Metabolites and bioactivities of Rhizophoraceae mangroves

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Abstract: This review examines the chemical compositions and bioactivities of mangrove plants belonging to the Rhizophoraceae family. The Rhizophoraceae family of true mangrove plants is the most common and is also widely distributed species. It consists of 24 species across four genera. Of the 24 species, 12 species remain unexamined for their phytochemical constituents. There have been 268 metabolites reported from 16 species. The key phytochemical constituents identified across the family are the diterpenoids and triterpenoids. The major diterpenoids include pimaranes, beyeranes, kaurenes, dolabranes and labdanes whereas the significant triterpenoids are lupanes, dammaranes and oleananes. Disulphides, dolabranes and labdanes are considered to be the chemotaxonomic markers of the genera *Bruguiera, Ceriops* and *Rhizophora* respectively.

Keywords: Rhizophoraceae, Bruguiera, Rhizophora, terpenoids, Ceriops

Introduction

Mangrove plants are potential sources of biologically active chemicals that are discernible from their wide spread application in ethnopharmaceutical practices. There habitat exists under stressful conditions and serve as a bridging ecosystem between freshwater and marine systems. These plants have specially adapted their own morphological structures and physiological mechanisms to their harsh natural surroundings. Pneumatophores, stilt roots and buttresses, with salt-excreting glands found in their leaves, and viviparous propagules are some of the several highly specialized adaptations of this group. The path of photosynthesis in mangroves is different from other glycophytes. Furthermore, there are alterations in other physiological processes such as carbohydrate metabolism or polyphenol synthesis. These plants survive under extreme conditions of salinity, temperature gradients, tidal fluctuations and anoxic soil conditions, with these plants possessing many chemical compounds, which protect them from these destructive elements¹. Even though extracts from mangroves and mangrove-dependent species possess therapeutic activity against humans, animal and plant pathogens, the specific metabolites responsible for these bioactivities remains to be elucidated.

1 Rhizophoraceae

The global mangrove plant have 84 species belonging to 24 genera and 16 families². Among them, 70 species are true mangroves pertaining to sixteen genera and eleven families whereas fourteen species are semi mangroves belonging to eight genera and five families. According to Wu et. al, suggests the family Rhizophoraceae belongs to true mangrove family, which contains 21 species in four genera². In contrast three more species; *Rhizophora annamalayana*³, *Kandelia obovata*⁴ and Ceriops zeppeliana blume5 become 24 species in four genera in the Rhizophoraceae family of true mangroves. Thus the family Rhizophoraceae include: Bruguiera which contains seven species, Ceriops (five species), Kandelia (two species) and Rhizophora (ten species). The distribution of species in Rhizophoraceae family is detailed in Table 1. 54 studies can achieve the validity of ethnomedicines as well as apply the use of mangrove plants in the development of new drugs.

In this review, the compounds identified from this family were listed, and their reported biological activities were compiled. Also chemotaxonamy and importance of further phytochemical research is discussed.

2 Chemical Constituents

2.1 Bruguiera: The genus Bruguiera has six species and one hybrid species which are derived from *B. sexangula*, including *B. cylindrica*, *B. exarista*, *B. gymnorrhiza*, *B. hainessi*, *B. parviflora*, *B. sexangula* and *B. sexangula* var.



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Table 1. Mangroves of Rhizophoraceae family of true mangroves

Bruguiera	Ceriops	Kandelia	Rhizophora
B. cylindrica	C. decandra	K. candel	R. apiculata
B. exarista	C. tagal	K. obovata	R. harrisoni
B. gymnorrhiza	C. tagal var. australasica		R. lamarckii
B. hainessi	C. tagal var. typical		R. mangle
B. parviflora	C. zeppeliana		R. mucronata
	blume		
B. sexangula			R. racemosa
B. sexangula var.			R. samoesis
rhynchopetala			
			R. selala
			R. stylosa
			R. annamalayana

rhynchopetala. The metabolic pattern of this genus has been extensively characterised by a suite of diterpenes and triterpenes. In addition, these species also produce flavonoids, tropane derivatives and cyclic polysulphides. These include 22 metabolites from B. cylindrica, 54 metabolites from B. gymnorrhiza, nine metabolites from B. exaristata, six metabolites from B. parviflora, two metabolites from B. sexangula and 40 metabolites from B. sexangula var rhynchopetala were identified so far. A total of 114 metabolites have been reported form this genus including. A detailed list of chemical compounds identified from Bruguiera is recorded in Table 2.

Three sulphur compounds along with an alkaloid brugine were reported from the stem and bark of B. cylindrica by Japanese scientists of during 1975-1976. 20 years later, a number of oleananes and lupanes (triterpenoids) and one kaurane (diterpenoid) were reported. The first report on the chemical constituents of B. gymnorrhiza dates back to 1978 in which Sarkar and Ganguly reported a new triterpenoid called gymnorhizol (3-epi- δ -amyrin). Since then various triterpenoids (lupanes, oleananes, ursanes, dammaranes) and diterpenoids (kauranes, pimaranes, beyaranes) along with sulphur compounds, sterols and aromatic compounds were reported from this plant.

Only two reports are available regarding the chemical constituents of B. exaristata. As part of their investigation on tumor inhibitory plants, in 1969 Loder and Russell identified the presence of alkaloids (brugine, tropine and tropine esters of acetic, iosbutyric, iosvaleric, propionic, n-butyric and benzoic acids) in the bark extracts of *B. exaristata*⁶ while a pronounced accumulation of 1-d-1-O-methyl-muco-inositiol in the young leaves of B. exaristata was reported later by Richter and his team⁷.

In a continuing search for bioactive constituents from Thai medicinal plants, Chumkaew and his team isolated and elucidated a new triterpenoid ester 3-(Z)-caffeoyllupeol along





with five other tritepenoids; lupeol caffeate, 3-(Z)coumaroyllupeol, dioslupecin A, lupeol and lupenone from the fruits of *B. Parviflora*⁸. The earliest work regarding the chemical constituents of mangroves deals with the isolation and characterization of the tropine 1,2-dithiolane-3carbonylate named as bruguine from the stem bark of B. sexangula⁹. Later the same team identified additional alkaloids from the same plant as part of their investigation on tumor inhibitory plants⁶. In a study focusing on the marine fauna and flora from Chinese coasts, Li and his coworkers collected samples of the mangrove B. sexangula from Hainan Province, China. On separation of an EtOAc-soluble fraction of a methanol extract of the title plant, they isolated a new triterpene, named sexangulic acid¹⁰.

Investigation of Chinese mangrove plants led to the isolation and characterisation of 13 compounds; three new diterpenes; six known diterpenes, a new dithiobenzoquinone two cyclic disulfides and 2,6-dimethoxy-1,4-benzoquinone from the EtOH extract of the stem of B. sexangula var. *rhynchopetala*¹¹. Further several triterpenoids and sterols were reported¹². Recently, a continuous investigation for chemical diversity of B. sexangula var. rhynchopetala led to the isolation and characterization of six new phenolic constituents named rhyncosides A-F, together with twelve known compounds including two phenolic glycosides, four flavonoids, and six lignan derivatives¹³

Alkaloids



Figure 1. Alkaloids from Rhizophoraceae mangroves

2.2 Ceriops: The genus of Ceriops has two species and two varieties, namely C. tagal (Perr.), C. decandra, C. tagal var. australasica and C. tagal var. typical. These plants are valued for their rich tannin content and are a rich source of pentacyclic triterpenoids¹⁷. To date, 30 metabolities from C. decandra and 72 metabolites from C. tagal are known. Thus a total of 92 metabolites including 45 diterpenoids (23 dolabranes, six dimeric diterpenoids, four beyeranes, five kauranes, and seven pimaranes) and 45 tritepenoids (35 lupanes, seven dammaranes, one oleanane, one ursane, and one abietane) along with two steroids have been reported so far from this genus.

On examination of the roots of C. decandra collected from the Kauvery estuary (Parangipettai coast), Anjaneyulu and his team isolated and characterised twelve diterpenoids^{30,31,32} Subsequently, two novel triterpene esters were isolated from the leaves of C. decandra in addition to 16 known triterpenes³³

by scientists from Thailand. Dolabranes (diterpenoids) are the marker metabolites of C. tagal. These compounds can be used as chemotaxonomic markers of this plant. Dimeric diterpenoids (tetraterpenoids) and triterpenoids of lupane, dammarane, pimarane groups are also found in this plant. One abietane and an oleanane triterpenoid were also isolated from the stems and twigs of C. tagal. The chemical constituents identified from *ceriops* are listed in Table 3.

Table 2. Chemical constituents from the genus Bruguiera				
Compound Class and Name	Plan	nt C C C	Plant Part	References
Alkaloids				
brugine (1)	<i>B. cy</i>	ylindrica	stem and bark	14
	В. ех	xaristata	stem bark	6
tropine (2)	В. ех	xaristata	stem bark	6
	B. se	exangula	stem bark	6
tropine acetate (3)	B. ex	xaristata	stem bark	6
	B. se	exangula	stem bark	6
tropine benzoate (4)	В. ех	xaristata	stem bark	6
	B. se	exangula	stem bark	6
tropine isobutyrate (5)	B. ex	xaristata	stem bark	6
	B. se	exangula	stem bark	6
tropine isovalerate (6)	В. ех	xaristata	stem bark	6
	B. se	exangula	stem bark	6
tropine n-butyrate (7)	В. ех	xaristata	stem bark	6
	B. se	exangula	stem bark	6
tropine propionate (8)	В. ех	xaristata	stem bark	6
	B. se	exangula	stem bark	6
D-Friedooleananes (Triterpenoids)				
3α -taraxerol (9)	<i>B. c</i> y	ylindrica	fruits	15
3α - <i>E</i> -caffeoyltaraxerol (10)	B. cy	ylindrica	fruits and hypocotyls	16
3α - <i>E</i> -coumaroyltaraxerol (11)	B. cy	ylindrica	fruits	15
3α -E-feruloyltaraxerol (12)	B. cy	ylindrica	fruits	15
3α -Z-coumaroyltaraxerol (13)	B. cy	ylindrica	fruits	15
3α -Z-feruloyltaraxerol (14)	B. cy	ylindrica	fruits	15
3β -taraxerol (26)	B. cy	ylindrica	fruits	15
3β -E-ferulovltaraxerol (17)	B. cv	vlindrica	fruits	15
3β -Z-feruloyltaraxerol (23)	B. cy	ylindrica	fruits	15
taraxerone (25)	B. se	exangula var. rhynchopetala	stem	12
Lupanes (Triterpenoids)				
3α -lupenol (32)	В. сч	vlindrica	fruits and hypocotyls	16
3α - <i>E</i> -coumaroyllupeol (34)	B. cv	vlindrica	fruits and hypocotyls	16
3α -Z-coumarovIlupeol (37)	B. cv	vlindrica	fruits and hypocotyls	16
3β -E-caffeovllupeol B (44)	B. cv	vlindrica	fruits and hypocotyls	16
3β -Z-caffeovllupeol (38)	B. pc	arviflora	fruits	8
3β - <i>E</i> -coumaroyllupeol (45)	B. cy	ylindrica	fruits and hypocotyls	16
3β -Z-coumaroyllupeol (55)	B. cv	vlindrica	fruits and hypocotyls	16
	B. pc	arviflora	fruits	8
betulin (57)	B. g	vmnorrhiza	leaves	17
lupenone (64)	B. cv	vlindrica	fruits and hypocotyls	16
	B. se	exangula var. rhvnchopetala	stem	12
	B. no	arviflora	fruits	8
lupeol (65)	B. cv	vlindrica	fruits and hypocotyls	16
1	B. 91	ymnorrhiza	leaves	17
	B. se	exangula var. rhvnchopetala	stem	12
	B. pa	arviflora	fruits	8
trans-hydroxy-cinnamoyl lupeol (66)	B. se	exangula var. rhvnchonetala	stem	12
dioslupecin A (57)	B. n	arviflora	fruits	8
Oleanane (Triterpenoids)	pt	······		-





oleanolic acid (70)	B gymnorrhiza	leaves	17
β -amyrin (71)	B. gymnorrhiza	leaves	17
β -amyril palmitate (72)	<i>B. sexangula</i> var. <i>rhvnchopetala</i>	stem	12
Ursanes (Triterpenoids)			
ursolic acid (73)	B. gymnorrhiza	leaves	17
α -amyrin (74)	B. gymnorrhiza	leaves	17
Dammaranes (Triterpenoids)			
bruguierin A (75)	B. gymnorrhiza	flowers	18
bruguierin B (76)	B. gymnorrhiza	flowers	18
bruguierin C (77)	B. gymnorrhiza	flowers	18
Triterpene alcohol			
gymnorhizol (3- <i>eni</i> - δ -amyrin) (85)	B. gymnorrhiza	leaves	19
Lanostanes (Triterpenoids)			
sexangulic acid (86)	R sexangula	stem	10
squalene (Triternenoid)	D. Sexungulu	Stelli	10
squalene (87)	R sexangula var rhvnchopetala	stem	12
Fatty acids	D. sexungula val. mynenopelala	Stelli	12
linoleic acid (88)	R symnorrhiza	leaves	20
linolenic acid (89)	B. gymnorrhiza R gymnorrhiza	leaves	20
nalmitic acid (90)	B. gymnorrhiza R gymnorrhiza	leaves	20
Steroids	D. Symnorrinza	icuves	20
3-0-a-L-rhamponyranosyl-(+)-catechin-	R commorrhiza	bark	21
$(4q \rightarrow 2)$ phloroglucinol (91)	D. gymnorrni2u	ourk	21
campesterol (92)	R commorrhiza	leaves	17
cholesterol (93)	B. gymnorrhiza R. gymnorrhiza	leaves	17
daucosterol (94)	B. gymnormizu R. sevangula var rhvnchonetala	stem	17
$\mathcal{B}_{\text{-sitosterol}}(94)$	B. sexangula var rhynchopetala	stem	12
p -situsteror (\mathbf{y})	B. sexungulu val. mynenopelala B. gymnorrhiza	leaves	12
stigmaste-7-en-3B-ol (98)	B. gymnorrhiza B. gymnorrhiza	leaves	17
stigmasterol (90)	B. gymnorrhiza	leaves	17
a-hydroxy-sitesterol (100)	B. sevangula var rhvnchonetala	stem	17
Kauranes (Diterpenoids)	D. sexungulu val. mynenopelalu	Stelli	12
(16 <i>R</i>)-13,17-epoxy-16-hydroxy- <i>ent</i> -kaur-9(11)-en-	B. sexangula var. rhynchopetala	stem	11
19-al (101)	- · ·		
13,16,17-trihydroxy- <i>ent</i> -9(11)-kaurene-19-oic acid	B. gymnorrhiza	stem	22
(102)	D 1.		22
13-hydroxy-16-ent-kauren-19-al (103)	B. gymnorrhiza	stem	22
	B. gymnorrhiza	bark	23
16,17-dihy-droxy-ent-9(11)-kaurene-19-al (107)	B. gymnorrhiza	stem	22
	B. sexangula var. rhynchopetala	stem	11
16,17-dihydroxy-ent-9(11)-kauren-19-oic acid (108)	B. gymnorrhiza	stem	22
16,17-dihydroxy-19-nor- <i>ent</i> -kaur-9(11)-en-3-one (109)	B. sexangula var. rhynchopetala	stem	11
16-ent-kaurene-13,19-diol (115)	B. gymnorrhiza	stem	22
	B. gymnorrhiza	bark	23
	B. cylindrica	roots	24
16-ent-kauren-19-ol (110)	B. gymnorrhiza	stem	22
16H-17,19-ent-kauranediol (104)	B. gymnorrhiza	stem	22
16H-17-hydroxy-ent-kauran-19-oic acid (105)	B. gymnorrhiza	stem	22
16,17-dihydroxy- <i>ent</i> -kauran-19-al (106)	B. gymnorrhiza	stem	22
17-chloro-13,16-dihydroxy-ent-kauran-19-al (111)	B. gymnorrhiza	stem	22
ceriopsin F (113)	B. sexangula var. rhynchopetala	stem	11
steviol (120)	B. gymnorrhiza	bark	23





$\begin{array}{c} B. \ gymnorrhiza \\ methyl-16a, 17-dihydroxy-ent-kaur-9(11)-en-19-oate \\ (116) \\ B. \ gymnorrhiza \\ B. \ gymnorrhiza \\ B. \ gymnorrhiza \\ B. \ gymnorrhiza \\ Stem \\ 22 \\ \hline \\ Pimaranes (Diterpenoids) \\ 15(S)-isopimar-7-en-15, 16-diol (123) \\ B. \ gymnorrhiza \\ Stem \\ 25 \\ B. \ gymnorrhiza \\ Stem \\ 25 \\ B. \ gymnorrhiza \\ Stem \\ 25 \\ ent-8(14)-pimarene-15R, 16-diol (128) \\ B. \ gymnorrhiza \\ Stem \\ 25 \\ ent-8(14)-pimarene-1a, 15R, 16-triol (129) \\ Isopimar-7-ene-1\beta, 15R, 16-triol (129) \\ B. \ gymnorrhiza \\ Stem \\ 25 \\ (5R,9S, 10R, 13S, 15S)ent-8(14)-pimarene-1-oxo- \\ 15R, 16-diol (122) \\ (1aH, 15R)-ent-pimar-8(14)-ene-1, 15, 16-triol (121) \\ B. \ gymnorrhiza \\ Stem \\ 25 \\ Stem \\ 21 \\ Stem \\ 22 \\ Stem \\ 21 \\ Stem \\ 21 \\ Stem \\ 21 \\ Stem \\ 22 \\ Stem \\ 21 \\ Stem \\ 21 \\ Stem \\ 21 \\ Stem \\ 22 \\ Stem \\ 21 \\ Stem \\ 22 \\ Stem \\ 22 \\ Stem \\ 22 \\ Stem \\ 21 \\ Stem \\ 21 \\ Stem \\ 21 \\ Stem \\ 22 \\ Stem \\ 31 \\ Stem \\$
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isopimar-7-ene-1 β ,15R,16-triol (130)B. gymnorrhizastem25(5R,9S,10R,13S,15S)ent-8(14)-pimarene-1-oxo- 15R,16-diol (122)B. gymnorrhizastem25(1 α H,15R)-ent-pimar-8(14)-ene-1,15,16-triol (121)B. sexangula var. rhynchopetalastem11Beyeranes (Diterpenoids)B. gymnorrhizastem22(4R,5S,8R,9R,10S,13S)-ent-17-hydroxy-16- oxobeyeran-19-al (135)B. gymnorrhizastem22ent-17-hydroxy-16-oxobeyer-9(11)-en-19-al (136)B. sexangula var. rhynchopetalastem11
$\begin{array}{cccccccccccccccccccccccccccccccccccc$
15R,16-diol (122) Image: 15R,16-diol (121) B. sexangula var. rhynchopetala stem 11 Beyeranes (Diterpenoids) Image: 15R,16-triol (121) B. sexangula var. rhynchopetala stem 12 (4R,55,8R,9R,10S,13S)-ent-17-hydroxy-16- B. gymnorrhiza stem 22 oxobeyeran-19-al (135) Image: 11 Image: 11 ent-17-hydroxy-16-oxobeyer-9(11)-en-19-al (136) B. sexangula var. rhynchopetala stem 11
$\begin{array}{c c c c c c c c c c c c c c c c c c c $
Beyeranes (Diterpenoids)B. gymnorrhizastem22(4R,5S,8R,9R,10S,13S)-ent-17-hydroxy-16- oxobeyeran-19-al (135)B. gymnorrhizastem11ent-17-hydroxy-16-oxobeyer-9(11)-en-19-al (136)B. sexangula var. rhynchopetalastem11
(4R,5S,8R,9R,10S,13S)-ent-17-hydroxy-16- B. gymnorrhiza stem 22 oxobeyeran-19-al (135) ent-17-hydroxy-16-oxobeyer-9(11)-en-19-al (136) B. sexangula var. rhynchopetala stem 11
oxobeyeran-19-al (135) ent-17-hydroxy-16-oxobeyer-9(11)-en-19-al (136) B. sexangula var. rhynchopetala stem 11
ent-17-hydroxy-16-oxobeyer-9(11)-en-19-al (136) B. sexangula var. rhynchopetala stem 11
Sulphur compounds
4-hydroxy-1,2-dithiolane (144)B. cylindricastem and bark14
brugierol (145) B. cylindrica stem and bark 14
B. gymnorrhiza flowers, leaves and 26, 27
stem
B. sexangula var. rhynchopetala stem 11
bruguiesulfurol (146) B. gymnorrhiza Flowers, leaves and 26, 27
stem
<i>cis</i> -3,30-dihydroxy-1,5,10,50-tetrathiacyclodecane <i>B. gymnorrhiza</i> leaves and stem 27
(147)
gymnorrhizol (148) B. gymnorrhiza leaves and stem 27
isobrugierol (149) B. cylindrica stem and bark 14
<i>B. gymnorrhiza</i> flowers, leaves and 26, 27
stem
B. sexangula var. rhynchopetala stem 11
neogymnorrhizol (150) B. gymnorrhiza leaves and stem 27
<i>trans-3,30-dihydroxy-1,5,10,50-tetrathiacyclodecane B. gymnorrhiza</i> leaves and stem 27
(151)
(-)-3,4-dihydro-3-hydroxy-/-methoxy-2H-1,5- B. sexangula var. rhynchopetala stem 11
Anometic company de
Aromatic compounds
1-(5-hydroxyphenyl)-hexane-2,5-diol (155) B. gymnorrniza stem 28
2,5-dimethoxy-5-propylphenol (154) B. gymnorrniza Dranch 29
3-(3-nyaroxybuty1)-1,1-almetinyiisochroman-6,8-aloi B. gymnorrniza stem 28
(155) hereanised A (156) B commonwhing stom 29
bruguierol A (150) B. gymnorrhiza stem 28
bruguierol G (157) D. gymnorrhiza stem 28
bruguierol D (150) D. gymnorrhiza bronch 20
Carbohydrates
Lad-1-Q-methyl much inositial (161) B gravistata leaves 7
Renzoquinone
2 6-dimethoxy-1 4-henzoquinone (162) B sexangula var rhynchonetala stem 11
Phenolic glycosides





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1-[<i>α</i> -L-rhamnopyranosyl-(1→6)- <i>β</i> -D-	B. sexangula var. rhynchopetala	stem	13
glucopyranosyloxy]-3,4,5-trimethoxybenzene (163)			
3,4,5-trimethoxyphenyl- β -D-glucopyranoside (164)	B. sexangula var. rhynchopetala	stem	13
rhyncoside A (168)	B. sexangula var. rhynchopetala	stem	13
rhyncoside B (169)	B. sexangula var. rhynchopetala	stem	13
rhyncoside C (170)	B. sexangula var. rhynchopetala	stem	13
rhyncoside D (171)	B. sexangula var. rhynchopetala	stem	13
Flavonoids			
myricetin-3-O-rutinoside (199)	B. sexangula var. rhynchopetala	stem	13
nicotiflorin (200)	B. sexangula var. rhynchopetala	stem	13
rutin (205)	B. sexangula var. rhynchopetala	stem	13
tricin (206)	B. sexangula var. rhynchopetala	stem	13
Lignans			
(+)-5'-methoxyisolariciresinol-9'-β-D-	B. sexangula var. rhynchopetala	stem	13
xylopyranoside (207)			
(+)-lyoniresinol- 3α - O - α -L-rhamnopyranoside (208)	B. sexangula var. rhynchopetala	stem	13
brugunin A (209)	B. gymnorrhiza	branch	29
hedyotisols A (210)	B. sexangula var. rhynchopetala	stem	13
hedyotisols B (211)	B. sexangula var. rhynchopetala	stem	13
hedyotisols C (212)	B. sexangula var. rhynchopetala	stem	13
lyoniside (213)	B. sexangula var. rhynchopetala	stem	13
rhynocoside E (214)	B. sexangula var. rhynchopetala	stem	13
rhynocoside F (215)	B. sexangula var. rhynchopetala	stem	13

Table 3. Chemical constituents of Ceriops

Compound Class and Name	Plant	Plant Part	References
Dolabranes (Diterpenoids)			
tagalsin A (223)	Ceriops tagal	stem and twigs	34
		aerial parts	35
tagalsin B (224)	C. tagal	stem and twigs	34
		aerial parts	35
tagalsin C (225)	C. tagal	stem and twigs	34
		aerial parts	35
		roots	36
tagalsin D (226)	C. tagal	stem and twigs	34
tagalsin E (227)	C. tagal	stem and twigs	34
		aerial parts	35
tagalsin F (228)	C. tagal	stem and twigs	34
		aerial parts	35
		roots	37, 38
tagalsin G (229)	C. tagal	stem and twigs	34
		aerial parts	35
tagalsin H (230)	C. tagal	stem and twigs	34
tagalsin O (231)	C. tagal	aerial part	35
tagalsin P (232)	C. tagal	stems and twigs	38
tagalsin Q (233)	C. tagal	stems and twigs	38
tagalsin R (234)	C. tagal	stems and twigs	38
tagalsin S (235)	C. tagal	stems and twigs	38
tagalsin T (236)	C. tagal	stems and twigs	38
tagalsin U (237)	C. tagal	stems and twigs	38
(5 <i>S</i> *,8 <i>S</i> *,9 <i>S</i> *,10 <i>R</i> *,13 <i>S</i> *)-3-hydroxy-16-nor-2-	C. tagal	stems and twigs	38
oxodolabr-3-en-15-oic acid (219)			
(5 <i>S</i> *,8 <i>S</i> *,9 <i>S</i> *,10 <i>R</i> *,13 <i>S</i> *)-3,16-dihydroxydolabar-3-	C. tagal	stems and twigs	38
ene-2,15-dione (218)			





(5 <i>S</i> *,8 <i>S</i> *,9 <i>S</i> *,10*,13 <i>S</i> *)-2-hydroxy-16-nor-3-	C. tagal	stems and twigs	38
oxodolabr-1,4(18)dien-15-oic acid (217)			
$(5S^*, 8S^*, 9S^*, 1R^*, 13S^*)$ -dolabr-3-ene-15-, 16-diol	C. tagal	stems and twigs	38
(220)			
(5 <i>S</i> *,8 <i>S</i> *,9 <i>S</i> *,10 <i>R</i> *,13 <i>S</i> *)-dolabr-4(18)-ene-15,16-	C. tagal	stems and twigs	38
diol (216)			
erythroxydiol Y (222)	C. tagal	stems and twigs	38
dolabr-4(17),15(16)-dien-3-one (238)	C. tagal	roots	39
7-glycoloyl-2-hydroxy-1,4b,7,10a-tetramethyl-	C. tagal	roots	40
4a,4b,5,6,7,8,8a,9,10,10a-decahydrohenanthren-			
3(4 <i>H</i>)-one (239)			
Dimeri diterpenoids	~ .		
tagalsns I (244)	C. tagal	stems and twigs	41
	C. tagal	roots	36
tagalsins J (245)	C. tagal	stems and twigs	41
tagalsin L (241)	C. tagal	roots	42
tagalsin M (242)	C. tagal	roots	42
tagalsin N (243)	C. tagal	roots	42
8(14)-enyl-pimar-2'(3')-en-4'(18')-en-	C. tagal	roots	36
15'(16')endolabr-16,15,2',3'-oxoan-16-one (240)			
Beyranes (Diterpenoids)			
ceropsin A (137)	C. tagal	roots	30
ceiopsin B (138)	C. tagal	roots	30
criopsin G (139)	C. tagal	roots	31
isosteviol (140)	C. tagal	roots	31
Auranes (Diterpenoids)			
cerioprin E (112)	C. tagal	roots	32
ceriopsin F (113)	C. tagal	roots	31
steviol (120)	C. tagal	roots	31
methyl-ent- 16β ,17-dihydroxy-9(11)-kauren-19-oate	C. tagal	roots	31
(118)			
<i>ent</i> -16 β ,17-dihydroxy-9(11)-kauren-19-oic acid	C. decandra	roots	31
Dammarane (Triterpenoids)			10
cereotagalol A (78)	C. tagal	hypocotyls and fruits	43
cereotagalol B (79)	C. tagal	hypocotyls and fruits	43
cereotagaloperoxide (80)	C. tagal	hypocotyls and fruits	43
dammarenediol II (81)	C. tagal	hypocotyls and fruits	43
fouquierol (82)	C. tagal	hypocotyls and fruits	43
isofouquierol (83)	C. tagal	hypocotyls and fruits	43
	C. tagal	hypocotyls and fruits	43
Oleananes (Triterpenoids)	~ .		10
oleanolic acid (70)	C. tagal	hypocotyls and fruits	43
ursane (Triterpenoid)			
ursolic acid (73)	C. decandra	leaves	33
Lupanes (Triterpenoids)			
28-hydroxylup-20(29)-en-3-one (27)	C. tagal	aerial parts	44
	C. tagal	roots	36
3- <i>epi</i> -betulin (28)	C. tagal	aerial parts	44
$30\text{-nor-lup-}3\beta\text{-ol-}2\text{-one}(29)$	C. decandra	leaves	33
3-epi-betulinic acid (30)	C. tagal	aerial parts	44
3-oxo-lup-20(29)-en-28-oic acid (31)	C. tagal	roots	36
3α -betulinic acid (33)	C. tagal	hypocotyls and fruits	43
	C. decandra	leaves	33
3 <i>a-O-trans</i> -coumaroylbetulinic acid (35)	C. tagal	aerial parts	44





3α -O-trans-feruloylbetulinic acid (36)	C. tagal	aerial parts	44
3β - <i>E</i> -caffeoyllupeol (44)	C. decandra	leaves	33
3β -O-cis-coumaroylbetulin (39)	C. tagal	aerial parts	44
3β ,20-dihydroxylupane (40)	C. decandra	leaves	33
3β -acetylbetulinic acid (41)	C. tagal	hypocotyls and fruits	43
3β - <i>E</i> -caffeoylbetulin (42)	C. tagal	hypocotyls and fruits	43
3β - <i>E</i> -caffeoylbetulinic acid (43)	C. tagal	hypocotyls and fruits	43
3β - <i>E</i> -coumaroyllupeol (45)	C. decandra	leaves	33
	C. tagal	hypocotyls and fruits	43
3β -E-feruloylbetulin (46)	C. decandra	leaves	33
	C. tagal	hypocotyls and fruits	43
3β - <i>E</i> -ferulovlbetulinic acid (47)	C. tagal	hypocotyls and fruits	43
3β - <i>E</i> -ferulovllupeol (48)	C. decandra	leaves	33
	C. tagal	hypocotyls and fruits	43
3β -hydroxylunan-29-oic acid (49)	C decandra	leaves	33
3β - <i>O</i> - <i>cis</i> -coumarovlbetulinic acid (50)	C tagal	aerial parts	44
3β -O-trans-ferulovlbetulin (51)	C tagal	aerial parts	44
3β -O-cis-ferulovlbetulin (52)	C tagal	aerial parts	44
3 <i>B-O-trans</i> -coumaroylbetulinic acid (53)	C tagal	aerial parts	44
3B-O-trans-coumaroylbetulin (54)	C tagal	aerial parts	44
$3B_{2}$ -Coumarovllupeol (55)	C. decandra	leaves	33
3β -Z-ferulovllupeol (56)	C. decandra	leaves	33
sp 2 fordioynapeor (so)	C tagal	hypocotyls and fruits	43
hetulin (57)	C. decandra	leaves	33
	C tagal	hypocotyls and fruits	43
	C tagal	aerial parts	43
betulinaldehyde (58)	C. decandra	leaves	33
betulinic acid (59)	C. decandra	leaves	33
betalline acta (59)	C tagal	hypocotyle and fruits	13
betulonic acid (60)	C. tagal	hypocotyls and fruits	43
$\lim_{n \to \infty} 20(29) = en_3 \beta 28 = diol(61)$	C. tagal	roots	30
rup-20(2))-cn-3p,20-dioi (01)	C. tagal	roots	36
$\lim_{n \to \infty} 20(20) = 38.30 \text{ dial}(62)$	C. lagai C. dacandra	leaves	30
$1 \text{ up -} 20(29) \text{ en } 3\beta$ hydroxy 28 oic (63)	C. tagal	roots	30
$\frac{1}{10000000000000000000000000000000000$	C. lugui C. docandra	leaves	33
lupcal (65)	C. decandra	leaves	33
lupeol (03)	C. tecentru	hymosotyle and fruits	33
	C. tagal	aprial parts	43
	C. tagal	aeriai parts	44 20
Dimanan og (Ditann on oidg)	C. lagai	10015	39
Primaranes (Diterpenoias) § 15 renovumimoron 16 ol (125)	C docandra	roota	21
(125)	C. decandra	roots	31
certopsin C (120)	C. decanara	Tools	30
$\frac{15}{12} \frac{15}{12} \frac{15}{12} \frac{11}{12} 11$	C. aecanara	roots	30
ent-8(14)-pimarene-15K,16-diol (128)	C. tagal	roots	45
$\frac{1}{1}$	C. tagal	stems and twigs	38
1 sopimar-8(14)-en-15,16-diol(131)	C. tagal	roots	39
150 pinar-8(14)-en-10-nydroxy-15-one (132)	C. tagal	roots	39 45
metnoxy-ent-8(14)-pimarenely-15-one (133)	C. tagal	roots	45
Abietane (Triterpenoid)			20
abieta-8,11,13-trien-18-oic acid (246)	C. tagal	stems and twigs	38
Steroids	<i>a</i> .		
stigmasterol (99)	C. tagal	roots	45
β-sitosterol (96)	C. tagal	roots	45





	Table 4. Chemical constituents of the bark of Kandelia candel	
Compound Class	Compound Name	
	afzelechin- $(4\alpha \rightarrow 8)$ - afzelechin (180)	
Propelargonidin dimers	afzelechin- $(4\alpha \rightarrow 8)$ - catechin (181)	
	afzelechin- $(4\alpha \rightarrow 8)$ - epicatechin (182)	
Procyanidin trimers	epicatechin- $(4\beta \rightarrow 6)$ -epicatechin- $(4\beta \rightarrow 6)$ -epicatechin (188)	
	epicatechin-($4\beta \rightarrow 6$)-epicatechin-($4\beta \rightarrow 8$)-catechin (189)	
	epicatechin-($4\beta \rightarrow 6$)-epicatechin-($4\beta \rightarrow 8$)-epicatechin (190)	
	cinchonain Ia (184)	
	cinchonain Ib (185)	
Proanthocynadins	cinchonain IIa (186)	
	cinchonain IIb (187)	
	kandelins A-1, A-2, B-1, B-2, B-3, B-4 (193-198)	
	proanthicyanidin B-1, B-2, C-1 (201–203)	
	proanthicyanidin trimer (204)	
	(–)-epicatechin (172)	
	(+)-afzeleczhin (173)	
Flavan-3-ols	(+)-catechin (174)	
	(+)-gallocatechin (176)	

Table 5. Chemical constituents of the genus Rhizophora				
Compound Class and Name	Plant	Plant Part	References	
D-Friedooleananes (Triterpenoids)				
3β -O-E-coumaroyl-taraxerol (15)	R. stylosa	stems and twigs	48	
3β - <i>E</i> -caffeoyltaraxerol (16)	R. mucronata	fruits	49	
3β -O-Z)coumaroyl-taraxerol (19)	R. stylosa	stems and twigs	48	
3β -taraxerol acetate (20)	R. stylosa	stems and twigs	48	
3β -taraxerol formate (21)	R. stylosa	stems and twigs	48	
3β -Z-caffeoyltaraxerol (22)	R. mucronata	fruits	49	
3β -Z-p-coumaroyltaraxerol (24)	R. mucronata	fruits	49	
3β - <i>E</i> - <i>p</i> -coumaroyltaraxerol (18)	R. mucronata	fruits	49	
careaborin- $(3\beta$ - <i>E</i> - <i>p</i> -coumaroyltaraxerol) (18)	R. apiculata	leaves	47	
	R. stylosa	leaves	50	
	R. stylosa	leaves	50	
taraxerol (28)	R. apiculata	leaves	47	
	R. mangle	leaves and stems	51	
	R. stylosa	leaves	50	
	R. stylosa	stems and twigs	48	
	R. mucronata	fruits	49	
taraxerone (26)	R. stylosa	leaves	50	
taraxeryl-cis-p-hydroxycinnamate (24)	R. apiculata	leaves	47	
Lupanes (Triterpenoids)				
trans-hydroxycinnamoyllupeol (66)	R. mangle	leaves and stems	51	
lupeol (65)	R. apiculata	stem	52	
	R. mucronata	leaves	17	
	R. mucronata	stem bark	53	
Oleananes (Triterpenoids)				
15α-hydroxy- β -amyrin (67)	R. stylosa	stems and twigs	48	
3β -O-(E)-(4-methoxy)-cinnamoyl- 15α -hydroxyl- β -amyrin	R. mucronata	stem bark	53	
(68)				
3β -O-(E)-coumaroyl- 15α -hydroxy- β -amyrin (69)	R. stylosa	stems and twigs	48	
oleanolic acid (70)	R. mucronata	leaves	17	
β -amyrin (71)	R. mucronata	root bark	54	
	R. mucronata	leaves	17	

Ursanes (Triterpenoids)





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			1.7
ursolic acid (73)	R. mucronata	leaves	17
a-amyrin (74)	R. mucronata	root bark	54
Aliphatic alcohols		1 (1	
dotriacontanol (247)	R. apiculata	heartwood	55 55
nentriacontanoi (248)	R. apiculata	heartwood	55 55
nonacosanol (249)	R. apiculata	heartwood	55
octacosanol (250)	R. apiculata	heartwood	55
triacontanol (251)	R. apiculata	heartwood	33
Aliphatic saturated carboxylic acids		1 (1	
doicosanoic (252)	R. apiculata	heartwood	55
henicosanoic (253)	R. apiculata	heartwood	55
hentriacontanoic (254)	R. apiculata	heartwood	55
heptacosanoic (255)	R. apiculata	heartwood	55
hexatriacontanoic (256)	R. apiculata	heartwood	55
octacosanoic (257)	R. apiculata	heartwood	55
pentacosanoic (258)	R.a apiculata	heartwood	55
tetracosanoic (259)	R. apiculata	heartwood	55
tetratriacontanoic (260)	R. apiculata	heartwood	55
triacontanoic (261)	R. apiculata	heartwood	55
tritriacontanoic (262)	R. apiculata	heartwood	55
Steroids			
campesterol (92)	R. apiculata	heartwood	55
daucosterol (94)	R. mucronata	root bark	54
	R. stylosa	leaves	50
ergosta-7,22-dien-3 β -ol (95)	R. apiculata	stem	52
sitosterol (96)	R. apiculata	heartwood	55
	R. mucronata	root bark	54
	R. stylosa	leaves	50
sitosteryl-3-glucoside (97)	R. apiculata	heartwood	55
stigmasterol (99)	R. apiculata	heartwood	55
Aromatic compound			
syringaldehyde (160)	R. apiculata	heartwood	55
Benzoquinone			
2,6-dimethoxy-p-benzoquinone (162)	R. apiculata	heartwood	55
Labdanes(Diterpenoids)			
apiculol (263)	R. apiculata	roots	56
rhizophorin A (264)	R. mucronata	roots	57
	R. mucronata	roots	58
Beyeranes (Diterpenoids)			
rhizophorin B (141)	R. mucronata	roots	58
rhizophorin C (142)	R. mucronata	roots	58
rhizophorin D (143)	R. mucronata	roots	58
Pimaranes (Diterpenoids)			
15(S)-isopimar-7-en-1-oxo-15,16-diol (124)	R. apiculata	stem	52
rhizophorin E (134)	R. mucronata	roots	58
Kauranes (Diterpenoids)			
13,16α,17-trihydroxy- <i>ent</i> -9(11)-kauren-19-oic acid (102)	R. apiculata	stem	52
16R-13,17-epoxy-16-hydroxy-ent-kaur-9(11)-en-19-al	R. apiculata	stem	52
(101)	-		
<i>ent</i> -12,17-epoxy-16β-hydroxy-9(11)-kauren-19-oate (112)	R. apiculata	stem	52
mathyl ant 16β 17 dihydroxy $9(11)$ kayran 10 oots (119)	P aniculata	stem	52
$methyl_{ent} = 0(11)_{ent} = 13, 17$ evpoys 16 hydroxy 10	R apiculata	stem	52 52
$\frac{110}{110} = \frac{110}{110} = \frac{110}{110} = \frac{110}{100} = \frac{1100}{100} = \frac{1100}{1$	к. ирісшиш	515111	52
Valle (117)			

Sesquiterpene





mucronatone (265)	R. mucronata	fruits	49
Carbohydrate			
1-d-O-methyl-muco-inositol (161)	R. mucronata	roots	7
Hopanoid			
adian-5-en-3-ol (266)	R. mucronata	stem bark	53
Phenolic compounds			
atranorin (165)	R. mucronata	root bark	54
protocatechuic acid (167)	R. stylosa	leaves	50
isovanillic acid (166)	R. stylosa	leaves	50
Xanthone(aromatic ketone)			
lichixanthone (267)	R. mucronata	root bark	54
Aliphatic ketone			
palmitone (268)	R. mucronata	root bark	54
Flavonoids			
rutin (205)	R. stylosa	leaves	50
astilbin (183)	R. stylosa	leaves	50
(-)-3,7-O-diacetyl-epicatechin (178)	R. stylosa	stems and twigs	59
(–)-epicatechin (172)	R. stylosa	stems and twigs	59
	R. stylosa	stems	60
(-)-3-O-acetyl-epicatechin (179)	R. stylosa	stems and twigs	59
(-)-3,3',4',5,7-O- pentaacetyl-epicatechin (177)	R. stylosa	stems and twigs	59
(+)-afzelechin (173)	R. stylosa	stems and twigs	59
(+)-catechin (174)	R. stylosa	stems and twigs	59
	R. stylosa	stems	60
proanthocyanidin B2 (202)	R. stylosa	stems and twigs	59
glabraoside A (191)	R. stylosa	stems	60
glabraoside B (192)	R. stylosa	stems	60
cinchonain IIa (186)	R. stylosa	stems	60
cinchonain IIb (187)	R. stylosa	stems	60
(+)-catechin-3- O - α -L-rhamnoside (175)	R. stylosa	stems	60
cinchonain Ia (186)	R. stylosa	stems	60
	R. stylosa	stems	60
cinchonain Ib (185)	R. stylosa	stems and twigs	59
	R. stylosa	stems	60

2.3 *Kandelia*: There are two species in the mangrove genus *Kandelia*: *K. candel* and *K. obovata*. Only one report is available regarding the chemical constituents of plants of this genus. A few tannin compounds have been reported from *K. candel*. Investigation of *K. obovata* for its chemical constituents remains to be observed. 24 phenolic compounds including three propelargonidin dimmers, three procyanidin trimers, fourteen proanthocyanidins and four flavan-3-ols have been isolated from the bark of *K. candel Druce*⁴⁶.

2.4 *Rhizhophora*: The mangrove genus *Rhizophora* has ten species: *R. apiculata, R. harrisonii, R. lamarckii, R. mangle, R. mucronata, R. racemosa, R. samoesis, R. selala, R. stylosa* and *R. annamalayana*. Of these ten species, chemical constituents have only been reported in *R. apiculata, R. mangle, R. mucronata*, and *R. stylosa*. These reports reveal a total of 34 metabolites from *R. apiculata*, two metabolites from *R. mangle*, 23 metabolites from *R. mucronata* and 25 metabolites from *R. stylosa*, thus a total of 81 different metabolites from the genera *Rhizophora*, with details shown in Table 5.

The chemical investigation carried out by Majumdar and Patra in 1976 resulted in the isolation of β -amyrin, β -amyrone, taraxerol, β -sitosterol, and triacantanol from *R. apiculata*⁴⁷. Later in early nineties Kokpol and his team had identified three terpenoids, five long chain aliphatic alcohols, eleven long chain aliphatic saturated carboxylic acids, three steroids, 2,6-dimethoxy-*p*-benzoquinone, syringaldehyde and sitosteryl 3-glucoside from this plant species. Also, five kauranes, one labdane and one pimarane diterpenoids and one lupane triterpenoid are reported so far from *R. apiculata*.

The study conducted by Williams et. al^{51} reported the isolation and chemical characterisation of taraxerol and cinnamoyllupeol, two triterpenoids from the leaves and stems of *Rhizophora mangle* L. A variety of steroids, diterpenoids and triterpenoids were reported from the leaves and bark of *R. mucronata*. A few beyeranes (diterpenoids) were identified from this plant, and are unique to this species. Only triterpenoids of the classes oleananes and D-friedooleananes from *R. stylosa* are reported to date.



D-Friedooleananes



Figure 2. D-friedooleananes (triterpenoids) from Rhizophoraceae mangroves

3 Bioactivities

3.1 Bioactivities of Compounds Identified: With stably transfected HepG2 cells, three new dammarane triterpenes; bruguierins A–C and a new cyclic 4-hydroxy-dithiosulfonate-bruguiesulfurol as well as two known 4-hydroxydithiolane-1-oxides; brugierol and isobrugierol, were isolated from the flowers of *Bruguiera gymnorrhiza*. These phytochemicals activated an antioxidant response element (ARE luciferase activation) with EC₅₀ values of 7.8, 9.4, 15.7, 56.7, 3.7 and 1.8 μ M, respectively. Furthermore, bruguierin A, brugierol and isobrugierol also inhibited phorbol ester-induced NF κ B (nuclear factor- κ B) luciferase activation with an IC₅₀ value of 1.4, 85.0 and 14.5 μ M respectively, while bruguierin A and brugierol selectively inhibited cyclooxygenase-2 (COX-2) activity with an IC₅₀ value of 0.37 and 6.1 μ M respectively^{18,26}.

The compounds 16α -17,19-*ent*-kauranediol; 13-hydroxy-16-*ent*-kaurene-19-ol and 16-*ent*-kaurene-19-ol showed promising activity against K-562 (human chronic myeloid leukemia) and L-929 (mouse fibroblasts) of which 16-*ent*kaurene-19-ol showed the greatest selectivity for K-562 (IC₅₀ 6.8 µg/mL)²². (5*R*,9*S*,10*R*,13*S*,15*S*)*ent*-8(14)-pimarene-1-oxo-15,16-diol showed moderate cytotoxic activities against L-929²⁵.

The 15 membered macrocyclic polysulfide, gymnorrhizol, possesses an novel carbon skeleton which was isolated from *B*. *gymnorrhiza* and exhibited potent inhibitory activity against



protein tryrosine phosphatase 1B (PTP1B). PTP1B is an enzyme involved in the regulation of insulin signaling and which is regarded as a key for treatment of type III diabetes and obesity²⁷. One of the aromatic compounds extracted from the stem of *B. gymnorrhiza*, bruguierol C showed moderate activity against gram-positive and gram-negative bacteria including mycobacteria and resistant strains (MICs 12.5 $\mu g/mL$)²⁸.

The lupane caffeoyl ester, 3-(Z)-caffeoyllupeol extracted from *B. parviflora* exhibited antimalarial activity with an EC_{50} value of 8.6 μ g/mL⁸. Sexangulic acid obtained from *B*. sexangula showed moderate in vitro cytotoxicity against human lung cancer (A-549) and human luekaemic (H-L60) cell lines at a concentration of 5 µg/ml¹⁰. Tagalsin C found in C. tagal was found to exhibit moderate cytotoxicity against HeLa human cervical carcinoma cell lines³⁵. The dimeric diterpenoid, 8(14)-enyl-pimar-2'(3')-en-4'(18')-en-15'(16')endolabr-16,15,2',3'-oxoan-16-one and the other terpenoids; tagalsin C, tagalsin I, lup-20(29)-ene-3*β*,28-diol, 3-oxolup-20(29)-en-28-oic acid and 28-hydroxy-lup-20(29)-en-3-one isolated from the roots of the mangrove plant Ceriops tagal exhibited antifouling activity against cyprid larvae of the barnacle without significant toxicity³⁶. The other nontoxic were antifouling compounds identified ethoxy-ent-8(14)-pimarenely-15-one, ent-8(14)-pimarene-15R,16-diol, stigmasterol and β -sitosterol⁴⁵. Tagalsins Q, R and U showed moderate antifeedant activity against the third instar larvae of



Figure 3-1. Lupanes from Rhizophoraceae mangroves

Brontispa longissima at a concentration of 1 mg/mL³⁸. Dolabr-4(17),15(16)-dien-3-one, isopimar-8(14)-en-15,16-diol, isopimar-8(14)-en-16-hydroxy-15-one, lupeol, lup-20(29)-en- 3β ,28-diol and lup-20(29)-en- 3β -hydroxy-28-oic acid were isolated from the roots of marine mangrove *C. tagal* which were evaluated for the activation of caspase-3 enzyme using caspase-3 colourimetric assay. Caspase-3 enzyme was activated by all compounds in cleaving *p*NA from Ac-DEVD*p*NA in the presence of caspase-3-inhibitor; Ac-DEVD-CHO³⁹.

2,6-dimethoxy-*p*-benzoquinone isolated from *R. apiculata* was identified as an active constituent component against fungi, bacteria and boll weevils⁵⁵. Taraxerol and cinnamoyllupeol, are two triterpenoids derived from the leaves

and stems of *Rhizophora mangle* L, were found exhibit insecticidal activity towards *Cylas formicarius*: one of the most destructive pests of the sweet potato⁵¹.

Among the compounds isolated from the leaves of *Rhizophora stylosa*, taraxerol has been confirmed to have growth inhibitory effects of Hela and BGC-823 with IC₅₀ of 73.4 μ mol/L and 73.3 μ mol/L, respectively, while *cis*-careaborin may inhibit the growth of BGC-823 and MCF-7 with IC₅₀ of 45.9 μ mol/L and 116.0 μ mol/L, respectively. Furthermore, the presence of astilbin and rutin were initially reported to stimulate the proliferation of mice splenic lymphocytes markedly in a dose-dependent manner⁵⁰. The





Figure 3-2. Lupanes from Rhizophoraceae mangroves

compounds, (–)-epicatechin, (–)-catechin, 3-O-acetyl-(–)epicatechin, 3,7-O-acetyl-(–)-epicatechin, (+)-afzelechin, cinchonain 1b and proanthocyanidin B2 were isolated from the same plant displayed DPPH radical scavenging activity which were comparable to that of the positive control butylated hydroxytoluene (BHT). Proanthocyaninidin B2 showed the strongest activity with IC₅₀ 4.3 µg/mL, being four fold greater than the positive control, BHT (IC₅₀ 18.0 µg/mL). The antioxidant flavan-3-ol glycosides from *R. stylosa* showed an increase in their radical scavenging activities with increase in number of catechol moieties present in the molecules⁶⁰.

3.2 Bioactivities of Mangrove Extracts: Various publications have reported the biological activities of



mangrove extracts. The components of crude alkaloid mixtures from *B. sexangula* and *B. exarista* were identified as tumor inhibitors⁶. A polysaccharide extracted from the leaves of *B. cylindrica*, *R. apiculata* and *R. mucronata* of Rhizophoraceae along with some other mangrove plants exhibited positive activity against human immunodefiency viruses (HIV)⁶¹. All parts of *Ceriops decandra* have proven antiviral activity⁶¹. It also possess promising antibacterial⁶², antiinflammatory⁶³, and antidiabetic activity⁶⁴. The leaves and bark extract of *C. tagal* shows antibacterial activity⁶⁵. Phenolics are important components of the leaf extract and hypocotyls of *K. candel* and show excellent antioxidant activities^{66,67}. Therefore, *K. candel* can be a good candidate for further development as an antioxidant medicine. During the study on the antibacterial activities of mangrove extracts against two antibiotic resistant

Oleananes and Ursanes









78 $R_1 = CH_2OH, R_2 = B$ **79** $R_1 = CH_2OH, R_2 = C$ **80** $R_1 = CH_3, R_2 = A$ **81** $R_1 = CH_3, R_2 = D$ **82** $R_1 = CH_3, R_2 = B$ **83** $R_1 = CH_3, R_2 = C$ **84** $R_1 = CH_3, R_2 = E$



70 $R_1 = OH, R_2 = H, R_3 = COOH$ **71** $R_1 = OH, R_2 = H, R_3 = CH_3$ **74** $R_1 = H, R_2 = OH, R_3 = CH_3$

OH

OH

Α

С

Ε

OH













Figure 5. Triterpene alcohol, lanostane, squalene and fatty acids from Rhizophoraceae mangroves

pathogenic bacteria *Staphylococcus aureus* and *Proteus* sp., it was observed that the ethyl acetate extract of *B. Sexangula* and *R. apiculata* also possessed promising antibacterial activity⁶⁸. This antibacterial activity was also reported in a study showing that gallic acid was extracted from hydrolysable tannin from



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the barks of R. apiculata. The gallic acid possessed a significant antiyeast (anticandidal) activity towards some yeast species of medical importance. It is anticipated that gallic acid from R. apiculata is a novel antiveast agent which may be useful in the treatment of candidiasis⁶⁹. Alcoholic extract of the leaves of Rhizophora apiculata from the mangrove forest of Sunderbans, West Bengal, India were prepared and displayed manglycemic/anti-hyperglycemic activity in streptozotocin induced diabetic rats fed a glucose bolus. The results of this study revealed that this plant extract had potential hypoglycemic action⁷⁰. The cholinesterase inhibition activity of R. lamarckii was established by Natarajan et. al 2009. The antihyperglycaemic effect of R. mangle was studied⁷¹. The leaf extracts of three mangrove plants of Rhizophoraceae family; Rhizophora mucronata, R. apiculata and R. annamalavana were found to have potential antidiabetic capacity due to the presence of an insulin-like protein⁷². The various studies mentioned, provide scientific support for the use of the mangroves in folklore medicine for the treatment of diabetes.

Various mangrove plants were tested for their antioxidant capacity^{65,73}. It was found that the Rhizophoraceae mangroves showed comparatively higher antioxidant capacity which can be attributed to their higher phenolic content. Additionally, the mangrove plants of Rhizophoraceae family are the source of potent antiviral substances⁶¹.

4 Chemotaxonomy

The chemical constituents of mangrove plants of the three true mangrove genera (Rhizophoraceae); *Bruguiera*, *Ceriops* and *Rhizophora*, are the diterpenoid class kauranes exist in the genera *Bruguiera* and *Ceriops*, however kauranes are absent in the genus *Rhizophora*. The genus *Bruguiera* is characterised by the presence of disulphides and polydisulphides which are unique to the genus. Thus they can be considered as significant chemotaxonamic markers of this genus. Also, it was observed that *ent*-pimarane coexists with isopimarane in the genus *Bruguiera*. Interestingly, dolabranes only exist in the genus *Ceriops* making it a significant chemotaxonomic marker. Similarly, labdane was found only in the genus *Rhizophora*, making it a significant chemotaxonomic marker of that specific genus.

Furthermore, extensive investigation is needed to identify and classify the chemical constituents of mangrove plants to construct a thorough basis for the chemotaxonamic studies of these versatile plants.

5 Conclusions

In this review, the chemistry and bioactivities of mangroves plants of Rhizophoraceae family have been summarised. Two types of diterpenoids; beyerane and pimarane, and three types of triterpenoids; lupane, oleanane and dammarane, are common chemical constituents ubiquitously found in this family, including 268 metabolites. To date, the chemical constituents from all the mangrove plants of this family have not been investigated. It is clear that mangrove plants can provide a new



Figure 6. Steroids from Rhizophoraceae mangroves



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Figure 7. Kauranes, pimaranes and beyeranes (diterpenoids) from Rhizophoraceae mangroves

Sulphur compounds

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Figure 8. Sulphur compounds from Rhizophoraceae mangroves

Aromatic compounds





Figure 9. Sulphur compounds from Rhizophoraceae mangroves



Figure 10. Carbohydrate and benzoquione identified from Rhizophoraceae mangroves

Phenolic compounds





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 $\begin{array}{l} \textbf{168} \ \textbf{R}_1 \text{=} \ \textbf{H}, \ \textbf{R}_2 \text{=} \ \textbf{OH} \\ \textbf{169} \ \textbf{R}_1 \text{=} \ \textbf{OMe}, \ \textbf{R}_2 \text{=} \ \textbf{OH} \\ \textbf{170} \ \textbf{R}_1 \text{=} \ \textbf{OMe}, \ \textbf{R}_2 \text{=} \ \textbf{OMe} \end{array}$

Figure 11. Phenolic compounds from Rhizophoraceae mangroves









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Figure 12. Flavonoids from Rhizophoraceae mangroves

Lignans





Figure 13. Lignans from Rhizophoraceae mangroves

κ,



219 $X = O, R_1 = OH, R_2 = COOH$

232 X = O, $R_1 = OH$, $R_2 = OH$

229 X = O, R₁ = OH, R₂ = H₂C=CH

R₁



217 R₁ = OH, R₂ = H, R₃ = COOH **225** $R_1 = OH$, $R_2 = H$, $R_3 = CH_2 = CH$ **231** $R_1 = H, R_2 = H, R_3 = CH_2 = CH$ **233** R₁ = H, R₂ = H, R₃ = OH



 R₁ =H, R₂ = H, R₃ = H₂C=CH R₁ = H, R₂ = OH, R₃ = H₂C=CH $R_1 = H$, $R_2 = OH$, $R_3 = COOH$ **235** $R_1 = H$, $R_2 = OH$, $R_3 = HO-CH_2C(O)$ $R_1 = H, R_2 = OH, R_3 = HO - CH_2C(O)$ $R_1 = H$, $R_2 = OH$, $R_3 = HO - CH_2C(OH)$







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Figure 14. Dolabranes and dimeric diterpenoids from Rhizophoraceae mangroves





Aliphatic acids











Figure 16. Terpenoids, abietane, labdanes, hopanoid, a sesquitepenoid, aromatic and aliphatic ketones in Rhizophoraceae mangroves

bank of phytochemical substances that are biologically active substances, with novel structures. It is essential to systematically conserve the biodiversity in the mangrove ecosystem and for the proper of this ecosystem for the future use of humanity.

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References

- [1] Bandaranayake, W. M. Wetl. Ecol. Manag. 2002, 10, 421–452.
- [2] Wu, J.; Xiao, Q.; Xu, J.; Li, M. Y.; Pan, J. Y.; Yang, M. H. Nat. prod. Rep. 2008, 25, 955–981.
- [3] Kathiresan, K. Environ. Ecol. 1995, 13, 240–241.
- [4] Sheue, C.; Liu, H.; Yong, J. W. H. Taxon 2003, 52, 287-294.
- [5] Sheue, C.; Rashid, S. M. A.; Yong, J. W. H.; Yang, Y. *Taiwania* **2010**, *55*, 72–77.
- [6] Loder, J. W.; Russell, G. B. Aust. J. Chem. 1969, 22, 1271-1275.
- [7] Richter, A.; Thonke, B.; Popp, M. Phytochemistry 1990, 29, 1785–1786.
- [8] Chumkaew, P.; Kato, S.; Chantrapromma, K. Chem. Pharm. Bull. 2005, 53, 95–96.
- [9] Loder, J. W.; Russell, G. B.; Tetrahedron Lett. 1966, 51, 6527– 6529.
- [10] Li, L.; Huang, C. G.; Wang, C. Y.; Guo, Y. W. Nat. Prod. Res. 2010, 24, 1044–1049.
- [11] Bao, S.; Deng, Z.; Fu, H.; Proksch, P.; Lin, W. Helv. Chim. Acta 2005, 88, 2757–2763.
- [12] SY, B.; Lin, W. Zhongguo Zhong Yao Za Zhi 2006, 31, 1168– 1171.
- [13] Bao, S.; Ding, Y.; Deng, Z., Proksch, P.; Lin, W. Chem. Pharm. Bull. 2007, 55, 1175–1180.
- [14] Katu, A.; Takahashi, J. Phytochemistry 1975, 5, 220-221.
- [15] Laphookhieo, S.; Karalai, C.; Ponglimanont, C.; Chantrapromma, K. J. Nat. Prod. 2004, 67, 886–888.
- [16] Karalai, C.; Laphookhieo, S. Aust. J. Chem. 2005, 58, 556–559.

- [17] Ghosh, A.; Misra, S.; Dutta, A. K.; Choudary, A. *Phytochemistry* 1985, 24, 1725–1727.
- [18] Homhual, S.; Bunyapraphatsara, N.; Kondratyuk, T.; Herunsalee, A.; Chaukul, W.; Pezzuto, J. M.; Fong, H. H. S.; Zhang, H. J. J. Nat. Prod. 2006, 69, 421–424
- [19] Ganguly, S. N.; Sarkar, A. Indian J. Chem. B. 1978, 16, 742-744.
- [20] Hogg, R. W.; Gillan, F. T. Phytochemistry 1984, 23, 93-97.
- [21] Achamadi, S.; Syahbirin, G.; Choong, E. T.; Hemingway, W. R. *Phytochemistry* 1994, 35, 217–219.
- [22] Han, L.; Huang, X.; Sattler, I.; Dahse, H. M.; Fu, H.; Lin, W.; Grabley, S. J. Nat. Prod. 2004, 67, 1620–1623.
- [23] Subrahmanyam, C.; Rao, B. V.; Ward, R. S.; Hursthouse, M. B.; Hibbs, D. E. *Phytochemistry* **1999**, *83*, 83–90.
- [24] Salae, A. W.; Chantrapromma, S.; Fun, H. K.; Ponglimanont, C. Acta Crystallogr. E. 2007, 63, 1899–1901.
- [25] Han, L.; Huang, X.; Sattler, I.; Dahse, H. M.; Fu, H.; Grabley, S.; Lin, W. Pharmazie 2005, 60, 705–707
- [26] Homhual, S.; Zhang, H. J.; Bunyapraphatsara, N.; Kondratyuk, T.; Santarsiero, B. D.; Mesecar, A. D.; Herunsalee, A.; Chaukul, W.; Pezzuto, J. M.; Fong, H. H. *Planta Med.* **2006**, *72*, 255–260.
- [27] Huang, X. Y.; Wang, Q.; Liu, H. L.; Zhang, Y.; Xin, G. R.; Shen, X.; Dong, M. L.; Guo, Y. W. *Phytochemistry* **2009**, *70*, 2096– 100.
- [28] Han, L.; Huang, X.; Sattler, I.; Moellmann, U.; Fu, H.; Lin, W.; Grabley, S.; *Planta Med.* **2005**, 71, 160–164.
- [29] Han, L.; Huang, X.; Sattler, I.; Moellmann, U.; Fu, H.; Lin, W.; Grabley, S. J. Asian Nat. Prod. Res. 2007, 9, 327–331.
- [30] Anjaneyulu, A. S. R.; Rao, V. L. Phytochemistry 2002, 60, 777– 782.
- [31] Anjaneyulu, A. S. R.; Rao, V. L. Phytochemistry 2003, 62, 1207–1211.
- [32] Anjaneyulu, A. S. R.; Rao, V. L.; Lobkovsky, E.; Clardy, J. J. Nat. Prod. 2002, 65, 592–594.
- [33] Ponglimanont, C.; Thongdeeying, P. Aust. J. Chem. 2005, 58, 615–618.
- [34] Zhang, Y.; Deng, Z.; Gao, T., Proksch, P.; Lin, W. Phytochemistry 2005, 66, 1465–1471.
- [35] Ouyang, X. W.; Wang, X. C.; Yue, Q. X.; Hu, L. H. Nat. Prod. Commun. 2010, 5, 9–12.
- [36] Chen, J. D.; Yi, R. Z.; Lin, Y. M.; Feng, D. Q.; Zhou, H. C.;



Wang, Z. C. Int. J. Mol. Sci. 2011, 12, 6517-6528.

- [37] Fun, H. K.; Pakhathirathien, C.; Chantrapromma, S.; Karalai, C.; Chantrapromma, K. Acta Crystallogr. E. 2006, 62, 5539–5541.
- [38] Hu, W. M.; Li, M. Y.; Li, J.; Xiao, Q.; Feng, G.; Wu, J. J. Nat. *Prod.* **2010**, *73*, 1701–1705.
- [39] Chacha, M. Int. J. Biol. Chem. Sci. 2011, 5, 402-409.
- [40] Chantrapromma, S.; Fun, H. K.; Pakhathirathien, C.; Karalai, C.; Chantrapromma, K. Acta Crystallogr. E. 2007, 63, 459–461.
- [41] Zhang, Y.; Lu, Y.; Mao, L.; Proksch, P.; Lin, W. Org. Lett. 2005, 7, 3037–3040.
- [42] Chen, J. D.; Qiu, Y.; Yang, Z. W.; Lin, P.; Lin, Y. M. Helv. Chim. Acta 2008, 91, 2292–2298.
- [43] Pakhathirathien, C.; Karalai, C.; Ponglimanont, C.; Subhadhirasakul, S.; Chantrapromma, K. J. Nat. Prod. 2005, 68, 1787– 1789.
- [44] Wang, X. C.; Ouyang, X. W.; Hu, L. H. J. Asian Nat. Prod. Res. 2010, 12, 576–581.
- [45] Chen, J. D.; Feng, D. Q.; Yang, Z. W.; Wang, Z. C.; Qiu, Y.; Lin, Y. M. Molecules 2008, 13, 212–219.
- [46] Hsu, F. L.; Nonaka, G. I.; Nishioka, I. Chem. Pharm. Bull. 1985, 33, 3142–3152.
- [47] Kokpol, U.; Chavasiri, W. J. Nat. Prod. 1990, 53, 953-955.
- [48] Li, D. L.; Li, X. M.; Wang, B. G. Nat. Prod. Res. 2008, 22, 808– 813.
- [49] Laphookhieo, S.; Karalai, C.; Ponglimanont, C. Chem. Pharm. Bull. 2004, 52, 883–885.
- [50] Yang, X. H.; Li, H. B.; Chen, H.; Li, P.; Ye, B. P. Yao Xue Xue Bao 2008, 43, 974–978.
- [51] Williams, L. Die Naturwissenschaften 1999, 86, 450-452.
- [52] Gao, M. Z.; Yuan, X. Y.; Cheng, M. C.; Xiao, H. B.; Bao, S. X. J. Asian Nat. Prod. Res. 2011, 13, 776–779.
- [53] Rohini, R. M.; Das, A. K. Nat. Prod. Res. 2010, 24, 197-202.
- [54] Rao, B. V.; Rao, C. V.; Subrahmanyam, C.; Jairaj, M. A. J. Indian Chem. Soc. 2005, 82, 155–157.
- [55] Kokpol, U.; Chavasiri, W.; Chittawong, V.; Bruce, M.; Cunningham, G. N.; Miles, D. H. *Phytochemistry* **1993**, *33*, 1129– 1131
- [56] Saxena, E.; Garg, H. S. Nat. Product Lett. 1994, 4, 149-154.

Nat. Prod. Bioprospect. 2013, 3, 207-232

- [57] Anjaneyulu, A. S. R.; Rao, V. L. Nat. Product Lett. 2001, 15, 13–19.
- [58] Anjaneyulu, A. S. R.; Anjaneyulu, V.; Rao, V. L. J. Asian Nat. Prod. Res. 2002, 4, 53–61.
- [59] Li, D. L.; Li, X. M.; Peng, Z. Y.; Wang, B. G. Molecules 2007, 12, 1163–1169.
- [60] Takara, K.; Kuniyoshi, A.; Wada, K.; Kinjyo, K.; Iwasaki, H. Biosci. Biotech. Biochem. 2008, 72, 2191–2194.
- [61] Premanathan, M.; Kathiresan, K.; Nakashima, H. South Pacific Study 1999, 19, 49–57.
- [62] Chandrasekaran, M.; Kannathasan, K.; Venkatesalu, V.; Prabhakar, K. World J. Microb. Biotech. 2008, 25, 155–160.
- [63] Hossain, H.; Moniruzzaman, Sk.; Nimmi, I.; Kawsar, H.; Hossain, A.; Islam, A.; Jahan, I. A. Oriental Pharm. Exp. Med. 2011, 11, 215–220.
- [64] Nabeel, M. A.; Kathiresan, K.; Manivannan, S. J. Diabetes 2010, 2, 97–103.
- [65] Arivuselvan, N.; Silambarasan, D.; Govindan, T.; Kathiresan, K. Adv. Biol. Res. 2011, 5, 251–254.
- [66] Zhang, L. L.; Lin, Y. M.; Zhou, H. C.; Wei, S. D.; Chen, J. H.; *Molecules* 2010, 15, 420–31.
- [67] Wei, S. D.; Zhou, H. C.; Lin, Y. M. Int. J. Mol. Sci. 2010, 11, 4080–4093.
- [68] Abeysinghe, P. D. Indian J. Pharm. Sci. 2010, 72, 167-172.
- [69] Hong, L. S.; Ibrahim, D.; Kassim, J.; Sulaiman, S. J. Appl. Pharm. Sci. 2011, 1, 75–79.
- [70] Sur, T. K.; Seal, T.; Pandit, S.; Bhattacharya, D. Nat. Prod. Sci. 2004, 10, 11–15.
- [71] Alarcon-Aguilara, F. J.; Roman-Ramos, R.; Perez-Gutierrez, S.; Aguilar-Contreras, A.; Contreras-Weber, C. C.; Flores-Saenz, J. L. J. Ethnopharmacol. 1998, 61, 101–110.
- [72] Nabeel, A. M.; Kathiresan, K.; Chinthamani, M.; Manivannan, S. Nat. Prod. Res. 2012, 26, 1161–1166.
- [73] Agoramoorthy, G.; Chen, F. A.; Venkatesalu, V.; Kuo, D. H.; Shea, P. C. Asian J. Chem. 2008, 20, 1311–1322.

