## **Chapter 26 Environmental Transfer of Carbon-14 in Japanese Paddy Fields**

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Abstract It has been recognized that carbon-14 (<sup>14</sup>C) is one of the dominant radionuclides affecting dose from transuranic (TRU) wastes. This radionuclide has a decay half-life of 5,730 years, and <sup>14</sup>C organic materials have very low sorption properties to clay and rock in the environment, which raises some concerns about the releases of <sup>14</sup>C to the biosphere from radioactive waste repositories. For the safety assessment of TRU waste disposal, we studied the behavior of <sup>14</sup>C in rice paddy field soils. We also determined key parameters such as soil-soil solution distribution coefficients ( $K_d$ s) and soil-to-rice plant transfer factors (TFs) of <sup>14</sup>C in the field soils. The TFs were obtained in laboratory and field experiments. In our laboratory experiments, we used [1,2-<sup>14</sup>C] sodium acetate as a source of <sup>14</sup>C because it has been suggested that low molecular weight organic-<sup>14</sup>C compounds are released from metallic TRU wastes. The results showed that <sup>14</sup>C-bearing sodium acetate in irrigated paddy soils was rapidly decomposed by indigenous bacteria. Although some of the <sup>14</sup>C was assimilated into the bacterial cells, most of the <sup>14</sup>C was released into the air as gaseous compounds. The main chemical species of <sup>14</sup>C gases was <sup>14</sup>CO<sub>2</sub>, and a part of the released <sup>14</sup>CO<sub>2</sub> gas was used by rice plants during photosynthesis. Only a negligible amount of  ${}^{14}C$  was absorbed through the roots. Therefore, the contamination of rice plants is mainly caused by gasification of <sup>14</sup>C, and microorganisms are responsible for driving this process. The activity of microorganisms is a key issue in the behavior of  ${}^{14}C$  in paddy fields.

**Keywords** Bacteria • Behavior • Degradation • Radiocarbon • Rice paddy fields • Safety assessment • TRU wastes

#### 26.1 Introduction

Transuranic (TRU) wastes contain a variety of radionuclides, for example, Np, Pu, and long-lived radionuclides such as <sup>14</sup>C and <sup>129</sup>I. In Japan these wastes are categorized into four groups in accordance with their physical properties and the

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concentration of radioactive materials. Group two waste includes hull and end piece wastes with relatively high amounts of <sup>14</sup>C, and leaching of low molecular weight <sup>14</sup>C organic materials from simulated hull wastes has been reported [1]. The <sup>14</sup>C organic materials have very few sorption properties to clay and rock, and <sup>14</sup>C has a relatively long half-life of 5,730 years. These properties raise concerns about releases of <sup>14</sup>C to the biosphere from radioactive waste repositories.

Rice is a major agricultural crop throughout Asia, and thus human exposure to <sup>14</sup>C through rice intake must be considered. To reduce the risk of the internal radiation dose from <sup>14</sup>C, it is important to clarify the behavior of <sup>14</sup>C in rice paddy fields. In this study, we determined transfer pathways of <sup>14</sup>C through the rice paddy fields to rice grains. Environmental parameters such as soil–soil solution distribution coefficients ( $K_{ds}$ ) and soil-to-rice plant transfer factors (TFs) of <sup>14</sup>C were also determined, because these parameters are often used in transfer models to predict the behavior of radionuclides in the environment. From a series of our experimental results, we describe the behavior of <sup>14</sup>C in rice paddy field soils and the importance of microbial activity.

### 26.2 Partitioning of <sup>14</sup>C into Solid, Liquid, and Gas Phases

We carried out batch sorption experiments using 63 Japanese rice paddy soil samples to clarify the transfer pathways of <sup>14</sup>C in rice paddy fields. The soil samples were collected throughout Japan and taken to our laboratory where they were air dried and sieved (<2 mm). These sieved soils were mixed with a [1,2-<sup>14</sup>C] sodium acetate solution at the ratio of soil : solution = 0.5 g : 5 ml, and the flooded soil samples were incubated at 25 °C for 7 days [2]. During the incubation period, the <sup>14</sup>C atoms of the sodium acetate were partitioned into solid, liquid, and gas phases. Each partitioning ratio is shown in Fig. 26.1. Approximately 63 % of the total <sup>14</sup>C on average was released into the air as gaseous compounds. Partitioning ratios into solid and liquid phases were 34 % and 3 %, respectively. These results suggest that gasification is an important pathway in the environmental transfer of <sup>14</sup>C in Japanese rice paddy fields.

When <sup>14</sup>C is released into the air, <sup>14</sup>C-bearing gases must pass through the soil solution. Because soil solution pH affects chemical reactions such as hydrolysis and degassing of CO<sub>2</sub>, chemical forms of <sup>14</sup>C-bearing gases may change in the soil solution. We, therefore, investigated relationships between pH and partitioning ratios of <sup>14</sup>C into the liquid phase at day 7 of incubation (Fig. 26.2). The partitioning ratio increased with increasing in pH, and a significant correlation (r = 0.7) was found. These data fit well with the solubility curve of total carbonic acid in water, which refers to the sum of dissolved carbon dioxide and the carbonic acid. This observation suggested that the dominant chemical species of <sup>14</sup>C in gas forms was carbon dioxide. To confirm the effect of pH on the partitioning of <sup>14</sup>C into the liquid phase, a soil sample was suspended in MES [2-(*N*-morpholino)ethanesulfonic acid] buffers with the initial pH value adjusted to 5.5, 6.5, and 7.5 (Fig. 26.3). A control





sample was prepared consisting of the soil and deionized water (pH unadjusted). The partitioning ratio also increased with increasing pH, suggesting that the partitioning ratio of <sup>14</sup>C into the liquid phase depended on the pH of the soil solution.

Soil-soil solution distribution coefficient ( $K_d$ ) is a commonly used parameter to evaluate behaviors of radionuclides in the environment. In our study, the  $K_d$  values were calculated from activities of the <sup>14</sup>C in the solid and liquid phases at the end of incubation, and the obtained  $K_d$  value was  $139 \pm 77$  ml g<sup>-1</sup> on average. Negatively charged anions generally have low  $K_d$  values because of simple electrostatic interaction. Our value, however, was higher than expected from the chemical form of <sup>14</sup>CH<sub>3</sub><sup>14</sup>COO<sup>-</sup>. For example, Kaneko et al. [1] obtained the  $K_d$  value of 9.5 ml g<sup>-1</sup> for the sorption test of acetic acid using cement materials. The reason for our high  $K_d$  value is explained next.

#### 26.3 Involvement of Microorganisms in the <sup>14</sup>C Behavior

Many microorganisms inhabit rice paddy fields, and they are responsible for nutrient cycling. We studied the involvement of microorganisms in environmental transfer of <sup>14</sup>C. Microorganisms in batch cultures were treated with autoclaving (121 °C, 15 min), mixing with glutaraldehyde [final concentration of 2.5 % (vol/vol)], and mixing with cycloheximide (final concentration, 250  $\mu$ g ml<sup>-1</sup>). Autoclaving and expose to glutaraldehyde inactivate bacteria and fungi, but exposure to cycloheximide only inhibits fungi. The partitioning ratios of <sup>14</sup>C into solid, liquid, and gas phases for each treatment sample are listed in Table 26.1. When microorganisms were treated by autoclaving and exposing to glutaraldehyde, almost all the <sup>14</sup>C added remained in the liquid phase; that is, negligible transformation of <sup>14</sup>C occurred. On the other hand, the <sup>14</sup>C atoms in the control and the cycloheximide-treated sample were similar between the control and the cycloheximide samples. We confirmed fungi made no contribution to partitioning of <sup>14</sup>C



based on these results. We concluded that environmental transfer of <sup>14</sup>C in rice paddy fields was driven by bacteria, not by fungi.

To confirm incorporation of <sup>14</sup>C into bacteria cells, bacteria that were isolated from a flooding water of a paddy soil sample were cultivated on agar plates containing [1,2-<sup>14</sup>C] sodium acetate [3]. After cultivation, bacterial colonies were formed, and their autoradiography images showed that all colonies had the ability to take up <sup>14</sup>C (Fig. 26.4). In our experimental procedure, bacterial cells were consequently partitioned into the solid phase, and thus the solid phase contains the <sup>14</sup>C incorporated by bacteria, which could be one of the reasons for the relatively high  $K_d$  values.

#### 26.4 Transfer of <sup>14</sup>C from Soil to Rice Plants

Soil-to-rice plant transfer factors (TFs) of <sup>14</sup>C, which was defined as <sup>14</sup>C concentration in rice grains (Bq/kg-dry) divided by that in soil (Bq/kg-dry), were determined by laboratory and field experiments. In the laboratory experiment using a

	Partitioning ratio (%)		
Treatment	Solid phase	Liquid phase	Gas phase
Control	27.9	4.5	67.5
Autoclaving	0	98.0	2.0
Glutaraldehyde exposure	0	96.8	3.2
Cycloheximide exposure	29.3	4.8	65.9

Table 26.1 The partitioning ratios of <sup>14</sup>C into solid, liquid, and gas phases for each treatment.



Fig. 26.4 Colonies of bacteria (a) and their autoradiography image (b). Heterotrophic bacteria have the ability to uptake  ${}^{14}$ C from an agar medium

growth chamber, we grew rice plants with addition of  $[1,2^{-14}C]$  sodium acetate. This <sup>14</sup>C compound was supplied once to rice plants in the flooding water just before blooming, and TF of  $6.8 \pm 2.4$  on average was obtained. In these tracer experiments, rice plants were also cultivated without  $[1,2^{-14}C]$  sodium acetate as negative controls in the same growth chamber as the <sup>14</sup>C-treated rice. Interestingly <sup>14</sup>C was detected even from the rice grains of negative control samples. These results suggested that the <sup>14</sup>C-bearing gas, which was released from bacterial cells in rice paddy soils, was fixed by the rice plants in the negative controls through photosynthesis.

We also examined the possibility of root uptake of <sup>14</sup>C by stable isotope techniques under field conditions [4]. If plant carbon originates from the atmospheric CO<sub>2</sub>, the  $\delta^{13}$ C values in crops can be calculated using the  $\delta^{13}$ C value, -8% in air [5], and the <sup>13</sup>C fractionation ratio in photosynthesis by rice plants of -18 to -20% [6, 7]. The calculated  $\delta^{13}$ C values in our study ranged from -28% to -26%, and the results implied that no soil carbon contribution occurred for white rice; however, by setting some conditions, for example, <sup>13</sup>C fractionation ratio of 19‰, we obtained the average TF value of  $0.11 \pm 0.04$  for white rice. To compare these TF values obtained in laboratory and field experiments, it is necessary to pay attention to the difference between  $[1,2-^{14}C]$  sodium acetate and the actual organic compounds present in the natural soil.



Fig. 26.5 A conceptual diagram for the behavior of <sup>14</sup>C in rice paddy fields

#### 26.5 Behavior of <sup>14</sup>C in Rice Paddy Fields

From the aforementioned results, the behavior of <sup>14</sup>C in rice paddy fields could be considered as follows (a conceptual diagram appears in Fig. 26.5). When irrigation water is contaminated by <sup>14</sup>C-bearing sodium acetate, the <sup>14</sup>C compound is taken up and metabolized by indigenous bacteria. A part of the <sup>14</sup>C is assimilated by the bacterial cells, and the rest of the <sup>14</sup>C is released as gaseous compounds from the cells as a result of dissimilation. The dominant chemical species of <sup>14</sup>C in gas forms is carbon dioxide, and thus some of the released <sup>14</sup>CO<sub>2</sub> is dissolved in soil solution depending on pH. For example, when the pH of the soil solution is less than 6.5, most of  ${}^{14}C$  in gas forms is released into the air. The released  ${}^{14}CO_2$  is eventually taken up by rice plants during photosynthesis. When the pH of the soil solution is between 6.5 and 10.5, <sup>14</sup>C-bearing bicarbonate ion dominates in the soil solution. In addition, once <sup>14</sup>CO<sub>2</sub> has been released into the air, a part of the <sup>14</sup>CO<sub>2</sub> gas may be redissolved in the soil solution again as bicarbonate ion. When the pH of the soil solution is greater than 10.5, although this is not probable in paddy fields, <sup>14</sup>Cbearing carbonate ion dominates in the soil solution. Carbonate ion is thermally unstable and thus precipitates as carbonate minerals such as CaCO<sub>3</sub>. In these alkaline situations, the ratio of <sup>14</sup>C in the solid phase may increase as a result of the precipitation of <sup>14</sup>C. Because the root uptake of <sup>14</sup>C by rice plants is negligible, gasification of <sup>14</sup>C is an important environmental transfer pathway for the safety assessment of TRU wastes, and bacteria are responsible for driving this pathway.

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