Czernekova et al. BMC Infectious Diseases 2014, 14(Suppl 2):P68 http://www.biomedcentral.com/1471-2334/14/S2/P68

POSTER PRESENTATION

BMC Infectious Diseases



Differential glycosylation of envelope gp120 affects reactivity with HIV-1 specific antibodies

Lydie Czernekova^{1*}, Milan Raska^{1,2}, Zina Moldoveanu², Katerina Zachova¹, Stacy Hall², Dita Badalova¹, Alzbeta Krcmarska¹, Hana Synkova¹, Michael Hoelscher³, Leonard Maboko⁴, Rhubell Brown², Zdenek Novak², Jiri Mestecky², Jan Novak²

From Abstracts from International Symposium HIV and Emerging Infectious Diseases 2014 Marseille, France. 21-23 May 2013

Introduction

Cellular entry of human immunodeficiency virus type 1 (HIV-1) depends on envelope glycoprotein (gp120/gp41) interactions with host-cell receptors. Approximately one-half of molecular mass of gp120 consists of N-glycans which may act as antigenic determinants as well as a shield against immune recognition. We have shown that glycosy-lation of subtype B HIV-1 gp120 varies according to the producing cell type and the differential glycosylation affects reactivities with serum antibodies of persons infected with HIV-1 subtype B. Here we studied reactivities of above proteins with panel of monoclonal gp120-specific broadly neutralizing antibodies and sera from persons infected with HIV-1 subtype A/C.

Materials and methods

Recombinant gp120 produced in different cell lines (HEK 293T, Jurkat, RD, HepG2, and CHO) were tested as a native and after partial removal of N-glycans by PNGase F under native conditions. Several methods (ELISA, Cytometric Bead Array - CBA, SDS-PAGE with western blot, and dot blot) were used for determination and comparison of reactivities with monoclonal gp120-specific antibodies (268-D IV, F425 B4e8, 257-D IV, 447-52D, 19b, 2G12, and b12) or with sera of HIV-1 subtype A/C-infected persons.

Results

After partial removal of N-glycans from gp120, the reactivity of most monoclonal antibodies increased, as did the reactivities of sera from HIV-1-infected persons. The largest increase in binding of polyclonal antibodies after PNGase F treatment was found for gp120 expressed in

¹Department of Immunology, Palacky University, Olomouc, Czech Republic Full list of author information is available at the end of the article HEK 293T and CHO cell lines, common producers of recombinant proteins for vaccination purposes. Conversely, the gp120 produced by T cells (Jurkat) displayed the least increase in reactivity after partial deglycosylation.

Conclusion

Changes in reactivity of selected monoclonal antibodies with native and PNGase F-treated proteins indicated that some glycans were resistant to deglycosylation and that this characteristic was dependent on producer cell type. Furthermore, CBA allowed more sensitive detection (about 40-times) of gp120-specific antibodies compared to ELISA.

Supported by CZ.1.07/2.3.00/20.0164 European Social Fund, UAB CFAR Developmental Grant (P30AI027767) and a Pilot Grant from UAB School of Medicine.

Authors' details

¹Department of Immunology, Palacky University, Olomouc, Czech Republic. ²University of Alabama, Birmingham, Alabama, USA. ³Clinic of the University of Munich, Munich, Germany. ⁴NIMR-Mbeya Medical Research Program, Mbeya, Tanzania.

Published: 23 May 2014

doi:10.1186/1471-2334-14-S2-P68 Cite this article as: Czernekova *et al.*: Differential glycosylation of envelope gp120 affects reactivity with HIV-1 specific antibodies. *BMC Infectious Diseases* 2014 14(Suppl 2):P68.



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