BMC Cancer

RESEARCH ARTICLE

Inari et al. BMC Cancer (2017) 17:160

DOI 10.1186/s12885-017-3154-3





Expression of enhancer of zeste homolog 2 correlates with survival outcome in patients with metastatic breast cancer: exploratory study using primary and paired metastatic lesions

Hitoshi Inari^{1*}, Nobuyasu Suganuma¹, Kae Kawachi³, Tatsuya Yoshida¹, Takashi Yamanaka¹, Yoshiyasu Nakamura², Mitsuyo Yoshihara², Hirotaka Nakayama⁴, Ayumi Yamanaka⁴, Katsuhiko Masudo⁴, Takashi Oshima⁴, Tomoyuki Yokose³, Yasushi Rino⁴, Satoru Shimizu¹, Yohei Miyagi^{2*} and Munetaka Masuda^{4*}

Abstract

Background: In metastatic breast cancer, the status of the estrogen receptor (ER), progesterone receptor (PR), and human epidermal growth factor receptor 2 (HER2), as well as the Ki-67 index sometimes change between primary and metastatic lesions. However, the change in expression levels of enhancer of zeste homolog 2 (EZH2) between primary and metastatic lesions has not been determined in metastatic breast cancer.

Methods: Ninety-six metastatic breast cancer patients had biopsies or resections of metastatic lesions between September 1990 and February 2014 at the Kanagawa Cancer Center. We evaluated ER, PR, HER2, Ki-67, and EZH2 in primary lesions and their corresponding metastatic lesions using immunohistochemistry. We examined the change in expression of EZH2 between primary and metastatic lesions, the correlation between the expression of EZH2 and the relationship between EZH2 expression and patient outcome in metastatic breast cancer.

Results: EZH2 expression was significantly higher in metastatic lesions compared with primary lesions. EZH2 expression was highly correlated with Ki-67 expression in primary and metastatic lesions. High-level expression of EZH2 was associated with poorer disease-free survival (DFS) outcomes in patients with primary lesions (P < 0.001); however, high-level expression of EZH2 was not associated with poorer DFS outcomes in patients with metastatic lesions (P = 0.063). High-level expression of EZH2 was associated with poorer overall survival (OS) postoperatively in patients with primary (P = 0.001) or metastatic lesions (P = 0.005). High-level expression of EZH2 was associated with poorer OS outcomes after recurrence in patients with metastatic lesions (P = 0.014); however, high-level expression of EZH2 was not associated with poorer OS outcomes after recurrence in patients with metastatic lesions (P = 0.014); however, high-level expression of EZH2 was not associated with poorer OS outcomes after recurrence in patients with metastatic lesions (P = 0.014); however, high-level expression of EZH2 was not associated with poorer OS outcomes after recurrence in patients with primary lesions (P = 0.096). High-level expression of EZH2 in metastatic lesions was independently associated with poorer OS outcomes after recurrence. (Continued on next page)

* Correspondence: inari-hitoshi@nifty.com; miyagi@gancen.asahi.yokohama.jp; mmasuda@yokohama-cu.ac.jp

²Molecular Pathology and Genetics Division, Kanagawa Cancer Center Research Institute, 2-3-2 Nakao, Asahi-ku, Yokohama 241-0815, Japan ⁴Department of Surgery, Yokohama City University, 3-9 Fukuura, Kanazawa-ku, Yokohama 236-0004, Japan

Full list of author information is available at the end of the article



© The Author(s). 2017 **Open Access** This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated.

¹Department of Breast and Endocrine Surgery, Kanagawa Cancer Center, 2-3-2 Nakao, Asahi-ku, Yokohama 241-0815, Japan

(Continued from previous page)

Conclusions: EZH2 expression was significantly increased in metastatic lesions compared with primary lesions. High-level expression of EZH2 in metastatic lesions was associated with poorer OS outcomes after primary surgery and recurrence.

Keywords: Metastatic breast cancer, EZH2, Ki-67, Prognostic factor, Immunohistochemistry, Epigenetics

Background

Metastatic breast cancer (MBC) is difficult to treat using currently available conventional therapies, and median long-term survival rates in MBC patients have been reported to be as little as 18–24 months or 2–4 years from the time of diagnosis [1, 2]. Following chemotherapy, 10-year survival rates are approximately 5% and 2–3% in patients with MBC and those who survive >20 years, respectively [3, 4].

Management of MBC generally consists of systemic treatment (chemotherapy and targeted therapy, including antiestrogen and anti-human epidermal growth factor receptor 2 [HER2] therapies). Treatment decisions for patients with MBC are usually based on estrogen receptor (ER), progesterone receptor (PR) and HER2 status of the primary tumor, the disease-free interval (DFI), site(s) of recurrence and performance status [5]. Because performing biopsies of metastatic lesions risks damaging vital organs and tissues, investigation of biomarkers in metastatic lesions is often challenging. Therefore, based on biomarkers in the primary tumor, the systemic treatment is often given to MBC patients. However, previously published reports show that, because biomarker levels change between primary and metastatic lesions, surgical biopsy of metastatic lesions followed by pathological confirmation for the investigation of biomarkers is occasionally proposed as an effective strategy in the treatment of MBC patients [6–12].

Enhancer of zeste homolog 2 (EZH2) is a well-known histone modifier protein that functions as a methyltransferase at lysine 27 of histone H3 [13]. EZH2 is a member of the polycomb group of genes [14] that is important for transcriptional regulation through chromatin remodeling, nucleosome modification and interactions with other transcription factors. It is assumed that EZH2 promotes breast cancer progression by transcriptional repression of tumor suppressors and by maintaining cells in a stem cell-like state [15, 16]. EZH2 has been demonstrated to be overexpressed in many types of malignancies, including breast, prostate and endometrial cancers, and has been suggested as a candidate for targeted treatment [17, 18]. In primary breast cancer (PBC), Kleer et al. [17] showed that EZH2 overexpression was further associated with a larger tumor size, ERand PR-negative status, an advanced stage of disease, and significantly reduced disease-free survival (DFS) and overall survival (OS). Other investigators have reported that EZH2 promotes neoplastic progression in the breast, and that downregulation in EZH2 expression reduces in vivo tumor growth of breast cancer cells [17, 19, 20]. EZH2 is important for the control of cell proliferation and invasion, and has recently been shown to regulate DNA repair pathways and genomic stability [19, 21–24]. However, few reports have examined EZH2 expression in metastatic lesions, changes in EZH2 expression levels between primary and metastatic lesions, and patient outcome measures in MBC in relation to EZH2 expression.

The purpose of this study was to examine the expression levels of EZH2 in 96 pairs of primary cancer tissues and metastatic lesions obtained from patients with MBC. To evaluate the clinicopathological significance of EZH2 expression in metastatic lesions, we examined the correlations and changes in ER, PR, HER2, Ki-67 and EZH2 expression between primary cancer tissues and metastatic lesions, and DFS and OS outcomes after primary surgery and recurrence in patients with MBC.

Methods

Patients and samples

We retrospectively studied surgical specimens of PBC tumors and their corresponding metastatic lesions from patients who underwent surgery for their PBC tumor at the Kanagawa Cancer Center, Yokohama, Japan, between December 1977 and March 2013. Of those who relapsed after primary surgery between September 1990 and February 2014, there were 96 consecutive patients from whom metastatic lesions were obtained, either by surgery or biopsy, and evaluated using immunohistochemistry (IHC). In all cases, archival hematoxylin and eosin-stained slides of the PBC tumor and its corresponding metastatic lesion were retrieved and reviewed for confirmation of pathological features, as well as to select suitable tissue blocks for IHC analysis. We constructed tissue microarrays (TMAs) using PBC tumors and metastatic lesions. In patients receiving neoadjuvant chemotherapy, we examined the PBC tumor using a core needle biopsy before treatment was commenced in order to avoid potential bias. The Ethics Committees of the Kanagawa Cancer Center, Yokohama, Japan, approved the study protocol.

TMAs

TMAs consisting of cores, each measuring 2 mm in diameter, were assembled from formalin-fixed, paraffin-

embedded blocks of surgically removed tissue from primary tumors and their metastatic lesions in breast cancer patients. We included tissue cores from each primary tumor, metastatic lesion and normal breast tissue, which was used as a control, in the array.

IHC analysis

IHC staining for biomarkers ER, PR, HER2, Ki-67 and EZH2 was performed in all cases. TMAs were cut into 4µm-thick sections and mounted onto pre-coated glass slides. All sections were stained using an autostainer (trade name Histostainer; Nichirei Biosciences Inc., Tokyo, Japan) using primary antibodies to ER (clone 1D5, dilution 1:80; Nichirei Biosciences Inc., Tokyo, Japan), PR (clone A9621A, dilution 1:100; Nichirei Biosciences Inc., Tokyo, Japan), HER2 (clone D8F12, dilution 1:800; Cell Signaling Technology Inc., Danvers, MA, USA), Ki-67 (clone SP-6, dilution 1:200; Nichirei Biosciences Inc., Tokyo, Japan) and EZH2 (clone D2C9, dilution 1:50; Cell Signaling Technology Inc., Danvers, MA, USA).

The results of the IHC analysis were assessed in a blinded fashion by a breast surgeon (H.I.) and pathologist (K.K.) who examined each slide independently. Unclear cases were discussed between the breast surgeon and pathologist. Each tumor was assessed twice and an average was calculated between the two scores. Nuclear immunoreactivity of each hormone receptor was scored independently by evaluating the percentage of positively stained cancer cells. ER and PR were defined as positive if there was staining of $\geq 1\%$ of tumor cell nuclei. HER2 expression was scored as 0, 1+, 2+ or 3+ in accordance with the guidelines of the American Society of Clinical Oncology/College of American Pathologists [25]. A HER2 score of 3+ was considered positive. IHC 2+ tumors were not analyzed using in situ hybridization techniques. A HER2 score of 2+ was considered negative (see Additional file 1).

Regardless of the staining intensity, nuclear immunoreactivity of EZH2 and Ki-67 expression were scored independently by evaluating the proportion of positively stained cancer cells: Score $1 = \le 1/100$ cells stained; Score $2 = \le 1/10$ cells stained; Score $3 = \le 1/3$ cells stained; Score $4 = \le 2/3$ cells stained; and Score 5 > 2/3 cells stained (Fig. 1). EZH2 expression scores of 4 and 5, and Ki-67 expression scores of 3, 4 and 5, were considered high expression. EZH2 expression scores of 1, 2 and 3, and Ki-67 expression. The median EZH2 score and Ki-67 expression score across all PBC tumors sampled were 4 and 3, respectively (see Additional file 2).

ER, PR, HER2 and Ki-67 expression were used to identify distinct molecular subtypes (Table 1). These were defined as follows: luminal A = ER and/or PR+, HER2–, and low Ki-67 expression; luminal B = ER and/or PR+, HER2–, and

high Ki-67 expression; luminal HER2 = ER and/or PR+, HER2+; HER2-type = ER and PR-, HER2+; and triplenegative breast cancer (TNBC) = ER and PR-, HER2 – .

Follow-up

Follow-up was performed using the KCCH Cancer Registry until October 31, 2015. Active follow-up was conducted by accessing hospital visit records, resident registration cards, and permanent domicile data. During the study period, no subject was lost to follow-up. The day of the biopsy of the metastatic lesions was defined as the date of diagnosis of recurrence. DFS was defined as the period from the day of primary surgery until the day of the biopsy of the metastatic lesions. OS after primary surgery was defined as the period from the day of primary surgery until the day of death. OS after recurrence was defined as the period from the day of biopsy of the metastatic lesions until the day of death. Median follow-up time was 96 months (range, 1-299 months) after the primary operation, and median follow-up time was 40 months (range, 0-231) after recurrence.

Statistical analyses

Relationships between biomarkers of the primary and metastatic breast cancer lesions and clinicopathological characteristics of the patients were analyzed using chisquare tests. Correlations between EZH2 expression and that of other biomarkers were evaluated using Pearson product-moment correlation coefficients (r). EZH2 and Ki-67 scores between primary and metastatic breast cancer lesions were compared using independent *t*-tests. DFS, survival rates after primary surgery, and survival rates after recurrence were analyzed using the Kaplan-Meier method, and any differences in survival rates were assessed using log-rank tests according to the expression of EZH2 in the primary and metastatic lesions. Cox proportional hazards models were applied to the multivariate analyses. Since we showed Ki-67 expression and EZH2 expression in primary and metastatic lesions to be strongly correlated (Table 2), we assessed prognostic factors (except for the Ki-67 expression) in multivariate analysis. For Pearson product-moment correlation coefficients (r), a P < 0.01, and for chi-square tests, independent t-tests, log-rank tests and Cox proportionalhazards models, a P < 0.05 was considered statistically significant. All statistical analyses were performed using SPSS version 20 (SPSS Inc., Chicago, IL, USA).

Results

Clinicopathological characteristics of all patients with MBC in this study

Patient characteristics are summarized in Table 3. Among 96 biopsies or resections of metastases, 26 (27.0%) were of



brain, eight (8.3%) of lung, one (1.0%) of liver, two (2.0%) of ovary, 13 (13.5%) of chest wall, 15 (15.6%) of lymph nodes, seven (7.3) of distant skin and 24 (25%) of bone.

Changes in ER, PR, HER2, Ki-67 and EZH2 expression between primary and metastatic lesions

Compared with primary lesions, metastatic lesions exhibited significantly higher levels of expression of Ki-67 (75.0% vs. 57.3% P = 0.010) and EZH2 (82.3% vs. 56.3% P < 0.0001). Conversely, no statistical differences in ER (42.7% vs. 53.1%, P = 0.149), PR (40.6% vs. 49.0%, P = 0.246) or HER2 status (14.6% vs. 16.7%, P = 0.691) were observed between primary and metastatic lesions (Table 1). We subsequently analyzed the scores of expression levels of Ki-67 and EZH2, which demonstrated a significant difference between the two groups (i.e., high vs. low expression) (Fig. 2). The means and standard deviations of the Ki-67 scores were 2.74 \pm 0.92 and 3.10 \pm 0.97 for primary and metastatic

lesions, respectively (P = 0.009) (Fig. 2a), while the means and standard deviations of the EZH2 expression scores were 3.56 ± 1.34 and 4.26 ± 1.08 for primary and metastatic lesions, respectively (P < 0.001) (Fig. 2b). Ki-67 and EZH2 expression scores were significantly higher in metastatic lesions compared with PBC lesions.

Correlation coefficients of ER, PR, HER2, Ki-67 and EZH2 expression scores in primary and metastatic lesions

In PBC lesions, ER (r = -0.103, P = 0.318) and PR (r = -0.111, P = 0.282) status were not significantly correlated with EZH2 expression, HER2 status exhibited a significant low correlation with EZH2 expression (r = 0.361, P < 0.001), and Ki-67 expression exhibited a significant high correlation with EZH2 expression (r = 0.722, P < 0.0001) (Table 2). Similarly, in metastatic lesions, ER (r = -0.099, P = 0.339) and PR (r = -0.190, P = 0.064) status were not significantly correlated with EZH2 expression, HER2 status exhibited a significant low correlation

Table 1 Compa	rison of estrogen r	eceptor, progesteror	ne				
receptor, human epidermal growth factor receptor 2, Ki-67, and							
enhancer of zeste homolog 2 biomarkers between primary lesions							
and metastatic lesions in breast cancer patients ($n = 96$)							
Biomarker	Primary lesions	Metastatic lesions	P-value				

ER status, n (%)			
positive	51 (53.1)	41 (42.7)	0.149
negative	45 (46.9)	55 (57.3)	
PR status, n (%)			
positive	47 (49.0)	39 (40.6)	0.246
negative	49 (51.0)	57 (59.4)	
HER2 status, <i>n</i> (%)			
positive	16 (16.7)	14 (14.6)	0.691
negative	80 (83.3)	82 (85.4)	
Ki-67 expression, n	(%)		
high	55 (57.3)	72 (75.0)	0.010*
low	41 (42.7)	24 (25.0)	
EZH2 expression, n	(%)		
high	54 (56.3)	79 (82.3)	< 0.0001
low	42 (43.7)	17 (17.7)	
Molecular subtype	, n (%)		
luminal A ^a	31 (32.3)	19 (19.8)	0.304
luminal B^{b}	22 (22.9)	29 (30.2)	
luminal HER2 ^c	5 (5.2)	5 (5.2)	
HER2-type ^d	11 (11.5)	9 (9.4)	
TNBC ^e	27 (28.1)	34 (35.4)	

Abbreviations: ER estrogen receptor, EZH2 enhancer of zeste homolog 2, HER2 human epidermal growth factor receptor 2, PR progesterone receptor, TNBC triple-negative breast cancer

^aLuminal A = ER and/or PR+, HER2–, and low Ki-67 expression ^bLuminal B = ER and/or PR+, HER2–, and high Ki-67 expression

^cLuminal HER2 = ER and/or PR+, HER2+

^dHER2-type = ER and PR-, HER2+ ^eTNBC = ER and PR-, HER2-

^{*}Indicates values that are statistically significant (P < 0.05)

Table 2 Correlation coefficient of biomarkers and EZH2 so	cores
-----------------------------------------------------------	-------

	EZH2	r	P-value
Primary ER status	Primary	-0.103	0.318
Primary PR status	Primary	-0.111	0.282
Primary HER2 status	Primary	0.361	< 0.001*
Primary Ki-67 expression	Primary	0.722	< 0.0001*
Metastatic ER status	Metastatic	-0.099	0.339
Metastatic PR status	Metastatic	-0.190	0.064
Metastatic HER2 status	Metastatic	0.306	0.002*
Metastatic Ki-67 expression	Metastatic	0.685	< 0.0001*

Abbreviations: ER estrogen receptor, EZH2 enhancer of zeste homolog 2, HER2 human epidermal growth factor receptor 2, *PR* progesterone receptor ^{*} Indicates values that are statistically significant (P < 0.01)

Table 3 Clinicopathological	Characteristics	of all	metastatic
patients in this study			

Characteristic	Patients ($n = 96$)	Percent
Age, years (mean ± SD)	51 ± 7.5	
Menopausal status		
pre-	44	45.8
post-	52	54.2
Tumor size		
≤ 20 mm	17	17.7
> 20 mm	77	80.2
unknown	2	2.1
LN status		
positive	63	65.6
negative	33	34.4
Histological type		
ductal	83	86.5
special ^a	13	13.5
LVI status		
positive	63	65.6
negative	25	26.1
unknown	8	8.3
Operation status		
partial mastectomy	17	17.8
mastectomy	79	82.2
Stage at the primary diagnosis		
1	11	11.5
2	48	50
3	34	35.4
4	1	1.0
unknown	2	2.0
Chemotherapy		
adjuvant	83	86.5
none	13	13.5
Hormone therapy		
adjuvant	50	52.1
none	46	47.9
Site of recurrence		
brain	26	27.0
lung	8	8.3
liver	1	1.0
ovary	2	2.1
chest wall	13	13.5
lymph node	15	15.6
distant skin	7	7.3
bone	24	25

Abbreviations: LN lymph node, LVI lymphovascular invasion, SD standard deviation ^aSpecial type is invasive breast carcinoma except invasive ductal carcinoma



with EZH2 expression (r = 0.306, P = 0.002), and Ki-67 expression scores exhibited a high correlation with EZH2 expression (r = 0.685, P < 0.001) (Table 2).

Relationship between EZH2 expression and patient clinicopathological characteristics

Patient clinicopathological characteristics and their correlation with EZH2 expression in primary and metastatic lesions are summarized in Table 4. Relationships between the expression level status of EZH2 and patient age, menopausal status, tumor size, lymph node status, histological type (i.e., ductal vs. special), lymphovascular invasion status, operation status (i.e., partial vs. full mastectomy), adjuvant chemotherapy and hormone therapy status, primary ER status, primary PR status, primary HER2 status, primary Ki-67 expression, metastatic ER status, metastatic PR status, metastatic HER2 status, metastatic Ki-67 expression, the site(s) of recurrence (i.e., viscera, soft tissue or bone) and DFI (disease-free interval) (i.e., ≤ 2 years, > 2 years, ≤ 10 years, > 10 years) were evaluated. Factors significantly associated with PBC lesions included lymph node status, histological type, primary ER status, metastatic PR status, primary and metastatic HER2 status, primary and metastatic Ki-67 expression, and DFI, whereas factors significantly associated with metastatic lesions included histological type, adjuvant chemotherapy status, primary PR status, primary HER2 status, primary and metastatic Ki-67 expression, and the site(s) of recurrence (Table 4).

Relationship between EZH2 expression and patient outcome

We examined DFS and OS outcomes after primary surgery and recurrence in patients with MBC according to primary EZH2 expression (Fig. 3a, b, c), and DFS and OS outcomes after primary surgery and recurrence in patients with MBC according to metastatic EZH2 expression (Fig. 3d, c, f). Patient clinicopathological characteristics and their correlation with EZH2 expression in primary and metastatic lesions are summarized in Table 4. First, DFS, survival rates after primary surgery, and survival rates after recurrence were analyzed according to the expression of EZH2 in PBC lesions. Low EZH2 expression in PBC lesions occurred in 42 patients and high EZH2 expression in PBC lesions in 54 patients. Median DFS time in patients with high expression levels of EZH2 in PBC lesions was 30 compared with 74 months in patients with low expression levels of EZH2 in PBC lesions (Fig. 3a). Median survival time after primary surgery in patients with high expression levels of EZH2 in PBC lesions was 55 compared with 133 months in patients with low expression levels of EZH2 in PBC lesions (Fig. 3b). Patients expressing high levels of EZH2 in PBC lesions had significantly poorer DFS and OS outcomes than patients expressing low levels of EZH2 in PBC lesions (P < 0.001, P = 0.001). Second, DFS, survival rates after primary surgery, and survival rates after recurrence were analyzed according to the expression of EZH2 in the metastatic lesions. Low EZH2 expression in metastatic lesions occurred in 17 patients and high EZH2 expression in metastatic lesions occurred in 79 patients. Median survival time after primary surgery in patients with high expression levels of EZH2 in metastatic lesions was 66 compared with 161 months in patients with low expression levels of EZH2 in metastatic lesions (Fig. 3e). Median survival time after recurrence in patients with high expression levels of EZH2 in metastatic lesions was 25 compared with 50 months in patients with low expression levels of EZH2 in metastatic lesions (Fig. 3f). Patients expressing high levels of EZH2 in metastatic lesions had significantly poorer OS outcomes after primary surgery and recurrence than patients expressing low levels of EZH2 in metastatic lesions (P = 0.005, P = 0.014).

characteristics of patients (n = 96)

etweenEZH2 expression in primary and metastatic breast cancer lesic	ons and the clinicopathological
etweenEZH2 expression in primary and metastatic breast cancer lesic	ons and the clinicopathological

low (n = 42)high (n = 54)low (n = 17)high (n = 79)Age, years (mean \pm 50) 52 ± 55 50 ± 98 0.198 50 ± 7.8 51 ± 97 0.241 Menopausal status, n (%)nc 7 (6.4) 0.353 10 (22.7) 34 (77.3) 0.236 post-25 (48.1)27 (51.9)7 (13.5) 45 (86.5) 10 (22.7) 34 (77.3) 0.236 post-25 (48.1)27 (51.9)7 (13.5) 45 (86.5) 15 (88.2) 20 nm 32 (49.1) 9 (52.9) 2 (11.8) 15 (88.2) 20 nm 32 (42.9) 44 (57.1) 0.936 14 (18.2) 63 (81.8) 0.396 unknown1 (50.0)1 (50.0)1 (50.0)1 (50.0)1 (50.0) 10 (50.0) 10 10 (70.0)IN status, n (%)megative19 (57.6)44 (42.4) 0.046^{**} 61.82 27 (81.8) 0.930 Histological type, n (%)megative19 (57.6)14 (42.4) 0.046^{**} 61.82 27 (81.8) 0.930 UN status, n (%)megative32 (50.8) 51 (61.4) 0.016^{**} 12 (14.5) 71 (85.5) 0.035^{**} special10 (76.9) 3 (23.1) 5 (88.5) 8 (61.5) 11 (17.5) 52 (82.5) 10 (14.60) 24 (84.0) 0.942 unknown2 (25.0) 6 (75.0) 2 (25.0) 6 (75.0) 27 (80.0) 0.942 unknown2 (25.0) 6 (75.0) 2 (25.0) 6 (75.0) 27 (80.5) 0.952 adjuvant 24 (41.0)<	Characteristic	Primary EZH2	Primary EZH2		Metastatic EZH2	Metastatic EZH2	
Age, years (mean \pm SD) 52 \pm 8.5 50 \pm 9.8 0.198 50 \pm 7.8 51 \pm 9.7 0.241 Menopausal status, n (%) pre- 17 (38.6) 27 (61.4) 0.353 10 (02.7) 34 (77.3) 0.236 post- 25 (48.1) 27 (51.9) 7 (13.5) 45 (86.5) 10 12.7 34 (77.3) 0.236 20 mm 8 (47.1) 9 (52.9) 2 (11.8) 15 (88.2) 396 14 (18.2) 63 (81.8) 0.396 20 mm 33 (42.9) 44 (57.1) 0.936 14 (18.2) 63 (81.8) 0.396 unknown 1 (50.0) 1 (50.0) 1 (50.0) 1 (50.0) 1 (50.0) 1 (50.0) 1 (50.0) 10 (75.0) 22 (82.5) negative 19 (57.6) 14 (42.4) 0.048° 6 (18.2) 27 (81.8) 0.930 Histological type, n (%) Utctal 32 (50.8) 51 (61.4) 0.010° 12 (14.5) 71 (85.5) 0.035° special 10 (76.9) 3 (23.1) 5 (38.5) 8 (61.5) 11 (17.5) 52 (82.5) negative 18 (16.0) 21 (84.0) 0.842 unknown 2 (25.0)		low $(n = 42)$	high (<i>n</i> = 54)		low $(n = 17)$	high (<i>n</i> = 79)	
Menopausal status, n (%)pre-17 (48.6)2 / 61.9)0.33310 (2.7)34 (7.3)0.236post-25 (48.1)2 / 51.9)70 (2.7)34 (7.3)0.2362 morn size, n (%)2211.1015 (88.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2)34 (8.2) <t< td=""><td>Age, years (mean ± SD)</td><td>52 ± 8.5</td><td>50 ± 9.8</td><td>0.198</td><td>50 ± 7.8</td><td>51 ± 9.7</td><td>0.241</td></t<>	Age, years (mean ± SD)	52 ± 8.5	50 ± 9.8	0.198	50 ± 7.8	51 ± 9.7	0.241
pre- 17 (38.6) 27 (61.4) 0.353 10 (22.7) 34 (77.3) 0.236 post- 25 (48.1) 27 (51.9) 7 (13.5) 45 (66.5) Tumor size, n (%) \$ \$ 21 (11.8) 15 (88.2) \$ \$ 20 mm 31 (42.9) 44 (57.1) 0.936 14 (18.2) 63 (81.8) 0.393 \$ 20 mm 150.0) 150.0) 150.00 1 (50.0) 1 (50.0) 1 (50.0) 1 (50.0) 1 (50.0) 1 (50.0) 1 (50.0) 0.048* 1 (17.5) 5 (82.5) \$ 1 (17.5) 5 (82.5) \$ 1 (17.5) 5 (82.5) \$ 1 (17.5) 5 (82.5) \$ 1 (17.5) 5 (82.5) \$ 1 (17.5) 5 (82.5) \$ 1 (17.5) 5 (82.5) \$ 1 (17.5) 5 (82.5) \$ 1 (17.5) 5 (82.5) \$ 1 (17.5) 5 (82.5) \$ 1 (17.5) 5 (82.5) \$ 1 (17.5) 5 (82.5) \$ 1 (17.5) 5 (82.5) \$ 1 (17.5) 5 (82.5) \$ <td>Menopausal status, n (%)</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td>	Menopausal status, n (%)						
<table-container>post-25 (48.1)27 (51.9)7 (13.5)45 (86.5)Turor size, n (%)≤20 mm8 (47.1)9 (52.9)2 (11.8)63 (81.8)9 (82.5)> 20 mm13 (34.2)4 (57.1)0.93614 (18.2)63 (81.8)9 (81.8)9 (81.8)> 20 mm150.001 (50.0)1 (50.0)1 (50.0)1 (50.0)1 (50.0)1 (50.0)LN status, n (%)1 (17.5)5 (82.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)Ustatus, n (%)1 (17.5)5 (82.5)1 (17.5)5 (82.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)Ustatus, n (%)5 (82.5)1 (17.5)5 (82.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (81.5)9 (</table-container>	pre-	17 (38.6)	27 (61.4)	0.353	10 (22.7)	34 (77.3)	0.236
<table-container>Tumor size, n (%)\$20 mm8 (47.1)9 (52.9)2 (1.8)15 (88.2)> 20 mm3 (40.2)4 (57.1)0.9361 (10.2)6 (81.2)0.936Inknown1 (50.0)1 (50.0)1 (50.0)1 (50.0)1N status, n (%)1 (17.5)5 (82.5)0.936positive23 (35.5)40 (63.5)1 (17.5)5 (82.5)0.936negative19 (57.6)14 (42.4)0.048*6 (18.2)27 (81.8)0.936*ductal10 (76.9)3 (31.0)0.10*5 (35.0)0.935*9 (31.4)1 (17.5)5 (82.5)0.935*special10 (76.9)3 (31.0)0.10*1 (31.6)1 (81.5)0.935*9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)9 (31.6)</table-container>	post-	25 (48.1)	27 (51.9)		7 (13.5)	45 (86.5)	
\$20 mm8 (47.1)9 (52.9)2 (11.8)1 5 (88.2)>20 mm3 (42.9)44 (57.1)0.93614 (18.2)63 (81.8)0.366unknown1 (50.0)1 (50.0)1 (50.0)1 (50.0)1 (50.0)1 (50.0)LN staus, n (%)5 (85.5)11 (17.5)5 (82.5)0.035'negative19 (57.6)14 (42.4)0.048'6 (18.2)2 (81.5)0.035'special10 (76.9)3 (23.1)5 (38.5)8 (61.5)0.035'special10 (76.9)3 (24.1)5 (38.5)8 (61.5)0.035'negative8 (32.0)17 (68.0)0.1484 (16.0)2 (48.0)0.842unknown2 (25.0)6 (75.0)11 (17.5)5 (82.5)0.035'negative8 (32.0)17 (68.0)0.1484 (16.0)2 (48.0)0.842unknown2 (25.0)6 (75.0)11 (17.5)5 (85.2)0.035'negative8 (32.0)17 (68.0)0.0644 (16.0)2 (48.0)0.842unknown3 (25.1)13 (76.5)0.06412 (11.8)15 (88.2)0.479matectomy8 (48.1)14 (15.9)0.06412 (11.8)15 (88.2)0.035'none8 (45.1)13 (76.5)0.06412 (11.8)16 (8.0)0.051unknotn3 (41.5)15 (85.2)15 (85.2)0.051'10.0110.01unknotn16 (45.2)13 (76.5)0.06412 (14.5)71 (85.5)0.035'none8 (61.5) <t< td=""><td>Tumor size, n (%)</td><td></td><td></td><td></td><td></td><td></td><td></td></t<>	Tumor size, n (%)						
> 20 mm 33 (429) 44 (57.1) 0.936 14 (18.2) 63 (81.8) 0.396 unknown 1 (50.0) 1 (50.0) 1 (50.0) 1 (50.0) 1 (50.0) LN status, n (%)	≤ 20 mm	8 (47.1)	9 (52.9)		2 (11.8)	15 (88.2)	
unknown 1 (500) 1 (500) 1 (500) 1 (500) LN status, n (%) 23 (36.5) 40 (63.5) 11 (17.5) 52 (82.5) negative 19 (57.6) 14 (42.4) 0.048 ³ 6 (18.2) 27 (81.8) 0.930 Histocycal type, n (%) 10 (76.9) 3 (16 1.4) 0.010 ⁴ 12 (14.5) 71 (85.5) 0.035 ⁴ goedal 10 (76.9) 3 (13 (12.2) 52 (82.5) 0.035 ⁴ 52 (82.5) 0.042 11 (17.5) 52 (82.5) 0.042 0.048 ⁴ 0.160.0 0.042 10 (60.0) 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.	> 20 mm	33 (42.9)	44 (57.1)	0.936	14 (18.2)	63 (81.8)	0.396
LN status, n (%) 23 (36.5) 40 (63.5) 11 (17.5) 52 (82.5) negative 19 (57.6) 14 (42.4) 0.048" 6 (18.2) 27 (81.8) 0.930 Histological type, n (%) 32 (38.6) 51 (61.4) 0.010" 12 (14.5) 71 (85.5) 0.035" gocial 10 (76.9) 32 (31.1) 5 (38.5) 8 (61.5) 0.035" LN status, n (%) 11 (17.5) 5 (28.25) 8 (32.0) 11 (17.5) 5 (28.25) unknown 2 (50.8) 31 (49.2) 11 (17.5) 5 (28.25) 4 (42.2) 0.042 10 (0.4) 0.422 unknown 2 (50.0) 17 (68.0) 0.148 4 (16.0) 2 (28.0) 0.424 unknown 2 (50.0) 13 (76.5) 0.064 2 (11.8) 15 (88.2) 0.479 mastectomy 3 (84.1) 13 (76.5) 0.064 2 (11.8) 15 (88.2) 0.403 chorenterapy, n (%) 3 (6.15) 5 (89.2) 0.015" 16 (19.2) 0.015" adjuvant 6 (50.2) 5 (38.2) 6 (19.2) 12 (40.9)	unknown	1 (50.0)	1 (50.0)		1 (50.0)	1 (50.0)	
positive 23 (36.5) 40 (63.5) 11 (17.5) 52 (82.5) negative 19 (57.6) 14 (42.4) 0.048* 6 (18.2) 27 (81.8) 0.930 Histological type, n (%) 32 (38.6) 51 (61.4) 0.010* 12 (14.5) 71 (85.5) 0.035* special 10 (76.9) 3 (23.1) 5 (38.5) 8 (61.5) 0.014* LVI status, n (%) 5 (38.5) 8 (61.5) 0.84 positive 32 (50.8) 31 (49.2) 11 (17.5) 52 (82.5) negative 8 (32.0) 17 (68.0) 0.148 4 (16.0) 21 (84.0) 0.842 unown 2 (25.0) 6 (75.0) 2 (25.0) 6 (75.0) 10 (17.8) 15 (88.2) 0.479 mastectomy 3 (4 (1.0) 13 (76.5) 0.064 12 (11.8) 15 (88.2) 0.479 none 3 (4 (1.0) 49 (59.0) 0.164 12 (14.5) 14 (8.0) 0.99 induvant 3 (4 (1.0) 49 (59.0) 0.164 12 (4.0) 38 (76.0) 0.92	LN status, n (%)						
negative 19 (57.6) 14 (42.4) 0.048* 6 (18.2) 27 (81.8) 0.930 Histological type, n (%) 32 (38.6) 51 (61.4) 0.010* 12 (14.5) 71 (85.5) 0.035* special 10 (76.9) 3 (23.1) 5 (38.5) 8 (61.5) 0.035* LVI status, n (%) 71 (85.0) 11 (17.5) 5 (28.5) 8 (61.5) positive 8 (32.0) 17 (68.0) 0.148 4 (16.0) 2 (28.0) 0.842 unknown 2 (25.0) 6 (75.0) 2 (25.0) 6 (75.0) 0.064 2 (11.8) 15 (88.2) 0.449 opartion status, n (%) 71 (85.5) 14 (15.9) 15 (19.0) 15 (19.0) 0.649 2 (11.8) 15 (88.2) 0.479 mastectomy 8 (43.1) 13 (76.5) 0.064 2 (11.8) 15 (80.2) 0.479 inone 8 (61.5) 5 (38.5) 5 (38.5) 8 (61.5) 10 (11.5) 10 (11.5) 10 (11.5) 10 (11.5) 10 (11.5) 10 (11.5) 10 (11.5) 10 (11.5) 10 (11.5)	positive	23 (36.5)	40 (63.5)		11 (17.5)	52 (82.5)	
Histological type, n (%) 32 (38.6) 51 (61.4) 0.010* 12 (14.5) 71 (85.5) 0.03* special 10 (76.9) 3 (23.1) 5 (38.5) 8 (61.5) LVI status, n (%) 11 (17.5) 5 (28.2) 1 negative 8 (32.0) 17 (68.0) 0.148 4 (16.0) 2 (84.0) 0.842 unknown 2 (25.0) 6 (75.0) 2 (25.0) 6 (75.0) 0.064 2 (11.8) 15 (88.2) 0.479 partial mastectomy 4 (23.5) 13 (76.5) 0.064 2 (11.8) 15 (88.2) 0.479 mastectomy 38 (84.1) 13 (76.5) 0.064 2 (11.8) 15 (88.2) 0.479 gluvant 8 (48.1) 13 (76.5) 0.064 2 (11.8) 15 (88.2) 0.479 ajuvant 9 (36.1) 15 (19.0) 16 (81.0) 16 (81.0) 16 (81.0) 16 (81.0) rome 16 (34.0) 0.052 5 (38.5) 16 (91.0) 16 (91.0) 17 (91.0) 17 (91.0) 17 (91.0) 17 (91.0) 17	negative	19 (57.6)	14 (42.4)	0.048*	6 (18.2)	27 (81.8)	0.930
ductal 32 (38.6) 51 (61.4) 0.010 [*] 12 (14.5) 71 (85.5) 0.03 [*] special 10 (76.9) 3 (23.1) 5 (38.5) 8 (61.5) LVI status, n (%) 11 (17.5) 5 (28.2.5) 8 (61.2) negative 8 (32.0) 17 (68.0) 0.148 4 (16.0) 21 (84.0) 0.842 unknown 2 (25.0) 6 (75.0) 2 (25.0) 6 (75.0) 0.064 2 (11.8) 15 (88.2) 0.479 partial mastectomy 4 (23.5) 13 (76.5) 0.064 2 (11.8) 15 (88.2) 0.479 mastectomy 3 (4 (1.0) 13 (76.5) 0.064 2 (11.8) 15 (88.2) 0.479 mastectomy 3 (4 (8.1) 13 (76.5) 0.064 2 (11.8) 15 (88.2) 0.479 mastectomy 8 (48.1) 4 (9 (59.0) 0.064 2 (18.0) 6 (15.0) 0.035* none 8 (61.5) 5 (38.5) 5 (38.5) 8 (61.5) 0.092* inversert 3 (6 (5.2) 5 (38.5) 5 (10.9) 14 (89.	Histological type, n (%)						
special 10 (76.9) 3 (23.1) 5 (38.5) 8 (61.5) LV status, n (%) 31 (49.2) 11 (17.5) 52 (82.5) negative 32 (50.8) 13 (49.2) 11 (17.5) 52 (82.5) 0.842 negative 8 (32.0) 17 (68.0) 0.148 4 (16.0) 21 (84.0) 0.842 unknown 2 (25.0) 6 (75.0) 2 (25.0) 6 (75.0) 2 (25.0) 6 (75.0) Operation status, n (%) y 31 (76.5) 0.064 2 (11.8) 6 (81.0) 0.47.0 mastectomy 4 (32.5) 13 (76.5) 0.064 2 (11.8) 6 (81.0) 0.47.0 mastectomy 4 (32.5) 13 (76.5) 0.064 2 (11.8) 6 (81.0) 0.47.0 chemoty, n (%) y 5 (38.5) 5 (18.9) 5 (82.0) 0.03* adjuvant 6 (65.0) 6 (36.2) 5 (0.89 3 (66.2) 6 (19.1) 9 (19.1) 10.0 rimary ER status, n (%) y 3 (65.2) 6 (19.1) 5 (19.9) 3 (19.1) 11.0 <t< td=""><td>ductal</td><td>32 (38.6)</td><td>51 (61.4)</td><td>0.010*</td><td>12 (14.5)</td><td>71 (85.5)</td><td>0.035*</td></t<>	ductal	32 (38.6)	51 (61.4)	0.010*	12 (14.5)	71 (85.5)	0.035*
LVI status, n (%) 32 (50.8) 31 (49.2) 11 (17.5) 52 (82.5) negative 8 (32.0) 17 (68.0) 0.148 4 (16.0) 2 (84.0) 0.842 unknown 2 (25.0) 6 (75.0) 2 (25.0) 6 (75.0) 6 (75.0) Operation status, n (%) 1 (17.6) 5 (82.0) 0.842 mastectomy 3 (84.1) 1 (17.9) 6 (81.0) 0.479 mastectomy 3 (84.1) 1 (17.9) 6 (81.0) 0.479 digluvant 3 (41.0) 4 (95.9) 0.644 1 (17.9) 6 (81.0) thrmore therapy, n (%) 3 (81.2) 5 (85.0) 6 (81.5) 0.035* gluvant 2 (6 52.0) 24 (48.0) 0.89 1 (24.0) 3 (76.0) 0.992 none 1 (6 (48.0) 0.089 1 (24.0) 3 (87.0) 0.992 indiguvant 2 (6 52.0) 2 (4 (48.0) 0.089 1 (24.0) 3 (87.0) 0.992 indiguvant 2 (6 52.0) 3 (0 (52.1) 5 (10.9) 3 (10.1)	special	10 (76.9)	3 (23.1)		5 (38.5)	8 (61.5)	
positive32 (50.8)31 (49.2)11 (17.5)52 (82.5)negative8 (32.0)17 (68.0)0.1484 (16.0)21 (84.0)0.842unknown2 (25.0)6 (75.0)2 (25.0)6 (75.0)2Operation status, <i>n</i> (%)131 (76.5)0.0642 (11.8)15 (88.2)0.479partial mastectomy4 (23.5)13 (76.5)0.0642 (11.8)15 (88.2)0.479mastectomy8 (48.1)13 (76.5)0.0642 (11.8)6 (40.1)0.479chemotherapy, <i>n</i> (%)4 (15.9)15 (19.0)6 (19.0)0.035*6 (35.0)0.035*adjuvant8 (41.0)49 (59.0)0.16412 (14.5)71 (85.5)0.035*none8 (61.5)5 (38.5)5 (38.5)6 (51.0)0.092adjuvant6 (52.0)24 (48.0)0.08912 (24.0)38 (76.0)0.092none16 (34.8)2 (65.2)5 (19.0)14 (89.1)1111positive2831 (65.2)5 (19.0)12 (24.0)38 (76.0)0.012*none16 (34.8)2 (34.0)0.019*12 (3.0)12 (3.0)0.112positive2831 (3.1)12 (3.0)350.049*rumary RY status, <i>n</i> (%)113 (19.0)12 (19.0)350.049*positive2423 (20.0)15 (20.0)12 (20.0)3510 (20.0)positive2831 (20.0)12 (20.0)36 (20.0)12 (20.0)12 (20.0)pos	LVI status, n (%)						
negative 8 (32.0) 17 (68.0) 0.148 4 (16.0) 2 (84.0) 0.842 unknown 2 (25.0) 6 (75.0) 2 (25.0) 6 (75.0) 2 operation status, n (%) 2 2 (11.8) 15 (88.2) 0.479 mastectomy 4 (23.5) 13 (76.5) 0.064 2 (11.8) 15 (88.2) 0.479 mastectomy 38 (48.1) 4 (51.9) 0.064 2 (11.8) 64 (81.0) 0.479 chemotherapy, n (%) 3 4 (51.9) 0.164 12 (14.5) 71 (85.5) 0.035* none 8 (61.5) 5 (38.5) 5 (38.5) 8 (61.5) 0.035* adjuvant 8 (61.5) 5 (38.5) 5 (38.5) 8 (61.5) 0.092* none 16 (34.8) 3 (65.2) 5 (0.90 12 (24.0) 38 (76.0) 0.092 none 16 (34.8) 3 (65.2) 5 (10.9) 4 (89.1) 12 positive 24 3 (3 (20.1) 5 (10.9) 3 (10.1) 12 negative 14	positive	32 (50.8)	31 (49.2)		11 (17.5)	52 (82.5)	
unknown 2 (25.0) 6 (75.0) 2 (25.0) 6 (75.0) Operation status, n (%) <t< td=""><td>negative</td><td>8 (32.0)</td><td>17 (68.0)</td><td>0.148</td><td>4 (16.0)</td><td>21 (84.0)</td><td>0.842</td></t<>	negative	8 (32.0)	17 (68.0)	0.148	4 (16.0)	21 (84.0)	0.842
Operation status, n (%) partial mastectomy 4 (23.5) 13 (76.5) 0.064 2 (11.8) 15 (88.2) 0.479 mastectomy 38 (48.1) 41 (51.9) 15 (19.0) 64 (81.0) Chemotherapy, n (%) adjuvant 34 (41.0) 49 (59.0) 0.164 12 (14.5) 71 (85.5) 0.033 [*] none 8 (61.5) 5 (38.5) 5 (38.5) 5 (38.5) 61.5) 0.035 [*] adjuvant 8 (61.5) 5 (38.5) 5 (38.5) 8 (61.5) 0.032 [*] none 16 (34.8) 24 (48.0) 0.089 12 (24.0) 38 (76.0) 0.092 none 16 (34.8) 30 (65.2) 5 (10.9) 41 (89.1) 12 (24.0) 14 (89.1) Primary ER status, n (%) 1 5 (10.9) 14 (89.1) 12 (14.0) 12 (14.0) 12 (14.0) 12 (14.0) 12 (14.0) 12 (14.0) 12 (14.0) 12 (14.0) 12 (14.0) 12 (14.0) 12 (14.0) 12 (14.0) 12 (14.0) 12 (14.0) 12 (14.0) 12 (14.0) <td>unknown</td> <td>2 (25.0)</td> <td>6 (75.0)</td> <td></td> <td>2 (25.0)</td> <td>6 (75.0)</td> <td></td>	unknown	2 (25.0)	6 (75.0)		2 (25.0)	6 (75.0)	
partial mastectomy 4 (23.5) 13 (76.5) 0.064 2 (11.8) 15 (88.2) 0.479 mastectomy 38 (48.1) 4 (51.9) 15 (19.0) 64 (81.0) Chemotherapy, n (%) 71 (85.5) 0.035* adjuvant 34 (41.0) 49 (59.0) 0.164 12 (14.5) 71 (85.5) 0.035* none 8 (61.5) 5 (38.5) 5 (38.5) 8 (61.5) 0.035* Hormone therapy, n (%) 38 (76.0) 0.092 38 (76.0) 0.092 none 16 (34.8) 30 (65.2) 5 (10.9) 31 (80.1) 0.092 positive 16 (34.8) 30 (65.2) 5 (10.9) 41 (89.1) 112 positive 28 23 0.019* 12 (3.0) 39.0 0.112 regative, n (%) 12 39.0 0.149* 114 114 114 114 114 114 114 114 114 114 114	Operation status, <i>n</i> (%)						
mastectomy 38 (48.1) 41 (51.9) 15 (19.0) 64 (81.0) Chemotherapy, n (%) 34 (41.0) 49 (59.0) 0.164 12 (14.5) 71 (85.5) 0.035 [*] none 8 (61.5) 5 (38.5) 5 (38.5) 5 (38.5) 8 (61.5) Hormone therapy, n (%)	partial mastectomy	4 (23.5)	13 (76.5)	0.064	2 (11.8)	15 (88.2)	0.479
Adjuvant 34 (41.0) 49 (59.0) 0.164 12 (14.5) 71 (85.5) 0.035 [*] none 8 (61.5) 5 (38.5) 5 (38.5) 5 (38.5) 8 (61.5) Hormone therapy, n (%) 44 (48.0) 0.089 12 (24.0) 38 (76.0) 0.092 none 16 (34.8) 30 (65.2) 5 (10.9) 41 (89.1) 0.092 Primary ER status, n (%) 5 (10.9) 12 (24.0) 39.0 0.112 positive 16 (34.8) 30 (65.2) 5 (10.9) 41 (89.1) 0.092 10.002 112 positive 16 (34.8) 30 (65.2) 5 (10.9) 40 0.112 112 negative n (%) 12 (24.0) 12 (24.0) 39.0 0.112 112 112 positive 16 (34.8) 30 (65.2) 5 (10.9) 12 (24.0) 39.0 0.112 negative n (%) 12 (10.000 12 (10.000 12 (10.000 12 (10.000 12 (10.000 primary PR status, n (%) 21 (20.000 12 (20.000 12 (20.000 12 (20.000 12 (20.000 12 (20.000 12 (20.000 <	mastectomy	38 (48.1)	41 (51.9)		15 (19.0)	64 (81.0)	
adjuvant 34 (41.0) 49 (59.0) 0.164 12 (14.5) 71 (85.5) 0.035 [*] none 8 (61.5) 5 (38.5) 5 (38.5) 5 (38.5) 8 (61.5) Hormone therapy, <i>n</i> (%) adjuvant 26 (52.0) 24 (48.0) 0.089 12 (24.0) 38 (76.0) 0.092 none 16 (34.8) 30 (65.2) 5 (10.9) 41 (89.1) 0.092 Primary ER status, <i>n</i> (%) 5 (30.9) 12 (24.0) 38 (76.0) 0.092 nope 16 (34.8) 30 (65.2) 5 (10.9) 41 (89.1) 0.092 Primary ER status, <i>n</i> (%) 12 5 (10.9) 41 (89.1) 0.112 negative 14 31 5 40 0.112 Primary PR status, <i>n</i> (%) 5 23 0.157 12 35 0.049 [*]	Chemotherapy, n (%)						
none 8 (61.5) 5 (38.5) 5 (38.5) 8 (61.5) Hormone therapy, n (%) - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - - -	adjuvant	34 (41.0)	49 (59.0)	0.164	12 (14.5)	71 (85.5)	0.035*
Hormone therapy, n (%) 26 (52.0) 24 (48.0) 0.089 12 (24.0) 38 (76.0) 0.092 none 16 (34.8) 30 (65.2) 5 (10.9) 41 (89.1) Primary ER status, n (%) 90 23 0.019* 12 24.00 39.00 0.112 negative 14 31 5 50.00 40 112 primary PR status, n (%) 90 9.157 12 Andread 35 0.049*	none	8 (61.5)	5 (38.5)		5 (38.5)	8 (61.5)	
adjuvant 26 (52.0) 24 (48.0) 0.089 12 (24.0) 38 (76.0) 0.092 none 16 (34.8) 30 (65.2) 5 (10.9) 41 (89.1) Primary ER status, n (%) 9 23 (24.0) 12 (24.0) 38 (76.0) 0.092 positive 28 23 0.019* 12 (24.0) 41 (89.1) 12 negative 14 31 5 40 112 Primary PR status, n (%) 9 23 0.157 12 35 0.049*	Hormone therapy, <i>n</i> (%)						
none 16 (34.8) 30 (65.2) 5 (10.9) 41 (89.1) Primary ER status, n (%) 39 0.112 positive 28 23 0.019* 12 39 0.112 negative 14 31 5 40 Primary PR status, n (%) 12 35 0.049*	adjuvant	26 (52.0)	24 (48.0)	0.089	12 (24.0)	38 (76.0)	0.092
Primary ER status, n (%) 28 23 0.019* 12 39 0.112 negative 14 31 5 40 Primary PR status, n (%) 24 23 0.157 12 35 0.049*	none	16 (34.8)	30 (65.2)		5 (10.9)	41 (89.1)	
positive 28 23 0.019* 12 39 0.112 negative 14 31 5 40 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 12 <td>Primary ER status, n (%)</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td>	Primary ER status, n (%)						
negative 14 31 5 40 Primary PR status, n (%) 24 23 0.157 12 35 0.049*	positive	28	23	0.019*	12	39	0.112
Primary PR status, n (%) 24 23 0.157 12 35 0.049*	negative	14	31		5	40	
positive 24 23 0.157 12 35 0.049*	Primary PR status, <i>n</i> (%)						
	positive	24	23	0.157	12	35	0.049*
negative 18 31 5 44	negative	18	31		5	44	
Primary HER2 status, n (%)	Primary HER2 status, <i>n</i> (%)						
positive 1 15 0.001 [*] 0 16 0.042 [*]	positive	1	15	0.001*	0	16	0.042*
negative 41 39 17 63	negative	41	39		17	63	
Primary Ki-67 expression, n (%)	Primary Ki-67 expression, n ((%)					
high 8 47 <0.001 [*] 2 53 <0.001 [*]	high	8	47	< 0.001*	2	53	< 0.001*
low 34 7 15 26	low	34	7		15	26	

33	0.689
46	
30	0.254
49	
14	0.060
65	
70	< 0.001*
9	
32 (86.5)	0.012*
32 (91.4)	
15 (62.5)	
27 (87.1)	0.394
52 (80)	
70 (84.3)	0.185
9 (69.2)	
	14 65 70 9 32 (86.5) 32 (91.4) 15 (62.5) 27 (87.1) 52 (80) 70 (84.3) 9 (69.2)

Table 4 Relationship between EZH2 expression in primary and metastatic breast cancer lesions and the clinicopathological characteristics of patients (n = 96) (*Continued*)

Abbreviations: ER estrogen receptor, EZH2 enhancer of zeste homolog 2, HER2 human epidermal growth factor receptor 2, LN lymph node, LVI lymphovascular invasion, PR progesterone receptor, SD standard deviation

^aSpecial type is invasive breast carcinoma except invasive ductal carcinoma

^bViscera includes brain, lung, liver and ovary

^cSoft tissues includes chest wall, lymph node, distant skin

^{*}Indicates values that are statistically significant (P < 0.05)

In the univariate analysis, compared with low EZH2 expression, high EZH2 expression was not a poor prognostic indicator of OS after recurrence outcome in PBC (hazard ratio [HR] 1.449; 95% confidence interval [CI] 0.930-2.258, P = 0.101) (Table 5), whereas in metastatic lesions, high EZH2 expression was a poor prognostic indicator of OS outcome after recurrence (HR 2.116; 95% CI 1.143-3.916, P = 0.017) (Table 5). Other poor prognostic indicators of OS outcome after recurrence in PBC and metastatic lesions from the univariate analysis included primary ER or primary PR status, primary and metastatic high Ki-67 expression, and ≤2 years of DFI (Table 5). A Cox proportional-hazards model using multivariate analysis but not including Ki-67 expression demonstrated that high EZH2 expression was independently associated with poorer OS outcomes after recurrence in patients with metastatic lesions (HR 2.047; 95% CI 1.074–3.902, *P* = 0.029) (Table 5). Multivariate analysis of prognostic factors related to OS after recurrence including Ki-67 expression is shown in Additional file 3: Table S1.

Discussion

In this study, using primary and paired metastatic lesions from patients with MBC, EZH2 expression scores correlated significantly with Ki-67 expression scores in both primary and metastatic lesions, and Ki-67 expression and EZH2 expression scores were significantly higher in metastatic lesions compared with PBC lesions. Because Ki-67 expression scores in metastatic lesions increased more than in PBC lesions, we considered that proliferation in metastatic lesions increased more than in PBC lesions. We expected that EZH2 expression in metastatic lesions would be higher than that in PBC lesions. We showed that EZH2 expression correlated significantly with the Ki-67 expression in both PBC and metastatic lesions. We thought that EZH2 promotes breast cancer progression by transcriptional repression of tumor suppressors; consequently, Ki-67 expression increased in breast cancer cells with high EZH2 expression. We showed that EZH2 expression levels in bone metastatic lesions were significantly lower than in viscera and soft tissue metastatic lesions. Expression of



EZH2 in primary tumors of patients with DFIs ≤ 2 years was higher than in those with DFIs >2 years, whereas expression of EZH2 in primary tumors of patients with DFIs >10 years was lower than in those with DFIs \leq 10 years. Given that bone metastasis occurs more frequently in ER-positive than ER-negative breast cancer [26] and in MBC patients with DFIs >10 years [27], we expected that EZH2 expression would be low in bone

Prognostic factor Patients (n = 96)	Patients	Univariate	Univariate analysis			Multivariate analysis		
	HR	95% CI	P-value	HR	95% CI	P-value		
Menopausal status, n (%)								
pre-	44 (45.8)	1						
post-	52 (54.2)	1.396	0.891-2.186	0.145				
Tumor size, n (%)								
≤ 20 mm	17 (17.7)	1						
> 20 mm	77 (80.2)	1.317	0.726-2.388	0.364				
unknown	2 (2.1)							
LN status, n (%)								
positive	63 (65.6)	1.309	0.829-2.066	0.248				
negative	33 (34.4)	1						
Histological type, n (%)								
ductal	83 (86.5)	1						
speciala	13 (13.5)	1.028	0.543-1.945	0.932				
LVI status, n (%)								
positive	63 (65.6)	1.287	0.769-2.156	0.337				
negative	25 (26.1)	1						
unknown	8 (8.3)							
Operation status, n (%)								
partial mastectomy	17 (17.7)	1						
mastectomy	79 (82.3)	0.894	0.500-1.596	0.704				
Chemotherapy, n (%)								
adjuvant	83 (86.5)	1.320	0.707-2.461	0.383				
none	13 (13.5)	1						
Hormone therapy, <i>n</i> (%)								
adjuvant	50 (52.1)	0.719	0.460-1.122	0.146				
none	46 (47.9)	1						
Primary ER status, n (%)								
positive	51 (53.1)	0.541	0.344-0.850	0.008*	0.724	0.382-1.372	0.322	
negative	45 (46.9)	1						
Primary PR status, n (%)								
positive	47 (49.0)	0.595	0.380-0.931	0.023*	0.911	0.484-1.714	0.773	
negative	49 (51.0)	1						
Primary HER2 status, n (%)								
positive	16 (16.7)	1.122	0.616-2.046	0.706				
negative	80 (83.3)	1						
Primary Ki-67 expression, n (%) ^b							
high	55 (57.3)	1.848	1.170-2.917	0.008*				
low	41 (42.7)	1						
Primary EZH2 expression, n	(%)							
high	54 (56.3)	1.449	0.930–2.258	0.101				
low	42 (43.7)	1						

Table 5 Univariate and multivariate analysis of prognostic factors related to overall survival after recurrence (n = 96)

Metastatic ER status, n ((%)						
positive	41 (42.7)	0.972	0.625-1.513	0.901			
negative	55 (57.3)	1					
Metastatic PR status, n	(%)						
positive	39 (40.6)	0.692	0.440-1.089	0.112			
negative	57 (59.4)	1					
Metastatic HER2 status,	n (%)						
positive	14 (14.6)	1.282	0.673-2.440	0.450			
negative	82 (85.4)	1					
Metastatic Ki-67 express	sion, <i>n</i> (%) ^b						
high	72 (75.0)	2.422	1.394-4.206	0.002*			
low	24 (25.0)	1					
Metastatic EZH2 expres	sion, <i>n</i> (%)						
high	59 (77.6)	2.116	1.143-3.916	0.017*	2.047	1.074-3.902	0.029*
low	17 (22.4)	1					
Metastatic sites, n (%)							
bones	24 (25.0)	0.899	0.548-1.476	0.674			
others	72 (75.0)	1					
Disease free interval							
≤ 2 years	28 (29.0)	1.713	1.086-2.702	0.021*	1.601	0.978-2.621	0.061
> 2 years	68 (71.0)	1					
≤ 10 years	83 (87.0)	1					
> 10 years	13 (13.0)	0.861	0.442-1.679	0.661			

Table 5 Univariate and multivariate analysis of prognostic factors related to overall survival after recurrence (n = 96) (Continued)

Abbreviations: CI confidence interval, ER estrogen receptor, EZH2 enhancer of zeste homolog 2, HER2 human epidermal growth factor receptor 2, HR hazard ratio, LN lymph node, LVI lymphovascular invasion, PR progesterone receptor

^aSpecial type is invasive breast carcinoma except invasive ductal carcinoma

^bKi-67 expression was excluded from the multivariate analysis because of an association with EZH2 expression in primary and metastatic breast cancer lesions ^{*}Indicates values that are statistically significant (P < 0.05)

metastasis and in MBC patients with DFIs >10 years. However, because MBC patients with DFIs ≤ 2 years had a poor prognosis, we expected that EZH2 expression levels would be higher. We showed that high EZH2 expression in primary lesions was shown to be independently associated with poorer DFS and OS outcomes after primary surgery in MBC, whether or not high EZH2 expression in primary lesions was shown to be associated with OS after recurrence. High EZH2 expression in metastatic lesions was not associated with DFS after primary surgery, even if high EZH2 expression in metastatic lesions was shown to be independently associated with poorer OS outcomes after primary surgery and recurrence. We had considered the proliferation of primary lesions associated with DFS and OS after primary surgery until recurrence; on the other hand, proliferation of metastatic lesions was more associated with OS after recurrence than PBC lesions.

Previous reports have demonstrated that in PBC, EZH2 expression was significantly increased in malignant tumors, and was associated with a larger tumor

size, ER- and PR-negative status, TNBC, advanced stage disease and reduced progression-free survival and OS [17, 28, 29]. In colon cancer and poorly differentiated synovial sarcomas, EZH2 expression was significantly related to increased tumor cell proliferation, as assessed using the Ki-67 expression [30, 31]. Nishimura et al. [12] reported that, in comparison to primary lesions, the Ki-67 expression score increased significantly in metastatic lesions. We found that in breast cancer patients, EZH2 expression scores correlated significantly with Ki-67 expression scores in both primary and metastatic lesions and Ki-67 expression and EZH2 expression scores were significantly higher in metastatic lesions compared with PBC lesions. Furthermore, high EZH2 expression in metastatic lesions was shown to be independently associated with poorer OS outcomes after recurrence in MBC. Few reports have examined the relationship between EZH2 expression in metastatic lesions and outcomes in patients with MBC.

MBC is difficult to treat using conventional therapies that are currently available on the market, and development of

new therapeutic approaches is needed. Considering the downstream effects of EZH2, silencing of the EZH2 gene in the ER+ MCF-7 cell line resulted in higher expression of ER and increased sensitivity to anti-estrogen therapy [32]. EZH2 gene silencing has also been reported to result in a significant reduction in tumor growth in the MB-231 TNBC orthotopic mouse model of breast carcinomas. High EZH2 expression was shown to be significantly associated with TNBC and reduced OS outcomes [28]. In our cohort of MBC patients, we demonstrated a significant correlation between EZH2 expression and Ki-67 expression scores in primary and metastatic lesions. Therefore, EZH2 may represent a potential therapeutic target for this aggressive breast cancer that exhibits high expression levels of Ki-67, thus warranting further investigation. Using data obtained in this study as a reference of expression of EZH2 status in metastatic lesions and the correlation between EZH2 and other biomarkers in MBC, biopsy of metastatic lesions may become less necessary, thereby avoiding risk of vital organ damage because of the biopsy procedure.

Recently, several EZH2 inhibitors have been developed and tested in multiple types of cell lines and xenografts [33, 34]. Both EPZ-6438 (E7438) and GSK126, selective small-molecule inhibitors of histone methyltransferase activity, have yielded promising results in small cell lung cancer cell lines and malignant rhabdoid tumors [33, 34]. In the second quarter of 2015, Epizyme^o Inc., Cambridge, MA, USA initiated a phase 2 monotherapy trial of EPZ-6438 in patients with relapsed or refractory non-Hodgkin lymphomas [35].

This study has some limitations. First, the retrospective nature of the study design was prone to selection bias. Patients in this study tended to have a more aggressive breast cancer with poorer prognosis in comparison with a group from the general breast cancer patient population, since all of those recruited were diagnosed with MBC. Second, this study could not consider the effects of adjuvant therapy, which differed according to each patient, owing to the fact that patients were recruited over a long period, from 1977 to 2013. Previous reports have demonstrated that examining change due to treatment based on Ki-67 expression, the number of responders to endocrine therapy as the neoadjuvant therapy declined, and the prognosis of patients exhibiting decreased levels of Ki-67 was good [36]. In addition, the prognosis of patients with decreased levels after chemotherapy was also reported to be good [37, 38]. Third, the EZH2 scoring method in this study was used in house as described in the Materials and Methods section. Forth, we could not assess EZH2 expression in this study according to subtype (luminal A, luminal B, luminal HER2, HER2-type and triple-negative breast cancer) due to sample limitation. We assessed the clinicopathological significance of EZH2 expression in MBC according to subtype; however, the number of breast cancer patients for each subtype was too small to show statistical significance. Increasing the number of MBC patients in this study was not possible since we had already collected samples from 96 patients between 1977 and 2013, which was the maximum that was achievable. Fifth, we used a HER2 immunostaining score of 2+ to designate a negative score, as described in the Materials and Methods section and Additional file 1.

As further investigations, we think that examination of the correlation between EZH2 expression levels and pathological response, and the correlation between EZH2 expression levels and prognosis in neoadjuvant chemotherapy and hormonal therapy should be considered.

Conclusion

Our results suggest that EZH2 expression levels correlate significantly with the Ki-67 expression score. Therefore, EZH2 may represent a potential therapeutic target for this aggressive breast cancer, especially for those with a high Ki-67 expression score, which warrants further investigation. EZH2 expression scores were significantly higher in metastatic lesions compared with PBC lesions. We also showed that high EZH2 expression levels in primary lesions were independently associated with poorer DFS and OS after primary surgery, and that high EZH2 expression levels in metastatic lesions were independently associated with poorer OS outcomes after primary surgery and recurrence. A part of the present work was precedingly reported at the 2016 ASCO Annual Meeting [39].

Additional files

Additional file 1: Reasons for assessing cases scored as HER2 2+ as negative. (DOC 23 kb)

Additional file 2: Evaluation of immunostaining of EZH2 and Ki67. (DOC 52 kb)

Additional file 3: Table S1. Univariate and multivariate analysis of prognostic factors related to overall survival after recurrence including Ki-67 expression. (DOC 111 kb)

```
Additional file 4: Dataset of this study. (XLSX 26 kb)
```

Abbreviations

CI: Confidence interval; DFI: Disease-free interval; DFS: Disease-free survival; ER: Estrogen receptor; EZH2: Enhancer of zeste homolog 2; HER2: Human epidermal growth factor receptor 2; HR: Hazard ratio; IHC: Immunohistochemistry; MBC: Metastatic breast cancer; OS: Overall survival; PBC: Primary breast cancer; PR: Progesterone receptor; TMA: Tissue microarray; TNBC: Triple-negative breast cancer

Acknowledgements

We would like to express our gratitude to the staff in the Department of Pathology, Kanagawa Cancer Center, for their technical assistance and for collecting cancer tissue.

Funding

This work was partly supported by the Platform of Supporting Cohort Study and Biospecimen Analysis, Grant-in-Aid for Scientific Research on Innovative Areas (16H06277).

Availability of data and materials

The dataset supporting the conclusions of this article is included within the article (and its Additional file 4).

Authors' contributions

HI, NS, TO, and YM designed the study. HI performed clinical, pathological and, statistical investigation, and drafted the manuscript. KK participated in the histological and immunohistochemical evaluation. HI, TY and MY constructed TMAs. HI, TY and YN performed the immunohistochemical staining. TY, HN, AY, and KM assisted the clinical investigation. TY, YR, SS, and MM participated in preparing and drafting the manuscript. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Consent for publication

Not applicable.

Ethics approval and consent to participate

All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards. The Research Ethics Committee, Kanagawa Cancer Center, Yokohama, Japan, approved this study protocol. The reference number of this study is EKI-31-2014. Informed and signed consent was obtained from all individual participants included in the study.

Author details

¹Department of Breast and Endocrine Surgery, Kanagawa Cancer Center, 2-3-2 Nakao, Asahi-ku, Yokohama 241-0815, Japan. ²Molecular Pathology and Genetics Division, Kanagawa Cancer Center Research Institute, 2-3-2 Nakao, Asahi-ku, Yokohama 241-0815, Japan. ³Department of Pathology, Kanagawa Cancer Center, 2-3-2 Nakao, Asahi-ku, Yokohama 241-0815, Japan. ⁴Department of Surgery, Yokohama City University, 3-9 Fukuura, Kanazawa-ku, Yokohama 236-0004, Japan.

Received: 27 February 2016 Accepted: 22 February 2017 Published online: 27 February 2017

References

- Kobayashi T. From improved survival to potential cure in patients with metastatic breast cancer. Breast Cancer. 2012;19(3):187–90. doi:10.1007/ s12282-011-0307-0.
- Hortobagyi GN. Can we cure limited metastatic breast cancer? J Clin Oncol. 2002;20(3):620–3.
- Greenberg PA, Hortobagyi GN, Smith TL, Ziegler LD, Frye DK, Buzdar AU. Long-term follow-up of patients with complete remission following combination chemotherapy for metastatic breast cancer. J Clin Oncol. 1996; 14(8):2197–205.
- Rahman ZU, Frye DK, Smith TL, Asmar L, Theriault RL, Buzdar AU, Hortobagyi GN. Results and long term follow-up for 1581 patients with metastatic breast carcinoma treated with standard dose doxorubicincontaining chemotherapy: a reference. Cancer. 1999;85:104–11.
- 5. Hortobagyi GN. Treatment of breast cancer. N Engl J Med. 1998;339:974-84.
- NCCN.org. (2016) NCCN clinical practice guidelines in oncology: breast cancer. Ver. 1. http://nccn.org/professionals/physician_gls/pdf/breast.pdf. Accessed 2 Jan 2016
- Cardoso F, Costa A, Norton L, Senkus E, Aapro M, André F, et al. ESO-ESMO 2nd international consensus guidelines for advanced breast cancer (ABC2). Ann Oncol. 2014;25:1871–88. doi:10.1093/annonc/mdu385.
- Liedtke C, Broglio K, Moulder S, Hsu L, Kau SW, Symmans WF, et al. Prognostic impact of discordance between triple-receptor measurements in primary and recurrent breast cancer. Ann Oncol. 2009;20(12):1953–8. doi:10. 1093/annonc/mdp263.
- Simmons C, Miller N, Geddie W, Gianfelice D, Oldfield M, Dranitsaris G, et al. Does confirmatory tumor biopsy alter the management of breast cancer patients with distant metastases? Ann Oncol. 2009;20(9):1499–504. doi:10.1093/annonc/mdp028.
- 10. Thompson AM, Jordan LB, Quinlan P, Anderson E, Skene A, Dewar JA, et al. Prospective comparison of switches in biomarker status between primary

and recurrent breast cancer: the breast recurrence in tissues study (BRITS). Breast Cancer Res. 2010;12(6):R92. doi:10.1186/bcr2771.

- Amir E, Miller N, Geddie W, Freedman O, Kassam F, Simmons C, et al. Prospective study evaluating the impact of tissue confirmation of metastatic disease in patients with breast cancer. J Clin Oncol. 2012;30(6):587–92. doi: 10.1200/JCO.2010.33.5232.
- Nishimura R, Osako T, Okumura Y, Tashima R, Toyozumi Y, Arima N. Changes in the ER, PgR, HER2, p53 and Ki-67 biological markers between primary and recurrent breast cancer: discordance rates and prognosis. World J Surg Oncol. 2011;9:131. doi:10.1186/1477-7819-9-131.
- Cao R, Wang L, Wang H, Xia L, Erdjument-Bromage H, Tempst P, et al. Role of histone H3 lysine 27 methylation in Polycomb-group silencing. Science. 2002;298:1039–43.
- 14. Schuettengruber B, Chourrout D, Vervoort M, Leblanc B, Cavalli G. Genome regulation by polycomb and trithorax proteins. Cell. 2007;128:735–45.
- Boyer LA, Plath K, Zeitlinger J, Brambrink T, Medeiros LA, Lee TI, et al. Polycomb complexes repress developmental regulators in murine embryonic stem cells. Nature. 2006;441:349–53.
- 16. Levine SS, Weiss A, Erdjument-Bromage H, Shao Z, Tempst P, Kingston RE. The core of the polycomb repressive complex is compositionally and functionally conserved in flies and humans. Mol Cell Biol. 2002;22: 6070–8.
- 17. Kleer CG, Cao Q, Varambally S, Shen R, Ota I, Tomlins SA, et al. EZH2 is a marker of aggressive breast cancer and promotes neoplastic transformation of breast epithelial cells. Proc Natl Acad Sci U S A. 2003;100:11606–11.
- Bachmann IM, Halvorsen OJ, Collett K, Stefansson IM, Straume O, Haukaas SA, et al. EZH2 expression is associated with high proliferation rate and aggressive tumor subgroups in cutaneous melanoma and cancers of the endometrium, prostate, and breast. J Clin Oncol. 2006;24:268–73.
- Gonzalez ME, Li X, Toy K, DuPrie M, Ventura AC, Banerjee M, et al. Downregulation of EZH2 decreases growth of estrogen receptor-negative invasive breast carcinoma and requires BRCA1. Oncogene. 2009;28:843–53. doi:10.1038/onc.2008.433.
- Cao Q, Yu J, Dhanasekaran SM, Kim JH, Mani RS, Tomlins SA, et al. Repression of E-cadherin by the polycomb group protein EZH2 in cancer. Oncogene. 2008;27:7274–84. doi:10.1038/onc.2008.333.
- Zeidler M, Varambally S, Cao Q, Chinnaiyan AM, Ferguson DO, Merajver SD, et al. The Polycomb group protein EZH2 impairs DNA repair in breast epithelial cells. Neoplasia. 2005;7:1011–9.
- Chang CJ, Yang JY, Xia W, Chen CT, Xie X, Chao CH, et al. EZH2 promotes expansion of breast tumor initiating cells through activation of RAF1-βcatenin signaling. Cancer Cell. 2011;19:86–100. doi:10.1016/j.ccr.2010.10.035.
- Gonzalez ME, DuPrie ML, Krueger H, Merajver SD, Ventura AC, Toy KA, et al. Histone methyltransferase EZH2 induces Akt-dependent genomic instability and BRCA1 inhibition in breast cancer. Cancer Res. 2011;71:2360–70. doi:10. 1158/0008-5472.CAN-10-1933.
- Bracken AP, Pasini D, Capra M, Prosperini E, Colli E, Helin K. EZH2 is downstream of the pRB-E2F pathway, essential for proliferation and amplified in cancer. EMBO J. 2003;22:5323–35.
- Wolff AC, Hammond ME, Hicks DG, Dowsett M, McShane LM, Allison KH, et al. Recommendations for human epidermal growth factor receptor 2 testing in breast cancer: American Society of Clinical Oncology/College of American Pathologists clinical practice guideline update. J Clin Oncol. 2013;31:3997–4013. doi:10.1200/JCO.2013.50.9984.
- Sihto H, Lundin J, Lundin M, Lehtimäki T, Ristimäki A, Holli K, et al. Breast cancer biological subtypes and protein expression predict for the preferential distant metastasis sites: a nationwide cohort study. Breast Cancer Res. 2011;13(5):R87. doi:10.1186/bcr2944.
- Kiba T, Inamoto T, Nishimura T, Ueno M, Yanagihara K, Teramukai S, et al. The reversal of recurrence hazard rate between ER positive and negative breast cancer patients with axillary lymph node dissection (pathological stage I-III) 3 years after surgery. BMC Cancer. 2008;8:323. doi:10.1186/1471-2407-8-323.
- Hussein YR, Sood AK, Bandyopadhyay S, Albashiti B, Semaan A, Nahleh Z, et al. Clinical and biological relevance of enhancer of zeste homolog 2 in triple-negative breast cancer. Hum Pathol. 2012;43:1638–44. doi:10.1016/j. humpath.2011.12.004.
- Reijm EA, Timmermans AM, Look MP, Meijer-van Gelder ME, Stobbe CK, van Deurzen CH, et al. High protein expression of EZH2 is related to unfavorable outcome to tamoxifen in metastatic breast cancer. Ann Oncol. 2014;25(11): 2185–90. doi:10.1093/annonc/mdu391. Epub 2014 Sep 5.

- Fluge Ø, Gravdal K, Carlsen E, Vonen B, Kjellevold K, Refsum S, Lilleng R, et al. Expression of EZH2 and Ki-67 in colorectal cancer and associations with treatment response and prognosis. Br J Cancer. 2009;101(8):1282–9. doi:10. 1038/sj.bjc.6605333.
- Changchien YC, Tátrai P, Papp G, Sápi J, Fónyad L, Szendrői M, et al. Poorly differentiated synovial sarcoma is associated with high expression of enhancer of zeste homologue 2 (EZH2). J Transl Med. 2012;10:216. doi:10. 1186/1479-5876-10-216.
- Reijm EA, Jansen MP, Ruigrok-Ritstier K, van Staveren IL, Look MP, van Gelder ME, et al. Decreased expression of EZH2 is associated with upregulation of ER and favorable outcome to tamoxifen in advanced breast cancer. Breast Cancer Res Treat. 2011;125:387–94. doi:10.1007/s10549-010-0836-9.
- Sato T, Kaneda A, Tsuji S, Isagawa T, Yamamoto S, Fujita T, et al. PRC2 overexpression and PRC2-target gene repression relating to poorer prognosis in small cell lung cancer. Sci Rep. 2013;3:1911. doi:10.1038/ srep01911.
- Knutson SK, Warholic NM, Wigle TJ, Klaus CR, Allain CJ, Raimondi A, et al. Durable tumor regression in genetically altered malignant rhabdoid tumors by inhibition of methyltransferase EZH2. Proc Natl Acad Sci U S A. 2013;110: 7922–7. doi:10.1073/pnas.1303800110.
- Epizyme[®], Inc. (2013). Epizyme[®] is a clinical stage biopharmaceutical company creating personalized therapeutics for patients with genetically defined cancers. http://epizyme.com. Accessed 20 Dec 2015
- Dowsett M, Dunbier AK. Emerging biomarkers and new understanding of traditional markers in personalized therapy for breast cancer. Clin Cancer Res. 2008;14:8019–26.
- Nishimura R, Osako T, Okumura Y, Hayashi M, Arima N. Clinical significance of Ki-67 in neoadjuvant chemotherapy for primary breast cancer as a predictor for chemosensitivity and for prognosis. Breast Cancer. 2010;17: 269–75. doi:10.1007/s12282-009-0161-5.
- Tanei T, Shimomura A, Shimazu K, Nakayama T, Kim SJ, Iwamoto T, et al. Prognostic significance of Ki67 expression after neoadjuvant chemotherapy in breast cancer. Eur J Surg Oncol. 2011;37:155–61. doi:10.1016/j.ejso.2010.10.009.
- Hitoshi I, Nobuyasu S, Kae K, Tatsuya Y, Hirotaka N, Takashi Y, et al. Correlation of expression of enhancer of zeste homolog 2 with the survival outcome in patients with metastatic breast cancer using primary and paired metastatic lesions. J Clin Oncol. 2016;34:e23227.

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at www.biomedcentral.com/submit

