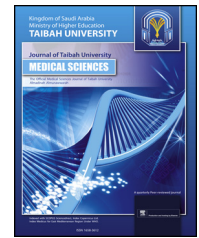




Taibah University

Journal of Taibah University Medical Sciences

www.sciencedirect.com



Experimental Article

Crocin mitigates carbon tetrachloride-induced liver toxicity in rats



Saleh Bahashwan, PhD^a, Memy H. Hassan, PhD^{a,*}, Hamdy Aly, PhD^b,
Mohamed M. Ghobara, PhD^c, Hesham A. El-Beshbishy, PhD^c and
Ibrahim Busati, B.Sc^a

^a Department of Pharmacology and Toxicology, College of Pharmacy, Taibah University, Almadinah Almunawwarah, KSA

^b Department of Pharmacology and Toxicology, Faculty of Pharmacy, King Abdel-Aziz University, Jeddah, KSA

^c Department of Medical Laboratories Technology, Faculty of Applied Medical Sciences, Taibah University, Almadinah Almunawwarah, KSA

Received 24 March 2014; revised 17 September 2014; accepted 24 September 2014; Available online 23 December 2014

المخلص

أهداف البحث: يعد رابع كلوريد الكربون واحدا من أخطر الملوثات البيئية التي تسبب سمية الكبد، لذلك تهدف هذه الدراسة إلى تقييم التأثير الوقائي المحتمل، وآلية عمل الكروسين ضد سمية الكبد المستحدثة برابع كلوريد الكربون.

طرق البحث: تم توزيع ٤٠ فأرا من الذكور للعلاج لمدة أسبوعين عن طريق إعطاء زيت الذرة، أو رابع كلوريد الكربون المذاب في زيت الذرة، أو الكروسين (١٠٠ مجم/كجم)، أو الكروسين ورابع كلوريد الكربون. تم استخراج الكبد أثناء القتل الرحيم، ووزنه ثم معالجته لتقييم الأنسجة، وتقدير محتويات الكبد من الكسباس النشط ٣، وبيروكسيد الدهون، والجلوتاثيون المختزل. كما تم تقييم أنشطة الإنزيمات المضادة للأكسدة ديسميوتاز الفائق، والجلوتاثيون بيروكسيداز والكاتالاز، وإنزيم مرحلة الأيض الأولى السيوكروم "ب" ٤٥٠. وإنزيم مرحلة الأيض الثانية، الجلوتاثيون -إس- ترانسفيراز في أنسجة الكبد. كما تم جمع عينات الدم لتقييم اختبارات وظائف الكبد، والسيوكينات الالتهابية إنترلوكين-٦ وعامل نخر الورم ألفا.

النتائج: أحدث رابع كلوريد الكربون زيادة ذات أهمية في كل من الوزن النسبي للكبد إلى وزن الجسم، ومحتوى بيروكسيد الدهون، والكسباس النشط ٣، ومستويات البلازما من إنترلوكين-٦ وعامل نخر الورم ألفا. كما أحدث رابع كلوريد الكربون اختلال في نسيج الكبد، وإنزيمات الأيض الكبدية، واختبارات وظائف الكبد. وأحدث رابع كلوريد الكربون انخفاضا ملحوظا في أنشطة الإنزيمات المضادة للأكسدة والجلوتاثيون المختزل. أدت إضافة الكروسين مع رابع كلوريد الكربون إلى تخفيف جميع العوامل المختلفة نتيجة رابع كلوريد الكربون والحفاظ على أنسجة الكبد قريبة من وضعها الطبيعي.

الاستنتاجات: قام الكروسين بتحسين الكبد من العطب الناجم من رابع كلوريد الكربون بواسطة تثبيط السيتوكينات الالتهابية، والكسباس ٣ والإجهاد التأكسدي بالإضافة إلى تعديل إنزيمات الأيض الكبدية لصالح القضاء على نواتج رابع كلوريد الكربون الأيضية السامة.

الكلمات المفتاحية: رابع كلوريد الكربون; سمية الكبد; الإجهاد التأكسدي; الكروسين

Abstract

Objectives: Carbon tetrachloride (CCl₄) is one of the most dangerous hepatotoxic environmental pollutants thus this study aimed at investigating the potential preventive effect and mechanism of crocin against CCl₄-induced hepatotoxicity.

Methods: Forty Male rats were allocated for two weeks treatment with; corn oil, CCl₄ in corn oil, crocin (100 mg/kg), or crocin plus CCl₄. At time of euthanasia liver was removed, weighted and processed for histopathological evaluation and estimation of liver contents of active caspase3, lipid peroxidation (MDA) and reduced glutathione (GSH). We also evaluated antioxidant enzymes activities [superoxide dismutase (SOD), glutathione peroxidase (GSH-Px) and catalase (CAT)], phase I metabolizing enzyme [cytochrome P450 sub family 2E1 (CYP2E1)] an Phase II metabolizing enzyme, [glutathione-S-transferase (GST)] in liver tissue. Blood samples were used for evaluation of liver function tests and inflammatory cytokines [interleukin 6 (IL-6) and tumor necrosis factor alpha (TNF-α)].

* Corresponding address: College of Pharmacy, Taibah University, P.O. Box 30001, Almadinah Almunawwarah, KSA.

E-mail: memymahmoud@yahoo.com (M.H. Hassan)

Peer review under responsibility of Taibah University.



Production and hosting by Elsevier

Results: CCl₄ induced significant ($p < 0.001$), increase in: relative liver weight to body weight, liver MDA content, liver active caspase-3 and plasma levels of IL-6 and TNF- α . In addition, CCl₄ disturbed liver histology, liver metabolizing enzymes (CYP2E1 and GST), and liver function tests (aspartate aminotransferase, alanine aminotransferase, total bilirubin and alkaline phosphatase). CCl₄ induced significant decrease in activities of SOD, CAT, GSH-Px and GSH content. Administration of crocin with CCl₄ mitigated all CCl₄-disturbed parameters and preserved liver histology close to normal.

Conclusion: Crocin ameliorated CCl₄-induced liver injury via inhibition of inflammatory cytokines, caspase3 and oxidative stress along with modulation of liver metabolizing enzymes favoring elimination of CCl₄ toxic metabolite.

Keywords: Carbon tetrachloride; Liver toxicity; Oxidative stress; Crocin

© 2014 The Authors.

Production and hosting by Elsevier Ltd on behalf of Taibah University. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Introduction

Carbon tetrachloride (CCl₄) is a common industrial solvent which is well known for its hepatotoxicity.^{1–3} CCl₄ is used in the synthesis of chlorinated organic compounds including chlorofluorocarbon refrigerants, agricultural fumigant, in the production of semiconductors, in the processing of fats, oils and rubber and in laboratory applications.^{4,5}

Occupational exposure to carbon tetrachloride may occur in the chemical industry, in laboratories, and during degreasing operations.^{6,7} Numerous poisonings and fatalities have occurred due to high exposure to carbon tetrachloride. The major pathological changes have been seen in the liver even at levels that were generally below 5 ppm (32 mg/m³).^{8,9} Liver damage can occur after 24 h and in serious cases resulted in hepatic coma and death.^{10,11} Through the investigation of acute CCl₄-induced liver damage in animal models, it is now generally accepted that CCl₄ toxicity resulted from free radical generation by cytochrome P450 system in liver microsomes and consequently causes lipid peroxidation of membranes that leads to liver injury.^{12–15}

One of the possible popular successful approaches for the protection and even treatment of liver damage due to oxidative stress could be achieved through use of phytochemicals with antioxidant properties.^{13,16,17} One such phytochemical is crocin, a main constituent of the *Crocus sativus* (*Saffron*), exhibits a variety of pharmacological effects including inhibition of skin tumor growth in mice,¹⁸ improvement of learning behavior previously impaired by ethanol,¹⁹ anti-hyperlipidemic effects,²⁰ anti-atherosclerotic effects,²¹ and anti-cancer effect.^{22,23} Interestingly, recent studies indicated, potential free radical

scavenging, antioxidant and lipid peroxidation inhibition properties of crocin.^{24,25}

Note worthily, study by Lin and Wang²⁶ pointed out a first clue for partial hepatoprotective effect of crocin against liver carcinogenesis induced by diethylnitrosamin and aflatoxin B₁. Our previous study documented a possible protective utility of crocin against beryllium chloride-induced brain and liver injury.²⁵ As a continuation for efforts to establish an effective hepatoprotective modality utilizing nature substance against CCl₄ an environmental toxin, it is a pertinent to study crocin in well-designed animal model for liver toxicity, which will answer questions regarding its protective utility and potential mechanism. Therefore, the global aim of this proposal is to evaluate the potential protective utility of crocin against hepatic toxicity induced by carbon tetrachloride. Moreover, exploring the possible mechanisms whereby this agent mediated its beneficial effects.

Materials and Methods

Animals and study protocol

Forty male Sprague–Dawley rats (200–250 g) were kept at 20–25°C in a 12 h light/12 h dark cycle with free access to food and water. The animals were feed standard chow and water *ad libitum*. The experimental protocols were approved and carried out according to guidelines for the use and care of experimental animals.

Design of the work

After a period of adaption, rats were classified into four groups ($n = 10$) as follow:

- > Group I: Negative control; treated intraperitoneal (i.p.) with corn oil in a dose of 0.2 ml/100 g animal.
- > Group II: crocin group; injected i.p. with crocin in a dose of 100 mg/kg/day.^{25,26}
- > Group III: CCl₄ group; Injected i.p. with CCl₄, dissolved 1:1 in sterile corn oil. In a dose of 0.2 mL/100 g animal for two consecutive days/week.^{13,28,29}
- > Group IV: Combination group; injected I.P with both crocin and CCl₄ using the same doses schedule mention before.

Animal treatment was continued for 2 weeks then the experiment was concluded and animals were killed under anesthesia, blood samples were collected and livers were rapidly removed then weighted to calculate relative liver weight to body weight. Liver and blood samples were processed for measuring the following parameters:

Biochemical analysis:

Each blood sample were placed in dry clean centrifuge tube, and then centrifuged for 10 min at 3000 revolutions per minute (rpm) to separate the serum. Serum was carefully separated into clean dry Wassermann tubes by using a Pasteur pipette and used for determination of serum liver function tests [(aspartate aminotransferase (AST), alanine aminotransferase (ALT) alkaline phosphatase (ALP) and total bilirubin,] using standard techniques.³⁰

Histological evaluation:

Liver samples were fixed in 4% buffered formaldehyde, embedded in paraffin, and stained with hematoxylin-eosin according to Bancroft and Stevens.³¹ Histologic evaluation was performed twice by two pathologists blinded to the protocol.

Estimation of liver metabolizing enzymes activities

Cytosol and microsomes fractions were prepared at 0–4 °C from liver tissue homogenate according to method of Benson et al.,³² and used for evaluation of metabolizing enzymes. Liver microsomal phase I metabolizing enzyme; CYP2E1 activity was assayed using *p*-nitrophenol as a substrate³³ Cytosolic liver phase II metabolizing enzyme; Glutathione –S-transferase (GST) activity was determined using 1-chloro 2,4 dinitrobenzene (CDNB) as a substrate according to method of Habig et al.³⁴ GST and CYP2E1 activities were normalized against protein content and presented as percentage of corresponding control values.

Measurement of liver oxidative stress status

Liver tissue malondialdehyde (MDA), an indicator of lipid peroxidations; superoxide dismutase (SOD); catalase (CAT); glutathione peroxidase (GSH-Px); and reduced glutathione (GSH) measurements were performed as described previously^{35–38} to determine the oxidative status in the liver specimens.

Evaluation of inflammatory cytokines

Plasma Interleukin-6 (IL-6) was determined colorimetrically using rat Elisa Kit (IBL Co., Ltd. Hamburg, Germany) in accordance with the manufacturer's instructions. Plasma tumor necrosis factor alpha (TNF- α) level was determined via a commercial ELISA kit (IBL Co., Ltd. Hamburg, Germany) using standard curve according to supplier protocol.

Evaluation of active Caspase3

The caspase-3 enzyme activity was measured in liver tissues collected from rats in the four treatment groups using the CaspACE assay system (Promega Corp., Madison, WI) based on the ability of the caspase 3 enzyme to release the yellow chromophore *p*-nitroaniline (pNA) from the colorimetric substrate (Ac-DEVD-pNA) provided in the CaspACE assay system. Relative caspase-3 activities for each sample and sample plus inhibitor were calculated from the standard curve as described previously.³⁹ Caspase-3 activity values were normalized against sample protein content and presented as percentage of control value.

Statistical analysis

Results were expressed as mean \pm SE, and the significance of differences were assessed by one-way ANOVA and Tukey's test as post hoc. The differences were accepted as statistically significant when *P* value was less than 0.05.

Results

Effect of treatment on body weight and relative liver weight to body weight

As illustrated in Table 1; the ratio of liver weight to 100 g body weight was significantly increased by sole administration of CCl₄ (5.8 ± 0.25 , $p < 0.01$) compared to control animals showing 4.5 ± 0.10 . Interestingly treatment with both crocin and CCl₄ exhibited liver weight/100 g body weight ratio of 4.7 ± 0.17 which is significantly lower than CCl₄ group ($p < 0.01$) and was close to normal value (Table 1). Comparing the animal total body weight at the end of experiment to its corresponding initial value, only, CCl₄ group exhibited a significant decrease compared to its corresponding initial weight (Table 1). Note northerly, the body weights exhibited by combination group had higher values compared to both it initial body weight and body weights exhibited by CCl₄-treated group however it is still less than the control values (Table 1).

Histological findings

Liver sections of control rats showed normal cellular architecture with distinct hepatic cells, sinusoidal spaces and central vein (Figure 1). Liver sections obtained from CCl₄ treated rat showed disarrangement of normal hepatic cells, pale stained hepatocytes around the central vein (C) with extensive vacuolization around the central vein massive fatty degeneration, multifocal centrilobular hepatic cell death, and cellular infiltration, and massive numbers of inflammatory cells infiltration (Figure 1). Conversely, mild vascular congestion (V) was detected in the crocin sections with no histopathological alteration appearing in hepatocytes. Compared to CCl₄ group, interestingly, livers of rats treated with crocin and CCl₄ revealed better preservation of the normal liver architecture and rare generalized vacuolization of the cytoplasm of hepatocytes, with apparently normal nuclei, very few inflammatory cells infiltration (Figure 1).

Effect of treatment on liver function tests

The serum levels of liver functions (ALP, ALT, AST and total bilirubin) are presented in Table 2. In the CCl₄ treated

Table 1: Effects of crocin on body and liver weights of carbon tetrachloride (CCL4) treated rats at the end of study (2 weeks).

	Body weight (% of initial)	Liver weight/100 g body weight
Control	116.64 \pm 3.7*	4.5 \pm 0.10
Crocin	113.5 \pm 4*	4.4 \pm 0.20
CCL4	94.17 \pm 2.6 ^{a,b,*}	5.8 \pm 0.25 ^{a,b}
Combination	100.47 \pm 3.2 ^{a,b}	4.7 \pm 0.17 ^c

- Data were calculated as relative weight of liver to 100 g animal body weight at the end of experiment.
- Data are presented as mean \pm standard error of 10 animals/group.
- ^{a, b or *, c} indicates significant difference from control, crocin or corresponding initial body weight respectively at $p \leq 0.01$ using Tukey's test as post ANOVA test.

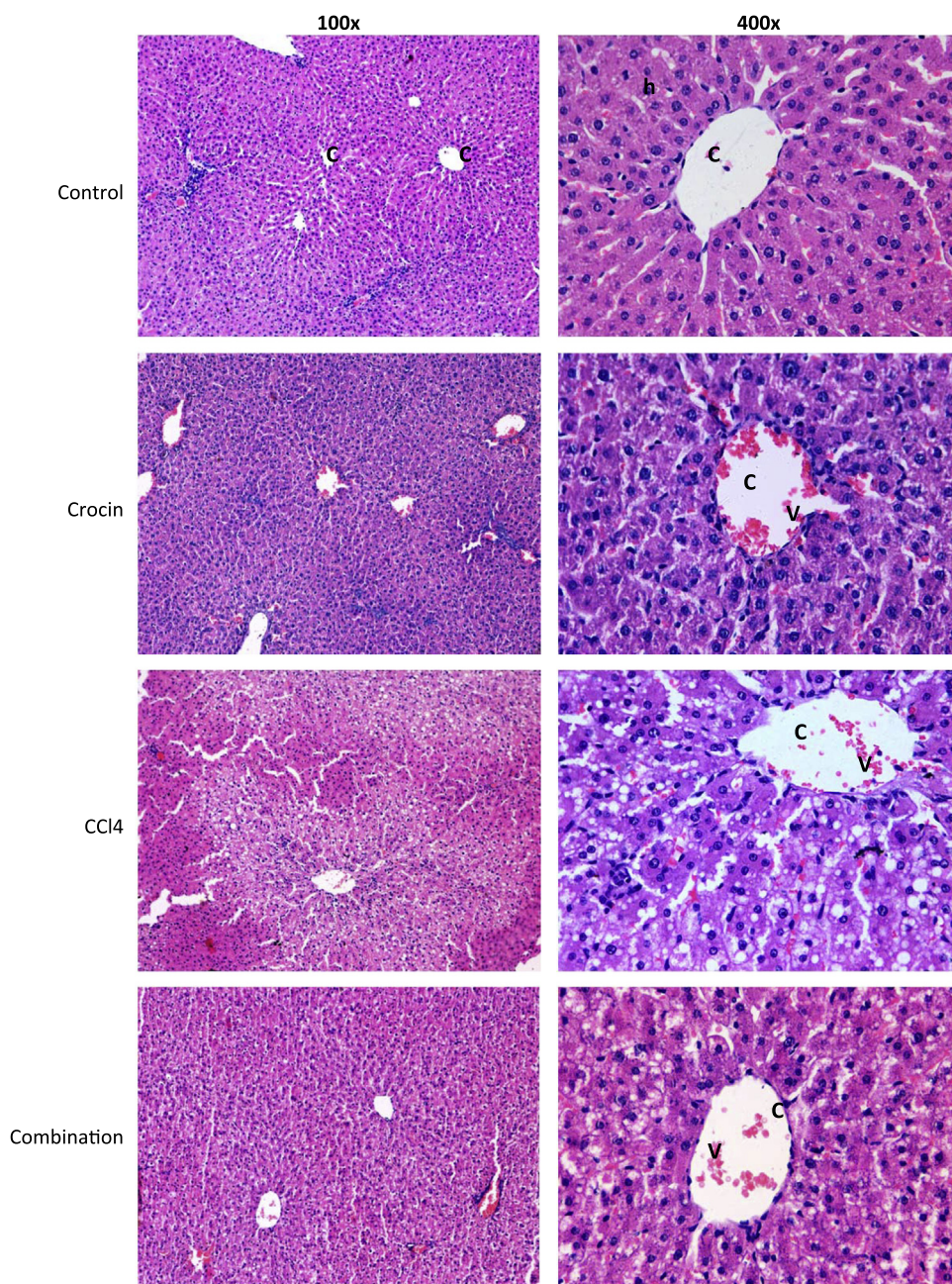


Figure 1: Representative H&E micrographs from liver tissues collected from rats treated with; corn oil (control), crocin, carbon tetrachloride (CCl₄), and combination of crocin and CCl₄. Control section showing normal histological structure of the central vein(c) and surrounding hepatocytes (h). CCl₄ treatment for two weeks showed pale stained hepatocytes around the central vein (C) with cytoplasmic vacuolization. CCl₄ induced histological changes manifested by massive number of inflammatory cells infiltration, fatty change, degeneration, necrosis and apoptosis in most of the hepatic parenchyma. Conversely, crocin treatment had no significant histopathological alteration except mild vascular congestion (V) while treatment with crocin plus CCl₄ restored normal liver histology, although showing slight inflammatory cells infiltration, few fatty changes and moderate generalized vacuolization of the cytoplasm of hepatocytes, with apparently normal nuclei, compared to CCl₄ alone.

group, the serum levels of ALP, ALT, AST and total bilirubin significantly, $p < 0.01$, increased to 369 ± 8.5 , 390 ± 6.9 , 461 ± 11.3 and 2.5 ± 0.007 respectively, compared to negative control group values of 66 ± 2.7 , 42 ± 3.9 , 36 ± 3.2 and 0.28 ± 0.005 respectively. Pretreatment of CCl₄-treated rats with crocin significantly, $p < 0.01$, decreased the CCl₄ induced elevation of these markers levels to 160 ± 7.4 , 183 ± 9.6 , 138 ± 8.5 and

0.5 ± 0.009 respectively (Table 2). Interestingly, sole crocin administration does not exhibit any significant change from control values of liver functions.

Effect of treatment on metabolizing enzymes in liver tissue

The effects of crocin and/or CCl₄ on activities of phase I metabolizing enzyme; (CYP2E1) and Phase II metabolizing

Table 2: Effect of crocin and or carbon tetrachloride on liver function test in rats after two weeks of treatment.

	Control	Crocin	CCL4	Combination
ALP (U/L)	66 ± 2.7	82 ± 3.8	369 ± 8.5 ^{a,b}	160 ± 7.4 ^{a,b,c}
ALT (U/L)	42 ± 3.9	44 ± 2.75	390 ± 6.9 ^{a,b}	183 ± 9.6 ^{a,b,c}
AST (U/L)	36 ± 3.2	38 ± 1.3	461 ± 11.3 ^{a,b}	138 ± 8.5 ^{a,b,c}
Total bilirubin (mg/dL)	0.28 ± 0.005	0.29 ± 0.004	2.5 ± 0.007 ^{a,b}	0.5 ± 0.009 ^{a,b,c}

- Data were presented as mean ± standard error of 10 animals/group.
- ^{a, b or c} indicates significant difference from control, crocin or CCl₄ respectively at $p \leq 0.01$ using Tukey's test as post ANOVA test.
- ALP: Alkaline phosphatase; AST: Aspartylaminotransferase; ALT: Alanine aminotransferase.

enzymes (GST) are compiled in Figure 2. Administration of CCl₄ resulted in a significant, $p < 0.01$, increase in CYP2E1 activity to 260% ± 6.8 of control value with concomitant significant decrease in GST activity to 45% ± 2.2 of control value respectively, $p < 0.01$. Conversely sole treatment with crocin exhibited an opposite effect. Crocin treated rats exhibited 60% ± 1.7 and 210% ± 8.1 of control values for CYP2E1, and GST respectively. Interestingly, addition of crocin to CCl₄ treatment abrogated CCl₄-induced disturbance of metabolizing enzymes and normalized all values.

Effect on lipid peroxidation and GSH level in liver tissue

CCl₄ caused a substantial increase in liver MDA content to 19.3 ± 0.5 with concomitant significant fall in liver GSH content 76 ± 2.2 compared to control group (Table 3). Administration of crocin alone showed a non-significant decrease in liver MDA content, 4.5 ± 0.5 (Table 3), while exhibited a significant increase in GSH content 168 ± 3.3 compared to control group (Table 3). Combined administration of crocin and CCl₄ resulted in a significant reduction in the liver MDA content (8.4 ± 0.75) with significant increase in liver GSH content (120 ± 4.4) compared to CCl₄ treated group (Table 3).

Effect of treatment on antioxidant enzymes activities in liver tissue

Table 3 shows the activities of enzymatic antioxidants (CAT, SOD, and GSH-Px) in the rats liver tissue. A significant decrease, $p < 0.01$, in the activities of the above mentioned enzymatic antioxidants activities was observed after CCl₄ administration (18 ± 1.3, 190 ± 12 and 8.25 ± 0.7 respectively) compared to control values of 32 ± 3.2, 380 ± 10 and 15.34 ± 0.3 respectively. In contrast, crocin alone treated group showed a significant increase in liver CAT, SOD and GSH-Px activities compared to control values. However, concomitant administration of crocin with CCl₄ significantly restored these enzyme activities back towards normalcy (27 ± 2.2, 360 ± 9 and 13.32 ± 0.9 for CAT, SOD and GSH-Px respectively) as shown in Table 3.

Results of plasma interleukin-6 and plasma tumor necrosis factor alpha

Table 4 showed that there was significant increase in plasma levels of IL-6 (110 ± 6.3) and TNF- α (188 ± 6.6) in

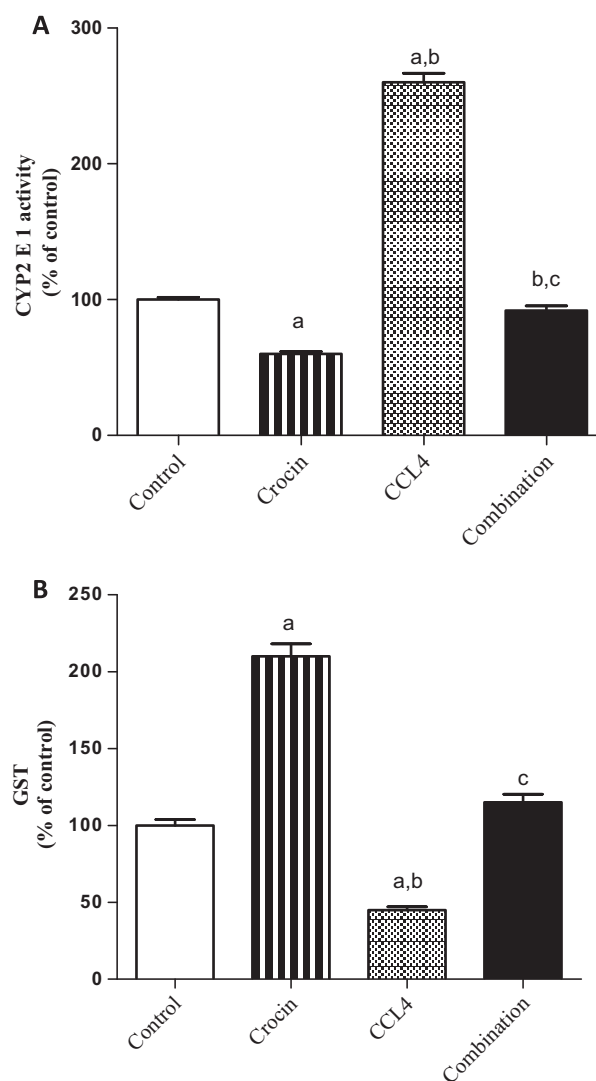


Figure 2: Effect of crocin and or carbon tetrachloride (CCl₄) on liver metabolizing enzymes activities in rat after two weeks of treatment.

- Enzymes are calculated as nmol of substrate/min/mg protein (normalized against tissue protein content) and presented as percentage of control value.
- Data are presented as mean ± standard error of 10 animals/group.
- ^{a, b or c} indicates significant difference from control, crocin or CCl₄, respectively, at $p \leq 0.01$ using Tukey's test as post ANOVA test.
- CYP 2 E1: cytochrom P450 subfamily 2 E1; GST: glutathione -S- transferase.

Table 3: Antioxidant enzymes activities, reduced glutathione content and lipid peroxidation content in liver tissue of rats treated with crocin and or carbon tetrachloride (CCL4).

	CAT (U/mg protein)	SOD (U/mg protein)	GSH-PX (U/mg protein)	GSH (umol/mg protein)	MDA (nmol/mg protein)
Control	32 ± 2.20	380 ± 10	15.34 ± 0.3	132 ± 3.5	4.75 ± 0.25
Crocin	39 ± 1.20 ^a	430 ± 10 ^a	18.39 ± 0.5 ^a	168 ± 3.3 ^a	4.5 ± 0.50
CCI4	18 ± 1.30 ^{a,b}	190 ± 12 ^{a,b}	8.25 ± 0.7 ^{a,b}	76 ± 2.2 ^{a,b}	19.3 ± 0.50 ^{a,b}
Combination	27 ± 2.25 ^{b,c}	360 ± 9 ^{b,c}	13.32 ± 0.9 ^{b,c}	120 ± 4.4 ^{b,c}	8.4 ± 0.75 ^{a,b,c}

- Data are presented as mean and standard error of 10 animals each group.
- ^{a, b or c} indicate significant change from control, crocin or CCL4 using Tukey's test as post ANOVA test at $p < 0.01$.
- Animals were treated for 2 weeks.
- CAT: Catalase; SOD: superoxide dismutase; GSH-PX: glutathione peroxidase; GSH: reduced glutathione; MDA: malondialdehyde.

CCI4 treated group as compared to normal control group exhibited; 39.7 ± 2.5 and 22 ± 1.4 for IL-6 and TNF- α respectively. On the contrary, crocin treated group had no significant change from control values. Interestingly, animals treated with both crocin and CCI4 exhibited significant mitigation of CCI4-induced changes in IL-6 and TNF- α (62 ± 3.6 and 55 ± 2.7 for IL-6 and TNF- α respectively).

Caspase-3 activity in liver tissue of rats treated with crocin and or CCI4

The CCI4 treated group exhibited a significant ($p < 0.01$) increase in active caspase-3 content to $416\% \pm 19.5$ of control value in liver tissue (Figure 3). Sole crocin administration had no significant effect on active caspase 3 activity ($98\% \pm 3.7$ of corresponding control value). While animals treated with crocin and CCI4 (combination group) showed a significant ($p < 0.01$) suppression in caspase-3 content, $143\% \pm 5$, compared to CCI4 treated group (Figure 3).

Discussion

This study aimed at evaluation of protective utility of crocin, a natural compound consumed in human diet, against CCI4-induced liver injury in rats. Since, it is well established that, CCI4-induced liver toxicity is mediated in part via free radical generation,⁴⁰ crocin was chosen in our

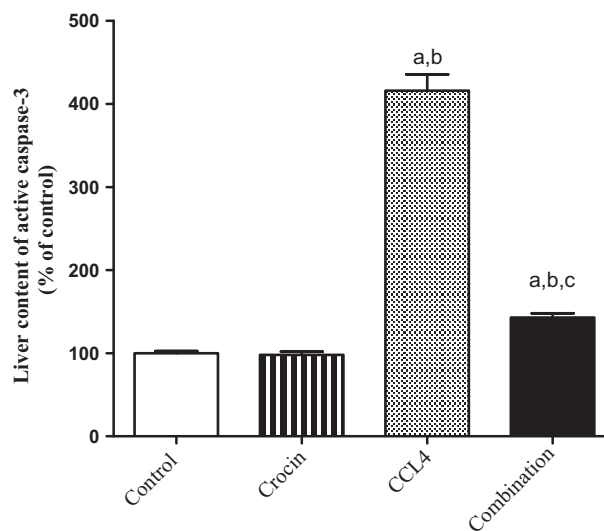
Table 4: Effect of treatment with crocin and or CCL4 on plasma inflammatory cytokines levels.

	Control	Crocin	CCI4	Combination
Plasma IL-6 (pg/ml)	39.7 ± 2.5	42 ± 2.5	110 ± 6.3 ^{a,b}	62 ± 3.6 ^{a,b,c}
Plasma TNF- α (pg/ml)	22 ± 1.4	24 ± 1.5	188 ± 6.6 ^{a,b}	55 ± 2.7 ^{a,b,c}

- Data are presented as mean and standard error of 10 animals each group.
- ^{a, b or c} indicate significant change from control, crocin or CCL4 using Tukey's test as post ANOVA test at $p < 0.01$.
- Animals were treated for 2 weeks.
- IL-6: interleukin 6; TNF- α : Tumor necrosis factor subfamily alpha.

study as a potential protective agent because of its antioxidant activity.²⁵

Per our results, CCI4 induced liver toxicity in rats which was evidenced by: increased liver weight to body weight elevated liver enzymes and total bilirubin, an indication of structural and functional defects in liver cells,^{41,42} along with histological disturbance of liver tissue. Our study indicated that, CCI4-induced liver toxicity are possibly mediated through: 1) generation of free radical and oxidative stress, indicated by: a) abrogation of antioxidant enzymes activity and glutathione content, b) elevation of MDA in liver tissues, 2) induction of phase I metabolizing enzymes leading to more production of CCI4-generated free radicals and concomitant inhibition of phase II metabolizing enzymes leading accumulation of CCI4-generated free radicals, 3) induction of inflammation confirmed by: a) elevation of: IL-6 and TNF- α and b) histopathological investigation, 4) activation of caspase cascade of apoptosis indicated by the reported increase

**Figure 3: Effect of carbon tetrachloride (CCI4), crocin or both crocin and CCI4 on active caspase- 3 activity in rat liver tissue.**

- Animals were treated for 2 weeks.
- Caspase-3 activity values were normalized against tissue protein content and presented as percentage of control value.
- Data are presented as mean and standard error of 10 animals each group.
- ^{a, b or c} indicate significant change from control, crocin or CCL4 using Tukey's test as post ANOVA test at $p < 0.01$.

in activity of active caspase-3 leading to damage of liver cells. These damaging effects could explain the elevated levels of liver enzymes and disturbance of liver functions reported in our results.^{13,42}

Of particular interest, crocin effectively protected against CCl₄-induced hepatotoxicity in rats. These protections are approved via: 1) restoration of relative liver weight to body weight (Table 1), 2) normalization of liver enzymes and serum total bilirubin (Table 2), 3) improvement of liver histological pictures, (Figure 1). According to our study these beneficial effects could be mediated through: 1) inhibition of CCl₄ induced disturbance of metabolizing enzymes (Figure 2) leading to reduction of CCl₄-induced free radical generation, 2) inhibition of lipid peroxidation manifested by decreased MDA content in liver (Table 3), 3) induction of antioxidant enzymes activity and elevation of reduced glutathione content (Table 3). Furthermore, 4) crocin inhibited CCl₄-induced inflammation indicated by abrogation of CCl₄-induced elevation of plasma IL-6 and TNF- α levels (Table 4); 5) inhibition of caspase 3 activity, an effect that protects liver cells from death (Figure 3)^{13,43} and 6) some or all of the above. These findings support our earlier report pointed out the hepatoprotective effect of crocin against bryllium chloride-induced liver injury via antioxidant activity.²⁵

CYP2E1, a phase I metabolizing enzyme, catalyzes the bioactivation of CCl₄ to its highly reactive trichlomethyl peroxy radical (CCl₃OO) and superoxide anion free radicals.^{13,44,45} These radicals initiate lipid peroxidation thereby contributing majorly to the pathogenesis of liver toxicity.^{46–48} In contrast to Phase I, Phase II metabolizing enzymes such as GST is involved in detoxification of CCl₄ and its reactive metabolites to facilitate their elimination from human body.^{46,49} Agents that induce Phase II enzymes have been reported to protect against toxic effects of chemicals in rats.^{50–52}

Therefore, the toxicity of CCl₄ depends, in part, on the balance between the activities of phase I and phase II enzymes. Thus protection could be achieved in part by inhibiting metabolism of CCl₄ and enhancing excretion of toxic metabolites secondary to induction of phase II detoxifying enzymes.⁵³ This principle has been supported by previous results.^{54–57}

Coping with this principle, our study showed significant induction of Phase I metabolizing enzyme (CYP2E1) and inhibition of Phase II metabolizing enzyme (GST) by CCl₄ leading to oxidative stress, confirmed by induction MDA content and abrogation of antioxidant enzymes and glutathione content, and hence liver damage confirmed by H&E staining and manifested by disturbed liver functions.^{46,47,58} Interestingly, treatment with crocin inhibited phase I and stimulated phase II metabolizing enzymes which in turn decreased the level of free radicals generation and abrogated CCl₄ induced oxidative stress which would preserve the integrity of liver cells and hence normalized liver histology, relative liver weight to body weight and liver enzymes.

Furthermore, in addition to modulation of metabolizing enzymes by crocin, the inhibition of CCl₄-induced oxidative stress by crocin in this study could be ascribed also to its antioxidant characters via induction of enzymatic antioxidant activities (SOD, CAT, GSH-PX) and non-enzymatic antioxidant namely GSH which in turn abrogated the lipid

peroxidation as evidenced by diminished MDA level compared to CCl₄-treated group.

Noteworthy to mention that, the amount of carbon tetrachloride metabolized in a given tissue is related to the CYP450 content of the tissue.^{59,60} In the liver, the greatest accumulation of carbon tetrachloride metabolites occurs in the centrilobular region, which has high CYP450 levels.^{44,59} These fact could explain the major histological disturbance in this region as illustrated in histological pictures. Furthermore the metabolic rate of CCl₄ in humans is more similar to the rate in rats than in other rodent species.⁴⁴

Tumor necrosis factor-alpha (TNF- α) and IL-6 are central regulators of inflammation⁶¹ and have been increased in many inflammatory conditions such as chemical toxicity.⁶² The anti-inflammatory effect of crocin has been studied before where crocin showed a suppressive activities on diverse proinflammatory mediators such as NO, IL-1 β , TNF- α , and reactive oxygen species.^{27,63,64} Our results are in line with these reports where crocin suppressed CCl₄-induced elevation of IL-6 and TNF- α , which could contribute for the protective effect of crocin against CCl₄-induced liver damage.

The increased level of caspase 3 in liver tissue collected from animal treated with CCl₄ might be attributed to oxidative stress and induction of inflammation.^{27,65} Caspase 3 plays the pivotal role in apoptosis where it mediates the virtual apoptotic effect.^{66,67} Administration of CCl₄ elevated the caspase 3 in rat liver resulting in liver cell death (Figure 3). Crocin ameliorated CCl₄-induced cell death as observed in the decreased centrilobular necrosis and fatty degenerations in combination group (Figure 1). It is plausible that antioxidant, radical scavenging effects and anti-inflammatory actions of crocin, reported in this study, interfered with CCl₄ induced elevation of caspase 3 activity and hence the liver cells were preserved, liver weight was normalized and functions were restored, this scenario has been supported by our collective results as well as previous results.^{68–70}

Conclusion

In conclusion, our findings revealed that crocin has encouraging protective properties against CCl₄-induced hepatotoxicity. These protective effects attributable to more than one mechanisms namely: modulation of metabolizing enzymes favoring low CCl₄ generated free radical accumulation, free radical scavenging potential, antioxidant activity, anti-inflammatory effect and inhibition of caspase 3 activity.

Conflict of interest

The authors have no conflict of interest to declare.

Funding

The authors disclosed receipt the full financial support and funding from the Deanship of Scientific Research, Taibah University, Almadinah Almunawwarah, Kingdom of Saudi Arabia (Grant number 1672–1433).

Authors' contributions

BS and HMH conceived idea, established design, interpreted data, performed statistical analysis, revised initial draft, performed the major physical work and addressed comments to reviewers.

AH was responsible for in vivo experiment.

GMM performed histopathological examination of liver tissues and its corresponding data analysis.

EHA contributed in evaluation of inflammatory cytokines and biochemical analysis.

BI purchased chemicals and kits and prepared important solutions.

All authors revised the manuscript critically for important intellectual content and approved for its final form.

Acknowledgment

We are deeply thankful to the animal house technician Mr. Islam Farouk, Pharm. B.Sc., department of Pharmacology, Faculty of Pharmacy, King Abdel-Aziz University, Jeddah, K.S.A. for his kind accommodation of laboratory animals and support during animal experiment.

References

- Abraham P, Wilfred G, Cathrine SP. Oxidative damage to the lipids and proteins of the lungs, testis and kidney of rats during carbon tetrachloride intoxication. *Clin Chim Acta* **1999**; 289: 177–179.
- Szymonik-Lesiuk S, Czechowska G, Stryjecka-Zimmer M, Slomka M, Madro A, Celinski K, et al. Catalase, superoxide dismutase, and glutathione peroxidase activities in various rat tissues after carbon tetrachloride intoxication. *J Hepatobiliary Pancreat Surg* **2003**; 10: 309–315.
- Güven A, Güven A, Gulmez M. The effect of kefir on the activities of GSHPx, GST, CAT, GSH and LPO levels in carbon tetrachloride-induced mice tissues. *J Vet Med B Infect Dis Vet Public Health* **2003**; 50: 412–416.
- Lewis Jr RJ. *Hawley's condensed chemical dictionary*. 12th ed. New York: Van Nostrand Reinhold; 1993. pp. 221–222.
- Kauppinen T, Toikkanen J, Pedersen D, Young R, Kogevinas M, Ahrens W, et al. Occupational exposure to carcinogens in the European Union. *Occup Environ Med* **2000**; 57(1): 10–18.
- United States International Trade Commission. *Synthetic organic chemicals: US production and sales*. 1991 (USITC Publ. 2607). Washington DC: United States Government Printing Office; 1993. pp. 3–85.
- Kauppinen T, Pukkala E, Saalo A, Sasco AJ. Exposure to chemical carcinogens and risk of cancer among Finnish laboratory workers. *Am J Ind Med* **2003**; 44: 343–350.
- Tomenson JA, Baron CE, O'Sullivan JJ, Edwards JC, Stonard MD, Walker RJ, et al. Hepatic function in workers occupationally exposed to carbon tetrachloride. *Occup Environ Med* **1995**; 52: 508–514.
- The International Agency for Research on Cancer (IARC). *IARC monographs on the evaluation of the carcinogenic risk of chemicals to humans* In **Some halogenated hydrocarbons**, vol. 20; 1979. pp. 371–399. Lyon.
- Agency for Toxic Substances and Disease Registry (ATSDR). *Toxicological profile for carbon tetrachloride*. Atlanta, US: US Department of Health and Human Services; 2005.
- International Programme on Chemical Safety (IPCS). *Carbon tetrachloride. Environmental health criteria 208*. Geneva: WHO; 1999.
- Miyazaki T, Karube M, Matsuzaki Y, Ikegami T, Doy M, Tanaka N, et al. Taurine inhibits oxidative damage and prevents fibrosis in carbon tetrachloride-induced hepatic fibrosis. *J Hepatol* **2005**; 43: 117–12515.
- Hassan MH, Edfawy M, Mansour A, Hamed AA. Antioxidant and antiapoptotic effects of capsaicin against carbon tetrachloride-induced hepatotoxicity in rats. *Toxicol Ind Health* **2012**; 28(5): 428–438.
- Slater TF. Free-radical mechanism in tissue injury. *Biochem J* **1984**; 222: 1–15.
- McCay PB, Lai EK, Poyer JL, DuBose CM, Janzen EG. Oxygen- and carbon-centered free radical formation during carbon tetrachloride metabolism. Observation of lipid radicals in vivo and in vitro. *J Biol Chem* **1984**; 259: 2135–2143.
- Hamadi N, Mansour A, Hassan MH, Khalifi-Touhami F, Badary O. Ameliorative effects of resveratrol on liver injury in streptozotocin-induced diabetic rats. *J Biochem Mol Toxicol* **2012**; 26(10): 384–392.
- Soylu AR, Aydogdu N, Basaran UN, Altaner S, Tarcin O, Gedik N, et al. Antioxidants vitamin E and C attenuate hepatic fibrosis in biliary-obstructed rats. *World J Gastroenterol* **2006**; 12: 6835–6841.
- Konoshima T, Takasaki M, Tokuda H, Morimoto S, Tanaka H, Kawata E, et al. Crocin and crocetin derivatives inhibit skin tumor promotion in mice. *Phytother Res* **1998**; 12: 400–404.
- Abe K, Saito H. Effects of saffron extract and its constituent crocin on learning behavior and long-term potentiation. *Phytother Res* **2000**; 14: 149–152.
- Lee IAH, Lee JH, Baek NI, Kim DH. Antihyperlipidemic effect of crocin isolated from the fructus of *Gardenia jasminoides* and its metabolite crocetin. *Biol Pharm Bull* **2005**; 28(11): 2106–2110.
- He SHY, Qian ZY, Tang FT, Wen N, Xu GL, Sheng L. Effects of crocin on experimental atherosclerosis in quails and its mechanisms. *Life Sci* **2005**; 77(8): 907–921.
- Bakshi H, Sam S, Feroz A, Ravesh Z, Shah GA, Sharma M. Crocin from Kashmiri saffron (*Crocus sativus*) induces in vitro and in vivo xenograft growth inhibition of Dalton's lymphoma. *Asian Pac J Cancer Prev* **2009**; 10(5): 887–890.
- Bakshi H, Sam S, Rozati R, Sultan P, Islam T, Rathore B, et al. DNA fragmentation and cell cycle arrest: a hallmark of apoptosis induced by crocin from Kashmiri saffron in a human pancreatic cancer cell line. *Asian Pac J Cancer Prev* **2010**; 11(3): 675–679.
- Asdaq SM, Inamdar MN. Potential of *Crocus sativus* (saffron) and its constituent, crocin, as hypolipidemic and antioxidant in rats. *Appl Biochem Biotechnol* **2010**; 162(2): 358–372.
- El-Beshbishy HA, Hassan MH, Aly HA, Doghish AS, Alghaithy AA. Crocin "saffron" protects against beryllium chloride toxicity in rats through diminution of oxidative stress and enhancing gene expression of antioxidant enzymes. *Ecotoxicol Environ Saf* **2012**; 83: 47–54.
- Lin JK, Wang CJ. Protection of crocin dyes on the acute hepatic damage induced by aflatoxin B1 and dimethylnitrosamine in rats. *Carcinogenesis* **1986**; 7(4): 595–599.
- Hariri AT, Moallem SA, Mahmoudi M, Memar B, Hosseinzadeh H. Sub-acute effects of diazinon on biochemical indices and specific biomarkers in rats: protective effects of crocin and safranal. *Food Chem Toxicol* **2010**; 48(10): 2803–2808.
- Tasci I, Mas MR, Vural SA, Deveci S, Comert B, Alcigir G, et al. Pegylated interferon-alpha plus taurine in treatment of rat liver fibrosis. *World J Gastroenterol* **2007**; 13(23): 3237–3244.

29. Mas N, Tasci I, Comert B, Ocal R, Mas MR. Ursodeoxycholic acid treatment improves hepatocyte ultrastructure in rat liver fibrosis. **World J Gastroenterol** 2008; 14(7): 1108–1111.
30. Reitman S, Frankel S. A colorimetric method for the determination of serum glutamic oxaloacetic and glutamic pyruvic transaminases. **Am J Clin Pathol** 1957; 28(1): 56–63.
31. Bancroft J, Stevens A. *Enzyme histochemistry: theory and practice of histological techniques*. New York: Churchill Livingstone; 1996.
32. Benson AM, Hunkeler MJ, Talalay P. Increase of NADPH, quinone reductase activity by dietary antioxidant: possible role in protection against carcinogenesis and toxicity. **Proc Natl Acad Sci USA** 1980; 77: 5216–5220.
33. Chang TK, Crespi CL, Waxman DJ. Spectrophotometric analysis of human CYP2E1-catalyzed p-nitrophenol hydroxylation. **Methods Mol Biol** 1998; 107: 147–152.
34. Habig WH, Pabst MJ, Jacob WB. Glutathione-S-transferase. The first enzymatic step in mercapturic acid formation. **J Biol Chem** 1974; 249: 7130e9.
35. Ohkawa H, Ohishi N, Yagi K. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. **Anal Biochem** 1979; 95(2): 351–358.
36. Marklund S, Marklund G. Involvement of superoxide anion radical in the autooxidation of pyrogallol and a convenient assay for superoxide dismutase. **Eur J Biochem** 1974; 47: 469–744.
37. Sinha AK. Colorimetric assay of catalase. **Anal Biochem** 1972; 47(2): 389–394.
38. Moron MS, Despierre JW, Minnervik B. Levels of glutathione, glutathione reductase and glutathione-S-transferase activities in rat lung and liver. **Biochim Biophys Acta** 1979; 582: 67–78.
39. Al-Hendy A, Lee EJ, Wang HQ, Copland JA. Gene therapy of uterine leiomyomas: adenovirus-mediated expression of dominant negative estrogen receptor inhibits tumor growth in nude mice. **Am J Obstet Gynecol** 2004; 191: 1621–1631.
40. Dahiru D, Mamman DN, Wakawa HY. Ziziphus mauritiana fruit extract inhibits CCl₄-induced hepatotoxicity in male rats. **Pak J Nutr** 2010; 9: 990–993.
41. Recknagel RO, Glende Jr EA, Dolak JA, Waller RL. Mechanisms of CCl₄ toxicity. **Pharmacol Ther** 1989; 43: 139–154.
42. Kalender S, Ogutcu A, Uzunhisarcikli M, Açikgoz F, Durak D, Ulusoy Y, et al. Diazinon-induced hepatotoxicity and protective effect of vitamin E on some biochemical indices and ultrastructural changes. **Toxicol** 2005; 211(3): 197–206.
43. Edfawy M, Hassan MH, Mansour A, Hamed AA, Amin HA. Meloxicam modulates oxidative stress status, inhibits prostaglandin E₂, and abrogates apoptosis in carbon tetrachloride-induced rat hepatic injury. **Int J Toxicol** 2012; 31(3): 276–286.
44. Zangar RC, Benson JM, Burnett Springer DL. Cytochrome P450 2E1 is the primary enzyme responsible for low-dose carbon tetrachloride metabolism in human liver microsomes. **Chem Biol Interact** 2000; 125(3): 233–243.
45. Brent JA, Rumack BH. Role of free radicals in toxic hepatic injury II. Are free radicals the cause of toxin-induced liver injury? **J Toxicol Clin Toxicol** 1993; 31: 173–196.
46. Ohnuma T, Anan E, Hoashi R, Takeda Y, Nishiyama T, Ogura K, et al. Dietary diacetylene falcarindiol induces phase 2 drug-metabolizing enzymes and blocks carbon tetrachloride-induced hepatotoxicity in mice through suppression of lipid peroxidation. **Biol Pharm Bull** 2011; 34(3): 371–378.
47. Hwang YP, Choi CY, Chung YC, Jeon SS, Jeong HG. Protective effects of puerarin on carbon tetrachloride-induced hepatotoxicity. **Arch Pharm Res** 2007; 30(10): 1309–1317.
48. Wong FW, Chan W, Lee SS. Resistance to carbon tetrachloride-induced hepatotoxicity in mice which lack CYP2E1 expression. **Toxicol Appl Pharmacol** 1998; 153: 109–118.
49. Ohnuma T, Nakayama S, Anan E, Nishiyama T, Ogura K, Hiratsuka A. Activation of the Nrf2/ARE pathway via S-alkylation of cysteine 151 in the chemopreventive agent-sensor Keap1 protein by falcarindiol, a conjugated diacetylene compound. **Toxicol Appl Pharmacol** 2010; 244(1): 27–36.
50. Dinkova-Kostova AT, Jenkins SN, Fahey JW, Ye L, Wehage SL, Liby KT, et al. Protection against UV-light-induced skin carcinogenesis in SKH-1 high-risk mice by sulforaphane-containing broccoli sprout extracts. **Cancer Lett** 2006; 240: 243–252.
51. Fahey JW, Haristoy X, Dolan PM, Kensler TW, Scholtus L, Stephenson KK, et al. Sulforaphane inhibits extracellular, intracellular, and antibiotic-resistant strains of *Helicobacter pylori* and prevents benzo[a]pyrene-induced stomach tumors. **Proc Natl Acad Sci USA** 2002; 99(11): 7610–7615.
52. Chung FL, Conaway CC, Rao CV, Reddy BS. Chemoprevention of colonic aberrant crypt foci in Fischer rats by sulforaphane and phenethyl isothiocyanate. **Carcinogenesis** 2000; 21(12): 2287–2291.
53. Presterla T, Zhang Y, Spencer SR, Wilczak CA, Talalay P. The electrophile counterattack response: protection against neoplasia and toxicity. **Adv Enzyme Regul** 1993; 33: 281–296.
54. Seidegård J, Pero RW, Markowitz MM, Roush G, Miller DG, Beattie EJ. Isozyme (s) of glutathione transferase (class m) as a marker for the susceptibility to lung cancer: a follow up study. **Carcinogenesis** 1990; 11(1): 33–36.
55. Hu Z, Wells PG. In vitro and in vivo biotransformation and covalent binding of benzo(a)pyrene in Gunn and Rha rats with a genetic deficiency in bilirubin uridine diphosphate UDP-glucuronosyltransferase. **J Pharmacol Exp Ther** 1992; 263: 334–342.
56. Lafuente A, Pujol F, Carretero P, Villa JP, Cuchi A. Human glutathione S-transferase m (GST m) deficiency as a marker for the susceptibility to bladder and larynx cancer among smokers. **Cancer Lett** 1993; 68: 49–54.
57. Kahl R. Synthetic antioxidants: biochemical actions and interference with radiation toxic compounds, chemical mutagens and chemical carcinogens. **Toxicol** 1984; 33: 185–228.
58. Halliwell B, Gutteridge JM. Role of free radicals and catalytic metal ions in human disease: an overview. **Methods Enzymol** 1990; 186: 1–85.
59. Bergman I, Loxley R. Two improved and simplified methods for the spectrophotometric determination of hydroxyproline. **Anal Chem** 1963; 35: 1961–1965.
60. Villarruel M, de Toranzo EGD, Castro JA. Carbon tetrachloride activation, lipid peroxidation and the mixed function oxygenase activity of various rat tissues. **Toxicol Appl Pharmacol** 1977; 41: 337–344.
61. Esposito E, Cuzzocrea S. Superoxide, NO, peroxynitrite and PARP in circulatory shock and inflammation. **Front Biosci (Landmark Ed)** 2009; 14: 263–296.
62. Ouyang W, Beckett O, Flavell RA, Li MO. An essential role of the forkhead-box transcription factor foxo1 in control of T cell homeostasis and tolerance. **Immunity** 2009; 30(3): 358–371.
63. Nam KN, Park YM, Jung HJ, Lee JY, Min BD, Park SU, et al. Anti-inflammatory effects of crocin and crocetin in rat brain microglial cells. **Eur J Pharmacol** 2010; 648(1–3): 110–116.
64. Yang R, Tan X, Thomas AM, Shen J, Qureshi N, Morrison DC, et al. Crocetin inhibits mRNA expression for tumor necrosis factor- α , interleukin-1 β , and inducible nitric oxide synthase in hemorrhagic shock. **JPEN J Parenter Enter Nutr** 2006; 30(4): 297–301.
65. Soeda S, Ochiai T, Paopong L, Tanaka H, Shoyama Y, Shimeno H. Crocin suppresses tumor necrosis factor- α -induced cell death of neuronally differentiated PC-12 cells. **Life Sci** 2001; 69(24): 2887–2898.
66. Ding WX, Ong NC. Role of oxidative stress and mitochondrial changes in cyanobacteria induced apoptosis and hepatotoxicity. **FEMS Microbiol Lett** 2003; 220: 1–7.

67. Shi J, Aisaki K, Ikawa Y, Wake K. Evidence of hepatocyte apoptosis in rat liver after the administration of carbon tetrachloride. *Am J Pathol* **1998**; 153(2): 515–525.
68. Dianat M, Esmailizadeh M, Badavi M, Samarbaf-Zadeh AR, Naghizadeh B. Protective effects of crocin on ischemia-reperfusion induced oxidative stress in comparison with vitamin E in isolated rat hearts. *J Nat Pharm Prod* **2014**; 9(2): e17187.
69. Sun Y, Yang J, Wang LZ, Sun LR, Dong Q. Crocin attenuates cisplatin-induced liver injury in the mice. *Hum Exp Toxicol* **2013**; 33(8): 855–862.
70. Tamaddonfard E, Farshid AA, Eghdami K, Samadi F, Erfanparast A. Comparison of the effects of crocin, safranal and diclofenac on local inflammation and inflammatory pain responses induced by carrageenan in rats. *Pharmacol Rep* **2013**; 65(5): 1272–1280.