitor treatment.

Sequence comparison from the 3' UTR of the *PM-HRGP* gene amplified from cDNA and genomic DNA of pearl millet showed the presence of a 127-nucleotide length intron downstream of the stop codon. A similar type of intron flanked by AGgt/agGT intron consensus sequences has been reported in maize and sorghum (Stiefel et al. 1990; Raz et al. 1991). In maize, this intron in the 3' UTR is reported to affect gene expression in a tissue- and promoter-dependent manner (Menossi et al. 2003). Our cDNA

cloning study showed that the intron flanking sequences in *PM-HRGP* are not similar to those found in maize and sorghum.

The predicted protein from the *PM-HRGP* gene showed a similarity of 54–67% with other monocot extensins. The presence of nine repeats of the hexapeptide PPTYTP and eight repeats of the pentapeptide TPKPT were observed, which were similar to those found in maize extensin (Stiefel et al. 1988). However, only a single SPKPP sequence was observed in PM-HRGP as compared to the maize extensin type of HRGP, which had 12 repeats of this sequence. Variants of all the above three kind of repeats were also observed in the PM-HRGP sequence. Several of these motifs are usually known to be present in other monocot extensins (Sommer-Knudsen et al. 1998). Proline-rich repetitive peptide motifs and their variants, singly or in combination present in the extensin gene, may constitute functional sites involved in various aspects of cell wall assembly (Kieliszewski and Lamport 1994). The post-translational modifications like hydroxylation of prolines, glycosylation and cross-linking establish the structural and functional role of this protein in the plant cell wall. It is now assumed that repetitive HRGP polypeptides themselves are the primary determinants for the hydroxylation and glycosylation (Jamet et al. 2008).

The predicted protein from the *PM-HRGP* gene starts with a putative signal peptide of 24 amino acids with a cleavage site between Ala and Asp, based on the rules of von Heijne (1986), indicating an extracellular localization of the gene product. This putative cleavage site has also been observed in maize and sorghum extensins (Stiefel et al. 1988; Raz et al. 1991). It has been reported that the N-terminal signal sequence of HRGPs helps in targeting the polypeptide to the ER/Golgi for post-translational processing (Kieliszewski 2001). The C-terminal domain with a single Ser-(Hyp₄) motif characteristic of all members of the monocot extensin group was also found in the PM-HRGP confirming the presence of a Gramineae HRGP subfamily belonging to the plant cell wall HRGP family. A phylogenetic tree constructed revealed that the PM-HRGP formed a separate clade along with other monocot extensin type of HRGP genes.

Tyrosines containing YXY repeats were observed in most of the extensin protein sequences. Such sequences are involved in cross-linking (Fry 1986; Held et al. 2004). It had been shown in vitro that coupling of extensin tyrosine residues, to form mainly isodityrosine (IDT) and pulcherosine, is a peroxidase-mediated process involving H₂O₂ (Brady et al. 1996,1998; Cannon et al. 2008). This may help in the formation of an extensin network and strengthen the cell wall in arresting pathogen ingress (Cassab 1998; Sommer-Knudsen et al. 1998; Jose-Estanyol and Puigdomenech 2000). Increased accumulation of IDT in the cell wall

hydrolysate has also been reported in pearl millet seedlings infected with *S. graminicola* (Deepak et al. 2007). The predicted protein sequence of the *PM-HRGP* gene characterized in the present study has only a single YGY motif near the N-terminus that is commonly involved in extensin cross-linking. On the other hand, several repeating tyrosine residues mainly in the motif PPTYTP and also its variants such as PPTYKP and PPSYTP are also found. It remains to be elucidated if these repeating motifs have a role in the formation of stable tyrosine cross-links between different polypeptides in the pearl millet cell walls.

A differential expression study provides insight into up-regulation of *PM-HRGP* transcripts, observed only in resistant cultivars of pearl millet during downy mildew infection

Accumulation of HRGP takes place in maize during wounding (Ludevid et al. 1990), arbuscular mycorrhizal symbiosis (Balestrini et al. 1997) and in response to fungal elicitors and infection (Showalter et al. 1985). This phenomenon was not observed during wounding in other monocots such as barley, oat, wheat or rice (Mazau and Esquerretugaye 1986; Li and McClure 1990; Sommer-Knudsen et al. 1998). The quantitative real-time PCR indicated an increased expression level of the *PM-HRGP* gene in the resistant cultivar from 4 to 8 h after inoculation with the pathogen, showing a fourfold increase at the 6th hour. The expression remained at basal level in the susceptible cultivar of pearl millet upon pathogen infection. The present study confirmed the putative role of the *PM-HRGP* gene in the defense reaction of pearl millet against downy mildew. Accumulation of the *PM-HRGP* transcripts was observed only in the resistant cultivar after inoculation with the pathogen.

Concluding remarks

The failure of pathogenic fungi to breach the plant cell wall constitutes a major component in plant defense. Our findings suggest that accumulation of an extensin class of HRGP in resistant cultivars of pearl millet in response to *S. graminicola* soon after inoculation and their possible cross-linking property in the presence of H₂O₂ may offer effective restriction for the downy mildew pathogen. Furthermore, we characterized a full-length gene coding for an extensin type of HRGP, *PM-HRGP* in pearl millet. The understanding of the mechanism of regulation of this gene will have importance in downy mildew disease management, especially to screen and identify resistant genotypes. In addition, biotechnological applications could improve pearl millet by providing increased resistance against downy mildew.

Compared to other important cereals such as rice, maize or sorghum, limited molecular studies have been carried out and only a few genes have been characterized in pearl millet with its huge genome size of 2,352 Mb. The complex array of defense-related genes playing a major role is yet to be characterized in this plant system. A pearl millet fosmid library covering four haploid genome equivalents in the inbred 7042S cultivar has been constructed in our study and is available on request. This library will be a valuable tool for several molecular marker-based studies in pearl millet and also to identify gene candidates involved in the plant defense mechanism.

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