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Constitutive patterns of gene expression regulated by RNA-binding proteins

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Abstract

Background: RNA-binding proteins regulate a number of cellular processes, including synthesis, folding, translocation, assembly and clearance of RNAs. Recent studies have reported that an unexpectedly large number of proteins are able to interact with RNA, but the partners of many RNA-binding proteins are still uncharacterized.

Results: We combined prediction of ribonucleoprotein interactions, based on catRAPID calculations, with analysis of protein and RNA expression profiles from human tissues. We found strong interaction propensities for both positively and negatively correlated expression patterns. Our integration of *in silico* and *ex vivo* data unraveled two major types of protein–RNA interactions, with positively correlated patterns related to cell cycle control and negatively correlated patterns related to survival, growth and differentiation. To facilitate the investigation of protein–RNA interactions and expression networks, we developed the catRAPID express web server.

Conclusions: Our analysis sheds light on the role of RNA-binding proteins in regulating proliferation and differentiation processes, and we provide a data exploration tool to aid future experimental studies.

Background

With the advent of high-throughput proteomic and transcriptomic methods, genome-wide data are giving previously unprecedented views of entire collections of gene products and their regulation. Recently, approaches based on nucleotide-enhanced UV cross-linking and oligo(dT) purification have shown that a number of proteins are able to bind to RNA [1,2].

RNA-binding proteins (RBPs) are key regulators of post-transcriptional events [3] and influence gene expression by acting at various steps in RNA metabolism, including stabilization, processing, storing, transport and translation. RBP-mediated events have been described using recognition and regulatory elements in RNA sequences [4,5] as well as expression profiles [6] that are tissue specific and conserved across species [7-9]. Although heterogeneity in gene regulation is responsible for phenotypic variation and evolution [10], very little is

known about constitutive expression patterns controlled by RBPs [11,12], which are the subject of this work.

Data from recent transcriptomic and proteomic studies [13,14] are becoming attractive for studying mechanisms of gene regulation [15,16]. Despite the increasing amount of genomic data, the development of computational methods for integrating, interpreting and understanding molecular networks remains challenging [17,18]. Here we combine our predictions of protein–RNA interactions, based on catRAPID calculations [19,20], with the information obtained from expression data to investigate constitutive regulatory mechanisms. The catRAPID approach has been previously employed to predict protein associations with non-coding RNAs [21,22] as well as ribonucleoprotein interactions linked to neurodegenerative diseases [23,24]. Our theoretical framework has been used to unravel self-regulatory pathways controlling gene expression [25]. The catRAPID omics algorithm, validated using photoactivatable-ribonucleoside-enhanced cross-linking and immunoprecipitation (PAR-CLIP) data, has been recently developed to predict protein–RNA associations at the transcriptomic and proteomic levels [26].

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Using comprehensive and manually annotated databases of expression profiles in human tissues, at both protein and RNA levels, we investigated the correlation between RBP activity and regulation. The link between interaction propensity and expression levels was exploited to reveal the fine-tuned functional sub-networks responsible for regulatory control. To explore the results further, we developed the catRAPID express web server [27].

Results

In this study, we focused on the mRNA interactomes of RBPs detected through nucleotide-enhanced UV cross-linking and oligo(dT) purification approaches [1,2]. Exploiting gene ontology (GO) annotations [28] for protein-coding genes, we systematically analyzed protein–RNA interactions and expression data for human tissues.

At present, few studies have investigated how altering protein expression affects the abundance of RNA targets. Interrogating the Gene Expression Omnibus (GEO) [29] and ArrayExpress databases [30], we found two human proteins, ELAV-like protein 1 (or human antigen R, HuR) [31] and Protein lin-28 homolog B (LIN28B) [32,33], whose knock-down has been shown to alter the expression of target genes identified by PAR-CLIP (see Materials and methods).

Our predictions, made using the catRAPID algorithm [26], identified experimentally validated interactions with high significance (HuR: $P = 10^{-8}$; LIN28B: $P = 10^{-3}$; Fisher's exact test; see Materials and methods). The interactions were effectively discriminated from non-interacting pairs using score distributions (LIN28B: $P = 10^{-4}$; HuR: $P = 10^{-16}$; Student's t-test; see Materials and methods). Hence, catRAPID is very good at predicting physical interactions between a protein and RNA partners (other

statistical tests are given in Materials and methods and Additional file 1).

To understand the regulation of HuR and LIN28B targets better, we studied the relation between interaction propensities and expression levels. We found that the expression of predicted HuR targets is altered (log-fold change, LFC) when HuR is knocked down ($P < 10^{-5}$; Kolmogorov–Smirnov test; Figure 1A), which is in agreement with experimental data [31]. Similarly, predicted LIN28B targets are downregulated upon protein depletion ($P < 10^{-2}$; Kolmogorov–Smirnov test; Figure 1B), as shown in a previous study [33]. Moreover, we compared the top 1% of predicted associations with the top 1% of experimental interactions and found the same enrichments for transcripts changing in expression levels upon protein depletion. Specifically, 62% of HuR experimental interactions and 63% of HuR predicted associations had $LFC > 0$. Similarly for LIN28B, 57% of experimental interactions and 56% of predicted associations had $LFC > 0$.

These HuR and LIN28B examples indicate that changes in protein expression influence the abundance of RNA targets, suggesting that a large-scale analysis of co-expression and interaction propensities could improve understanding of RBP-mediated regulatory mechanisms.

RNA-binding protein–mRNA interactions and relative expression profiles

Our predictions indicate that interacting molecules have both more correlated and anti-correlated expression patterns (see Materials and methods and Figure 2). By contrast, non-correlated expression is not associated with any enrichment in interaction propensity (Additional

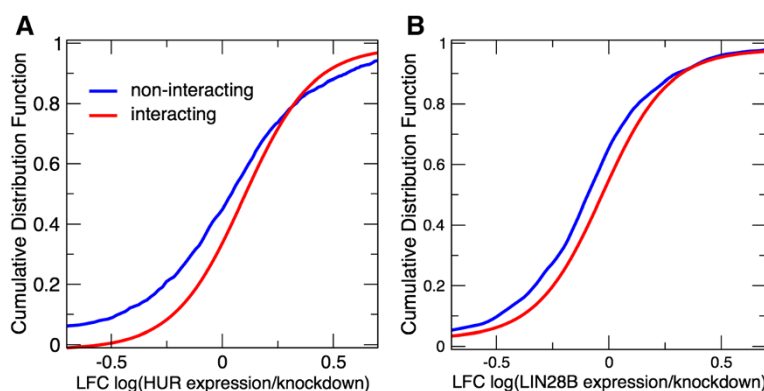
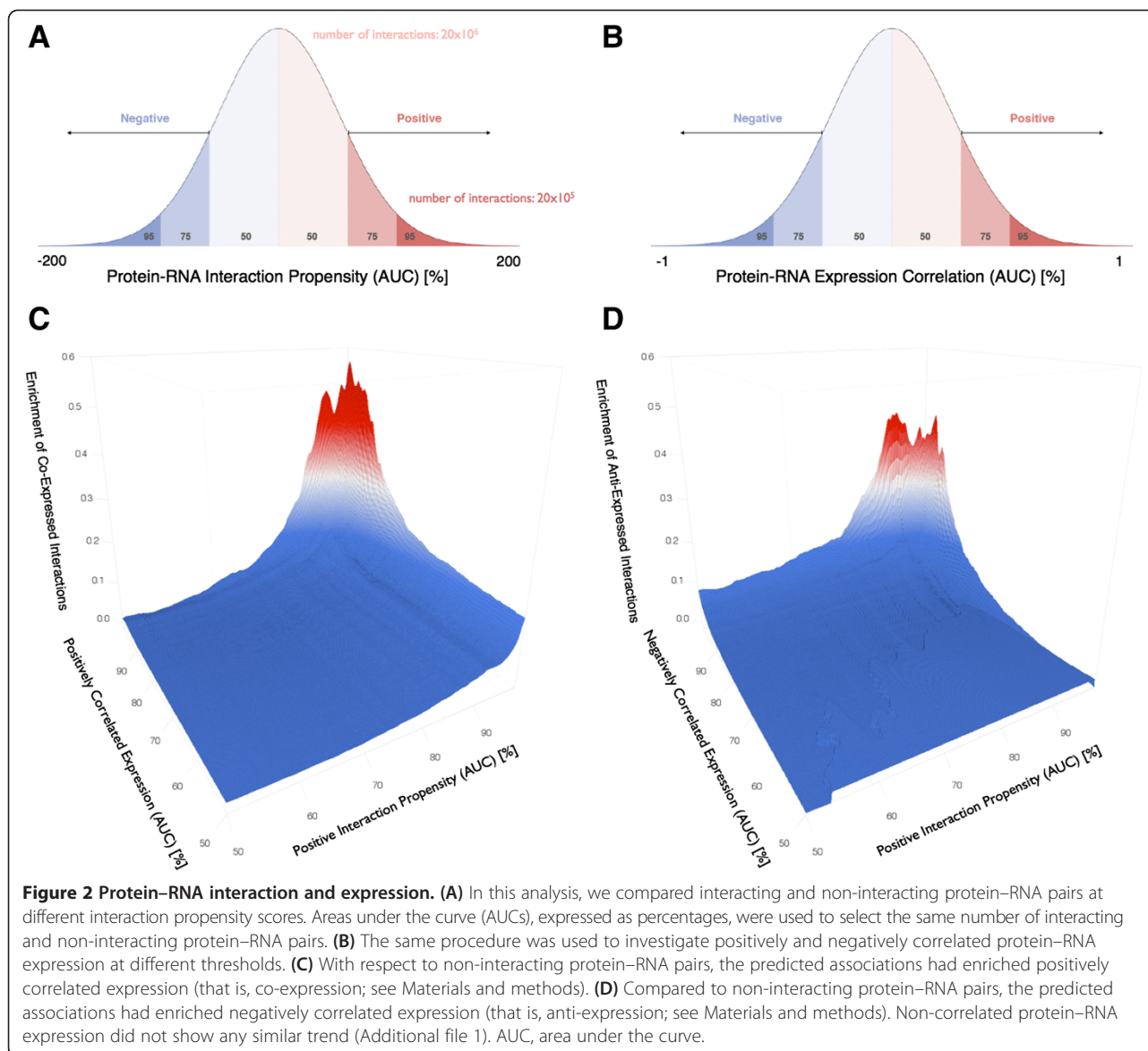


Figure 1 Relation between protein and RNA regulation. (A) HuR interactome: our predictions, made using catRAPID [26], indicate that expression levels of RNA targets change upon HuR knock-down (log-fold changes, LFC), in agreement with experimental evidence [31] ($P < 10^{-5}$; Kolmogorov–Smirnov test). (B) LIN28B interactome: RNA targets are downregulated upon LIN28B knock-down (LFC), as reported in a previous study [33] ($P < 10^{-2}$; Kolmogorov–Smirnov test). In this analysis, the prediction of the interactions was highly significant (HuR: $P < 10^{-8}$; LIN28B: $P < 10^{-3}$; Fisher's exact test). Our results indicate that changes in protein expression influence the abundance of RNA targets to a significant extent. HuR, human antigen R; LFC, log-fold change; LIN28B, lin-28 homolog B.



file 2: Figure S1A). We observed the same results using immunohistochemistry [34] and RNA sequencing data [6] to estimate protein abundances (Additional file 2: Figures S1B and S2; see Materials and methods). This finding is truly remarkable. Direct proportionality between protein and mRNA expression levels has been observed in bacteria and fungi [13,14] but post-transcriptional modification is known to influence the overall abundance of the protein product in higher eukaryotes [35]. Since immunohistochemistry only provides a qualitative estimate of the amount of protein (see Materials and methods) and the analysis is restricted to 612 proteins, we used RNA sequencing for our predictions (1,156 RBPs).

The enrichment shown in Figure 2 suggests that a good relation exists between interaction and expression of protein–RNA molecules, which should have co-

evolved to be either co-expressed or anti-expressed to exert a regulatory function (Figure 2C,D).

Conservation of expression pattern for functionally related genes

We classified protein–RNA associations into four categories: interacting and co-expressed (IC), interacting and anti-expressed (IA), non-interacting and co-expressed (NIC) and non-interacting and anti-expressed (NIA). We applied conditional tests on each subset to detect significantly over-represented gene ontology (GO) terms (see Materials and methods and Additional file 3: Table S1).

For high interaction propensities, transcripts in the IC subset have more processes associated with *cell cycle control*, in particular the negative regulation of proliferation (Discussion; Additional file 3: Table S1).

Transcripts interacting with anti-expressed proteins (IA subset) are involved in *survival, growth and differentiation* processes and have more regulative functions at the DNA level (Discussion; Additional file 3: Table S1).

No clear functional assignments and/or insufficiently populated GO terms were found for transcripts in non-interacting protein–RNA pairs (NIC and NIA subsets).

Intrinsic disorder and RNA-binding protein interaction propensity

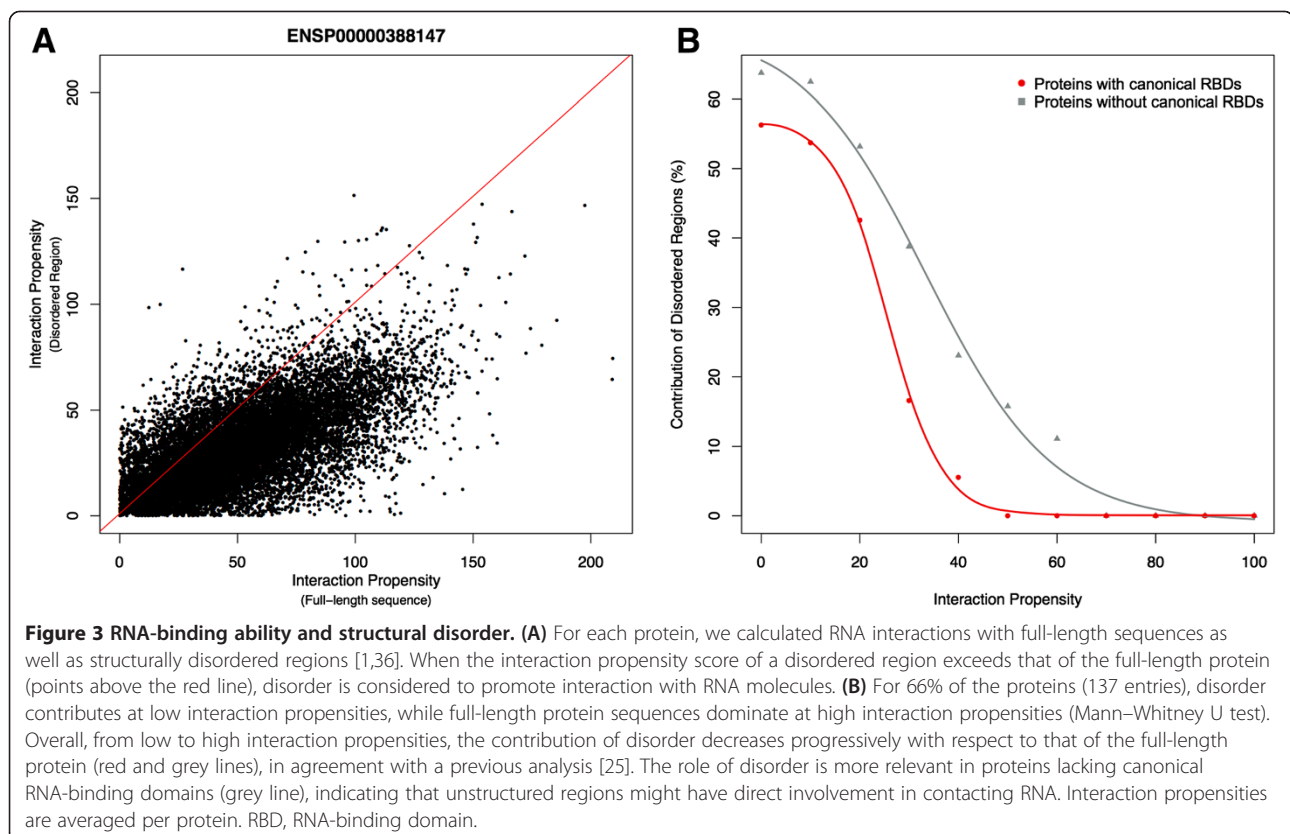
Recent findings suggest that RBPs have more structurally disordered regions [1]. To investigate the relation between disorder and RNA-binding ability, we used the IUPred algorithm [36]. For each protein, we extracted structurally disordered regions (IUPred score > 0.4 [1]) and calculated the interaction propensities with human transcripts. We considered both canonical RBPs (that is, containing RNA-binding domains) and putative RBPs (that is, lacking RNA-binding domains) [1]. With respect to the RNA-binding ability of full-length sequences, the contribution of disorder is higher at low interaction propensity scores and becomes negligible at high interaction propensities (see Materials and methods and Figure 3A). Nevertheless, the role of structural disorder is more pronounced in proteins lacking canonical RNA-binding

domains, indicating that unfolded regions might be able to promote interactions with RNA (Figure 3B).

In a previous study we observed that catRAPID scores correlate with chemical affinities [21], which suggests that the interaction propensity can be used to estimate the strength of association [21,26]. Hence, our results indicate that structural disorder might contribute to low-affinity interactions with RNA (Figure 3A,B), which is in agreement with what has been observed for protein–protein associations [37,38]. As a matter of fact, it has been reported that disorder regions are able to promote promiscuous and non-specific interactions [39].

Discussion

Because they are associated with transcriptional control of gene expression, RBPs play fundamental roles in health and disease. Indeed, by binding to their target mRNAs, RBPs can influence protein production at different levels (transcription, translation and protein/mRNA degradation). Protein–RNA complexes are very dynamic and can undergo extensive remodeling. Thus, they can control the spatiotemporal regulation of target gene expression and the overall switching *on and off* of the distinct sets of genes involved in biological processes such as cell cycle progression, cell differentiation, cell



response to metabolic *stimuli* and stress conditions, organ morphogenesis and embryonic development.

Co-expression and interaction propensity are features of cell cycle control

At high interaction propensities (AUC > 95%; see Materials and methods), the IC subset has more GO terms linked to cell cycle control and housekeeping functions such as nucleobase metabolism and purine biosynthesis (Figure 4 and Additional file 3: Table S1). In particular, mRNAs interacting with co-expressed RBPs code for negative regulators of cell proliferation and migration (translation, signaling and metabolite utilization). We found a number of tumor suppressors in the IC subset (AHRR, BAX, BRMS1, CDKN1A, CDKN2A, CTBP1, DAB2IP, DKK3, FLCN, FOXP1, GADD45G, GALR1, GTPBP4, HIC1, IGFBP3, IRF8, KLF4, MEN1, MLH1, NF2, NR0B2, PARK2, PAWR, PAX4, PAX5, PCGF2, PHB, PML, PPP1R1B, PPP2R4, PTPRJ, PYCARD, RHOA, SIRT2, TFAP2A, TNFAIP3, TRIM24, TSC2, TSG101, UCHL1). Interestingly, 90% of IC genes annotated with more functional categories (381 out of 422) are listed in the gene index of the National Institutes of Health's Cancer Genome Anatomy Project [40]. Terms associated with inhibition of cellular pathways (especially the negative regulation of phosphorylation and regulation of protein serine/threonine kinase activity) are also more prevalent in the IC subset when immunochemistry data are used.

As mutations altering tumor suppression lead to aberrant proliferative events, we speculate that downregulation of specific genes is a mechanism for preventing indiscriminate cellular growth. In agreement with this hypothesis, it has been reported that somatic loss of function of the tumor suppressor *tuberous sclerosis 2*

(*TSC-2*) leads to the development of benign and malignant lesions in the myometrium, kidney and other tissues sharing common features such as a low rate of renewal and defects in the mitochondrial respiratory chain associated with oncogenesis [41,42]. This gene is annotated in all the functional categories prevalent in the IC subset. Intriguingly, it is predicted that *TSC-2* mRNA interacts strongly with Nuclear Protein 5A (NOP56). The interaction propensity is 175 corresponding to an AUC of 99.5%. This protein is an essential component of the splicing machinery [43] that is differentially expressed in leiomyoma and downregulated in response to hypoxia [44]. It is possible that hypoxia-dependent repression of NOP56 expression [45-47] is a protective mechanism against fast growth and potential tumor progression. Indeed, it has been reported that *NOP56* and *TSC-2* are not differentially expressed in renal carcinomas and oncocytomas [48,49] (ArrayExpress: E-GEOD-12090; ArrayExpress: E-GEOD-19982), indicating loss of regulation during malignant progression.

Based on these observations, we propose that downregulation of RBPs promoting the translation of dysfunctional tumor suppressors can prevent indiscriminate cellular growth and that loss of control can destine a cell to malignancy (additional examples are reported in Additional file 1).

Anti-expression and interaction propensity are features of repressing processes

For AUC > 95%, the IA subset has more terms associated with cell differentiation processes (for example, proximal/distal pattern formation) as well as inflammation (for example, positive regulation of isotype switching), which are known to be tightly linked [50-52]. In fact, a

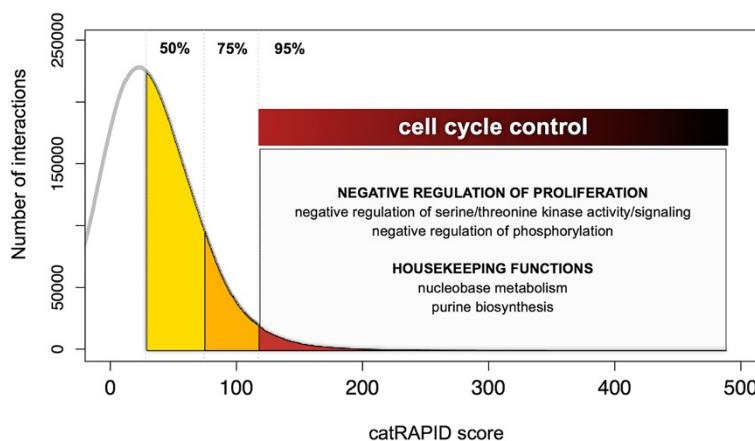


Figure 4 GO enrichment for interacting mRNA-RBP pairs correlated in expression (IC subset). Using the catRAPID score distribution, we counted mRNA GO enrichment associated with different areas under the curve (see Materials and methods). The color gradient (yellow to red) indicates the AUC values (number of interactions: 20,702,804 for AUC > 50%, 10,351,402 for AUC > 75%, 2,070,280 for AUC > 95%). We found that cell cycle processes have more highly interacting mRNA-RBP pairs (AUC > 95%) that are correlated in expression. AUC, area under the curve; GO, gene ontology; IC, interacting and co-expressed; RBP, RNA-binding protein.

number of differentiation cytokines (IL18, IL23 and EBI3/IL27) and stimulators of cytokine production (CD28 and CD80CCR2/CD192) are in the subset. Moreover, a large fraction of entries is also linked to protein-DNA complex assembly and regulation of transcription initiation from RNA polymerase II promoter (Figure 5 and Additional file 3: Table S1). It has been shown that 94% of genes in IA enriched functional categories (124 out of 132) are listed in the annotated gene index of the National Institutes of Health's Cancer Genome Anatomy Project [40]. Remarkably, terms clearly associated with cell differentiation and inflammation (especially regulation of embryonic development and B cell activation involved in immune response) are more prevalent in the IA subset when immunochemistry data are used.

IA genes share the common functional property of regulating survival, growth and differentiation processes. As RBPs play a crucial role in repressing gene expression [53,54], IA associations could be involved in the regulation of proliferative events. Indeed, adult tissues are constantly maintained at the steady state [13] but a dramatic reawakening of growth, survival and differentiation genes occur in either physiological conditions (for example, wound healing [50]) or pathological progression to cancer [55].

In the IA set, we found YTHDC1 (YT521-B), which is a ubiquitously expressed member of the novel RNA-binding YTH-domain family [56]. YTHDC1 represses gene expression by either sequestering splicing factors or directly binding to transcripts [57-59] (Additional file 2: Figure S5A). Among the transcripts that we predict to be potentially targeted by YTHDC1, we found several proto-oncogenes or tumor-associated genes such as *RET*, *PRMT2*, *RARG* and *HOXA9* (*RET*: interaction

propensity = 166; *PRMT2*: interaction propensity = 209; *RARG*: interaction propensity = 194; *HOXA9*: interaction propensity = 165; all corresponding to an AUC of 99.5%). In particular, alternatively spliced variants of *PRMT2* were related to survival and the invasiveness of breast cancer cells [60,61], while high expression of *RARG* and *HOXA9* has been observed in human hepatocellular carcinomas and acute leukemia [62,63]. We hypothesize that perturbation of the regulation by YTHDC1 of potentially oncogenic genes such as *RET*, *PRMT2*, *RARG* and *HOXA9* could be involved in the pathogenesis of related tumors. In fact, experimental studies support the implications for YTHDC1 in cancer progression with regard to angiogenesis, growth factor signaling, immortalization, genetic instability, tissue invasion and apoptosis [59,64,65].

Similarly, the translational silencer TIA-1, also reported to induce mRNA decay [66-68], is predicted to interact with the ubiquitously expressed *NAP1L1* transcript (interaction propensity = 113 corresponding to an AUC of 95%), consistent with iCLIP data for HeLa cells (ArrayExpress: E-MTAB-432) [69] (Additional file 4: Table S2). Deregulation of *NAP1L1* expression has been documented for several tumors such as small intestine carcinoid neoplasia [70], neuroendocrine tumors [71], ovarian cancer [72] and hepatoblastomas [73]. We hypothesize that TIA-1 plays a fundamental role in the post-transcriptional regulation of *NAP1L1* and that alteration of this regulatory process contributes to *NAP1L1*-associated tumor development.

We note that repression of aberrant interactions can be achieved by gene silencing, which prevents the potential stabilizing action of RBPs on specific transcripts (Additional file 2: Figure S5B). For instance, the *Nodal*

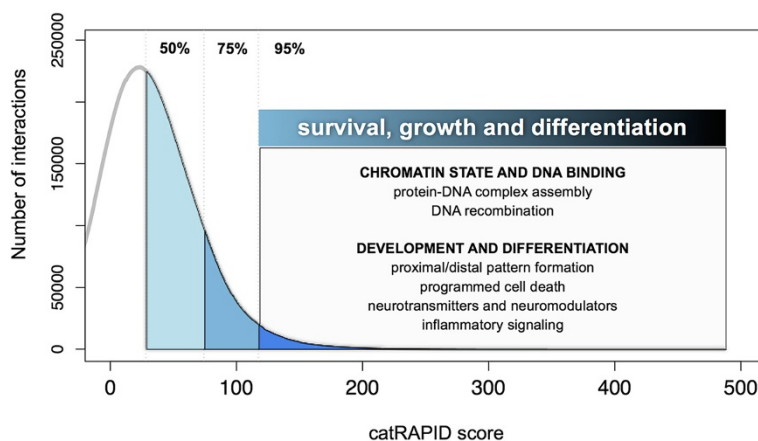


Figure 5 GO enrichment for interacting mRNA-RBP pairs anti-correlated in expression (IA subset). Using the catRAPID score distribution, we evaluated mRNA GO enrichment associated with different areas under the curve (see Materials and methods). A color gradient (cyan to blue) shows the AUC values (number of interactions: 20,702,804 for AUC > 50%, 10,351,402 for AUC > 75%, 2,070,280 for AUC > 95%). We found that cell differentiation processes are more prevalent in interacting mRNA-RBP pairs (AUC > 95%) that are anti-correlated in expression. AUC, area under the curve; GO, gene ontology; IA, interacting and anti-expressed; RBP, RNA-binding protein.

gene is normally silenced in adult tissues and its expression is associated with tumor progression [74]. Since *Nodal* is a member of the Transforming Growth Factor β (TGFB) superfamily and controls mesoderm formation and axial patterning during embryonic development [74], it is possible that *Nodal* interactions with specific RBPs lead to pathogenesis in adult tissues. Our predictions indicate that the transcript *Nodal* interacts with a number of anti-expressed RBPs (ADD1, API5, ARCN1, CANX, CAPRN1, CCT6A, DKFZP434I0812, GSPT1, HSP90AB1, PKM, PUF60, XRCC5, YTHDC1 and YWHAZ). Since the exact mechanism regulating *Nodal* is at present unknown, we generated a list of protein partners that could be exploited for future experimental studies (Additional file 5: Table S3).

Conclusions

Comparative expression studies provide important insights into biological processes and can lead to the discovery of unknown regulation patterns. While evolutionary constraints on tissue-specific gene expression patterns have been extensively investigated [7-9,75,76], the constitutive regulation of RBP-mediated interactions is still poorly understood [11,12]. It has been previously observed that cellular localization and gene expression levels impose stringent conditions on the physicochemical properties of both protein and RNA sequences [77,78], but large-scale computational analyses of constitutive RBP-mediated regulatory networks have never been attempted before. Our study shows for the first time that the integration of *in silico* predictions [19] with *ex vivo* expression profile data [6,34] can be used to discover distinct features of RBP biological functions.

We observed an enrichment of unique and functionally related GO terms for RBP-mRNA pairs associated with high interaction propensities and specific expression patterns. In our analysis, co-expression of interacting mRNA-RBP pairs (IC set) is linked to regulation of proliferation and cell cycle control, while anti-expression (IA set) is a characteristic feature of survival, growth and differentiation-specific processes. We do not exclude that RBP-mRNA associations displaying poor interaction propensities (NIC and NIA sets) might have important evolutionary implications as spatiotemporal separation and limited chemical reactivity could be ways to avoid aberrant associations [55].

We found that RNA-binding proteins are enriched in structurally disordered regions and that unfolded polypeptide fragments promote association with RNA molecules at low interaction propensities. As disordered proteins are highly reactive [37], it is reasonable to assume that interaction with RNA needs to be tightly regulated to avoid cellular damage [39]. In this regard, our results expand at the nucleic acid level what has been

previously observed for the general promiscuity of natively unfolded proteins [38,79].

In conclusion, we hope that our study of protein-RNA interaction and expression will be useful in the design of new experiments and for further characterizing ribonucleoprotein associations. A list of proposed interactions and a server for new inquiries are available at the catRAPID express webpage [27].

Materials and methods

Prediction for LIN28B and HuR interactions

We performed a number of tests to assess the quality of our calculations (see section on RNA-binding protein-mRNA interaction propensity) using PAR-CLIP data [31,33]. In this analysis, we used all the RNA interactions present in our dataset (positive set: 285 sequences for LIN28B and 579 for HuR) and, due to the unavailability of non-bound RNAs, the full list of human transcripts (negative set: 105,000 sequences).

For the *significance of interaction predictions*, we performed Fisher's exact test comparing the top 1% of predicted interactions with the remaining protein-RNA associations (HuR: $P = 10^{-8}$; LIN28B: $P = 10^{-3}$). Fisher's exact test was computed using equal amounts (that is, 1% of the total interactions) of randomly extracted negative subsets (HuR: $P = 10^{-7}$; LIN28B: $P = 0.0002$; Additional file 2: Figure S3).

For the *significance of score distributions*, we used Student's *t*-test to compare the score distribution of positives and negatives (HuR: $P = 10^{-16}$; LIN28B: $P = 10^{-4}$). We also performed Student's *t*-test using random extractions of negative subsets, each containing the same number of RNAs as positives (LIN28B: $P = 0.03$; HuR: $P < 10^{-8}$; Student's *t*-test).

Other statistical tests (receiver operating characteristics and precision/recall curves) are discussed in Additional file 1. The expression data for HuR and LIN28B were taken from the original manuscripts [31,33] and processed as indicated by the authors. The datasets were downloaded from GEO [29] (GSE29943) and ArrayExpress [80] (E-GEOD-44615 and E-GEOD-44613).

mRNA dataset: Human BodyMap

The Human BodyMap (HBM) 2.0 contains expression data generated using the HiSeq 2000 system and it has expression profiles for a number of human tissues [22]. The HBM RNA sequencing (RNA-seq) data was downloaded from ArrayExpress [81] under accession number E-MTAB-513. The final mRNA dataset contained 35,818 transcripts (11,584 genes) with expression levels for 14 human tissues (see section on RNA-binding protein-mRNA expression). We considered all human cDNAs from Ensembl release 68. Transcripts incompatible with the catRAPID size restrictions (that is, 50 to 1,200

nucleotides) or not expressed in at least one tissue were filtered out. In the analysis, we evaluated different CD-HIT [82] sequence similarity cutoff thresholds (see section on Gene ontology analysis).

RNA-binding protein dataset: Human Protein Atlas

We considered all the RBPs reported in two studies on RBPs binding to mRNAs [1,2]. The initial dataset consisted of 3,500 RBPs (832 genes). Proteins incompatible with catRAPID's size restrictions (that is, 50 to 750 amino acids) and above a CD-HIT [82] sequence similarity cutoff of 75% were filtered out. Similarly, proteins not present in the Human Protein Atlas (HPA) database (version 11.0) [34] and not expressed in at least one tissue were discarded. The final RBP (HPA) dataset contained 612 proteins (491 genes) with expression levels for 14 human tissues (see section on RNA-binding protein–mRNA expression). All protein sequences were retrieved from Ensembl release 68.

RNA-binding protein dataset: Human BodyMap

As for RBPs in the HPA, filters on sequence size and redundancy were applied. Proteins not present in the Human BodyMap database (version 2.0) [6] were discarded. The final RBP (HBM) dataset contained 1,156 proteins (543 genes) with expression levels for 14 human tissues (see section on RNA-binding protein–mRNA expression). All protein sequences were retrieved from Ensembl release 68.

RNA-binding protein–mRNA expression

We analyzed 14 human tissues for which both immunohistochemistry [34] and transcript abundances [6] were available. At present, the Human Protein Atlas is the largest collection of protein abundance data available [34]. Transcripts in the *mRNA dataset* and proteins in the *RBP dataset* were represented by vectors containing the normalized relative abundance of the following tissues: adrenal gland, brain, breast, colon, heart, kidney, liver, lung, lymph, muscle, lymph node, ovary, prostate and thyroid. For the immunohistochemistry data, the readouts 'no', 'low', 'intermediate' or 'high' expression were transformed into numbers (0, 1, 2, 3) and subject to Z-normalization per tissue. As for the transcript data, the vectors were Z-normalized using the average and standard deviation per tissue. For each RBP–mRNA combination we computed the pairwise Pearson's correlation coefficient of the vectors. As shown in Additional file 2: Figures S1 and S2, we observed the same trends using immunohistochemistry [34] and RNA-seq data [6] to estimate protein abundances in human tissues.

RNA-binding protein–mRNA interaction propensity

We used catRAPID [19,20] to compute the interaction propensity of each protein in the RBP dataset with each transcript in the mRNA dataset. catRAPID predicts protein–RNA associations by estimating the interaction propensity between amino acids and nucleotides using secondary structure information, hydrogen bonding and Van der Waals forces [19,20]. The approach was previously applied to predict associations between different types of proteins and RNA molecules [21,23]. Although each protein binds to distinct types of RNA structures [83], we observe that the contribution of hairpin loops accounts for 57% of the overall interaction propensity [19]. The catRAPID web server is publicly accessible from our webpage [84].

Protein–RNA interaction and expression

For a given protein, interacting (n_{int}) and non-interacting (n_{no-int}) protein–RNA pairs were compared at different AUCs (areas under the curve) of the interaction propensity distribution. The enrichment in positively correlated expression (Figure 2C) is calculated as:

$$\begin{aligned} & \text{enrichment (co-expressed interactions)} \\ &= \frac{n_{int}(r > r_{th}) - n_{no-int}(r > r_{th})}{n_{no-int}(r > r_{th})} \end{aligned} \quad (1)$$

In Equation (1), the correlation coefficient r follows the distribution of protein–RNA expression and the parameter $r_{th} > 0$ corresponds to an AUC spanning the range 50% to 99.5% (Figure 2B).

Similarly, for negatively correlated expressions (Figure 2D):

$$\begin{aligned} & \text{enrichment (anti-expressed interactions)} \\ &= \frac{n_{int}(r < l_{th}) - n_{no-int}(r < l_{th})}{n_{no-int}(r < l_{th})} \end{aligned} \quad (2)$$

In Equation (2), the parameter $l_{th} < 0$ corresponds to an AUC spanning the range 50% to 99.5% (Figure 2B).

Gene ontology analysis

For each area under the curve (AUC) of the catRAPID score distribution ($50\% < AUC < 99.5\%$), we created four subsets according to the correlation in tissue expression: (1) IC subset: positively correlating and interacting genes (expression correlation $\geq +0.7$ and positive interaction propensities); (2) IA subset: negatively correlating and interacting genes (expression correlation ≤ -0.7 and positive interaction propensities); (3) NIC subset: positively correlating and non-interacting genes (expression correlation $\geq +0.7$ and negative interaction propensities); (4) NIA subset: negatively correlating and non-interacting genes (expression correlation ≤ -0.7 and negative interaction propensities). The expression correlation of $|0.7|$ corresponds to AUC = 95% of the statistical distribution, for

which we found the highest enrichments (Figure 2C,D). We systematically applied conditional tests for GO term over-representation in each subset using the GOStats package (version 2.28.0) available from Bioconductor [85]. To assess the over-representation of a GO term in one particular subset at a certain AUC, we considered five criteria (Additional file 3: Table S1; Additional file 6: Table S4; Additional file 2: Figure S6):

1. The GO term must be reported for more than two genes.
2. The P value of the GO term must be significant ($P < 0.05$) in the subset of interest and non-significant ($P > 0.1$) in the others.
3. The enrichment must be conserved with respect to: (a) the entire human transcriptome (that is, including RNAs longer than 1,200 nucleotides and independently of expression data), (b) the complete set of analyzed genes (that is, including RNAs shorter than 1,200 nucleotides and with available expression) and (c) all genes under the same AUC (that is, considering both interacting and non-interacting pairs at the two tails of the distribution).
4. The P value of the GO term must be non-significant ($P > 0.1$) in: (a) the complete set of analyzed genes compared to the human transcriptome (significance would indicate enrichment irrespective of the subset assignment) and (b) the list of transcripts compatible with catRAPID length requirements compared to the human transcriptome (significance would indicate length bias in the statistics; see section on length bias statistics).
5. The enrichment must be conserved after sequence redundancy reduction to the 80% identity threshold.

Length bias statistics

Due to the conformational space of nucleotide chains, prediction of RNA secondary structures is difficult when RNA sequences are >1,200 nucleotides and simulations cannot be completed on standard processors (2.5 GHz; 4 to 8 GB memory). To see whether GO enrichment is biased by the catRAPID length restriction, we used a hypergeometric test (see section on the RNA-binding protein–mRNA interaction propensity). If a GO term is enriched in the length-restricted set, it is excluded *a priori* from the analysis because genes annotated in that GO term would be only selected for the length range. Thus, we imposed that GO terms must be non-significant ($P > 0.1$) in the length-restricted set of genes (see section on gene ontology analysis). This condition ensures that there is no bias due to length restrictions for any GO term enriched in a particular subset (Additional file 3: Table S1).

Analysis of RNA-binding protein sequence disorder

The content of disordered regions in the RBP sequences was computed using IUPred [36]. For each protein, we extracted structurally disordered regions (IUPred score higher than 0.4) and calculated their interactions against the reference transcriptome. We compared the interaction propensities of each disordered region with that of the full-length protein and assessed if there was an increase or decrease of the interaction propensity score (Figure 3A). The contribution of the disordered region was evaluated using a Mann–Whitney U test, where a significant increase ($P < 0.05$; $H_0 < H_1$) in the interaction propensity score is associated with a positive contribution. From low to high interaction propensities, the contribution of disorder decreases progressively with respect to that of the full-length proteins (Figure 3A). The role of disorder is more pronounced in proteins lacking canonical RNA-binding domains, indicating that unstructured regions have a direct involvement in contacting RNA (Figure 3B).

Web server

catRAPID express [27] is a publicly available implementation of catRAPID [19,20], which is used to study the relation between protein–RNA interaction propensity and expression in *Homo sapiens*. The tool has two components: (1) catRAPID predictions of protein–RNA interaction and (2) the computation of correlation using protein and RNA expression profiles [6,34]. A description of how catRAPID makes predictions can be found in the *Documentation, Tutorial and Frequently Asked Questions (FAQs)* on the webpage. Expression profiles of the RBP dataset and mRNA dataset are assigned respectively to input proteins and RNA using a homology-based criterion (ten top-ranked proteins with a BLAST [86] $e \leq 0.01$ and $\geq 75\%$ whole sequence similarity; ten top-ranked transcripts with a BLAST $e \leq 0.01$ and $\geq 95\%$ whole sequence similarity). Sequence similarity is evaluated using the Needleman–Wunsch algorithm [87].

Additional files

Additional file 1: Additional materials and methods.

Additional file 2: Figure S1. With respect to non-interacting protein–RNA pairs, non-correlated protein–RNA expression does not show enrichment using (A) RNA and (B) protein expression. Areas under the curve (AUCs) were used to select the same number of interacting/non-interacting and positively/negatively expressed protein–RNA pairs for the analysis.

Figure S2. Protein–RNA interaction and expression (immunohistochemistry expression data). (A) With respect to non-interacting protein–RNA pairs, predicted associations had enriched positively correlated expression.

(B) Compared to non-interacting protein–RNA pairs, predicted associations had enriched negatively correlated expression. **Figure S3.** P value distribution for HuR and LIN28B predictions. We compared P values (Fisher's exact test) for the catRAPID predictions for HuR and LIN28B RNA interactions (red arrow) using balanced bootstrap resampling

(random extractions of negative subsets with the same amount as the positive subset). The predicted interactions differ significantly from random associations. **Figure S4.** Receiver operating characteristic (ROC) and precision/recall (PR) curves for HuR and LIN28B predictions. We evaluated changes in the ROC and PR curves for the catRAPID predictions for the **(A)** HuR and **(B)** LIN28B RNA interactome for random samples using several ratios of positive and negative associations (pos/neg ratios).

Figure S5. Examples of protein–RNA anti-expression scenarios. **(A)** We propose that YTHDC1 represses the expression of tumor-associated genes by destabilizing mRNAs. **(B)** *Nodal* expression in adult tissues is associated with tumor progression, which might be due to transcript stabilization.

Figure S6. Nested representation of gene sets used in GO enrichment analysis. **Figure S7.** Changes in transcript and gene counts after sequence redundancy reduction. The mRNA database comprises 35,818 transcripts (11,584 genes). After redundancy filtering, the mRNA database is reduced to 33,936 transcripts (11,483 genes) at 95% sequence identity threshold; 32,700 transcripts (11,406 genes) at 90%; 31,287 transcripts (11,360 genes) at 85% and 29,673 transcripts (11,317 genes) at 80%.

Additional file 3: Table S1. mRNA GO-term enrichment analysis of interacting mRNA–RBP pairs (*P* values). Every GO term has been tested for over-representation for each subset (IC, IA, NIC and NIA) with respect to the human transcriptome, the complete set of analyzed genes (analyzed mRNA set) and the analyzed genes with the same AUC (relative AUC subset). Statistical control for the biased over-representation of GO terms in the complete set of analyzed genes and in a catRAPID length-restricted set of genes was used for the human transcriptome (see Materials and methods). Significant *P* values for the IC and IA subsets are shown in red. GOBP, gene ontology biological process; GOCC, gene ontology cellular component; GOMF, gene ontology molecular function; IA, interacting and anti-correlated in expression; IC, interacting and correlated in expression; NIC, not interacting and correlated in expression; Seq%id, sequence identity threshold used for redundancy reduction.

Additional file 4: Table S2. *NAP1L1* read counts from TIA-1 iCLIP data. The count of reads mapping into the *NAP1L1* gene and the relative cumulative distribution functions (cdf) are reported from the iCLIP experiment for controls and replicates of the TIA-1 protein [20]. The read count cdf was estimated after removal of genes with zero counts.

Additional file 5: Table S3. *Nodal* anti-expressed interacting (IA) RBPs.

Additional file 6: Table S4. mRNA GO-term enrichment analysis of interacting mRNA–RBP pairs (IC and IA gene counts are reported). Every GO term has been tested for over-representation in the IC and IA subsets with respect to the human transcriptome, the complete set of analyzed genes (analyzed mRNAs set) and the analyzed genes under the same AUC (relative AUC subset). GOBP, gene ontology biological process; GOCC, gene ontology cellular component; GOMF, gene ontology molecular function; IA, interacting and anti-correlated in expression; IC, interacting and correlated in expression; Seq%id, sequence identity threshold used for redundancy reduction.

Abbreviations

AUC: area under the curve; GEO: Gene Expression Omnibus; GO: gene ontology; HBM: Human BodyMap; HPA: Human Protein Atlas; HuR: human antigen R; IA: interacting and anti-expressed; IC: interacting and co-expressed; LFC: log-fold change; LIN28B: lin-28 homolog B; NIA: non-interacting and anti-expressed; NIC: non-interacting and co-expressed; NOP56: Nuclear Protein 5A; PAR-CLIP: photoactivatable-ribonucleoside-enhanced cross-linking and immunoprecipitation; RBP: RNA-binding protein; RNA-seq: RNA sequencing; TSC-2: tuberous sclerosis 2.

Competing interests

The authors declare that there are no competing financial interests.

Authors' contributions

GGT conceived this study. FA and GGT designed the *in silico* experiments. DC, FA and CML performed the computational analysis. DC, DM, TBO, FA and GGT analyzed the data. DM, DC and CML searched the literature on RNA networks. DC, DM and GGT wrote the manuscript. All authors read and approved the final version of the manuscript.

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References

- Castello A, Fischer B, Eichelbaum K, Horos R, Beckmann BM, Strein C, Davey NE, Humphreys DT, Preiss T, Steinmetz LM, Krijgsvelde J, Hentze MW: **Insights into RNA biology from an atlas of mammalian mRNA-binding proteins.** *Cell* 2012, **149**:1393–1406.
- Baltz AG, Munschauer M, Schwanhäusser B, Vasile A, Murakawa Y, Schueler M, Youngs N, Penfold-Brown D, Drew K, Milek M, Wyler E, Bonneau R, Selbach M, Dieterich C, Landthaler M: **The mRNA-bound proteome and its global occupancy profile on protein-coding transcripts.** *Mol Cell* 2012, **46**:674–690.
- Siomi H, Dreyfuss G: **RNA-binding proteins as regulators of gene expression.** *Curr Opin Genet Dev* 1997, **7**:345–353.
- Cook KB, Kazan H, Zuberi K, Morris Q, Hughes TR: **RBPDB: a database of RNA-binding specificities.** *Nucleic Acids Res* 2011, **39**:D301–D308.
- Dassi E, Malossini A, Re A, Mazza T, Tebaldi T, Caputi L, Quattrone A: **AURA: Atlas of UTR Regulatory Activity.** *Bioinform Oxf Engl* 2012, **28**:142–144.
- Harrow J, Frankish A, Gonzalez JM, Tapanari E, Diekhans M, Kokocinski F, Aken BL, Barrell D, Zadissa A, Searle S, Barnes I, Bignell A, Boychenko V, Hunt T, Kay M, Mukherjee G, Rajan J, Despacio-Reyes G, Saunders G, Steward C, Harte R, Lin M, Howald C, Tanzer A, Derrien T, Chrast J, Walters N, Balasubramanian S, Pei B, Tress M, et al: **GENCODE: the reference human genome annotation for The ENCODE Project.** *Genome Res* 2012, **22**:1760–1774.
- Merkin J, Russell C, Chen P, Burge CB: **Evolutionary dynamics of gene and isoform regulation in mammalian tissues.** *Science* 2012, **338**:1593–1599.
- Brawand D, Soumillon M, Necsulea A, Julien P, Csárdi G, Harrigan P, Weier M, Liechti A, Aximu-Petri A, Kircher M, Albert FW, Zeller U, Khaitovich P, Grützner F, Bergmann S, Nielsen R, Pääbo S, Kaessmann H: **The evolution of gene expression levels in mammalian organs.** *Nature* 2011, **478**:343–348.
- Chan ET, Quon GT, Chua G, Babak T, Trochesse M, Zirngibl RA, Aubin J, Ratcliffe MJH, Wilde A, Brudno M, Morris QD, Hughes TR: **Conservation of core gene expression in vertebrate tissues.** *J Biol* 2009, **8**:33.
- Wittkopp PJ, Haerum BK, Clark AG: **Evolutionary changes in cis and trans gene regulation.** *Nature* 2004, **430**:85–88.
- Masuda K, Kuwano Y, Nishida K, Rokutan K: **General RBP expression in human tissues as a function of age.** *Ageing Res Rev* 2012, **11**:423–431.
- Hogan DJ, Riordan DP, Gerber AP, Herschlag D, Brown PO: **Diverse RNA-binding proteins interact with functionally related sets of RNAs. Suggesting an extensive regulatory system.** *PLoS Biol* 2008, **6**:e255.
- Vogel C, Marcotte EM: **Insights into the regulation of protein abundance from proteomic and transcriptomic analyses.** *Nat Rev Genet* 2012, **13**:227–232.
- Maier T, Güell M, Serrano L: **Correlation of mRNA and protein in complex biological samples.** *FEBS Lett* 2009, **583**:3966–3973.
- Tartaglia GG, Pechmann S, Dobson CM, Vendruscolo M: **Life on the edge: a link between gene expression levels and aggregation rates of human proteins.** *Trends Biochem Sci* 2007, **32**:204–206.
- Greenbaum D, Colangelo C, Williams K, Gerstein M: **Comparing protein abundance and mRNA expression levels on a genomic scale.** *Genome Biol* 2003, **4**:117.
- Cox B, Kislinger T, Emili A: **Integrating gene and protein expression data: pattern analysis and profile mining.** *Methods San Diego Calif* 2005, **35**:303–314.
- Greenbaum D, Jansen R, Gerstein M: **Analysis of mRNA expression and protein abundance data: an approach for the comparison of the**

- enrichment of features in the cellular population of proteins and transcripts. *Bioinforma Oxf Engl* 2002, **18**:585–596.
19. Bellucci M, Agostini F, Masin M, Tartaglia GG: **Predicting protein associations with long noncoding RNAs.** *Nat Methods* 2011, **8**:444–445.
 20. Cirillo D, Agostini F, Tartaglia GG: **Predictions of protein–RNA interactions.** *Wiley Interdiscip Rev Comput Mol Sci* 2013, **3**:161–175.
 21. Agostini F, Cirillo D, Bolognesi B, Tartaglia GG: **X-inactivation: quantitative predictions of protein interactions in the Xist network.** *Nucleic Acids Res* 2013, **41**:e31.
 22. Iglesias-Platas I, Martin-Trujillo A, Cirillo D, Court F, Guillaumet-Adkins A, Campubi C, Bourc'his D, Hata K, Feil R, Tartaglia G, Arnaud P, Monk D: **Characterization of novel paternal ncRNAs at the Plagl1 locus, including Hymai, predicted to interact with regulators of active chromatin.** *PLoS One* 2012, **7**:e38907.
 23. Cirillo D, Agostini F, Klus P, Marchese D, Rodriguez S, Bolognesi B, Tartaglia GG: **Neurodegenerative diseases: quantitative predictions of protein–RNA interactions.** *RNA* 2013, **19**:129–140.
 24. Johnson R, Noble W, Tartaglia GG, Buckley NJ: **Neurodegeneration as an RNA disorder.** *Prog Neurobiol* 2012, **99**:293–315.
 25. Zanzoni A, Marchese D, Agostini F, Bolognesi B, Cirillo D, Botta-Orfila M, Livi CM, Rodriguez-Mulero S, Tartaglia GG: **Principles of self-organization in biological pathways: a hypothesis on the autogenous association of alpha-synuclein.** *Nucleic Acids Res* 2013, **41**:9987–9998.
 26. Agostini F, Zanzoni A, Klus P, Marchese D, Cirillo D, Tartaglia GG: **catRAPID omics: a web server for large-scale prediction of protein–RNA interactions.** *Bioinformatics* 2013, **29**:2928–2930.
 27. **catRAPID express.** [http://service.tartaglialab.com/page/catrapid_express_group]
 28. Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, Davis AP, Dolinski K, Dwight SS, Eppig JT, Harris MA, Hill DP, Issel-Tarver L, Kasarskis A, Lewis S, Matese JC, Richardson JE, Ringwald M, Rubin GM, Sherlock G: **Gene ontology: tool for the unification of biology. The gene ontology consortium.** *Nat Genet* 2000, **25**:25–29.
 29. Barrett T, Wilhite SE, Ledoux P, Evangelista C, Kim IF, Tomashevsky M, Marshall KA, Phillippy KH, Sherman PM, Holko M, Yefanov A, Lee H, Zhang N, Robertson CL, Serova N, Davis S, Soboleva A: **NCBI GEO: archive for functional genomics data sets – update.** *Nucleic Acids Res* 2012, **41**:D991–D995.
 30. Parkinson H, Kapushesky M, Kolesnikov N, Rustici G, Shojatalab M, Abeygunawardena N, Berube H, Dylag M, Emam I, Farne A, Holloway E, Lukk M, Malone J, Mani R, Piliicheva E, Rayner TF, Rezwan F, Sharma A, Williams E, Bradley XZ, Adamusiak T, Brandizi M, Burdett T, Coulson R, Krestyaninova M, Kurnosov P, Maguire E, Neogi SG, Rocca-Serra P, Sansone S-A, et al: **ArrayExpress update – from an archive of functional genomics experiments to the atlas of gene expression.** *Nucleic Acids Res* 2009, **37**:D868–D872.
 31. Lebedeva S, Jens M, Theil K, Schwanhäusser B, Selbach M, Landthaler M, Rajewsky N: **Transcriptome-wide analysis of regulatory interactions of the RNA-binding protein HuR.** *Mol Cell* 2011, **43**:340–352.
 32. Graf R, Munschauer M, Mastrobuoni G, Mayr F, Heinemann U, Kempa S, Rajewsky N, Landthaler M: **Identification of LIN28B-bound mRNAs reveals features of target recognition and regulation.** *RNA Biol* 2013, **10**:1146–1159.
 33. Hafner M, Max KEA, Bandaru P, Morozov P, Gerstberger S, Brown M, Molina H, Tuschl T: **Identification of mRNAs bound and regulated by human LIN28 proteins and molecular requirements for RNA recognition.** *RNA* 2013, **19**:613–626.
 34. Uhlen M, Oksvold P, Fagerberg L, Lundberg E, Jonasson K, Forsberg M, Zwahlen M, Kampf C, Wester K, Hober S, Wernerus H, Björling L, Ponten F: **Towards a knowledge-based human protein atlas.** *Nat Biotechnol* 2010, **28**:1248–1250.
 35. Lu P, Vogel C, Wang R, Yao X, Marcotte EM: **Absolute protein expression profiling estimates the relative contributions of transcriptional and translational regulation.** *Nat Biotechnol* 2007, **25**:117–124.
 36. Dosztányi Z, Csizmok V, Tompa P, Simon I: **IUPred: web server for the prediction of intrinsically unstructured regions of proteins based on estimated energy content.** *Bioinforma Oxf Engl* 2005, **21**:3433–3434.
 37. Gsponer J, Babu MM: **Cellular strategies for regulating functional and nonfunctional protein aggregation.** *Cell Rep* 2012, **2**:1425–1437.
 38. Babu MM, van der Lee R, de Groot NS, Gsponer J: **Intrinsically disordered proteins: regulation and disease.** *Curr Opin Struct Biol* 2011, **21**:432–440.
 39. Vavouri T, Semple JJ, Garcia-Verdugo R, Lehner B: **Intrinsic protein disorder and interaction promiscuity are widely associated with dosage sensitivity.** *Cell* 2009, **138**:198–208.
 40. Strausberg RL, Buetow KH, Emmert-Buck MR, Klausner RD: **The cancer genome anatomy project: building an annotated gene index.** *Trends Genet TIG* 2000, **16**:103–106.
 41. Cai S, Everitt JI, Kugo H, Cook J, Kleymenova E, Walker CL: **Polycystic kidney disease as a result of loss of the tuberous sclerosis 2 tumor suppressor gene during development.** *Am J Pathol* 2003, **162**:457–468.
 42. Simonnet H, Demont J, Pfeiffer K, Guenaneche L, Bouvier R, Brandt U, Schagger H, Godinot C: **Mitochondrial complex I is deficient in renal oncocytomas.** *Carcinogenesis* 2003, **24**:1461–1466.
 43. Wahl MC, Will CL, Lührmann R: **The spliceosome: design principles of a dynamic RNP machine.** *Cell* 2009, **136**:701–718.
 44. Crabtree JS, Jelinsky SA, Harris HA, Choe SE, Cotreau MM, Kimberland ML, Wilson E, Saraf KA, Liu W, McCampbell AS, Dave B, Broadus RR, Brown EL, Kao W, Skotnicki JS, Abou-Gharbia M, Winneker RC, Walker CL: **Comparison of human and rat uterine leiomyomata: identification of a dysregulated mammalian target of rapamycin pathway.** *Cancer Res* 2009, **69**:6171–6178.
 45. Francia G, Man S, Teicher B, Grasso L, Kerbel RS: **Gene expression analysis of tumor spheroids reveals a role for suppressed DNA mismatch repair in multicellular resistance to alkylating agents.** *Mol Cell Biol* 2004, **24**:6837–6849.
 46. Manalo DJ, Rowan A, Lavoie T, Natarajan L, Kelly BD, Ye SQ, Garcia JGN, Semenza GL: **Transcriptional regulation of vascular endothelial cell responses to hypoxia by HIF-1.** *Blood* 2005, **105**:659–669.
 47. Beyer S, Kristensen MM, Jensen KS, Johansen JV, Staller P: **The histone demethylases JMJD1A and JMJD2B are transcriptional targets of hypoxia-inducible factor HIF.** *J Biol Chem* 2008, **283**:36542–36552.
 48. Rohan S, Tu JJ, Kao J, Mukherjee P, Campagne F, Zhou XK, Hyjek E, Alonso MA, Chen Y-T: **Gene expression profiling separates chromophobe renal cell carcinoma from oncocytoma and identifies vesicular transport and cell junction proteins as differentially expressed genes.** *Clin Cancer Res Off J Am Assoc Cancer Res* 2006, **12**:6937–6945.
 49. Tan M-H, Wong CF, Tan HL, Yang XJ, Ditlev J, Matsuda D, Khoo SK, Sugimura J, Fujioka T, Furge KA, Kort E, Giraud S, Ferlicot S, Vihl P, Amsellem-Ouazana D, Debré B, Flam T, Thiounn N, Zerbib M, Benoît G, Droupy S, Molinié V, Vieillefond A, Tan PH, Richard S, Teh BT: **Genomic expression and single-nucleotide polymorphism profiling discriminates chromophobe renal cell carcinoma and oncocytoma.** *BMC Cancer* 2010, **10**:196.
 50. Martin P, Parkhurst SM: **Parallels between tissue repair and embry morphogenesis.** *Dev Camb Engl* 2004, **131**:3021–3034.
 51. Lu H, Ouyang W, Huang C: **Inflammation, a key event in cancer development.** *Mol Cancer Res MCR* 2006, **4**:221–233.
 52. Rider CC, Mulloy B: **Bone morphogenetic protein and growth differentiation factor cytokine families and their protein antagonists.** *Biochem J* 2010, **429**:1–12.
 53. Standart N, Jackson RJ: **Regulation of translation by specific protein/mRNA interactions.** *Biochimie* 1994, **76**:867–879.
 54. De Moor CH, Richter JD: **Translational control in vertebrate development.** *Int Rev Cytol* 2001, **203**:567–608.
 55. Quenneville S, Turelli P, Bojkowska K, Raclot C, Offner S, Kapopoulou A, Trono D: **The KRAB-ZFP/KAP1 system contributes to the early embryonic establishment of site-specific DNA methylation patterns maintained during development.** *Cell Rep* 2012, **2**:766–773.
 56. Zhang Z, Theler D, Kaminska KH, Hiller M, de la Grange P, Pudimat R, Rafalska I, Heinrich B, Bujnicki JM, Allain FH-T, Stamm S: **The YTH domain is a novel RNA binding domain.** *J Biol Chem* 2010, **285**:14701–14710.
 57. Harigaya Y, Tanaka H, Yamanaka S, Tanaka K, Watanabe Y, Tsutsumi C, Chikashige Y, Hiraoka Y, Yamashita A, Yamamoto M: **Selective elimination of messenger RNA prevents an incidence of untimely meiosis.** *Nature* 2006, **442**:45–50.
 58. Rafalska I, Zhang Z, Benderska N, Wolff H, Hartmann AM, Brack-Werner R, Stamm S: **The intranuclear localization and function of YT521-B is regulated by tyrosine phosphorylation.** *Hum Mol Genet* 2004, **13**:1535–1549.
 59. Zhang B, Zur Hausen A, Orłowska-Volk M, Jäger M, Bettendorf H, Stamm S, Hirschfeld M, Yiqin O, Tong X, Gitsch G, Sticker E: **Alternative splicing-related factor YT521: an independent prognostic factor in endometrial cancer.** *Int J Gynecol Cancer Off J Int Gynecol Cancer Soc* 2010, **20**:492–499.
 60. Baldwin RM, Moretton A, Paris G, Goulet I, Côté J: **Alternatively spliced protein arginine methyltransferase 1 isoform PRMT1v2 promotes the**

- survival and invasiveness of breast cancer cells. *Cell Cycle* 2012, **11**:4597–4612.
61. Zhong J, Cao R-X, Zu X-Y, Hong T, Yang J, Liu L, Xiao X-H, Ding W-J, Zhao Q, Liu J-H, Wen G-B: **Identification and characterization of novel spliced variants of PRMT2 in breast carcinoma.** *FEBS J* 2012, **279**:316–335.
 62. Yan T-D, Wu H, Zhang H-P, Lu N, Ye P, Yu F-H, Zhou H, Li W-G, Cao X, Lin Y-Y, He J-Y, Gao W-W, Zhao Y, Xie L, Chen J-B, Zhang X-K, Zeng J-Z: **Oncogenic potential of retinoic acid receptor-gamma in hepatocellular carcinoma.** *Cancer Res* 2010, **70**:2285–2295.
 63. Li D-P, Li Z-Y, Sang W, Cheng H, Pan X-Y, Xu K-L: **HOXA9 gene expression in acute myeloid leukemia.** *Cell Biochem Biophys* 2013, **67**:935–938.
 64. Hirschfeld M, Zhang B, Jaeger M, Stamm S, Erbes T, Mayer S, Tong X, Stickeler E: **Hypoxia-dependent mRNA expression pattern of splicing factor YT521 and its impact on oncological important target gene expression.** *Mol Carcinog* 2013. in press.
 65. Harris AL: **Hypoxia – a key regulatory factor in tumour growth.** *Nat Rev Cancer* 2002, **2**:38–47.
 66. Pieczyk M, Wax S, Beck AR, Kedersha N, Gupta M, Maritim B, Chen S, Gueydan C, Krusys V, Streuli M, Anderson P: **TIA-1 is a translational silencer that selectively regulates the expression of TNF-alpha.** *EMBO J* 2000, **19**:4154–4163.
 67. Dixon DA, Balch GC, Kedersha N, Anderson P, Zimmerman GA, Beauchamp RD, Prescott SM: **Regulation of cyclooxygenase-2 expression by the translational silencer TIA-1.** *J Exp Med* 2003, **198**:475–481.
 68. Yamasaki S, Stoeklin G, Kedersha N, Simarro M, Anderson P: **T-cell intracellular antigen-1 (TIA-1)-induced translational silencing promotes the decay of selected mRNAs.** *J Biol Chem* 2007, **282**:30070–30077.
 69. Wang Z, Kayikci M, Briesse M, Zarnack K, Luscombe NM, Rot G, Zupan B, Curk T, Ule J: **iCLIP predicts the dual splicing effects of TIA-RNA interactions.** *PLoS Biol* 2010, **8**:e1000530.
 70. Kidd M, Modlin IM, Mane SM, Camp RL, Eick G, Latich I: **The role of genetic markers – NAP1L1, MAGE-D2, and MTA1 – in defining small-intestinal carcinoid neoplasia.** *Ann Surg Oncol* 2006, **13**:253–262.
 71. Drozdov I, Kidd M, Nadler B, Camp RL, Mane SM, Hauso O, Gustafsson BI, Modlin IM: **Predicting neuroendocrine tumor (carcinoid) neoplasia using gene expression profiling and supervised machine learning.** *Cancer* 2009, **115**:1638–1650.
 72. Guidi F, Puglia M, Gabbiani C, Landini I, Gamberi T, Fregona D, Cinellu MA, Nobili S, Mini E, Bini L, Modesti PA, Modesti A, Messori L: **2D-DIGE analysis of ovarian cancer cell responses to cytotoxic gold compounds.** *Mol Biosyst* 2012, **8**:985–993.
 73. Nagata T, Takahashi Y, Ishii Y, Asai S, Nishida Y, Murata A, Koshinaga T, Fukuzawa M, Hamazaki M, Asami K, Ito E, Ikeda H, Takamatsu H, Koike K, Kikuta A, Kuroiwa M, Watanabe A, Kosaka Y, Fujita H, Miyake M, Mugishima H: **Transcriptional profiling in hepatoblastomas using high-density oligonucleotide DNA array.** *Cancer Genet Cytogenet* 2003, **145**:152–160.
 74. Lawrence MG, Margaryan NV, Loessner D, Collins A, Kerr KM, Turner M, Seftor EA, Stephens CR, Lai J, Postovit L-M, Clements JA, Hendrix MJC, APC BioResource: **Reactivation of embryonic nodal signaling is associated with tumor progression and promotes the growth of prostate cancer cells.** *Prostate* 2011, **71**:1198–1209.
 75. Ravasi T, Suzuki H, Cannistraci CV, Katayama S, Bajic VB, Tan K, Akalin A, Schmeier S, Kanamori-Katayama M, Bertin N, Carninci P, Daub CO, Forrest ARR, Gough J, Grimmond S, Han J-H, Hashimoto T, Hide W, Hofmann O, Kamburov A, Kaur M, Kawaji H, Kubosaki A, Lassmann T, van Nimwegen E, MacPherson CR, Ogawa C, Radovanovic A, Schwartz A, Teasdale RD, *et al*: **An atlas of combinatorial transcriptional regulation in mouse and man.** *Cell* 2010, **140**:744–752.
 76. Wu L, Candille SI, Choi Y, Xie D, Jiang L, Li-Pook-Than J, Tang H, Snyder M: **Variation and genetic control of protein abundance in humans.** *Nature* 2013, **499**:79–82.
 77. Tartaglia GG, Vendruscolo M: **Correlation between mRNA expression levels and protein aggregation propensities in subcellular localisations.** *Mol Biosyst* 2009, **5**:1873–1876.
 78. Tartaglia GG, Pechmann S, Dobson CM, Vendruscolo M: **A relationship between mRNA expression levels and protein solubility in *E. coli*.** *J Mol Biol* 2009, **388**:381–389.
 79. Olzscha H, Schermann SM, Woerner AC, Pinkert S, Hecht MH, Tartaglia GG, Vendruscolo M, Hayer-Hartl M, Hartl FU, Vabulas RM: **Amyloid-like aggregates sequester numerous metastable proteins with essential cellular functions.** *Cell* 2011, **144**:67–78.
 80. ArrayExpress. [www.ebi.ac.uk/arrayexpress]
 81. ArrayExpress. [http://www.ebi.ac.uk/arrayexpress/experiments/E-MTAB-513]
 82. Li W, Godzik A: **Cd-hit: a fast program for clustering and comparing large sets of protein or nucleotide sequences.** *Bioinformatics* 2006, **22**:1658–1659.
 83. Li X, Kazan H, Lipshitz HD, Morris QD: **Finding the target sites of RNA-binding proteins.** *Wiley Interdiscip Rev RNA* 2014, **5**:111–130.
 84. Tartaglia's group web servers. [http://service.tartagliolab.com]
 85. Falcon S, Gentleman R: **Using GOSTats to test gene lists for GO term association.** *Bioinforma Oxf Engl* 2007, **23**:257–258.
 86. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ: **Basic local alignment search tool.** *J Mol Biol* 1990, **215**:403–410.
 87. Rice P, Longden I, Bleasby A: **EMBOSS: the European molecular biology open software suite.** *Trends Genet TIG* 2000, **16**:276–277.

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