Journal of Molecular and Cellular Cardiology 83 (2015) 112–121

Contents lists available at ScienceDirect



Journal of Molecular and Cellular Cardiology

journal homepage: www.elsevier.com/locate/yjmcc



CrossMark

Review article Vascular biology of ageing—Implications in hypertension

Adam Harvey, Augusto C. Montezano, Rhian M. Touyz *

Institute of Cardiovascular and Medical Sciences, BHF Glasgow Cardiovascular Research Centre, University of Glasgow, UK

ARTICLE INFO

Article history: Received 20 January 2015 Received in revised form 30 March 2015 Accepted 9 April 2015 Available online 17 April 2015

Keywords: Vascular remodeling Endothelial dysfunction Oxidative stress Mitochondria

ABSTRACT

Ageing is associated with functional, structural and mechanical changes in arteries that closely resemble the vascular alterations in hypertension. Characteristic features of large and small arteries that occur with ageing and during the development of hypertension include endothelial dysfunction, vascular remodelling, inflammation, calcification and increased stiffness. Arterial changes in young hypertensive patients mimic those in old normotensive individuals. Hypertension accelerates and augments age-related vascular remodelling and dysfunction, and ageing may impact on the severity of vascular damage in hypertension, indicating close interactions between biological ageing and blood pressure elevation. Molecular and cellular mechanisms underlying vascular alterations in ageing and hypertension are common and include aberrant signal transduction, oxidative stress and accular changes could ameliorate vascular damage associated with hypertension. An overview on the vascular biology of ageing and hypertension is presented and novel molecular mechanisms contributing to these processes are discussed. The complex interaction between biological ageing and blood pressure elevation on the vasculature is highlighted. This article is part of a Special Issue entitled: CV Ageing.

© 2015 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/).

Contents

1	La ford d	11	2
1.		uction	
2.	Structu	ural and mechanical changes in the ageing vasculature	3
3.	Vascul	lar calcification \ldots	4
4.		g associated vascular inflammation	
5.	0 0		
		lar contractility and ageing	
6.	Endotl	helial function and ageing \ldots \ldots \ldots 14	4
7.	Vascul	lar signalling in ageing \ldots \ldots \ldots 14	4
	7.1.	Sirtuins	4
	7.2.	ΡGC-1α	5
	7.3.	FoxO transcription factors	5
	7.4.	p66shc	5
	7.5.	Cell cycle regulators, senescence and autophagy	6
	7.6.	Mitogen-activated protein kinases (MAPK)	6
	7.7.	Oxidative stress in vascular ageing	6
	7.8.	Endoplasmic reticulum stress in vascular ageing 110	6
	7.9.	Vascular changes in hypertension recapitulate those in ageing	7
	7.10.	Effects of pro-hypertensive stimuli on vascular ageing: the renin-angiotensin-aldosterone system (RAAS)	7
	7.11.	Effects of novel anti-hypertensive factors on vascular ageing	8
	7.12.	Vascular damage in hypertension may be independent of ageing	8
	7.13.	Lessons learned from children with hypertension	8
8.	Summ	nary and conclusions	9
0.	Summ		Э

E-mail address: Rhian.Touyz@glasgow.ac.uk (R.M. Touyz).

http://dx.doi.org/10.1016/j.yjmcc.2015.04.011

0022-2828/© 2015 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/).

^{*} Corresponding author at: Institute of Cardiovascular and Medical Sciences, BHF Glasgow Cardiovascular Research Centre, University of Glasgow, 126 University Place, Glasgow, G12 8TA, UK. Tel.: + 44 141 330 7775/7774; fax: + 44 141 330 3360.

Disclosures 119 Acknowledgements 119 References 119

1. Introduction

Clinical studies show a significant relationship between ageing and increased blood pressure, with advancing age being a major non-modifiable risk factor in the development of hypertension [1]. This is due, in part, to changes that occur in the vasculature, including endothe-lial dysfunction, vascular remodelling, increased vascular stiffness and inflammation. These functional and structural changes define the 'vascular phenotype' of hypertension, features that are also found during ageing [2] (Fig. 1). At the cellular level, there is endothelial cell damage, increased vascular smooth muscle cell growth, cell migration, inflammation, contraction, extracellular matrix deposition, fibrosis, and calcification [3].

Young patients with elevated blood pressure exhibit arterial changes similar to those in older individuals with normal blood pressure, and accordingly the concept of 'premature' or 'early' vascular ageing in hypertension has been proposed [4]. Hypertension accelerates agerelated vascular changes, processes that are attenuated when blood pressure is normalised. The direct relationship between ageing and vascular health is evident in progeria syndrome, where patients exhibit accelerated ageing, endothelial dysfunction, accelerated atherosclerosis and die prematurely from complications of cardiovascular disease, such as stroke and myocardial infarction [5]. Considering the fact that the population is ageing and that the major chronic disease of ageing is hypertension and associated cardiovascular complications, the potential health and economic burden in our modern society is enormous. Accordingly it is important to understand how vascular function changes with ageing and how this impacts on hypertension, so that targeted strategies could be developed to prevent and repair damaged 'aged' arteries and thereby reduce the risk of hypertension and target organ damage. In the present review, we discuss the vascular changes that occur with ageing and during the development of hypertension and focus on some molecular mechanisms that underlie these vascular changes.

2. Structural and mechanical changes in the ageing vasculature

Physiological changes to the vascular wall are dynamic and occur throughout life [6,7]. Endothelial cell turnover occurs over years, whereas that of vascular smooth muscle cells seems to occur over a shorter time period. Many structural and mechanical alterations have been observed in the aged vasculature including increased intimal-to-media (IM) thickness, evidenced by the finding that the IM thickness of the carotid artery increases two- to three-fold between 20 and 90 years of age [8,9]. Subclinical IM thickening is strongly associated with ageing and is also a predictor of future cardiovascular events [8,9]. Both aortic length and circumference gradually increase with advancing age [10–12]. Associated with these structural alterations are mechanical changes, characterised by a reduction in compliance, reduced elasticity/ distensibility and increased stiffness [8,9]. Stiffening of the large conduit arteries due to fracture of elastin fibres within the tunica media and collagenous remodelling, results in increased aortic pulse pressure and pulse wave velocity (PWV). Increased PWV, a non-invasive measure of vascular stiffness, increases in both sexes with ageing and is determined by the mean arterial pressure and the intrinsic stress/strain relationship (stiffness) of the arterial wall. As arterial wall stiffness increases, central systolic pressure increases and diastolic pressure decreases, leading to increased pulse pressure, an independent risk factor for future cardiovascular events [13]. Processes underlying these structural and mechanical changes involve growth and migration of vascular smooth muscle cells

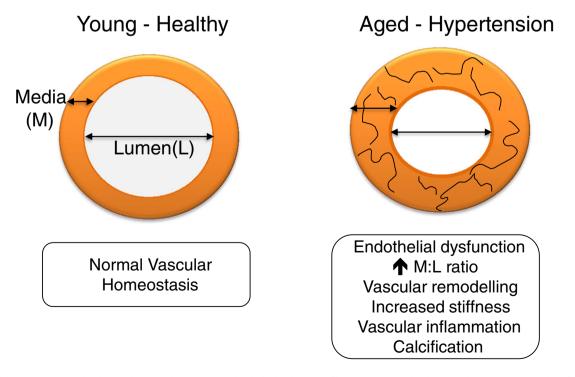


Fig. 1. Schematic demonstrating vascular changes that occur during ageing and with the development of hypertension. Vascular changes in hypertension mimic those found in arteries observed with ageing.

within the media, vascular calcification and changes in the ratio of collagen and elastin in the vascular wall. Physiologically, the ratio of collagen and elastin remains constant due to their gradual production and degradation. In aged rodents the absolute elastin content of the aorta was shown not to differ to young counterparts. However, increased collagen content in 30-month-old animals compared to 6-month-old animals meant that the relative elastin content was decreased [14]. Collagen and elastin are regulated by catabolic matrix metalloproteinases (MMPs). Throughout ageing the balance between MMPs and their inhibitors (TIMPs) changes. For example, increased MMP-2 expression and activity in the vessels of old rats and non-human primates is increased compared to young counterparts [15,16].

3. Vascular calcification

Vascular calcification is a tightly controlled process similar to bone formation, where mineralization of the internal elastic lamina and elastic fibres in the media results in vascular stiffening. Calcification of the vascular media is a hallmark of vascular ageing [17]. Upregulation of transcription factors such as cbfa1 (core-binding factor 1α)/Runx2, MSX-2 and bone morphogenetic protein 2 (BMP-2), are involved in normal bone development and vascular calcification by regulating the expression of osteogenic proteins, including osteocalcin, osteonectin, alkaline phosphatase, collagen-1, and bone sialoprotein [18,19]. Another mechanism contributing to vascular mineralization is loss of calcification inhibitors, such as fetuin-A, matrix Gla protein, pyrophosphate, and osteopontin [19-21]. Molecular processes underlying this remain to be fully defined but increased expression of BMP2 and the osteoblast transcription factor Runx2/Cbfa1 [22], and modulation of Ca²⁺ and Mg²⁺ transport through cation channels, such as TRPM7 may represent important mediators in this process [23,24]. A correlation between age and vascular calcification has been described from 5% in individuals younger than 50 years to > 12% in individuals older than 80 years [25]. Ageing-associated vascular calcification has been reported in the aorta of rodents where the associated mechanisms include dysregulation of Matrix Gla protein [26]. Further possible mechanisms contributing to increased calcification with ageing includes dysregulation of vascular pyrophosphate [27,28]. Human studies have shown a weak inverse correlation between age and plasma pyrophosphate [29,30].

4. Ageing associated vascular inflammation

With ageing there is a shift towards a proinflammatory vascular phenotype with upregulation of inflammatory cytokines, chemokines and adhesion molecules in the vascular wall [8,9,31-38]. Proinflammatory transcription factors and proteins that have been identified in the ageing vascular media include MCP-1, TGF- β 1, MMP-2, AP-1 and NF-kB [8,9,39]. Expression and activation of these molecules increases with ageing, processes that are usually associated with increased generation of reactive oxygen species (ROS). In aged arteries, there is downregulation of the transcription factor, nuclear factor (erythroid-derived 2)-like 2 (Nrf2), which stimulates expression of antioxidant enzymes, thereby leading to decreased anti-oxidant potential and increased ROS bioavailability with consequent oxidative stress [40]. Oxidative stress is a potent inducer of redox-sensitive proinflammatory signalling pathways, further contributing to inflammation and vascular damage with ageing [36-40].

5. Vascular contractility and ageing

Functionally, vascular contraction is altered during ageing and is determined in large part by changes in vascular smooth muscle cell cytoskeletal organisation and impaired contractile signalling. Mesenteric arteries from aged rats demonstrate hypercontractility in response to phenylephrine compared to young controls [40] an effect which is mirrored in the aorta [41]. These findings are paralleled in studies utilising the senescence-accelerated mouse (SAM-P8), which demonstrate increased vascular contractility in response to phenylephrine [42]. Conversely, studies performed on carotid vessels from aged guinea pigs displayed reduced contractile response to both phenylephrine and endothelin-1 (ET-1) compared to younger controls [43]. Thus it appears that differential responses during ageing may differ between species.

At the cellular level, with ageing, vascular smooth muscle cells, which are normally contractile, undergo phenotypic changes to become stiff and pro-migratory. Subsets of apoptotic, senescent and proliferative cells as well as hyper-contractile cells may co-exist in the vascular media. A major trigger for these functional changes is an increase in intracellular free Ca^{2+} concentration ($[Ca^{2+}]_i$), which occurs following activation of phopholipase C (PLC) leading to the generation of second messengers insitol trisphosphate (IP3) and diacylglycerol (DAG) [39,44]. Ca²⁺ binds to calmodulin facilitating an interaction with myosin light chain kinase (MLCK) leading to its activation. Activated MLCK triggers phosphorylation of the regulatory light chains of myosin (MLC20) promoting cycling of myosin cross-bridges with actin and consequent contraction. Dephosphorylation of MLC20 by myosin light chain phosphatase (MLCP) results in VSMC relaxation. As such, the relative activities of MLCK and MLCP determine vascular smooth muscle tone by influencing the degree of MLC20 phosphorylation. Arteries from aged animals display altered responses to various contractile agents including norepinephrine, serotonin and KCL [44,45]. Mechanisms for this are incompletely understood, but the percentage of phosphorylated MLC20 induced by vasoactive agonists is different in young versus aged rats and may play a role in altered age-related contractile responses [45].

6. Endothelial function and ageing

The vascular endothelium is a monolayer of cells that lines blood vessels and plays a key role in arterial function through synthesis and release of biologically active molecules that can influence endothelial function in an autocrine or paracrine fashion. The healthy endothelium is characterised by a vasodilatory, anti-inflammatory and antithrombotic phenotype. Endothelial dysfunction is characterised by reduced vasodilatory responses to flow or agonists and is proinflammatory. Independent of the occurrence of other pathologies, ageing results in altered endothelium-dependent relaxation of both the aorta and resistance arteries in rodents [46,47]. These findings have been corroborated in human studies that suggest that endothelial function is gradually compromised with ageing [48,49]. A primary mechanism responsible for the deterioration of endothelial function with ageing is thought to be reduced bioavailability of the endothelium derived relaxing factor, nitric oxide (NO), due to its interaction with ROS to form peroxynitrite. Peroxynitrite oxidises BH4, an essential cofactor for NO synthesis by endothelial nitric oxide synthase (eNOS), to its inactive form resulting in reduced NO production. Furthermore, reduced BH4 can result in eNOS uncoupling whereby superoxide is produced in preference to NO. Reductions in BH4 levels have been reported in aged rodents [50].

7. Vascular signalling in ageing

Molecular mechanisms and cell signalling events underlying the structural and functional alterations observed during ageing are similar to those that occur in hypertension (Fig. 2). Many age/longevity-related molecules and signalling cascades have been described, of which a few of the novel systems are highlighted below.

7.1. Sirtuins

Sirtuins (SIRTs) are a family of NAD-dependent protein deacetylases and ribosyl transferases consisting of 7 members which are localised in the cytoplasm (SIRT1 and SIRT2), nucleus (SIRT1, SIRT2, SIRT 6 and SIRT 7) or mitochondria (SIRT3, SIRT4 and SIRT 5). SIRTs have been

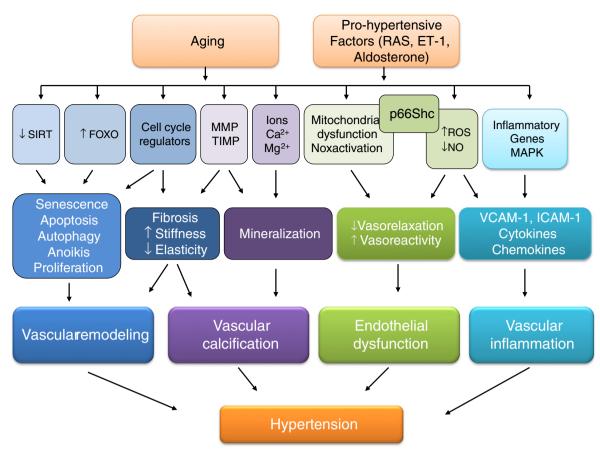


Fig. 2. Molecular and cellular mechanisms associated with vascular changes in ageing and hypertension. Activation of pro-fibrotic, pro-inflammatory, redox-sensitive and growth/apoptotic signalling pathways lead to changes in vascular structure, mechanics and function with resultant arterial remodelling, calcification, inflammation, stiffness and impaired vasoreactivity. These vascular alterations are common features during ageing and in hypertension. VCAM-1, vascular cell adhesion molecule-1; ICAM-1, intercellular adhesion molecule-1; MMP, matrix metalloproteinase; TIMP, tissue inhibitor of metalloproteinase; RAS, renin angiotensin system; ET-1, endothelin-1; NO, nitric oxide.

implicated in various cellular processes associated with ageing, including, apoptosis, inflammation and mitochondrial biogenesis. SIRTs are able to modulate the ageing process in a number of species [51–53]. This is highlighted by the following: 1) SIRT1 protects against phosphate-induced arterial calcification, possibly due to the inhibition of osteoblastic transdifferentiation [54]; 2) mitochondrial localised SIRT3 regulates many proteins that are important in the regulation of mitochondrial function including pyruvate dehydrogenase, SOD2 and cyclophilin D; 3) SIRT3 –/– mice exhibit accelerated cardiovascular ageing [55] and 4) SIRT3 has vasoprotective effects through interaction with FOXO3, which enhances mitochondrial antioxidant defence systems [56].

7.2. PGC-1α

Another emerging candidate implicated in age-related signalling in the vasculature, is peroxisome proliferator-activated receptor gamma coactivator-1 α (PGC-1 α), which plays an important role in regulating mitochondrial biogenesis and turnover [57]. Because mitochondria require continuous recycling and regeneration throughout the lifespan and are subject to continuous damage over time, regulation of mitochondrial biogenesis and turnover is critical for maintained energy production and prevention of oxidative damage, and the promotion of healthy ageing. Impaired mitochondrial biogenesis is an important inducer of age-related changes in the endothelium and vascular smooth muscle [58–60]. The aged vasculature displays reduced levels of PGC-1 α with consequent mitochondrial proteins leading to oxidative stress and vascular injury [60]. Decreased AMPK activity may contribute to reduced PGC-1 α activation and impaired mitochondrial function associated with ageing [61].

7.3. FoxO transcription factors

The FoxO family of Forkhead transcription factors are involved in tumour suppression, energy metabolism, and longevity. Mammals express four FoxO isoforms, FoxO1, FoxO3, FoxO4 and FoxO6. FoxO1, FoxO3 and FoxO4 are phosphorylated in an Akt-dependent manner that promotes FoxO export from the nucleus to the cytoplasm, thereby repressing FoxO transcriptional function. FoxO targets include genes that have pivotal roles in cell cycle progression (p21, p27) and ROS detoxification (MnSOD) and thus may be important in regulation of the ageing phenotype in the vasculature [62,63]. FoxO3 is a direct target of SIRT3 deacetylation protecting mitochondria against age-related oxidative stress and promoting upregulation of genes that are essential for mitochondrial homeostasis [64]. Several reports have suggested that FoxO3 may be a determinant of ageing, due to the fact that single-nucleotide polymorphisms in the FoxO3 gene are associated with longevity in humans [65,66]. FoxO3 knockout mice however do not exhibit reduced lifespan [67], and as such, the exact role of FoxO3 in longevity and ageing still remains unclear.

7.4. p66shc

Mitochondrial dysfunction and increased mitochondrial-derived ROS have been implicated in vascular changes in ageing [68]. An important mediator of mitochondrial ROS production and thus regulator of the intracellular pathways that govern oxidative stress, apoptosis, and cell growth/survival is the adapter protein p66shc. p66Shc is phosphorylated at serine 36 by PKC β and VEGF, resulting in recognition by the prolyl isomerase Pin1, allowing translocation and entrance into mitochondria where it interacts with cytochrome C resulting in production of H₂O₂. Levels of p66shc in heart, kidney and vascular smooth muscle increase with ageing [69]. Mice lacking p66shc gene display a 30% increase in lifespan compared to wild-type controls due to prevention of oxidative stress and improved endothelial function [70,71].

7.5. Cell cycle regulators, senescence and autophagy

In culture, vascular cells respond to prolonged series of replication and stresses by eventually entering an irreversible growth arrest or senescent state [72]. After the Hayflick limit, cells enter an irreversible cell cycle arrest in the G1 phase of the cell cycle and no longer respond to growth stimuli. This phenomenon is called replicative senescence and occurs in vascular ageing [73]. Senescent cells have a distinct phenotype—they are large and flattened, express specific markers (β -galactosidase), overexpress cell cycle molecular markers (p16 and p21), form heterochromatic foci (yH2AX) and accumulate lipofuscin, a non-degradable fluorescent compound [74]. Whilst the molecular mechanisms underlying cellular senescence have been the focus of numerous studies, the impact of senescence in vivo has yet to be fully established, especially since some studies show increased rates of vascular cell proliferation in ageing and longevity [75,76].

Considering the remarkable plasticity of vascular smooth muscle cells, there is a requirement for tight control of transcriptional, metabolic and ultrastructural processes, events that are coordinated through autophagy. Autophagy is the basic cellular mechanism that involves cell degradation of unnecessary or dysfunctional molecules through lysosomes [77]. In the vasculature, changes in autophagy have been observed in experimental ageing [78].

7.6. Mitogen-activated protein kinases (MAPK)

Protein kinases are major regulators of signal transduction that catalyse the phosphorylation of other proteins, thus regulating their activity. Primary targets of protein kinases include transcription factors which modulate intracellular signalling via specific alteration of downstream gene expression/activity [79]. A key group of protein kinases in the vasculature are the serine/threonine sub-family, which act by promoting phosphorylation of the OH group of serine or threonine residues on target proteins [80]. Mitogen activated protein kinases (MAPKs) represent a large family of proteins important in signal transduction within the cardiovascular system, where they are involved in regulation of a number of biological processes, such as cell migration, survival, apoptosis, proliferation, contraction and differentiation. MAPK signalling is promoted by many stimuli including GPCR activation, receptor tyrosine kinases, oxidative stress and growth factors, and comprises a number of sequentially acting kinases which ultimately result in phosphorylation and activation of terminal effector kinases, thereby transducing specific cellular actions [80,81]. Several MAPK family subgroups have been identified, of which the major mammalian types appear to be ERK1/2, c-Jun NH2-terminal kinases (JNK1, 2 and 3) and p38MAPK (α , β , δ and γ), which play key roles during cardiovascular development and vascular function [82,83]. Several studies have demonstrated an age-dependent increase in MAPK activation in vascular tissue [84,85].

7.7. Oxidative stress in vascular ageing

Common to many of the molecular and cellular processes described above that underlie changes in the vasculature with ageing is oxidative stress [86]. The concept that ROS are linked to ageing was suggested in 1956 by Harman when he proposed the Free Radical Theory of Ageing, stating that the accumulation of free radicals during ageing causes the damage of biomolecules by these ROS and the development of pathological disorders promoting cell senescence and organism ageing [87,88]. Such processes are evident in vessels associated with ageing and with hypertension [87,88]. Excessive production of ROS and reactive nitrogen species (RNS) leads to oxidative modification of proteins, DNA and lipids, which accumulate in cells leading to impaired cellular and vascular function. In addition increased vascular ROS levels, together with decreased eNOS-generated NO, compromise the vasodilatory actions of NO and promote the formation of injurious peroxynitrite, processes observed in aorta of aged rodents [89]. Oxidative stress is critically involved in many of the molecular events of vascular ageing, including: (1) increased pro-inflammatory responses in vascular cells, (2) vascular dysfunction through oxidative modification of structural and functional proteins regulating vascular contraction/relaxation, fibrosis and calcification, (3) altered calcium homeostasis in vascular cells, 4) activation of redox-sensitive pro-inflammatory and profibrotic transcription factors, and (4) activation of molecular mechanisms leading to senescence and autophagy in endothelial and vascular smooth muscle cells (Fig. 3). The fact that SOD mimetics, such as tempol, normalise endothelial dysfunction in old rodents supports a role for increased superoxide anion levels in age-related endothelial impairment [90].

Changes in cellular anti-oxidant systems are also important. The expression and activity of antioxidant enzymes, including SOD, decline as tissues age. Decreased anti-oxidant capacity is further promoted by downregulation of Nrf2, the master transcription factor regulating anti-oxidant genes [91]. These processes are accompanied by chronic low-grade inflammation mediated by redox-sensitive NFkB, which is upregulated in aged vessels [92].

Multiple oxidases generate ROS in the vascular wall and endothelium, including NADPH oxidases (Nox), xanthine oxidase, uncoupled NOS and mitochondrial oxidases. Of these, mitochondria seem to play a major role in processes related to ageing. Noxs, of which there are 7 isoforms (Nox1-5, Duox1, Duox2), have also been shown to contribute to oxidative stress in vascular ageing [93–95]. In particular, in aged spontaneously hypertensive rat aortas, expression of Nox1 and Nox2, but not of Nox4, was increased. This Nox upregulation was associated with endothelial dysfunction, which was reversed by VAS2870, a Nox inhibitor [96]. Noxs appear to be more important in pathological vascular remodelling associated with hypertension and cardiovascular diseases [97–99]. Vascular xanthine oxidase and cytochrome P450 epoxygenases seem to be less important, since expression and activity of these systems is not altered with ageing in humans [100].

With biological ageing, mitochondria become dysfunctional leading to reduced energy production and increased ROS formation. Mechanisms related to mitochondrial dysfunction during ageing include decreased ATP synthesis, increased apoptosis and mutations of mitochondrial DNA by oxidation [101]. During ageing, the electron flow in mitochondria decreases, altering the oxygen consumption and inducing ROS generation [101]. The pro-oxidative environment increases mitochondrial DNA damage, leading to further dysfunction of the respiratory chain and more ROS production. Consequently, the rate of apoptosis increases, releasing an excessive amount of ROS into the cytosol, further contributing to oxidative stress and vascular cell damage.

7.8. Endoplasmic reticulum stress in vascular ageing

Prolonged perturbation of the endoplasmic reticulum (ER) leads to ER stress and unfolded protein response (UPR) and contributes to pathogenic processes associated with vascular damage and endothelial dysfunction [102]. The ER is an important site where proteins are folded and post-translation modifications occur. It is also a site for Ca²⁺ storage and cholesterol/lipid biosynthesis. Due to the large amount of unfolded protein in the ER, a control system that avoids protein aggregation and accumulation of unfolded proteins is necessary. In experimental models of ageing, the expression and activity of ER chaperones or folding enzymes decay, whilst oxidative damage, such as carbonylation, is

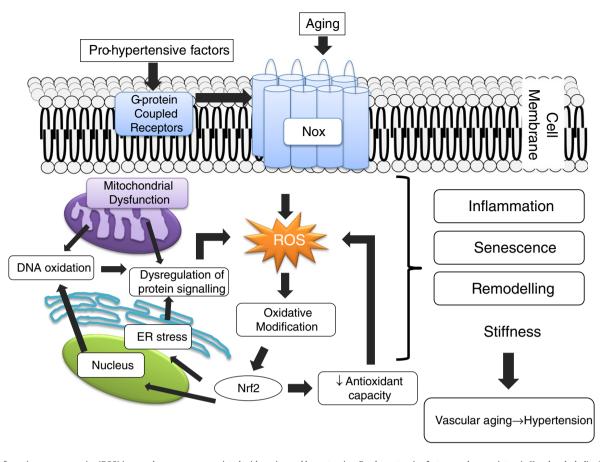


Fig. 3. Role of reactive oxygen species (ROS) in vascular processes associated with ageing and hypertension. Pro-hypertensive factors, such as angiotensin II and endothelin-1, and biological ageing, increase ROS production in vascular cells. An increase in the levels of ROS lead to oxidation of proteins and DNA, affecting cell signalling and inducing injurious responses, such as inflammation, senescence, fibrosis, calcification, and hypertrophy in the vasculature. Oxidation of transcription factors that regulate the anti-oxidant capacity in vascular cells, such as Nrf2, are also affected by oxidation leading to decreased activity. Sources responsible for the increase in ROS generation and oxidative modification of cellular molecules are the mitochondria, NADPH oxidases (Nox) and endoplasmic reticulum (ER) stress.

exacerbated, leading to accumulation of misfolded/unfolded proteins and ER stress. This activates signalling mechanisms that are part of the UPR. Induction of ER stress leads to endothelial cell apoptosis, but not senescence, implicated in endothelial dysfunction in ageing [103,104]. Inhibition of ER stress has been suggested as a novel therapeutic strategy to ameliorate vascular dysfunction during ageing [105]. However such approaches still require further investigation.

7.9. Vascular changes in hypertension recapitulate those in ageing

Many of the signalling pathways associated with vascular changes during ageing are also activated in hypertension leading to endothelial dysfunction, vascular inflammation, remodelling and increased arterial stiffness. With normal physiological ageing the process is gradual and regulated but in itself represents a strong and independent risk factor for hypertension and future cardiovascular events [106]. In susceptible individuals, due to genetic, environmental or in-utero factors 9fetal programming), processes underlying vascular changes are accelerated leading to 'early vascular ageing', which predisposes to cardiovascular disease. Numerous risk factors amplify the process of arterial ageing, including atherosclerosis, smoking, increased sodium intake and hypertension, due, in part, to increased oxidative stress, activation of proinflammatory and pro-fibrotic signalling pathways and upregulation of the renin-angiotensin-aldosterone system. As with ageing, experimental and human hypertension show a reduction in endotheliumdependent vasodilation, decreased NO bioavailability, NO synthase uncoupling, increased oxidative stress, telomere shortening and associated endothelial dysfunction. In arteries from aged humans, non-human primates and rodents, expression of the AT₁R is increased and sensitivity of the mineralocorticoid receptor to aldosterone is enhanced, phenomena that are also observed in hypertension [107–109]. Ang II promotes vascular calcification, inflammation, cell proliferation and fibrosis and mimics age-associated vascular remodelling in young rodents [110–112]. In large arteries these molecular and cellular processes manifest as increased arterial stiffness, which is a major contributor to elevated central blood pressure leading to isolated systolic hypertension, common in the elderly. Exactly what triggers these cellular and vascular events remains unclear, and it is difficult to dissect out the 'ageing effect' from the 'blood pressure and ageing' has recently been reviewed by AlGhatrif and Lakatta [113], who concluded that vascular properties depend on the net effect of multiple factors that are interdependent and which change with ageing over a lifetime.

7.10. Effects of pro-hypertensive stimuli on vascular ageing: the reninangiotensin-aldosterone system (RAAS)

The RAAS plays an important role in functional, structural and mechanical changes of the vasculature that occur with ageing and hypertension [114]. This occurs through increased signalling via the AT₁ receptor. Expression of various components of the RAS, including angiotensinogen, chymase, angiotensin converting enzyme (ACE) and the AT₁ receptor is increased in arteries of aged rodents and humans [8,9]. To further support a role for the RAAS in the ageing process and during hypertension are studies showing that ACE inhibitors and AT₁ receptor blockers decrease ageing-associated vascular damage. Mice

treated with enalapril or losartan demonstrated vasoprotection and an increase in life span [115,116]. Processes associated with these effects involve upregulation of NOS activity and increased NO production. An increase in antioxidant defences, such as SOD and glutathione, is another mechanism involved in the anti-ageing effects of the inhibition of the RAAS system, leading to an increase in NO bioavailability [117]. Moreover, lifelong treatment of young stroke-prone spontaneously hypertensive rats, with AT₁ receptor blockers doubles the lifespan by improving endothelial function and alleviating complications of hypertension [118]. To further support a role for Ang II/AT₁ receptor in oxidative stress, vascular injury and ageing, studies in mice with targeted disruption of the Agtr1a gene, which encodes the AT₁A receptor, resulted in prolonged life span [119]. Agtr1a -/- mice developed less cardiac and vascular injury and oxidative damage was reduced compared with wild-type counterparts. The longevity phenotype was associated with an increased number of mitochondria and upregulation of prosurvival genes.

Clinical and experimental studies demonstrate that many prohypertensive systems influence processes of vascular ageing, including aldosterone, ET-1 and growth factors [120–124]. Arteries from aged rodents demonstrate upregulation of these systems, leading to stimulation of signalling pathways, oxidative stress, and activation of proinflammatory transcription factors, which promote a shift of endothelial and vascular smooth muscle cells to an ageing phenotype. On the other hand, infusion of Ang II, aldosterone or ET-1 in young animals, recapitulates arterial changes observed in aged animals.

7.11. Effects of novel anti-hypertensive factors on vascular ageing

NO is a potent vasodilator produced by endothelial cells that mediates vascular relaxation and thus plays a critical role in the regulation of blood pressure. Abnormalities in endothelial production of NO occur in hypertension and are due, in large part, to decreased eNOS activity [125]. NO donors such as glyceryl trinitrate (GTN) have been shown to possess anti-hypertensive properties [126] and evidence is emerging that NO and NO donors could confer beneficial effects on the phenotypic alterations that occur in the vasculature with ageing. For example, NO prevents differentiation of VSMCs into osteoblastic cells by inhibiting TGF- β [127]. The NO donor *S*-nitroso-penicillamine significantly reduces endothelial cell senescence and age-dependent inhibition of telomerase activity [128,129].

The gaseous messenger hydrogen sulphide (H₂S) produced by cystathionine g-lyase (CSE) or cystathionine b-synthase (CBS) has recently emerged as a novel antihypertensive factor based on the observations that exogenous H₂S is vasoprotective in pulmonary hypertension [130] and that it reduces systemic blood pressure by improving endothelial function [131]. CSE-deficient mice, have increased blood pressure and impaired endothelial function [132]. Mouse embryonic fibroblasts from CSE knockout mice display accelerated cellular senescence and increased expression of p53 and p21, processes which were prevented by NaHS treatment. NaHS also enhanced Nrf2 nuclear translocation, and stimulated mRNA expression of Nrf2-targeted downstream anti-oxidant genes in this system, highlighting an important interplay between cellular ageing, senescence and oxidative stress [133]. Attenuation of endothelial cell senescence by H2S occurs through modulation of SIRT1 activity [134].

Plasma levels of H_2S in humans decline with age [135] and several studies have shown that H_2S protects against free radical-induced damage and exerts beneficial effects on age-associated diseases [136]. Several lines of evidence indicate that these beneficial effects may extend to vascular ageing, characterised by positive effects on many of the phenotypic vascular alterations that occur with advancing age. For example, the production of H_2S is decreased in a rodent model of vascular calcification with the addition of H_2S ameliorating this phenotype [137]. Further, in vascular smooth muscle cells, H_2S was found to inhibit

calcium deposition in the extracellular matrix and suppress induction of osteoblastic transformation genes [138].

In endothelial cells stimulated with TNF- α , NaHS (H2S donor) suppressed pro-inflammatory responses by reducing the TNF-induced increase in expression of intercellular adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1), P-selectin and E-selectin. Furthermore, TNF- α -induced NF- κ B was decreased in the presence of NaHS [139]. H₂S donors (NaHS and Na₂S) can inhibit leukocyte adherence in mesenteric venules whilst inhibition of endogenous H₂S synthesis promotes leukocyte adhesion and vascular inflammation [140].

7.12. Vascular damage in hypertension may be independent of ageing

Although there are many signalling pathways and functional and structural characteristics that are common in vessels during ageing and hypertension, these processes are dynamic and change throughout life and as such may not necessarily be superimposable. For example, with advancing age arterial stiffness and blood pressure start to diverge rather than parallel [141]. Also, circulating markers of inflammation including sVCAM-1, IL-6 and MCP-1 increase with age but do not necessarily correlate with elevations in blood pressure [142]. Furthermore, with ageing, aortic calcification is independently predictive of subsequent vascular morbidity and mortality beyond established risk factors with no evident correlation between calcification and systolic BP [143].

There is also some evidence to show that structural alterations in the vascular wall occur before the development of hypertension. For example rates of pulse wave velocity (PWV) increase are accelerated with both advancing age and elevated blood pressure. However, the effect of blood pressure on PWV increase occurs primarily during the prehypertensive phase and suggests that these vascular alterations precede the phase of established hypertension [8,144].

7.13. Lessons learned from children with hypertension

Vascular changes that occur with ageing may be independent of biological ageing in hypertension. This is highlighted in studies that have examined vascular function and arterial structure in children with hypertension. Endothelial dysfunction, arterial stiffening and structural alterations of the arterial wall may precede evidence of high blood pressure, as quantified by systolic and diastolic blood pressure, and may be independent of the ageing process [145]. This is evidenced by the findings that vascular injury is already present in children with mild hypertension, processes that are exaggerated as hypertension becomes more severe [146,147]. Functional alterations, including reduced endothelium-dependent vasorelaxation and decreased elasticity, seem to precede vascular structural changes. In obese pre-pubertal children, impaired brachial endothelial and vascular smooth muscle function is present without concomitant increase in carotid intima-to-media thickness. Functionally these changes lead to decreased vascular distensibility and increased rigidity or stiffness. Arterial stiffness, as assessed by measurement of PWV, is increased in children with type 1 diabetes and in children with hypertension. Increased arterial stiffness in childhood hypertension is an important risk factor for severe hypertension and cardiovascular complications later in life. Results from the Amsterdam Growth and Health Longitudinal Study indicate that individuals with stiffer carotid arteries at 36 years of age were characterised during adolescence by increased blood pressure and increased PWV [148]. Factors that have been implicated in vascular dysfunction in childhood hypertension include activation of the sympathetic nervous system, adipokines, upregulation of the RAAS and increased oxidative stress, processes that also underlie physiological vascular ageing and EVA in adult hypertension [149–152].

Taken together, emerging experimental and clinical evidence indicates that although the molecular and cellular processes that characterise the vascular phenotype in hypertension resemble those that occur with normal healthy ageing, age per se may not be a critical factor. However, ageing may be a compounding factor that amplifies vascular injury that occurs with blood pressure elevation. In the presence of other co-morbidities, such as diabetes, dyslipidemia, smoking and obesity, these processes may be further exaggerated.

8. Summary and conclusions

Ageing is associated with a progressive deterioration in endothelial function, vascular remodelling, inflammation and increased arterial stiffness. Processes underlying these processes include activation of pro-inflammatory transcription factors, oxidative stress, cell senescence and apoptosis, aberrant signalling cascades and a shift from a vasoconstrictor to a proliferative vascular cell phenotype. Many of these phenomena are also relevant in the pathophysiology of hypertension, which is characterised by a vascular phenotype of impaired endothelium-dependent vasorelaxation, arterial remodelling, increased stiffness and vascular inflammation. Through such vascular changes, ageing and hypertension are closely interlinked: ageing promotes hypertension and pro-hypertensive factors promote vascular ageing. Whilst many of the molecular processes and signalling pathways contributing to vascular dysfunction are common in ageing and in hypertension, biological age per se, may not be a fundamental factor, since vascular damage is already present in children and young adults with hypertension. A better understanding of the vascular biology of ageing will facilitate development of strategies to promote healthy vessels and suppress age-associated changes, especially in pathological conditions. Such approaches could prevent or ameliorate vascular damage in hypertension and hence reduce cardiovascular diseases, commonly linked to ageing.

Disclosures

There are no disclosures to declare.

Acknowledgements

Work from the author's laboratory was supported by grants 44018 and 57886, from the Canadian Institutes of Health Research (CIHR) and grants from the British Heart Foundation (BHF) RG/13/7/30099. RMT is supported through a BHF Chair CH/12/4/29762. ACM is supported through a Leadership Fellowship from the University of Glasgow.

References

- Wang M, Monticone RE, Lakatta EG. Arterial aging: a journey into subclinical arterial disease. Curr Opin Nephrol Hypertens 2010;19(2):201–7.
- [2] Barja G. Updating the mitochondrial free radical theory of aging: an integrated view, key aspects, and confounding concepts. Antioxid Redox Signal 2013; 19(12):1420–45.
- [3] Bachschmid MM, Schildknecht S, Matsui R, Zee R, Haeussler D, Cohen RA, et al. Vascular aging: chronic oxidative stress and impairment of redox signalling consequences for vascular homeostasis and disease. Ann Med 2013;45(1): 17–36.
- [4] Kotsis V, Stabouli S, Karafillis I, Nilsson P. Early vascular aging and the role of central blood pressure. | Hypertens 2011;29(10):1847–53.
- [5] Gerhard-Herman M, Smoot LB, Wake N, Kieran MW, Kleinman ME, Miller DT, et al. Mechanisms of premature vascular aging in children with Hutchinson–Gilford progeria syndrome. Hypertension 2012;59(1):92–7.
- [6] Collins JA, Munoz JV, Patel TR, Loukas M, Tubbs RS. The anatomy of the aging aorta. Clin Anat 2014;27(3):463-6.
- [7] Donato AJ, Gano LB, Eskurza I, Silver AE, Gates PE, Jablonski K, et al. Vascular endothelial dysfunction with aging: endothelin-1 and endothelial nitric oxide synthase. Am J Physiol Heart Circ Physiol 2009;297:H425–32.
- [8] Lakatta EG, Levy D. Arterial and cardiac aging: major shareholders in cardiovascular disease enterprises: Part I: aging arteries: a "set up" for vascular disease. Circulation 2003;107(1):139–46.
- [9] Lakatta EG. The reality of aging viewed from the arterial wall. Artery Res 2013;7(2): 73–80.
- [10] Gerstenblith G, Frederiksen J, Yin FC, Fortuin NJ, Lakatta EG, Weisfeldt ML. Echocardiographic assessment of a normal adult aging population. Circulation 1977;56(2): 273–8.

- [11] Virmani R, Avolio AP, Mergner WJ, Robinowitz M, Herderick EE, Cornhill JF, et al. Effect of aging on aortic morphology in populations with high and low prevalence of hypertension and atherosclerosis. Comparison between occidental and Chinese communities. Am J Pathol 1991;139(5):1119–29.
- [12] Sugawara J, Hayashi K, Yokoi T, Tanaka H. Age-associated elongation of the ascending aorta in adults. JACC Cardiovasc Imaging 2008;1(6):739–48.
- [13] Nilsson PM, Khalili P, Franklin SS. Blood pressure and pulse wave velocity as metrics for evaluating pathologic ageing of the cardiovascular system. Blood Press 2014; 23(1):17–30.
- [14] Michel J, Heudes D, Michel O, Poitevin P, Philippe M, Scalbert E, et al. Effect of chronic ANG I-converting enzyme inhibition on aging processes. II. Large arteries. Am J Physiol 1994;267:R124-R124.
- [15] Wang M, Takagi G, Asai K, Resuello RG, Natividad FF, Vatner DE, et al. Aging increases aortic MMP-2 activity and angiotensin II in nonhuman primates. Hypertension 2003; 41(6):1308–16.
- [16] Li Z, Froehlich J, Galis ZS, Lakatta EG. Increased expression of matrix metalloproteinase-2 in the thickened intima of aged rats. Hypertension 1999; 33(1):116–23.
- [17] Jiang L, Zhang J, Monticone RE, Telljohann R, Wu J, Wang M, et al. Calpain-1 regulation of matrix metalloproteinase 2 activity in vascular smooth muscle cells facilitates age-associated aortic wall calcification and fibrosis. Hypertension 2012; 60(5):1192–9.
- [18] Shanahan CM. Mechanisms of vascular calcification in CKD—evidence for premature ageing? Nat Rev Nephrol 2013;9(11):661–70.
- [19] Alam MU, Kirton JP, Wilkinson FL, Towers E, Sinha S, Rouhi M, et al. Calcification is associated with loss of functional calcium-sensing receptor in vascular smooth muscle cells. Cardiovasc Res 2009;81(2):260–8.
- [20] Rosito GA, Massaro JM, Hoffmann U, Ruberg FL, Mahabadi AA, Vasan RS, et al. Pericardial fat, visceral abdominal fat, cardiovascular disease risk factors, and vascular calcification in a community-based sample: the Framingham Heart Study. Circulation 2008;117(5):605–13.
- [21] McCarty MF, DiNicolantonio JJ. The molecular biology and pathophysiology of vascular calcification. Postgrad Med 2014;126(2):54–64.
- [22] Trebak M, Ginnan R, Singer HA, Jourd'heuil D. Interplay between calcium and reactive oxygen/nitrogen species: an essential paradigm for vascular smooth muscle signaling. Antioxid Redox Signal 2010;12(5):657–74.
- [23] Louvet L, Büchel J, Steppan S, Passlick-Deetjen J, Massy ZA. Magnesium prevents phosphate-induced calcification in human aortic vascular smooth muscle cells. Nephrol Dial Transplant 2013;28(4):869–78.
- [24] Montezano AC, Zimmerman D, Yusuf H, Burger D, Chignalia AZ, Wadhera V, et al. Vascular smooth muscle cell differentiation to an osteogenic phenotype involves TRPM7 modulation by magnesium. Hypertension 2010 Sep;56(3):453–62.
- [25] McClelland RL, Chung H, Detrano R, Post W, Kronmal RA. Distribution of coronary artery calcium by race, gender, and age: results from the Multi-Ethnic Study of Atherosclerosis (MESA). Circulation 2006;113(1):30–7.
- [26] Sweatt A, Sane D, Hutson S, Wallin R. Matrix Gla protein (MGP) and bone morphogenetic protein-2 in aortic calcified lesions of aging rats. J Thromb Haemost 2003; 1(1):178–85.
- [27] Rutsch F, Nitschke Y, Terkeltaub R. Genetics in arterial calcification: pieces of a puzzle and cogs in a wheel. Circ Res 2011;109(5):578–92.
- [28] Leopold JA. Vascular calcification: an age-old problem of old age. Circulation 2013; 127(24):2380–2.
- [29] Lomashvili KA, Khawandi W, O'Neill WC. Reduced plasma pyrophosphate levels in hemodialysis patients. J Am Soc Nephrol 2005;16(8):2495–500.
- [30] O'Neill WC, Sigrist MK, McIntyre CW. Plasma pyrophosphate and vascular calcification in chronic kidney disease. Nephrol Dial Transplant 2010;25(1):187–91.
- [31] Wang M, Monticone RE, Lakatta EG. Proinflammation of aging central arteries: a mini-review. Gerontology 2014;60(6):519–29.
- [32] Csiszar A, Ungvari Z, Edwards JG, Kaminski P, Wolin MS, Koller A, et al. Aging induced phenotypic changes and oxidative stress impair coronary arteriolar function. Circ Res 2002;90(11):1159–66.
- [33] Bruunsgaard H, Skinhøj P, Pedersen AN, Schroll M, Pedersen B. Ageing, tumour necrosis factor-alpha (TNF-α) and atherosclerosis. Clin Exp Immunol 2000; 121(2):255–60.
- [34] Csiszar A, Ungvari Z. Synergistic effects of vascular IL-17 and TNFα may promote coronary artery disease. Med Hypotheses 2004;63(4):696–8.
- [35] Csiszar A, Labinskyy N, Smith K, Rivera A, Orosz Z, Ungvari Z. Vasculoprotective effects of anti-tumor necrosis factor-α treatment in aging. Am J Pathol 2007; 170(1):388–98.
- [36] Arenas IA, Xu Y, Davidge ST. Age-associated impairment in vasorelaxation to fluid shear stress in the female vasculature is improved by TNF-alpha antagonism. Am | Physiol Heart Circ Physiol 2006;290(3):H1259–63.
- [37] Ungvari Z, Orosz Z, Labinskyy N, Rivera A, Xiangmin Z, Smith K, et al. Increased mitochondrial H2O2 production promotes endothelial NF-kappaB activation in aged rat arteries. Am J Physiol Heart Circ Physiol 2007;293(1):H37–47.
- [38] Paneni F, Costantino S, Cosentino F. Molecular pathways of arterial aging. Clin Sci (Lond) 2015;128(2):69–79.
- [39] Zicha J, Behuliak M, Pintérová M, Bencze M, Kuneš J, Vaněčková I. The interaction of calcium entry and calcium sensitization in the control of vascular tone and blood pressure of normotensive and hypertensive rats. Physiol Res 2014; 63:S19–27.
- [40] Davidge ST, Hubel CA, McLaughlin MK. Impairment of vascular function is associated with an age-related increase of lipid peroxidation in rats. Am J Physiol Regul Integr Comp Physiol 1996;271(6):R1625–31.
- [41] Reyes-Toso CF, Obaya-Naredo D, Ricci CR, Planells FM, Pinto JE, Linares LM, et al. Exp Gerontol 2007;42(4):337–42.

- [42] Lloréns S, de Mera Raquel M, Melero-Fernandez, Pascual A, Prieto-Martín A, Mendizábal Y, et al. The senescence-accelerated mouse (SAM-P8) as a model for the study of vascular functional alterations during aging. Biogerontology 2007; 8(6):663–72.
- [43] de Andrade CR, Correa FM, de Oliveira AM. Aging and total stenosis triggers differential responses of carotid and basilar arteries to endothelin-1 and phenylephrine. J Smooth Muscle Res 2009;45(6):307–21.
- [44] Marchand A, Abi-Gerges A, Saliba Y, Merlet E, Lompré A. Calcium signaling in vascular smooth muscle cells: from physiology to pathology. Adv Exp Med Biol 2012;740:795–810.
- [45] Kitamura-Sasaka F, Ueda K, Kawai Y. Effects of aging on contraction and Ca2 mobilization in smooth muscle cells of the rat coronary artery. Yonago Acta Med 2001; 44(1):61–8.
- [46] Celermajer DS, Sorensen KE, Spiegelhalter DJ, Georgakopoulos D, Robinson J, Deanfield JE. Aging is associated with endothelial dysfunction in healthy men years before the age-related decline in women. J Am Coll Cardiol 1994;24(2):471–6.
- [47] Dohi Y, Kojima M, Sato K, Lüscher TF. Age-related changes in vascular smooth muscle and endothelium. Drugs Aging 1995;7(4):278–91.
- [48] Favero G, Paganelli C, Buffoli B, Rodella LF, Rezzani R. Endothelium and its alterations in cardiovascular diseases: life style intervention. Biomed Res Int 2014:1–28.
- [49] Taddei S, Virdis A, Mattei P, Ghiadoni L, Gennari A, Fasolo CB, et al. Aging and endothelial function in normotensive subjects and patients with essential hypertension. Circulation 1995;91(7):1981–7.
- [50] Sindler AL, Delp MD, Reyes R, Wu G, Muller-Delp JM. Effects of ageing an exercise training on eNOS uncoupling in skeletal muscle resistance arterioles. J Physiol 2009;587(Pt 15):3885–97.
- [51] Mostoslavsky R, Chua KF, Lombard DB, Pang WW, Fischer MR, Gellon L, et al. Genomic instability and aging-like phenotype in the absence of mammalian SIRT6. Cell 2006; 124(2):315–29.
- [52] Vakhrusheva O, Braeuer D, Liu Z, Braun T, Bober E. Sirt7-dependent inhibition of cell growth and proliferation might be instrumental to mediate tissue integrity during aging. J Physiol Pharmacol 2008;59(Suppl. 9):201–12.
- [53] Zu Y, Liu L, Lee MY, Xu C, Liang Y, Man RY, et al. SIRT1 promotes proliferation and prevents senescence through targeting LKB1 in primary porcine aortic endothelial cells. Circ Res 2010;106(8):1384–93.
- [54] Takemura A, Iijima K, Ota H, Son BK, Ito Y, Ogawa S, et al. Sirtuin 1 retards hyperphosphatemia-induced calcification of vascular smooth muscle cells. Arterioscler Thromb Vasc Biol 2011;31(9):2054–62.
- [55] Hafner AV, Dai J, Gomes AP, Xiao CY, Palmeira CM, Rosenzweig A, et al. Regulation of the mPTP by SIRT3-mediated deacetylation of CypD at lysine 166 suppresses age-related cardiac hypertrophy. Aging (Albany NY) 2010;2(12):914–23.
- [56] Tseng P, Hou S, Chen R, Peng H, Hsieh C, Kuo M, et al. Resveratrol promotes osteogenesis of human mesenchymal stem cells by upregulating RUNX2 gene expression via the SIRT1/FOXO3A axis. J Bone Miner Res 2011;26(10):2552–63.
- [57] Mukherjee S, Basar MA, Davis C, Duttaroy A. Emerging functional similarities and divergences between Drosophila Spargel/dPGC-1 and mammalian PGC-1 protein. Front Genet 2014;5:216.
- [58] Mohamed SA, Hanke T, Erasmi AW, Bechtel MJ, Scharfschwerdt M, Meissner C, et al. Mitochondrial DNA deletions and the aging heart. Exp Gerontol 2006; 41(5):508–17.
- [59] Ungvari Z, Parrado-Fernandez C, Csiszar A, de Cabo R. Mechanisms underlying caloric restriction and lifespan regulation: implications for vascular aging. Circ Res 2008;102(5):519–28.
- [60] Zinovkin RA, Romaschenko VP, Galkin II, Zakharova VV, Pletjushkina OY, Chernyak BV, et al. Role of mitochondrial reactive oxygen species in age-related inflammatory activation of endothelium. Aging (Albany NY) 2014;6(8):661–74.
- [61] Reznick RM, Zong H, Li J, Morino K, Moore IK, Yu HJ, et al. Aging-associated reductions in AMP-activated protein kinase activity and mitochondrial biogenesis. Cell Metab 2007;5(2):151–6.
- [62] van den Berg Maaike CW, Burgering BM. Integrating opposing signals toward Forkhead box O. Antioxid Redox Signal 2011;14(4):607–21.
- [63] Oellerich MF, Potente M. FOXOs and sirtuins in vascular growth, maintenance, and aging. Circ Res 2012;110(9):1238–51.
- [64] Tseng AH, Shieh S, Wang DL, SIRT3 deacetylates FOXO3 to protect mitochondria against oxidative damage. Free Radic Biol Med 2013;63:222–34.
- [65] Willcox BJ, Donlon TA, He Q, Chen R, Grove JS, Yano K, et al. FOXO3A genotype is strongly associated with human longevity. Proc Natl Acad Sci U S A 2008; 105(37):13987–92.
- [66] Anselmi CV, Malovini A, Roncarati R, Novelli V, Villa F, Condorelli G, et al. Association of the FOXO3A locus with extreme longevity in a southern Italian centenarian study. Rejuvenation Res 2009;12(2):95–104.
- [67] Castrillon DH, Miao L, Kollipara R, Horner JW, DePinho RA. Suppression of ovarian follicle activation in mice by the transcription factor Foxo3a. Science 2003; 301(5630):215–8.
- [68] Remmen HV, Richardson A. Oxidative damage to mitochondria and aging. Exp Gerontol 2001;36(7):957–68.
- [69] Lebiedzinska M, Duszynski J, Rizzuto R, Pinton P, Wieckowski M. Age-related changes in levels of p66Shc and serine 36-phosphorylated p66Shc in organs and mouse tissues. Arch Biochem Biophys 2009;486(1):73–80.
- [70] Migliaccio E, Giorgio M, Mele S, Pelicci G, Reboldi P, Pandolfi PP, et al. The p66shc adaptor protein controls oxidative stress response and life span in mammals. Nature 1999;402(6759):309–13.
- [71] Francia P, delli Gatti C, Bachschmid M, Martin-Padura I, Savoia C, Migliaccio E, et al. Deletion of p66shc gene protects against age-related endothelial dysfunction. Circulation 2004;110(18):2889–95.

- [72] Minamino T, Yoshida T, Tateno K, Miyauchi H, Zou Y, Toko H, et al. Ras induces vascular smooth muscle cell senescence and inflammation in human atherosclerosis. Circulation 2003;108(18):2264–9.
- [73] Campisi J, Robert L. Cell senescence: role in aging and age-related diseases. Interdiscip Top Gerontol 2014;39:45–61.
- [74] Lauri A, Pompilio G, Capogrossi MC. The mitochondrial genome in aging and senescence. Ageing Res Rev 2014;18C:1–15.
- [75] Rivard A, Principe N, Andres V. Age-dependent increase in c-fos activity and cyclin A expression in vascular smooth muscle cells. A potential link between aging, smooth muscle cell proliferation and atherosclerosis. Cardiovasc Res 2000;45(4): 1026–34.
- [76] Vazquez-Padron RI, Lasko D, Li S, Louis L, Pestana IA, Pang M, et al. Aging exacerbates neointimal formation, and increases proliferation and reduces susceptibility to apoptosis of vascular smooth muscle cells in mice. J Vasc Surg 2004:40(6):1199–207.
- [77] Salabei JK, Hill BG. Autophagic regulation of smooth muscle cell biology. Redox Biol 2014;4C:97–103.
- [78] Gao L, Qi HB, Kc K, Zhang XM, Zhang H, Baker PN. Excessive autophagy induces the failure of trophoblast invasion and vasculature: possible relevance to the pathogenesis of preeclampsia. J Hypertens 2015;33(1):106–17.
- [79] Force T, Hajjar R, Del Monte F, Rosenzweig A, Choukroun G. Signaling pathways mediating the response to hypertrophic stress in the heart. Gene Expr 1999;7(4-6):337.
- [80] Cheng H, Force T. Why do kinase inhibitors cause cardiotoxicity and what can be done about it? Prog Cardiovasc Dis 2010;53(2):114–20.
- [81] Robinson MJ, Cobb MH. Mitogen-activated protein kinase pathways. Curr Opin Cell Biol 1997;9(2):180–6.
- [82] Bueno OF, De Windt LJ, Tymitz KM, Witt SA, Kimball TR, Klevitsky R, et al. The MEK1–ERK1/2 signaling pathway promotes compensated cardiac hypertrophy in transgenic mice. EMBO J 2000;19(23):6341–50.
- [83] Zhang S, Weinheimer C, Courtois M, Kovacs A, Zhang CE, Cheng AM, et al. The role of the Grb2-p38 MAPK signaling pathway in cardiac hypertrophy and fibrosis. J Clin Invest 2003;111(6):833–42.
- [84] Rice KM, Kinnard R, Harris R, Wright G, Blough E. Effects of aging on pressureinduced MAPK activation in the rat aorta. Pflugers Arch 2005;450(3):192–9.
- [85] Chung HY, Cesari M, Anton S, Marzetti E, Giovannini S, Seo AY, et al. Molecular inflammation: underpinnings of aging and age-related diseases. Ageing Res Rev 2009;8(1):18–30.
- [86] Montezano AC, Touyz RM. Reactive oxygen species, vascular Noxs, and hypertension: focus on translational and clinical research. Antioxid Redox Signal 2014; 20(1):164–82.
- [87] Harman D. Aging: a theory based on free radical and radiation chemistry. J Gerontol 1956;11:298–300.
- [88] Harman D. Free radical theory of aging: an update: increasing the functional life span. Ann N Y Acad Sci 2006;1067:10–21.
- [89] Fan Q, Chen L, Cheng S, Li F, Lau WB, Wang Le F, et al. Aging aggravates nitrate-mediated ROS/RNS changes. Oxid Med Cell Longev 2014;2014:376515.
- [90] Tatchum-Talom R1, Martin DS. Tempol improves vascular function in the mesenteric vascular bed of senescent rats. Can J Physiol Pharmacol 2004;82(3):200–7.
- [91] Ungvari Z, Bailey-Downs L, Sosnowska D, Gautam T, Koncz P, Losonczy G. Vascular oxidative stress in aging: a homeostatic failure due to dysregulation of NRF2mediated antioxidant response. Am J Physiol Heart Circ Physiol 2011;301:H363–72.
- [92] Spencer NF, Poynter ME, İm SY, Daynes RA. Constitutive activation of NF-kappa B in an animal model of aging. Int Immunol 1997;9(10):1581–8.
- [93] Krause KH. Aging: a revisited theory based on free radicals generated by NOX family NADPH oxidases. Exp Gerontol 2007;42(4):256–62.
- [94] Brandes RP, Weissmann N, Schröder K. Nox family NADPH oxidases: molecular mechanisms of activation. Free Radic Biol Med 2014;76C:208–26.
- [95] Cencioni C, Spallotta F, Martelli F, Valente S, Mai A, Zeiher AM, et al. Oxidative stress and epigenetic regulation in ageing and age-related diseases. Int J Mol Sci 2013; 14(9):17643–63.
- [96] Wind S, Beuerlein K, Armitage ME, Taye A, Kumar AH, Janowitz D, et al. Oxidative stress and endothelial dysfunction in aortas of aged spontaneously hypertensive rats by NOX1/2 is reversed by NADPH oxidase inhibition. Hypertension 2010; 56(3):490–7.
- [97] Touyz RM, Briones AM, Sedeek M, Burger D, Montezano AC. NOX isoforms and reactive oxygen species in vascular health. Mol Interv 2011;11(1):27–35.
- [98] Montezano AC, Touyz RM. Molecular mechanisms of hypertension-reactive oxygen species and antioxidants: a basic science update for the clinician. Can J Cardiol 2012;28(3):288–95.
- [99] Montezano AC, Burger D, Ceravolo GS, Yusuf H, Montero M, Touyz RM. Novel Nox homologues in the vasculature: focusing on Nox4 and Nox5. Clin Sci (Lond) 2011; 120(4):131–41.
- [100] Eskurza I, Kahn ZD, Seals DR. Xanthine oxidase does not contribute to impaired peripheral conduit artery endothelium-dependent dilatation with ageing. J Physiol 2006;571(Pt 3):661–8.
- [101] Kokoszka JE, Coskun P, Esposito LA, Wallace DC. Increased mitochondrial oxidative stress in the Sod2 (+/-) mouse results in the age-related decline of mitochondrial function culminating in increased apoptosis. Proc Natl Acad Sci U S A 2001;98: 2278–83.
- [102] Naidoo N, Ferber M, Master M, Zhu Y, Pack AI. Aging impairs the unfolded protein response to sleep deprivation and leads to proapoptotic signaling. J Neurosci 2008; 28:6539–48.
- [103] Lenna S, Han R, Trojanowska M. Endoplasmic reticulum stress and endothelial dysfunction. IUBMB Life 2014;66(8):530–7.
- [104] Panganiban RA, Mungunsukh O, Day RM. X-irradiation induces ER stress, apoptosis, and senescence in pulmonary artery endothelial cells. Int J Radiat Biol 2013;89(8): 656–67.

- [105] Galán M, Kassan M, Kadowitz PJ, Trebak M, Belmadani S, Matrougui K. Mechanism of endoplasmic reticulum stress-induced vascular endothelial dysfunction. Biochim Biophys Acta 2014;1843(6):1063–75.
- [106] Najjar SS, Scuteri A, Lakatta EG. Arterial aging: is it an immutable cardiovascular risk factor? Hypertension 2005;46(3):454–62.
- [107] Hinojosa-Laborde C, Craig T, Zheng W, Ji H, Haywood JR, Sandberg K. Ovariectomy augments hypertension in aging female Dahl salt-sensitive rats. Hypertension 2004;44(4):405–9.
- [108] Savoia C, Burger D, Nishigaki N, Montezano A, Touyz RM. Angiotensin II and the vascular phenotype in hypertension. Expert Rev Mol Med 2011;13:e11.
- [109] Chugh G, Lokhandwala MF, Asghar M. Altered functioning of both renal dopamine D1 and angiotensin II type 1 receptors causes hypertension in old rats. Hypertension 2012 May;59(5):1029–36.
- [110] Osako MK, Nakagami H, Shimamura M, Koriyama H, Nakagami F, Shimizu H, et al. Cross-talk of receptor activator of nuclear factor-kappaB ligand signaling with renin–angiotensin system in vascular calcification. Arterioscler Thromb Vasc Biol 2013;33(6):1287–96.
- [111] Gonzalez GE, Rhaleb NE, D'Ambrosio MA, Nakagawa P, Liu Y, Leung P, et al. Deletion of interleukin-6 prevents cardiac inflammation, fibrosis and dysfunction without affecting blood pressure in angiotensin II-high salt-induced hypertension. J Hypertens 2015;33(1):144–52.
- [112] Imanishi M, Tomita S, Ishizawa K, Kihira Y, Ueno M, Izawa-Ishizawa Y, et al. Smooth muscle cell-specific Hif-1alpha deficiency suppresses angiotensin II-induced vascular remodelling in mice. Cardiovasc Res 2014;102(3):460–8.
- [113] AlGhatrif M, Lakatta EG. The conundrum of arterial stiffness, elevated blood pressure, and aging. Curr Hypertens Rep 2015;17(2):1–9.
- [114] Montezano AC, Nguyen Dinh Cat A, Rios FJ, Touyz RM. Angiotensin II and vascular injury. Curr Hypertens Rep 2014;16(6):43.
- [115] de Cavanagh EMV, Piotrkowski B, Basso N, Stella I, Inserra F, Ferder L, et al. Enalapril and losartan attenuate mitochondrial dysfunction in aged rats. FASEB J 2003;17: 1096–8.
- [116] Gohlke P, Linz W, Schölkens BA, Wiemer G, Unger T. Cardiac and vascular effects of long-term losartan treatment in stroke-prone spontaneously hypertensive rats. Hypertension 1996;28(3):397–402.
- [117] Brown KA, Didion SP, Andresen JJ, Faraci FM. Effect of aging, MnSOD deficiency, and genetic background on endothelial function: evidence for MnSOD haploinsufficiency. Arterioscler Thromb Vasc Biol 2007;27:1941–6.
- [118] Basso N, Paglia N, Stella I, de Cavanagh EM, Ferder L, del Rosario Lores Arnaiz M, et al. Protective effect of the inhibition of the renin–angiotensin system on aging. Regul Pept 2005;128(3):247–52.
- [119] Benigni A, Corna D, Zoja C, Sonzogni A, Latini R, Salio M, et al. Disruption of the Ang Il type 1 receptor promotes longevity in mice. J Clin Invest 2009;119(3):524–30.
- [120] Meyer MR, Fredette NC, Barton M, Prossnitz ER. Endothelin-1 but not angiotensin II contributes to functional aging in murine carotid arteries. Life Sci 2014;118(2): 213–8.
- [121] Barton M. Aging and endothelin: determinants of disease. Life Sci 2014;118(2): 97–109.
- [122] Hannemann A, Wallaschofski H, Lüdemann J, Völzke H, Markus MR, Rettig R, et al. Plasma aldosterone levels and aldosterone-to-renin ratios are associated with endothelial dysfunction in young to middle-aged subjects. Atherosclerosis 2011; 219(2):875–9.
- [123] Paar M, Pavenstädt H, Kusche-Vihrog K, Drüppel V, Oberleithner H, Kliche K. Endothelial sodium channels trigger endothelial salt sensitivity with aging. Hypertension 2014;64(2):391–6.
- [124] Brown JM, Underwood PC, Ferri C, Hopkins PN, Williams GH, Adler GK, et al. Aldosterone dysregulation with aging predicts renal vascular function and cardiovascular risk. Hypertension 2014;63(6):1205–11.
- [125] Panza JA, Casino PR, Kilcoyne CM, Quyyumi AA. Role of endothelium-derived nitric oxide in the abnormal endothelium-dependent vascular relaxation of patients with essential hypertension. Circulation 993;87(5):1468–1474.
- [126] Sumimoto T, Hamada M, Kawakami H, Suzuki M, Abe M, Matsuoka H, et al. Effects of glyceryl trinitrate on blood pressure and arterial compliance. Angiology 1993; 44(12):951–7.
- [127] Kanno Y, Into T, Lowenstein CJ, Matsushita K. Nitric oxide regulates vascular calcification by interfering with TGF-signalling. Cardiovasc Res 2008;77(1):221–30.
- [128] Vasa M, Breitschopf K, Zeiher AM, Dimmeler S. Nitric oxide activates telomerase and delays endothelial cell senescence. Circ Res 2000;87(7):540–2.
- [129] Hayashi T, Matsui-Hirai H, Miyazaki-Akita A, Fukatsu A, Funami J, Ding QF, et al. Endothelial cellular senescence is inhibited by nitric oxide: implications in atherosclerosis associated with menopause and diabetes. Proc Natl Acad Sci U S A 2006;103(45):17018–23.

- [130] Chunyu Z, Junbao D, Dingfang B, Hui Y, Xiuying T, Chaoshu T. The regulatory effect of hydrogen sulfide on hypoxic pulmonary hypertension in rats. Biochem Biophys Res Commun 2003;302(4):810–6.
- [131] Ahmad FuD, Sattar MA, Rathore HA, Abdullah MH, Tan S, Abdullah NA, et al. Exogenous hydrogen sulfide (H2S) reduces blood pressure and prevents the progression of diabetic nephropathy in spontaneously hypertensive rats. Ren Fail 2012; 34(2):203–10.
- [132] Yang G, Zhao K, Ju Y, Mani S, Cao Q. Puukila S, et al. Hydrogen sulfide protects against cellular senescence via S-sulfhydration of Keap1 and activation of Nrf2. Antioxid Redox Signal 2013;18(15):1906–19.
- [133] Yang G, Wu L, Jiang B, Yang W, Qi J, Cao K, et al. H2S as a physiologic vasorelaxant: hypertension in mice with deletion of cystathionine gamma-lyase. Science 2008; 322(5901):587–90.
- [134] Suo R, Zhao Z, Tang Z, Ren Z, Liu X, Liu L, et al. Hydrogen sulfide prevents H2O2induced senescence in human umbilical vein endothelial cells through SIRT1 activation. Mol Med Rep 2013;7(6):1865–70.
- [135] Chen Y, Yao W, Geng B, Ding Y, Lu M, Zhao M, et al. Endogenous hydrogen sulfide in patients with COPD. CHEST J 2005;128(5):3205–11.
- [136] Zhang Y, Tang ZH, Ren Z, Qu SL, Liu MH, Liu LS, et al. Hydrogen sulfide, the next potent preventive and therapeutic agent in aging and age-associated diseases. Mol Cell Biol 2013;33(6):1104–13.
- [137] Wu S, Pan C, Geng B, Zhao J, Yu F, Pang Y, et al. Hydrogen sulfide ameliorates vascular calcification induced by vitamin D3 plus nicotine in rats. Acta Pharmacol Sin 2006;27(3):299–306.
- [138] Zavaczki E, Jeney V, Agarwal A, Zarjou A, Oros M, Katkó M, et al. Hydrogen sulfide inhibits the calcification and osteoblastic differentiation of vascular smooth muscle cells. Kidney Int 2011;80(7):731–9.
- [139] Pan L, Liu X, Gong Q, Wu D, Zhu Y. Hydrogen sulfide attenuated tumor necrosis factor-α-induced inflammatory signaling and dysfunction in vascular endothelial cells. PLoS One 2011;6(5):e19766.
- [140] Zanardo RC, Brancaleone V, Distrutti E, Fiorucci S, Cirino G, Wallace JL. Hydrogen sulfide is an endogenous modulator of leukocyte-mediated inflammation. FASEB J 2006;20(12):2118–20.
- [141] Franklin SS, Gustin IV W, Wong ND, Larson MG, Weber MA, Kannel WB, et al. Hemodynamic patterns of age-related changes in blood pressure. The Framingham Heart Study. Circulation 1997;96(1):308–15.
- [142] Miles EA, Rees D, Banerjee T, Cazzola R, Lewis S, Wood R, et al. Age-related increases in circulating inflammatory markers in men are independent of BMI, blood pressure and blood lipid concentrations. Atherosclerosis 2008;196(1):298–305.
- [143] Blacher J, Guerin AP, Pannier B, Marchais SJ, London GM. Arterial calcifications, arterial stiffness, and cardiovascular risk in end-stage renal disease. Hypertension 2001;38(4):938–42.
- [144] AlGhatrif M, Strait JB, Morrell CH, Canepa M, Wright J, Elango P, et al. Longitudinal trajectories of arterial stiffness and the role of blood pressure: the Baltimore Longitudinal Study of Aging. Hypertension 2013;62(5):934–41.
- [145] Litwin M, Feber J, Niemirska A, Michałkiewicz J. Primary hypertension is a disease of premature vascular aging associated with neuro-immuno-metabolic abnormalities. Pediatr Nephrol Feb. 28 2015. <u>http://dx.doi.org/10.1007/s00467-015-3065-y</u> [Epub ahead of print].
- [146] Feber J, Ahmed M. Hypertension in children: new trends and challenges. Clin Sci (Lond) 2010;119(4):151–61.
- [147] Falkner B. Hypertension in children and adolescents: epidemiology and natural history. Pediatr Nephrol 2010;25(7):1219–24.
- [148] Ferreira I, Twisk JW, van Mechelen W, Kemper HC, Seidell JC, Stehouwer CD. Current and adolescent body fatness and fat distribution: relationships with carotid intima-media thickness and large artery stiffness at the age of 36 years. J Hypertens 2004;22(1):145–55.
- [149] Wang M, Zhang J, Jiang LQ, Spinetti G, Pintus G, et al. Proinflammatory profile within the grossly normal aged human aortic wall. Hypertension 2007;50:219–27.
- [150] Wang M, Takagi G, Asai K, Resuello RG, Natividad FF, et al. Aging increases aortic MMP-2 activity and angiotensin II in nonhuman primates. Hypertension 2003; 41:1308–16.
- [151] Alpert BS, Bain HH, Balfe JW, Kidd BL, Olley PM. Role of the renin-angiotensin-aldosterone system in hypertensive children with coarctation of the aorta. Am J Cardiol 1979;43(4):828–34.
- [152] Rimoldi SF, Sartori C, Rexhaj E, Cerny D, Von Arx R, Soria R, et al. Vascular dysfunction in children conceived by assisted reproductive technologies: underlying mechanisms and future implications. Swiss Med Wkly 2014;144:w13973.