

available at [www.sciencedirect.com](http://www.sciencedirect.com)journal homepage: [www.elsevier.com/locate/rmed](http://www.elsevier.com/locate/rmed)

## REVIEW

# Aging and induced senescence as factors in the pathogenesis of lung emphysema

Stefan Karrasch <sup>a,b</sup>, Olaf Holz <sup>c</sup>, Rudolf A. Jörres <sup>b,\*</sup>

<sup>a</sup> *Institute for Inhalation Biology, Helmholtz Zentrum München – German Research Center for Environmental Health, Neuherberg/Munich, Germany*

<sup>b</sup> *Institute and Outpatient Clinic for Occupational, Social and Environmental Medicine, Ludwig-Maximilians-University, Munich, Germany*

<sup>c</sup> *Hospital Großhansdorf, Center for Pneumology and Thoracic Surgery, Großhansdorf, Germany*

Received 16 December 2007; accepted 4 April 2008

Available online 9 July 2008

## KEYWORDS

p53;  
HDAC;  
HAT;  
Methylation;  
Acetylation;  
SIRT1

## Summary

Classically, the development of emphysema in chronic obstructive pulmonary disease is believed to involve inflammation induced by cigarette smoke and leukocyte activation, including oxidant-antioxidant and protease-antiprotease imbalances. While there is substantial evidence for this, additional aspects have been suggested by a number of clinical and experimental observations.

Smokers exhibit signs of premature aging, particularly obvious in the skin. The link between aging and chronic disease is well-known, e.g., for the brain and musculoskeletal or cardiovascular system, as well as the clinical link between malnutrition and emphysema, and the experimental link to caloric restriction. Interestingly, this intervention also increases lifespan, in parallel with alterations in metabolism, oxidant burden and endocrine signaling.

Of special interest is the observation that, even in the absence of an inflammatory environment, lung fibroblasts from patients with emphysema show persistent alterations, possibly based

**Abbreviations:**  $\alpha_1$ -AT, alpha-1-antitrypsin; Akt, serine-threonine-kinase, member of the protein kinase B (PKB) family; ALT, alternative mechanisms of telomere lengthening; Bad, Bcl-2-associated death promoter, member of the B-cell leukemia/lymphoma 2 (Bcl-2) family; Bax, member of the Bcl-2 family; CDK, cyclin-dependent kinase; COPD, chronic obstructive pulmonary disease; DHEAS, dehydroepiandrosterone sulphate; DNA, deoxyribonucleic acid; FOXO, forkhead box O; GM-CSF, granulocyte/macrophage-colony-stimulating factor; H<sub>2</sub>O<sub>2</sub>, hydrogen peroxide; HAT, histone acetyltransferase; HDAC, histone deacetylase; IL, interleukin; IGF-1, insulin-like growth factor 1; MMP, matrix metalloproteinase; PCNA, proliferating cell nuclear antigen; PI3K, phosphatidylinositol 3-kinase; pRb, retinoblastoma protein; PPAR- $\gamma$ , peroxisome proliferator-activated receptor gamma; PTEN, phosphatase and tensin homolog, tyrosine and lipid phosphatase; RNA, ribonucleic acid; ROS, reactive oxygen species; SA- $\beta$ -gal, senescence-associated  $\beta$ -galactosidase; SIPS, stress-induced proliferative senescence; Sir2, silent information regulator 2; SIRT1, homolog of Sir2 in mammals; TNF- $\alpha$ , tumor necrosis factor alpha; mTOR, mammalian target-of-rapamycin kinase; UV, ultraviolet.

\* Corresponding author. Institute and Outpatient Clinic for Occupational, Social and Environmental Medicine, Ludwig-Maximilians-University Munich, Ziemssenstr. 1, Munich, Bavaria D-80336, Germany. Tel.: +49 89 5160 2466; fax: +49 89 5160 3957.

E-mail address: [rudolf.joerres@med.uni-muenchen.de](mailto:rudolf.joerres@med.uni-muenchen.de) (R.A. Jörres).

on epigenetic mechanisms. The importance of these mechanisms for cellular reprogramming and response patterns, individual risk profile and therapeutic options is becoming increasingly recognized. The same applies to cellular senescence. Recent findings from patients and experimental models open novel views into the arena of gene-environment interactions, including the role of systemic alterations, cellular stress, telomeres, CDK inhibitors such as p16, p21, pRb, PI3K, mTOR, FOXO transcription factors, histone modifications, and sirtuins.

This article aims to outline this emerging picture and to stimulate the identification of challenging questions. Such insights also bear implications for the long-term course of the disease in relation to existing or future therapies and the exploration of potential lung regeneration.

© 2008 Elsevier Ltd. All rights reserved.

## Contents

Introduction	1216
Aging and senescence: basic characteristics	1216
Aging as a contributor to chronic disease	1217
Aging and COPD	1217
Mechanisms of cellular senescence and evidence in COPD	1218
Telomere loss	1218
Inflammation, role of proteases and oxidative stress, mitochondrial dysfunction	1219
Major cell cycle regulators	1219
Role of PI3K and mTOR	1220
Epigenetic mechanisms and senescence	1221
Sirtuins and FOXOs as integrators of multiple pathways	1222
Links to energy metabolism/caloric restriction	1222
Conclusion and outlook	1224
Conflict of interest	1224
References	1224

## Introduction

Lung emphysema is a major phenotype of COPD<sup>1</sup> and represents a significant health burden. Even more so since no causal therapy is available to restore lung architecture. A formidable body of evidence has been accumulated regarding the role of inflammatory factors in the pathogenesis of the disease. It comprises the now classical concept of protease-antiprotease and oxidant-antioxidant imbalances.<sup>2,3</sup> Tissue destruction driven by neutrophils and macrophages via these compounds undoubtedly plays an important role.<sup>4</sup> This is supported by data for hereditary  $\alpha_1$ -antitrypsin (AT)-deficiency, which typically leads to severe emphysema relatively early in life, particularly in the presence of noxious agents.<sup>5</sup> Moreover, instillation of elastase into the lung is a well-known technique for inducing experimental emphysema in animals.<sup>6</sup> Intimately linked to the protease-antiprotease disturbance, oxidative stress originating from compounds of cigarette smoke or inflammatory cells can overcharge the antioxidative capacity of pulmonary tissue and further diminish the anti-protease defense.<sup>7</sup>

In combination with proinflammatory cytokine production and a host of other responses, these influences lead to apoptosis, necrosis, compensatory proliferation and an imbalance in the maintenance of cells, ultimately resulting in alveolar destruction and airway remodeling. While the importance of inflammation is undeniable, clinical and cell biological observations suggest mechanisms beyond

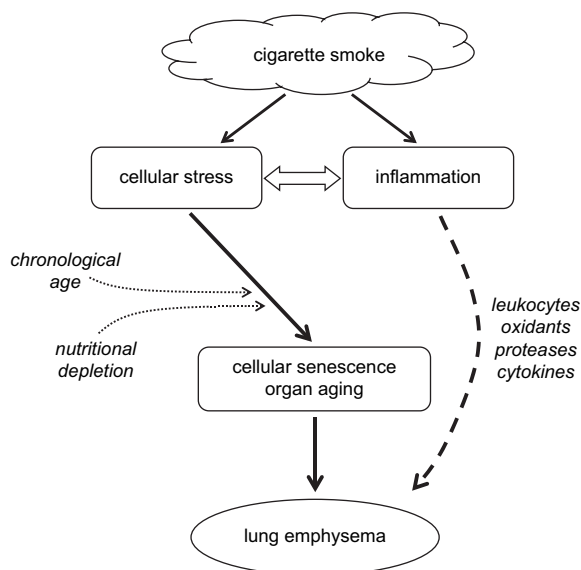
inflammation, albeit linked to it, to play a critical role as well (Fig. 1). These mechanisms include cellular senescence and epigenetic control, which appear of particular interest in view of the observed systemic alterations and chronicity of the disease.

The present article aims to provide the clinician with a comprehensive overview of the multiple facets presenting from the viewpoint of senescence and epigenetics, as well as to delineate major mechanistic aspects of this novel view. In order to truly appreciate the complexity of the molecular networks involved we refer to specialized reviews. A large number of missing links still need to be identified in this area, while the generation of clinically tractable research questions remains an exciting challenge.

## Aging and senescence: basic characteristics

Biological aging involves a variety of cellular, molecular and structural alterations based on several mechanisms.<sup>8</sup> Although normally linked to chronological age, biological aging can occur earlier in life, being partially independent from an individual's chronological age (premature aging). Interestingly, many markers that are used to describe biological aging are related to (chronic) inflammation, e.g., the serum levels of IL-6, IL-1 $\beta$  or TNF- $\alpha$ .<sup>9</sup> As a result, there seems to be no single, comprehensive or easily available marker of biological age.

In this article, the term "senescence" is used to describe aging on the cellular level (cellular aging), which



**Figure 1** Diagram illustrating major factors of aging and senescence that are involved in the development of lung emphysema. The contribution originating as a more or less direct result of inflammation is summarized by the dashed line.

comprises a series of cell morphological and functional alterations including the loss of proliferative activity in otherwise viable cells (Table 1). This loss is observed *in vitro*<sup>10,11</sup> and is in all likelihood also of relevance *in vivo*.<sup>12,13</sup> To distinguish the cellular process from the aging of whole organs or organisms, it is denoted more specifically by “proliferative/replicative senescence.” The cell arrest not only occurs after exhausting the predetermined proliferative capacity (intrinsic senescence), but is also inducible by external stressors administered in sublethal doses (extrinsic or stress-induced proliferative senescence, SIPS).<sup>14</sup> Thus, there are different routes leading to a senescent phenotype, though potentially sharing common signaling pathways. It should be kept in mind that senescence can be interpreted as an evolutionary protective mechanism against tumor development<sup>15</sup> which is particularly relevant in pre-malignant cells.<sup>16</sup> It circumvents eventual irreversible structural losses due to apoptosis, while avoiding the

**Table 1** Major molecular and cellular mechanisms associated with cellular senescence and aging

#### Mechanisms of aging

Telomere attrition <sup>243</sup>
Cumulative DNA damage <sup>244</sup>
Impairment of DNA repair <sup>245</sup>
Epimutations in nuclear DNA <sup>246</sup>
Mutations in mitochondrial DNA <sup>112</sup>
Increased rigidity of cytoskeleton <sup>247</sup>
Increased cross-linking of extracellular matrix <sup>248</sup>
Protein damage <sup>249,250</sup>
Increased production of free radicals <sup>251</sup>
Accumulation of waste products <sup>252</sup>

For each of the listed factors, a reference is given that provides either a comprehensive overview or an experimental example.

risk associated with the generation of replacement cells from aged, potentially damaged progenitors. This survival strategy, however, may still come at the price of impaired organ function.

## Aging as a contributor to chronic disease

Age represents a risk factor for the development of many diseases, including cardiovascular and metabolic disorders. Moreover, if a chronic disease is already established, it constitutes a factor contributing to mortality, e.g., in COPD and chronic hypercapnic respiratory failure.<sup>17</sup>

Aging comprises the accumulation of damage from exogenous causes as well as intrinsic, systemic susceptibilities determining the responses. These factors also determine its relation to the development of chronic diseases. Aging is known to be involved in neurodegenerative disorders<sup>18</sup> and cardiovascular diseases in which accelerated vascular aging and senescence of endothelial cells<sup>19</sup> seem to play a role through both telomere-dependent (see below) and -independent mechanisms.<sup>20</sup> Osteoarthritis<sup>21</sup> and the impact of aging on bone marrow-related therapies<sup>22</sup> are further examples. However, time-dependent losses might occur dissociated from age, as demonstrated for periodontal disease as a chronic, age-related disorder,<sup>23</sup> again emphasizing the concept of biological *versus* chronological age.

For a variety of diseases, mechanistic evidence already exists which indicates a link to (induced) aging. Moreover, patients with diabetic nephropathy showed signs of aging of skin fibroblasts.<sup>24</sup> In COPD, some of the comorbidities<sup>25</sup> can be considered as consequences of the lung disease, whereas others might be based on common susceptibility traits and linked to aging. COPD itself is a risk factor for other disorders including cardiovascular disease,<sup>26</sup> type II diabetes,<sup>27</sup> or cognitive and functional deteriorations,<sup>28</sup> all of which are age-related. Especially the association between arterial stiffness, osteoporosis and the severity of airflow obstruction<sup>29</sup> has provided supportive evidence on premature aging in COPD.<sup>30</sup> Moreover, reductions in lung function have been shown to be associated with systemic inflammation *per se* and thus potentially with aging, in addition to smoking.<sup>31</sup>

## Aging and COPD

Structure and function of the human lung show a variety of alterations as part of the normal aging process (Table 2).<sup>32,33</sup> Of particular interest seems to be the rarification of alveolar structures that is known to occur in older never-smokers.<sup>34</sup> Although the structural changes of the senile lung<sup>32</sup> are considered to be nondestructive<sup>35</sup> and are rather homogeneous compared to the more focal alterations in emphysema, the overall result appears to be similar with regard to the loss of tissue renewal and regenerative potential. Additionally, it should be noted that diffuse (senile) emphysema is difficult to diagnose by lung function indices and its occurrence rate might therefore be underestimated.

Genetically modified mice have provided additional support for a relationship between aging and emphysema, while at the same time demonstrating differences between phenotypes of the disease, e.g., homogeneous *versus* focal alterations.<sup>36</sup> The

**Table 2** Changes in structure and function of the human lung and respiratory system that occur with age

## Age-related alterations of the lung

Rarification of alveolar architecture/enlargement of air spaces
Vascular remodeling
Altered composition of extracellular matrix
Reduced strength of respiratory muscles
Impaired respiratory mechanics/increased stiffness of chest wall
Reduction of lung function reserves (volumes, flows)
Heterogeneity of ventilation
Impaired gas exchange capacity

Detailed accounts of the functional aspects can be found in excellent review articles.<sup>32,33</sup>

parallel between both processes has been emphasized before,<sup>37</sup> in view of defects in vascular maintenance in patients with emphysema<sup>38</sup> and genetically modified rodents. When interpreting the findings, particularly animal data, it seems prudent to keep in mind that aging is a multi-faceted process involving many intimately interwoven factors (Table 1) and that it might not be evaluated from a single aspect.<sup>37,39</sup>

The major extrinsic factor in COPD is smoking, promoting alterations of tissue and organ architecture that resemble those of aging. Prominent manifestations are cardiovascular or cerebrovascular diseases<sup>40</sup> and premature skin aging<sup>41,42</sup> as compared to normal<sup>41</sup> or UV-induced aging.<sup>43</sup> Skin aging includes skin wrinkling<sup>44</sup> which, interestingly enough, has been reported to exhibit a weak but significant association with pulmonary emphysema.<sup>45</sup> It also involves an increased proportion of elastic fibers,<sup>46,47</sup> associated with lung function impairment.<sup>48</sup> Intriguingly, there are associations between COPD and periodontitis,<sup>49</sup> the latter also being a disease of connective tissue mediated by inflammation and promoted by smoking, as in COPD. The association between periodontitis and age *per se* is not particularly strong,<sup>50</sup> but this seems to be analogous to the discrete alterations occurring in the senile lung in the absence of noxious agents.

Cultured lung parenchyma fibroblasts from patients with emphysema also showed elevated expression of the senescence-associated  $\beta$ -galactosidase (SA- $\beta$ -gal) compared to control smokers.<sup>51</sup> SA- $\beta$ -gal, a common marker of cellular senescence,<sup>52</sup> is regularly expressed in senescent cells, though probably as an indicator of stress in general.<sup>53</sup> Corresponding to this finding, *in vitro* exposure of human cells to cigarette smoke extract led to increased expression of SA- $\beta$ -gal,<sup>54,55</sup> similar to exposure of primary lung fibroblasts.<sup>56</sup> Available data suggest that cellular senescence is limited to lung fibroblasts and not present in skin fibroblasts of patients with lung emphysema,<sup>57</sup> which might underline the importance of local exposure levels.

In addition, proliferation rate and capacity, as major markers of senescence, were reduced in parenchymal lung fibroblasts of patients with emphysema compared to control smokers, although cells were grown under standardized conditions in the absence of inflammation.<sup>51,58,59</sup> While in these experiments the culture medium contained serum and thus a mixture of growth factors, the response

to defined stimulation by TGF- $\beta$  and IL-1 $\beta$  was also found to be altered in emphysema.<sup>60</sup> Fibroblasts also showed a dysregulation of decorin production,<sup>61</sup> a molecule involved in collagen assembly and related to aging.<sup>62</sup>

## Mechanisms of cellular senescence and evidence in COPD

### Telomere loss

For protective purposes, the ends of chromosomes carry noncoding DNA repeats called telomeres. In each DNA reduplication, about 35–100 base pairs are lost (end replication problem) implying a countdown mechanism and loss of replicative potential as telomeres shorten. Telomere shortening is a major determinant of cellular senescence, although currently it is not clear whether by the shortest telomere or mean telomere length.<sup>63</sup>

Many studies have demonstrated that telomere length of human skin fibroblasts<sup>64</sup> and blood leukocytes<sup>65</sup> decreases with age, with considerable variability inter- and intra-individually<sup>66–68</sup> and within cell populations.<sup>69</sup> The rate of telomere erosion can also differ between organs or cell populations, such as lymphocytes,<sup>70</sup> as well as between males and females.<sup>67</sup> Since telomere length appears to be a heritable trait,<sup>71</sup> it could be a key factor for the individual rate of aging and the disposition to develop age-related diseases.<sup>72</sup> Importantly, short telomeres can limit tissue renewal capacity<sup>73</sup> and thus are likely to affect the maintenance of organs. Based on this, telomeres rank among the most suitable markers of biological age, which integrate both intrinsic and extrinsic aging.

In addition to the end replication problem and sporadic telomere deletion, telomeres can be directly damaged by free radicals that target G-triplets<sup>74</sup> and induce single strand breaks. Apparently, oxidants enhance telomere loss primarily during mitosis<sup>75</sup>; it is not clear whether this also occurs in the absence of cell division.

Immortalized cells often, though not always, show an increase in the activity of telomerase, a ribonucleoprotein that can restore telomeres.<sup>76,77</sup> Alternative mechanisms of telomere lengthening (ALT) exist, and segments of telomeres can be copied from neighboring DNA strands.<sup>77,78</sup> Moreover, proteins involved in chromosome recombination were detected in telomerase-negative tumor cells, which obviously utilized this mechanism to gain unlimited replicative capacity. Conversely, senescent fibroblasts often though not always show shortened telomeres.<sup>79</sup>

Regarding their association with disease, telomere length in blood leukocytes has been found to be related to disease activity or chronicity e.g., in kidney diseases<sup>12</sup> including chronic renal insufficiency.<sup>80,81</sup> Their dysfunction also seems to be a predisposing factor for renal cancer.<sup>82</sup> Telomere dysfunction may also affect the immune system<sup>83</sup> and thus have implications beyond a single organ. It is also closely linked to other mechanisms that control cellular aging (see below). As a result of premature aging induced by external factors, telomere length can be reduced in response to accumulated stress, as shown for blood leukocytes and the oxidative stress of smoking,<sup>84</sup> or chronic psychological stress.<sup>85</sup>



With specific regard to COPD, telomere length in alveolar type II cells and endothelial cells *in situ* has been found to differ between emphysema and control patients.<sup>86</sup> In contrast, cultured parenchymal lung fibroblasts from patients with emphysema did not show altered telomere lengths despite unequivocal signs of cellular senescence.<sup>51</sup> Thus, as a point calling for methodological caution, different mechanisms of senescence could be active in the majority of cells found in histological sections of lung parenchyma as opposed to cells obtained by outgrowth cultures from such samples. Hence, the contribution of local or cell-specific telomere shortening to senescence in emphysema is currently difficult to quantify. It is also unknown whether telomere shortening is directly due to cellular stress and to what extent it is a consequence of increased cellular turnover due to inflammatory processes.

### Inflammation, role of proteases and oxidative stress, mitochondrial dysfunction

According to the common view, protease-antiprotease imbalances are involved in the development of COPD.<sup>2,3,87</sup> In line with this, smokers show upregulation of the fibroblast collagenase MMP-1 in the skin<sup>88</sup> which can mediate the degradation of interstitial collagen. In addition to the observations in patients, exposures of animals and cell cultures provided valuable insights. The upregulation of MMP-1 could be mimicked by *in vitro* exposure to cigarette smoke<sup>89,90</sup> or UV light, inducing a senescent cellular phenotype.<sup>91</sup> Cigarette smoke elicited further effects in cultured human lung fibroblasts, such as an increase in the activity of MMP-2,<sup>92</sup> as well as induction of cyclooxygenase-2 and microsomal prostaglandin E<sub>2</sub> synthase.<sup>93</sup> It also affected fibroblasts in terms of their ability to contract<sup>94</sup> and deteriorated epithelial cell repair capacity,<sup>95</sup> leading to the hypothesis that a disturbance in repair underlies the development of emphysema.<sup>96</sup> There is a close link to aging in which prostanoids<sup>97,98</sup> are involved, as well as MMPs which are implicated, e.g., in the age-related remodeling of vascular walls.<sup>99</sup> Age also implies a general decrease in the ability for tissue repair, as demonstrated, in liver regeneration for example.<sup>100</sup>

In addition to protease-antiprotease imbalance, oxidative stress originating from reactive oxygen species (ROS) is believed to drive chronic obstruction and emphysematous changes.<sup>87,101</sup> Oxidants arise from cigarette smoke and from inflammatory cells which might be additionally stimulated by recurrent respiratory tract infections.<sup>102</sup> From *in vitro* exposures, it is well established that oxidants such as H<sub>2</sub>O<sub>2</sub> can induce proliferative senescence in fibroblasts,<sup>103,104</sup> which is, however, not necessarily driven by telomeres.<sup>105</sup> Parallel results have been obtained for cigarette smoke exposure, showing a reduction in proliferation rate or capacity as one requisite of cellular senescence.<sup>106</sup> This can be induced *in vitro* by continuous or repeated,<sup>54</sup> or even a single, temporary exposure of human primary lung fibroblasts.<sup>56</sup> Hence, there is evidence from various sources that cigarette smoke-induced effects observed *in vitro* or *in vivo* resemble those of aging. Moreover, cigarette smoke might exert parallel effects in different organs, possibly on the basis of an intrinsic susceptibility that differs

between individuals, since only a minority of smokers develop clinically relevant emphysema.

ROS or ultraviolet (UV) radiation are known to particularly affect DNA integrity<sup>107</sup> and DNA damage signaling cascades.<sup>108</sup> They can also induce multiple other changes,<sup>109</sup> including cellular reprogramming and epigenetic mechanisms. For example, oxidative stress can induce cellular senescence via forkhead box O (FOXO) transcription factors and the deacetylase SIRT1, but the balance in this response can also be turned towards apoptosis<sup>110</sup> (see below). Notably, systemic inflammation and oxidant-antioxidant imbalance could favor changes in cellular phenotype throughout the organism, implying impaired maintenance in more than one organ.

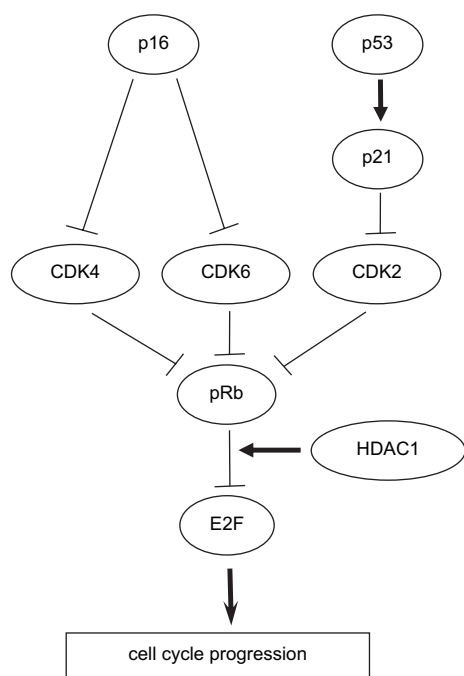
Mitochondrial dysfunction involves the production of ROS within the respiratory chain. These can directly damage proteins, RNA, and genomic or mitochondrial DNA,<sup>111</sup> which is generally considered an important contributor to aging.<sup>112</sup> Moreover, mitochondria exert indirect effects on cell survival, e.g., by mediating apoptosis. Though considered particularly important in neurodegenerative disorders, they are also probably involved in the aging of other organs, and accumulation of mutations in mitochondrial DNA leads to a decline in respiratory chain function.<sup>113</sup>

It is, however, not fully clear to what extent the loss of mitochondrial fidelity is causative for aging and to what extent it results from a decline in other functions. A causal contribution is suggested by the existence of mouse strains bearing mutations of mitochondrial DNA and showing the phenotype of premature aging. Currently, there seem to be no detailed data regarding mitochondrial dysfunction in COPD. Interestingly, *in vitro* exposure of lung epithelial cells to the supernatants from senescent lung fibroblasts reduced their protective capacity against mitochondrial dysfunction and increased ROS production.<sup>114</sup> This suggests that senescent cells can exert detrimental effects on other cells, a finding which is underscored, for example, by the observation that senescent fibroblasts can enhance the formation of tumors in mice.<sup>115</sup>

### Major cell cycle regulators

The progress of the cell cycle is basically controlled via cyclins, cyclin-dependent kinases (CDK) and their inhibitors, as well-known from tumor biology. Senescent dermal fibroblasts, and even more so lung fibroblasts, exhibit increased expression of the CDK inhibitor p16<sup>INK4a</sup> and the effector protein pRb. During the normal cell cycle, p16<sup>INK4a</sup> seems to serve as a constant braking mechanism.<sup>116</sup> Prior to senescence, cells exhibit an increase in p21<sup>Cip1/Waf1</sup> expression, another important CDK inhibitor. This is controlled by the tumor suppressor and transcription factor p53, which plays a major role in the induction of cellular senescence (Fig. 2). When reaching senescence, p21<sup>Cip1/Waf1</sup> expression decreases, while that of p16<sup>INK4a</sup> increases. In accordance with this, primarily p16<sup>INK4a</sup> and its pathway are considered to be responsible for the final irreversible proliferation stop.<sup>117,118</sup>

Thus, p21<sup>Cip1/Waf1</sup> can initiate senescence – primarily telomere-dependent – which is then maintained and established by p16<sup>INK4a</sup>. Additionally, pRb is involved in the



**Figure 2** Links between major cell cycle regulators involved in cellular senescence, with emphasis on cyclin-dependent kinases (CDK) and their inhibitors such as p16 and p21. Details on the multiple relationships and an explanation of abbreviations can be found in the text.

control of genes responsible for cell cycle progression and other functions, by recruitment of the histone deacetylase (HDAC) 1<sup>119</sup> and HDAC complexes<sup>120</sup> (see below). This is apparently related to heterochromatin formation as an epigenetic mechanism that permanently suppresses crucial growth-promoting genes.<sup>121</sup> Conversely, suppression of p16<sup>INK4a</sup> expression can increase proliferative capacity, provided that cells contain functional p16<sup>INK4a</sup>.<sup>122</sup> Further regulators of p16<sup>INK4a</sup>, e.g., transcription factors,<sup>122</sup> can act directly on the p16<sup>INK4a</sup> promoter.

Another protein controlling mitotic activity and encoded by the gene locus *INK4a*, ARF (p14<sup>ARF</sup>), can prevent p53 degradation and act as a link between diverse pathways of senescence. Accordingly, enhanced expression of p16<sup>INK4a</sup> and p14<sup>ARF</sup> was found in aging mammals including humans,<sup>123</sup> e.g., in pancreas islets, kidney, spleen, skin and lung.<sup>121</sup> While playing a central role in senescence induction, *INK4a* itself is subject to various control mechanisms, and there are links to metabolic status, since caloric restriction (see below) could, e.g., increase the expression of another CDK inhibitor, p27<sup>kip1</sup>.<sup>124</sup> The involvement of p16<sup>INK4a</sup> and p53 in maintaining senescence is underscored by the observation that pre-malignant lesions show a loss of these and other senescence markers when turning into malignancy.<sup>125</sup>

CDK inhibitors might well be involved in the development of cellular senescence in emphysema. Evidence on this has been provided by the observation that endothelial cells and alveolar type II cells from patient lungs showed increased expression of p16<sup>INK4a</sup> and p21<sup>Cip1/Waf1</sup>. Moreover, p16<sup>INK4a</sup> expression was opposite to that of the proliferation marker PCNA.<sup>86</sup> These markers were also linked to the

impairment of lung function, as a cumulative result of destruction and remodeling. Similar evidence arose from exposures of mice *in vivo* and human cells *in vitro* to cigarette smoke causing increased expression of p21<sup>Cip1/Waf1</sup>.<sup>55</sup> The expression of p16<sup>INK4a</sup> and p21<sup>Cip1/Waf1</sup> could also be increased by cigarette smoke extract in a human fibroblast cell line.<sup>54</sup> Such effects might, however, not be specific to cigarette smoke, since e.g., alveolar epithelial type II cells of rats exposed to bleomycin also exhibited signs of cellular senescence, e.g., p21<sup>Cip1/Waf1</sup> and SA- $\beta$ -gal expression.<sup>126</sup> It might be of interest that induced cellular senescence, probably by reperfusion ischemia, can also affect the function of transplanted organs,<sup>127,128</sup> and increased expression of p16<sup>INK4a</sup> has been observed in transplanted kidneys.<sup>129</sup> Cellular senescence seems to be a part of many disorders, as underlined by the fact that bleomycin is established for the induction of fibrotic lesions but not emphysema.<sup>130</sup> It might be speculated that an overshooting injury repair after bleomycin originates from a subpopulation of resistant cells which lack control by the other, now senescent, cells.

This also underlines a further important issue in studying cellular senescence, namely the heterogeneity of cell populations.<sup>131</sup> Single cell analysis has revealed that cell populations exhibit a broad range of activities at a single time point. Critical decisions, such as between apoptosis and senescence, depend on a balance between opposing factors that is likely to result in stochastic behavior of cells within the population (see, e.g., the discussion on FOXOs below). Even more importantly, the known heterogeneity of pulmonary fibroblasts<sup>132</sup> is likely to imply different responses to environmental stress.<sup>133</sup> It is therefore not unexpected that emphysema, as lung fibrosis, often starts as a spatially heterogeneous disorder which only attains a more homogeneous pattern of damage in later stages.<sup>134</sup> Heterogeneity in early stages might also have implications for regenerative therapy by being linked to the reversibility of structural changes.

### Role of PI3K and mTOR

Throughout a wide range of cell types, including pulmonary cells, phosphatidylinositol 3-kinase (PI3K) is one of the key regulators of survival and mitosis. PI3K has stimulated much interest, particularly in the understanding of tumor development. It controls longevity and robustness by phosphorylating the proapoptotic enzyme Bax.<sup>135,136</sup> Correspondingly, apoptosis is inducible via inhibition of the PI3K/Akt pathway,<sup>137</sup> which is mediated by FOXO type transcription factors (see below). A further level of control is exerted by the proapoptotic Bad,<sup>138</sup> a relative of Bax, which is influenced by cytokines such as GM-CSF and TNF- $\alpha$ . Interestingly, in lung epithelial cells PI3K could be activated by low concentrations of nicotine,<sup>139</sup> and the concomitant anti-apoptotic effect might be linked to malignant neoplasia. Importantly, PI3K is antagonized by the phosphatase PTEN, acting as a tumor suppressor.<sup>140</sup>

PI3K plays a central role in the increase in cell size that typically precedes mitosis, and in cell cycle initiation.<sup>141</sup> Among the various effects of PI3K<sup>142</sup> those mediating the combination of cellular hypertrophy and blocked mitotic activity appear to be especially relevant for senescent cells.<sup>143</sup> In the absence of mitogenic signals cellular growth

leads to cellular hypertrophy.<sup>144</sup> With regard to PI3K, the relation between senescence, aging and longevity is extremely complex and might depend on the species studied.<sup>145</sup> It is, however, clear that cell cycle arrest and a senescent phenotype can be induced by PI3K inhibitors.<sup>146,147</sup>

The Janus-faced position of PI3K in the control of mitosis is evident in the distinction between its anti-apoptotic action potentially entailing abnormal mitotic activity and tumorigenesis, and the induction of cellular senescence as characterized by abolished mitotic activity. Thus, switching of PI3K from metastable cellular states to opposing directions could also provide a link between senescence and tumorigenesis, particularly since cell populations are probably more heterogeneous in the disease rather than the healthy state.<sup>131</sup> This also appears interesting in view of the association between emphysema and lung cancer.<sup>148</sup>

Moreover, since PI3K is activated via the insulin receptor,<sup>149</sup> it offers a direct link to cell metabolism and nutrition which are known to be relevant factors in emphysema and COPD.<sup>150</sup> Interaction between the insulin pathway and PI3K or their homologs is essential for metabolic homeostasis.<sup>151,152</sup> PI3K also mediates mechanisms by which insulin adjusts the activity of FOXO transcription factors<sup>153</sup> (see below), thereby affecting senescence induction on many levels.<sup>154</sup>

A further important player in growth and cell cycle control is the kinase mTOR.<sup>155</sup> It is related to CDK inhibitors (see above), and its inhibition causes increased expression of p16<sup>INK4a</sup>.<sup>156</sup> Of particular interest among its multiple functions seem to be its action on adipocyte differentiation via the transcription factor PPAR- $\gamma$ <sup>157</sup> and its involvement in cellular stress responses including ROS.<sup>158</sup> These mechanisms could provide links between local lung disease and systemic alterations such as cachexia, a common phenomenon in emphysema.<sup>159</sup> Blocking of mTOR has been proposed as a therapeutic option to attenuate age-related malfunction,<sup>160</sup> but at present it is not clear whether this is feasible in emphysema, or whether interference with such a central enzyme results in significant negative side-effects.

## Epigenetic mechanisms and senescence

Epigenetics is increasingly recognized as a key to the understanding of gene-environment interactions, including the ontogenetic and (partially) transgenerational memory of gene expression patterns. The impact of epigenetic mechanisms on the senescent phenotype and altered regulation of mitotic activity has not yet been illuminated in much detail, but current evidence suggests that this regulation might, at least in part, rely on such mechanisms.

In its proper sense, the term "epigenetics" designates heritable changes in gene expression without changes in DNA sequence. In a broader sense, it is often used to describe a variety of regulatory mechanisms, basically including histone modifications and DNA methylation irrespective of their degree of heritance. While DNA sequences are essentially identical in all cells of an individual, patterns of epigenetic modification occur in multiple variants, some of them short-lived, some of them long-lived, e.g., in the determination of cell type or in imprinting.<sup>161</sup> Much research is currently devoted to a detailed understanding of these modes of control and the corresponding epigenetic codes.<sup>162</sup>

There are different types and levels of epigenetic control. On its lowest level, DNA methylation represents a mechanism of gene expression regulation<sup>163</sup> that is thought to be particularly important in tumor cells characterized by virtually unlimited mitotic capacity. Correspondingly, in lung tumors suppression of p16<sup>INK4a</sup> expression (see above) via methylation of the promoter DNA is regularly found.<sup>164</sup> DNA methylation and histone modification are intimately intertwined.<sup>165</sup> For example, silencing of p16<sup>INK4a</sup> could be reversed through cooperative action between histone deacetylase (HDAC) inhibitors and inhibitors of DNA methylation,<sup>166</sup> and cell proliferation could be antagonized.<sup>167</sup>

DNA is arranged on nucleosomes in a form resembling a series of pearls on a bead. The building blocks of nucleosomes are histones, while the entire chromatin scaffold comprises many other proteins. Each nucleosome is an octamer assembled from histones H2A, H2B, H3 and H4. The DNA double-helix is wound around this histone core, and nucleosomes are connected by so-called "linker DNA." The tails of the histones that protrude from the globular nucleosome can be altered by post-translational modifications, such as acetylation, methylation, phosphorylation, and their multiple combinations.<sup>168</sup> Their complex local state controls the binding of regulator proteins and the accessibility of DNA for transcription, probably being the key for short- and long-term gene regulation.<sup>169</sup>

For example, acetylation of lysine residues in histone tails generally facilitates the access to DNA, while deacetylation corresponds to a nonaccessible, silenced state. Specific enzymes mediate the different modifications, such as histone acetyltransferases (HAT) and HDACs which can partially control each other. According to the present view, histone acetylation represents a dynamic balance between HATs and HDACs, while it seems that certain methylations have the greatest potential for mediating persistent alterations.<sup>170</sup>

The involvement of histone modifications in induced senescence is currently not fully understood. Links are provided by the finding that the expression of p16<sup>INK4a</sup> and p21<sup>Cip1/Waf1</sup> is, at least partially, controlled through histone acetylation within promoter regions.<sup>171,172</sup> For example, increased p21<sup>Cip1/Waf1</sup> transcription through HDAC inhibition is linked to increased H3 acetylation in that region.<sup>173–175</sup> Such effects are thought to play a role in the anti-tumor effects of HDAC inhibitors, which are currently under investigation.<sup>176</sup>

In addition to nucleosomes, there are further levels of control, such as chromatin remodeling machines and complexes,<sup>177</sup> which can interact with HAT complexes to mediate gene expression.<sup>178</sup> This also seems to play a role in senescence by controlling both p16<sup>INK4a</sup> and p21<sup>Cip1/Waf1</sup> expression regardless of telomere shortening.<sup>178,179</sup> Regulation is also mediated by histone variants<sup>180</sup> substituting for other histones. All of these factors contribute to the partitioning between transcriptionally silent heterochromatin and transcriptionally competent, though not necessarily active, euchromatin. Silencing of DNA in cellular senescence can even be recognized macroscopically in terms of condensed, senescence-associated heterochromatic foci (SAHF) which comprise genes relevant for proliferation.<sup>181</sup>

HDAC inhibitors arrest the cell cycle not as a result of telomere shortening or uncapping. Their effect requires

functional p16<sup>INK4a</sup>, but is hardly influenced by p53.<sup>182</sup> Possibly, stabilization of DNA damage signals underlies this arrest. The reason is that telomere uncapping occurs physiologically in every mitosis, where it might induce transient activation of DNA damage signals. These are controlled by histone acetylation and could be perpetuated in the presence of HDAC inhibitors. In accordance with this, cells often resume proliferation after removal of the HDAC inhibitor.

Since various agents including cigarette smoke can induce a persistent cell cycle arrest associated with increased acetylation, the high degree of reversibility of acetylation calls for further lock-like mechanisms. While *in vivo* persistence might be supported through chronic inflammation, this would not explain increased acetylation in cultured cells outside the inflammatory environment.<sup>183</sup> It is currently not clear according to which timeframes senescence-relevant modifications of histones take place and which of them survive the disassembly of nucleosomes upon DNA replication, thus being transmitted to daughter cells.

Irrespective of these uncertainties, the involvement of epigenetic memory mechanisms is strongly suggested by the persistently altered gene expression profiles of experimental animals exposed to cigarette smoke<sup>184</sup> as well as airway of epithelial cells of ex-smokers.<sup>185</sup> Correspondingly, elevated global acetylation of H3 has been detected in the lung epithelium of ex-smokers with COPD and of H4 in current smokers,<sup>186</sup> specifically covering the IL-8 promoter region in COPD.<sup>187</sup> The same is true for cultured lung fibroblasts of patients with emphysema.<sup>183</sup> These findings are supplemented by data from rats after cigarette smoke exposure<sup>188</sup> and from cultured human lung fibroblasts studied immediately after *in vitro* exposure to cigarette smoke extract,<sup>183</sup> although the acetylation had disappeared 2 days after exposure. At present, the observed patterns of histone modifications do not allow safe conclusions as to whether they represent an intermediate state and cellular senescence is maintained by other mechanisms, or whether they basically underlie the persistence of senescence.

### Sirtuins and FOXOs as integrators of multiple pathways

Within the group of HDACs, sirtuins maintain a special position since they are structurally different from other HDACs and are inhibited by different compounds.<sup>189</sup> Sirtuins act on histones similarly to other HDACs thereby mediating gene silencing. Of particular interest in mammals is SIRT1 with its yeast homolog Sir2 that is known to deacetylate defined lysines in histones for gene silencing.<sup>190,191</sup> Importantly, SIRT1 also targets other proteins than histones, especially transcription factors. It can, for example, down-regulate p53-mediated senescence via deacetylation of p53.<sup>192</sup> Such effects seem relevant, since depletion of the pool of renewable cells by p53-mediated apoptosis and senescence could contribute to organismal aging.<sup>193</sup> Deactivation of p53 by Sir2 was also capable of raising cellular resistance to oxidative stress. However, cells without functional p53 still showed increased resistance after Sir2 activation, e.g., by resveratrol.<sup>194,195</sup> Obviously, there are several different mechanisms of stress response regulation controlled by sirtuins.

Sirtuin action depends on NAD<sup>+</sup> and is therefore coupled to the energy or redox state. This link might be relevant for emphysema, which is associated with metabolic imbalances. In addition to its supply function, NADH plays major regulatory roles. This is reflected in the fact that its energy is not needed for deacetylation, but probably spent for chromatin remodeling.<sup>196</sup> At present, the question remains open whether NAD<sup>+</sup> itself is decisive in the control of sirtuins, as studies have not detected appreciable changes in the NAD<sup>+</sup>/NADH ratio in senescent cells or cells studied under various stress conditions.<sup>191</sup> Nicotinamide, a source of NAD<sup>+</sup> metabolism, is an inhibitor of Sir2, suggesting that possibly the overall put-through rate of NAD<sup>+</sup> processing is crucial.<sup>189</sup>

Among the most noteworthy recent findings appears to be the link between SIRT1 and FOXO transcription factors which are key players in the determination of cell fate. Depending on the type of activation, FOXOs can exert diverse, even opposite, effects including induction of cell cycle arrest, cell differentiation, removal of ROS, activation of DNA repair, and induction of apoptosis.<sup>197,198</sup> FOXOs are negatively regulated by PI3K (see above), which activates the kinase Akt that phosphorylates FOXOs. This is accompanied by the export of FOXOs from the nucleus and loss of their transcriptional activity.<sup>199</sup> SIRT1 can reduce FOXO3a and FOXO4 activity and their ability to induce apoptosis.<sup>198</sup> It intervenes with FOXO3a on different levels, either indirectly suppressing the induction of proapoptotic genes,<sup>197</sup> or directly by deacetylating histones at the promoter regions of FOXO3a target genes.<sup>198</sup> At the same time SIRT1 can inhibit growth through p27<sup>kip1</sup> (see above) in connection with FOXO-induced stress responses, such as induction of manganese superoxide dismutase.<sup>195</sup> For this reason, SIRT1 may occupy a critical position in switching stress responses executed by FOXOs, ranging from upregulation of cellular defense through cell cycle arrest towards eventual apoptosis.<sup>197</sup> Pathways involving critical, possibly stochastic, decisions between divergent outcomes are of particular interest in COPD when considering the heterogeneity of the disease within the lung and between patients, as well as the systemic manifestations.

Recent data have indicated a role for SIRT1 in the regulation of inflammatory cells, in particular macrophages, when exposed to components of cigarette smoke<sup>200</sup> and SIRT1 activity in lung tissue has been found to be reduced in COPD and smokers.<sup>201</sup> Thus sirtuins, particularly SIRT1, are of great interest not only as general integrators of aging, longevity, stress responses, metabolic state, insulin signaling, and epigenetic mechanisms but also specifically due to their altered expression in COPD. Moreover, their regulation by specific compounds – e.g., resveratrol<sup>202</sup> – might provide an opportunity to move cells, and possibly organisms, into a state of higher stress resistance and increased lifespan. The multiple links also suggest that sirtuins may be effective candidates for mediating the relationship between lung disorder and systemic alterations, which are of major clinical importance.<sup>203</sup>

### Links to energy metabolism/caloric restriction

The term “caloric restriction” designates reduced energy intake without malnutrition, a condition of great interest as



it is linked to longevity, aging and disease development or progression. At least in humans, however, the underlying molecular network is still far from being elucidated in sufficient detail to draw safe conclusions. Caloric restriction elicits multiple changes in terms of a regulated response of cellular metabolism and signaling. Among other effects, it leads to an increase in SIRT1 activity<sup>204</sup> and attenuates insulin or insulin-like growth factor (IGF-1) signaling, while promoting longevity in many species including probably primates.<sup>205,206</sup> These effects are intimately linked to the action of FOXO transcription factors, PI3K and Akt (see above, Fig. 3).<sup>207</sup> Conversely, enhanced insulin signaling is thought to be associated with aging.<sup>208</sup>

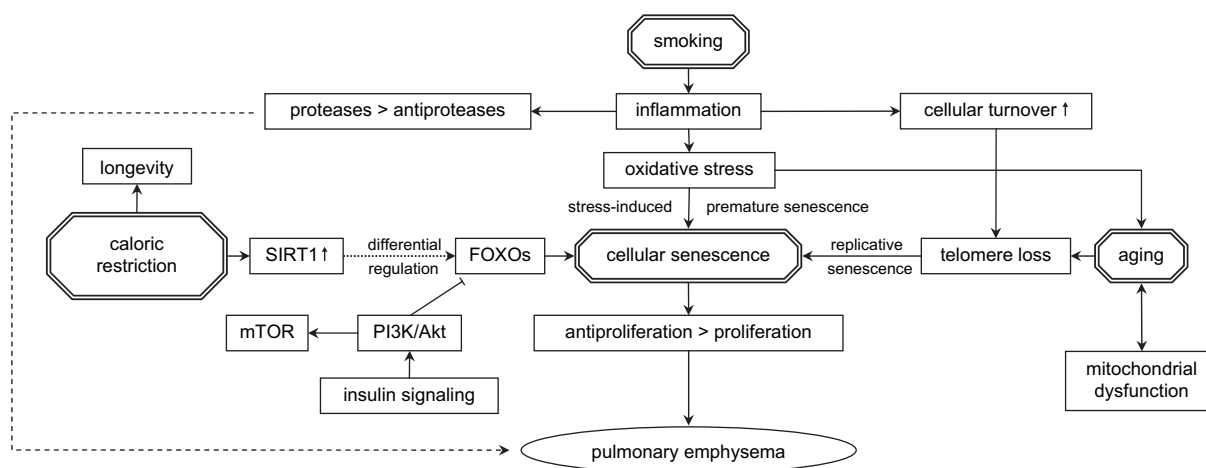
On the cellular level, caloric restriction is capable of antagonizing the loss of replicative capacity that occurs with increasing age both *in vivo* and *in vitro*.<sup>209,210</sup> Moreover, it can provide cellular protection by interfering with basic mechanisms involved in senescence, such as oxidative damage, telomere shortening, and changes in the hormone system.<sup>211</sup> However, the question, whether and under which conditions the effects of caloric restriction can enhance instead of attenuate those of cigarette smoke, has not yet been clarified.

Despite its ability to increase lifespan, caloric restriction has long been known to lead to alveolar rarification within a short time in rodents,<sup>212,213</sup> although it is not fully clear to what extent this is equivalent to diffuse lung emphysema.<sup>213</sup> Remarkably, the findings in animals are in agreement with the CT-morphological rarification of pulmonary tissue reported in young patients with *Anorexia nervosa*.<sup>214,215</sup> Despite unresolved issues regarding the interpretation of this observation,<sup>216</sup> it remains challenging, since it does not easily fit into the concept of the inflammatory origin of emphysema. Even more interesting is that the emphysema-like alterations induced by caloric restriction may disappear in mice within a short time after refeeding.<sup>217</sup>

The fact that caloric restriction can lead to reversible emphysema in rodents emphasizes the distinction between temporary, potentially reversible and chronic, in all likelihood, irreversible effects. The similarity – at least superficially –

between pulmonary alterations in hunger and cigarette smoke-induced emphysema raises the question about shared *versus* divergent pathways. In humans, there are currently no comparative data on this; results from mice suggest that at least elastase- and hunger-induced emphysema bear different biochemical and functional characteristics.<sup>218</sup> Intriguingly, this difference parallels that between cellular quiescence, as a reversible state of cell arrest, and cellular senescence, understood as an irreversible state. One of the most challenging issues appears to be whether the quiescent state associated with enhanced stress resistance can be converted into senescence by an unfortunate combination of external factors including inflammation. Possibly, such conversion occurs in emphysema, thus turning a somewhat beneficial response into a deleterious one. In addition, it might be that the inevitable chronological aging facilitates the process, as indicated by the observation that clinically relevant emphysema is generally a disease of the elderly.

Although the progression of emphysema in humans seems to be accelerated under malnutrition,<sup>219</sup> the predictive power of body weight for mortality in severe COPD<sup>17,220</sup> primarily reflects the overall physical reserves. The same is true for fat-free mass,<sup>221</sup> a phenomenological measure of lung function.<sup>224</sup> Cachexia is often viewed as a consequence of systemic processes including TNF- $\alpha$ -driven inflammation,<sup>225</sup> not as a factor which itself might promote lung destruction. In advanced disease, it is certainly extremely difficult to disentangle the threads corresponding to cause and effect. In this regard, it is, however, striking that smokers with slightly elevated body weight have been reported to exhibit a lower relative risk for developing COPD.<sup>226</sup> Energy depletion can be caused by starving or by wasting, and energy consumption might be raised in smokers, at least during light activity.<sup>227</sup> The evidence on a raised total energy expenditure in COPD is mixed, but the wasting of skeletal muscles and other tissues is likely to induce an imbalance between energy intake and need.<sup>228</sup> Sirtuins are more likely to be induced by starving than by wasting. Under caloric restriction, a reduction in cell cycle-inhibiting compounds could counteract an extrinsically induced senescence but



**Figure 3** Diagram providing details on the links between important regulators of cellular senescence. Information on specific interactions and an explanation of abbreviations are given in the text.

this mechanism seems to be secondary in the lung,<sup>123</sup> which might therefore be especially susceptible to the effects of malnutrition.

Converse to caloric restriction, in overweight persons accelerated aging and shortened telomeres have been described,<sup>229</sup> potentially indicating an elevated rate of cell turnover. Among the factors linked to the metabolic (catabolic *versus* anabolic) state, especially in aging and disease, are dehydroepiandrosterone sulphate (DHEAS) and cortisol levels.<sup>230,231</sup> Particularly for DHEAS, multiple associations with longevity and age-related changes have been reported.<sup>232,233</sup> The interplay between nutrition, body weight, aging and disease risk also seems to be manifest in the serum level of leptin, a multipotent adipokine.<sup>234</sup> Adipokines are of interest with regard to their pleiotropic effects beyond a single organ, and it is meanwhile recognized that adipose tissue can influence lifespan and aging through cell non-autonomous regulation.<sup>235</sup> Noteworthy is that the differentiation and function of adipocytes is partially controlled by SIRT1,<sup>236</sup> and that adipocyte performance might be hampered by cigarette smoke.<sup>237</sup>

## Conclusion and outlook

In recent years, aging and cellular senescence have come into focus as contributors to disease development and organ dysfunction that could provide considerable insight into the processes involved. The parallel between clinical signs of aging in smokers or patients with emphysema and findings on the cellular and biochemical level is striking, although the database is still limited. Animal experiments and *in vitro* exposure of human cells, including cells from the lung, have provided evidence that cigarette smoke compounds are capable of exerting effects that bear the signature of cellular senescence. Corresponding findings are available from immunohistological examinations of samples obtained from patients with emphysema, or from the analysis of lung cells taken in culture. The altered expression of key cell cycle regulators and mediators of growth arrest or proliferative responses, and the reduction in proliferative capacity indicate multi-faceted changes in the maintenance of cell and organ integrity. It is important to acknowledge that the notion of cellular senescence covers a multitude of causes and consequences, requiring a careful analysis in each individual condition.

Although it is impossible to draw a coherent picture at present, the multiple links between aging, longevity, stress responses, metabolic state, insulin signaling, sirtuins and epigenetic mechanisms render it likely that senescence based on these factors adds to, if not even underlies the pathogenesis of emphysema. Owing to its integrative capacity, the concept of senescence will substantially improve the understanding of the development and progression of this chronic disease. This is especially true since there seems to be a close link between senescence and epigenetic mechanisms. Such mechanisms are, among others, also of eminent importance in lung cancer, whose risk is associated with COPD. It is also worth considering the potential interference between cellular senescence and the action of pharmacological compounds. This should be one of the keys for unravelling their impact on long-term structural

changes in the lung. Recent work, for example, on the anti-inflammatory efficacy of corticosteroids in relation to gene expression control by histone acetylation<sup>238</sup> has the potential to be extended in the direction of senescence and aging. This fascinating perspective is reinforced by the fact that multipotent molecular players are involved, providing a link between environment, individual disposition, local lung disease, and systemic alterations.

These factors might also bear implications for the regenerative biology of the lung. Whether targeted interference with cell cycle and differentiation control, either by activation of resident cells or by indirect effects of stem cells, is a therapeutic option in lung emphysema remains a topic for future investigations, particularly since additional morphogenetic guidance is likely to be needed. Interventions involving epigenetic, senescence-related mechanisms might at least be an option to help slow down the disease progression, and sirtuins are particularly interesting in view of their broad networking abilities. If regenerative therapies would be feasible in future, it is likely that the (partial) reconstitution of lung architecture has to deal not only with unfavorable inflammatory, structural and mechanical<sup>239</sup> conditions in the diseased organ, but also with a potential inherent resistance of the resident cells to support reconstitution. Some of these limitations might be of evolutionary origin and manifested as genetic module-dependent ontogenetic constraints, and some might be due to (induced) cellular senescence. That regeneration in terms of recallable complex morphogenetic programs, in contrast to simple growth, is not entirely switched off in humans, is illustrated by the possibility of nearly complete restoration of fingertips in children, which also provides valuable insight into the contrast between wound healing and regeneration.<sup>240</sup> It is noteworthy that some species exhibit impressive persistent abilities of morphogenesis throughout much of their life, as demonstrated by the growth of antlers in deer, or of limbs<sup>241</sup> and even complete lung lobes in newts.<sup>242</sup> In this regard, it seems sensible to perform detailed comparisons between the morphogenetic programs and regulatory modules across species of different evolutionary position. This could provide crucial information for the induction of regeneration *in situ* in humans and for the (extracorporeal) generation of replacement organs, as well as for counteracting the constraints caused by cellular senescence.

## Conflict of interest

The authors declare that none of them has any conflict of interest related to the article or the research described.

## References

1. Rabe KF, Hurd S, Anzueto A, et al. Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease: GOLD executive summary. *Am J Respir Crit Care Med* 2007;176:532–55.
2. Barnes PJ, Shapiro SD, Pauwels RA. Chronic obstructive pulmonary disease: molecular and cellular mechanisms. *Eur Respir J* 2003;22:672–88.

3. Shapiro SD. Proteinases in chronic obstructive pulmonary disease. *Biochem Soc Trans* 2002;**30**:98–102.
4. Snider GL. Clinical relevance summary: collagen vs elastin in pathogenesis of emphysema; cellular origin of elastases; bronchiolitis vs emphysema as a cause of airflow obstruction. *Chest* 2000;**117**:244S–6S.
5. Mahadeva R, Lomas DA. Genetics and respiratory disease. 2. Alpha-1-antitrypsin deficiency, cirrhosis and emphysema. *Thorax* 1998;**53**:501–5.
6. Senior RM, Tegner H, Kuhn C, Ohlsson K, Starcher BC, Pierce JA. The induction of pulmonary emphysema with human leukocyte elastase. *Am Rev Respir Dis* 1977;**116**:469–75.
7. Wright JL, Churg A. Current concepts in mechanisms of emphysema. *Toxicol Pathol* 2007;**35**:111–5.
8. Balcombe NR, Sinclair A. Ageing: definitions, mechanisms and the magnitude of the problem. *Best Pract Res Clin Gastroenterol* 2001;**15**:835–49.
9. Johnson TE. Recent results: biomarkers of aging. *Exp Gerontol* 2006;**41**:1243–6.
10. Hayflick L. The limited in vitro lifetime of human diploid cell strains. *Exp Cell Res* 1965;**37**:614–36.
11. Hayflick L, Moorhead PS. The serial cultivation of human diploid cell strains. *Exp Cell Res* 1961;**25**:585–621.
12. Famulski KS, Halloran PF. Molecular events in kidney ageing. *Curr Opin Nephrol Hypertens* 2005;**14**:243–8.
13. Trueb RM. Aging of skin and hair. *Ther Umsch* 2005;**62**:837–46.
14. Toussaint O, Medrano EE, von Zglinicki T. Cellular and molecular mechanisms of stress-induced premature senescence (SIPS) of human diploid fibroblasts and melanocytes. *Exp Gerontol* 2000;**35**:927–45.
15. Campisi J. Senescent cells, tumor suppression, and organismal aging: good citizens, bad neighbors. *Cell* 2005;**120**:513–22.
16. Collado M, Serrano M. The senescent side of tumor suppression. *Cell Cycle* 2005;**4**:1722–4.
17. Budweiser S, Jörres RA, Riedl T, et al. Predictors of survival in COPD patients with chronic hypercapnic respiratory failure receiving noninvasive home ventilation. *Chest* 2007;**131**:1650–8.
18. Dröge W, Schipper HM. Oxidative stress and aberrant signaling in aging and cognitive decline. *Aging Cell* 2007;**6**:361–70.
19. Erusalimsky JD, Kurz DJ. Cellular senescence *in vivo*: its relevance in ageing and cardiovascular disease. *Exp Gerontol* 2005;**40**:634–42.
20. Matthews C, Gorenne I, Scott S, et al. Vascular smooth muscle cells undergo telomere-based senescence in human atherosclerosis: effects of telomerase and oxidative stress. *Circ Res* 2006;**99**:156–64.
21. Aigner T, Soder S, Gebhard PM, McAlinden A, Haag J. Mechanisms of disease: role of chondrocytes in the pathogenesis of osteoarthritis – structure, chaos and senescence. *Nat Clin Pract Rheumatol* 2007;**3**:391–9.
22. Beausejour C. Bone marrow-derived cells: the influence of aging and cellular senescence. *Handb Exp Pharmacol* 2007:67–88.
23. Ship JA, Beck JD. Ten-year longitudinal study of periodontal attachment loss in healthy adults. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 1996;**81**:281–90.
24. Morocutti A, Earle KA, Rodemann HP, Viberti GC. Premature cell ageing and evolution of diabetic nephropathy. *Diabetologia* 1997;**40**:244–6.
25. Fabbri LM, Rabe KF. From COPD to chronic systemic inflammatory syndrome? *Lancet* 2007;**370**:797–9.
26. Sin DD, Man SF. Chronic obstructive pulmonary disease as a risk factor for cardiovascular morbidity and mortality. *Proc Am Thorac Soc* 2005;**2**:8–11.
27. Rana JS, Mittleman MA, Sheikh J, et al. Chronic obstructive pulmonary disease, asthma, and risk of type 2 diabetes in women. *Diabetes Care* 2004;**27**:2478–84.
28. Ozge C, Ozge A, Unal O. Cognitive and functional deterioration in patients with severe COPD. *Behav Neurol* 2006;**17**:121–30.
29. Sabit R, Bolton CE, Edwards PH, et al. Arterial stiffness and osteoporosis in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2007;**175**:1259–65.
30. Vogelmeier C, Bals R. Chronic obstructive pulmonary disease and premature aging. *Am J Respir Crit Care Med* 2007;**175**:1217–8.
31. Gan WQ, Man SF, Sin DD. The interactions between cigarette smoking and reduced lung function on systemic inflammation. *Chest* 2005;**127**:558–64.
32. Janssens JP, Pache JC, Nicod LP. Physiological changes in respiratory function associated with ageing. *Eur Respir J* 1999;**13**:197–205.
33. Sprung J, Gajic O, Warner DO. Review article: age-related alterations in respiratory function – anesthetic considerations. [Article de synthèse: Les modifications de fonction respiratoire liées à l'âge – considerations anesthésiques]. *Can J Anaesth* 2006;**53**:1244–57.
34. Pinkerton KE, Green FHY. Normal aging of the lung. In: Harding R, Pinkerton KE, Plooper CG, editors. *The Lung: development, aging and the environment*. San Diego: Elsevier Academic Press; 2004.
35. Verbeken EK, Cauberghs M, Mertens I, Clement J, Lauweryns JM, Van de Woestijne KP. The senile lung. Comparison with normal and emphysematous lungs. 2. Functional aspects. *Chest* 1992;**101**:800–9.
36. Sato A, Hirai T, Imura A, et al. Morphological mechanism of the development of pulmonary emphysema in *kltho* mice. *Proc Natl Acad Sci USA* 2007;**104**:2361–5.
37. Tudor RM, Yoshida T, Arap W, Pasqualini R, Petrache I. State of the art. Cellular and molecular mechanisms of alveolar destruction in emphysema: an evolutionary perspective. *Proc Am Thorac Soc* 2006;**3**:503–10.
38. Kasahara Y, Tudor RM, Cool CD, Lynch DA, Flores SC, Voelkel NF. Endothelial cell death and decreased expression of vascular endothelial growth factor and vascular endothelial growth factor receptor 2 in emphysema. *Am J Respir Crit Care Med* 2001;**163**:737–44.
39. Teramoto S, Ishii M. Aging, the aging lung, and senile emphysema are different. *Am J Respir Crit Care Med* 2007;**175**:197–8 [author reply 8].
40. Bernhard D, Moser C, Backovic A, Wick G. Cigarette smoke – an aging accelerator? *Exp Gerontol* 2007;**42**:160–5.
41. Aizen E, Gilhar A. Smoking effect on skin wrinkling in the aged population. *Int J Dermatol* 2001;**40**:431–3.
42. Freiman A, Bird G, Metelitsa AI, Barankin B, Lauzon GJ. Cutaneous effects of smoking. *J Cutan Med Surg* 2004;**8**:415–23.
43. Kennedy C, Bastiaens MT, Bajdik CD, Willemze R, Westendorp RG, Bouwes Bavinck JN. Effect of smoking and sun on the aging skin. *J Invest Dermatol* 2003;**120**:548–54.
44. Koh JS, Kang H, Choi SW, Kim HO. Cigarette smoking associated with premature facial wrinkling: image analysis of facial skin replicas. *Int J Dermatol* 2002;**41**:21–7.
45. Patel BD, Loo WJ, Tasker AD, et al. Smoking related COPD and facial wrinkling: is there a common susceptibility? *Thorax* 2006;**61**:568–671.
46. Frances C, Boisnic S, Hartmann DJ, et al. Changes in the elastic tissue of the non-sun-exposed skin of cigarette smokers. *Br J Dermatol* 1991;**125**:43–7.
47. Just M, Ribera M, Monso E, Lorenzo JC, Ferrandiz C. Effect of smoking on skin elastic fibres: morphometric and immunohistochemical analysis. *Br J Dermatol* 2007;**156**:85–91.
48. Just M, Monso E, Ribera M, Lorenzo JC, Morera J, Ferrandiz C. Relationships between lung function, smoking and morphology of dermal elastic fibres. *Exp Dermatol* 2005;**14**:744–51.
49. Scannapieco FA, Ho AW. Potential associations between chronic respiratory disease and periodontal disease: analysis

- of National Health and Nutrition Examination Survey III. *J Periodontol* 2001;**72**:50–6.
50. Burt BA. Periodontitis and aging: reviewing recent evidence. *J Am Dent Assoc* 1994;**125**:273–9.
  51. Müller KC, Welker L, Paasch K, et al. Lung fibroblasts from patients with emphysema show markers of senescence *in vitro*. *Respir Res* 2006;**7**:32.
  52. Dimri GP, Lee X, Basile G, et al. A biomarker that identifies senescent human cells in culture and in aging skin *in vivo*. *Proc Natl Acad Sci USA* 1995;**92**:9363–7.
  53. Cristofalo VJ. SA beta Gal staining: biomarker or delusion. *Exp Gerontol* 2005;**40**:836–8.
  54. Nyunoya T, Monick MM, Klingelhut A, Yarovinsky TO, Cagley JR, Hunninghake GW. Cigarette smoke induces cellular senescence. *Am J Respir Cell Mol Biol* 2006;**35**:681–8.
  55. Tsuji T, Aoshiba K, Nagai A. Cigarette smoke induces senescence in alveolar epithelial cells. *Am J Respir Cell Mol Biol* 2004;**31**:643–9.
  56. Jörres RA, Kronseder A, Uhlmann S, et al. Replicative senescence of lung fibroblasts after exposure to hydrogen peroxide or cigarette smoke extract. *Eur Respir J* 2005;**26**:102s.
  57. Müller KC, Paasch K, Feindt B, et al. In contrast to lung fibroblasts – no signs of senescence in skin fibroblasts of patients with emphysema. *Exp Gerontol*; 2008.
  58. Holz O, Zühlke I, Jaksztat E, et al. Lung fibroblasts from patients with emphysema show a reduced proliferation rate in culture. *Eur Respir J* 2004;**24**:575–9.
  59. Nobukuni S, Watanabe K, Inoue J, Wen FQ, Tamaru N, Yoshida M. Cigarette smoke inhibits the growth of lung fibroblasts from patients with pulmonary emphysema. *Respirology* 2002;**7**:217–23.
  60. Noordhoek JA, Postma DS, Chong LL, et al. Different proliferative capacity of lung fibroblasts obtained from control subjects and patients with emphysema. *Exp Lung Res* 2003;**29**:291–302.
  61. Noordhoek JA, Postma DS, Chong LL, et al. Different modulation of decorin production by lung fibroblasts from patients with mild and severe emphysema. *COPD* 2005;**2**:17–25.
  62. Nomura Y. Structural change in decorin with skin aging. *Connect Tissue Res* 2006;**47**:249–55.
  63. Graakjaer J, Bischoff C, Korsholm L, et al. The pattern of chromosome-specific variations in telomere length in humans is determined by inherited, telomere-near factors and is maintained throughout life. *Mech Ageing Dev* 2003;**124**:629–40.
  64. Geigl JB, Langer S, Barwisch S, Pfliegerhaer K, Lederer G, Speicher MR. Analysis of gene expression patterns and chromosomal changes associated with aging. *Cancer Res* 2004;**64**:8550–7.
  65. Weng N. Interplay between telomere length and telomerase in human leukocyte differentiation and aging. *J Leukoc Biol* 2001;**70**:861–7.
  66. Baird DM, Rowson J, Wynford-Thomas D, Kipling D. Extensive allelic variation and ultrashort telomeres in senescent human cells. *Nat Genet* 2003;**33**:203–7.
  67. Cherif H, Tarry JL, Ozanne SE, Hales CN. Ageing and telomeres: a study into organ- and gender-specific telomere shortening. *Nucleic Acids Res* 2003;**31**:1576–83.
  68. Goronzy JJ, Fujii H, Weyand CM. Telomeres, immune aging and autoimmunity. *Exp Gerontol* 2006;**41**:246–51.
  69. Baird DM. Telomere dynamics in human cells. *Biochimie*; 2007.
  70. Mariani E, Meneghetti A, Formentini I, et al. Different rates of telomere shortening and telomerase activity reduction in CD8 T and CD16 NK lymphocytes with ageing. *Exp Gerontol* 2003;**38**:653–9.
  71. Aviv A. Telomeres, sex, reactive oxygen species, and human cardiovascular aging. *J Mol Med* 2002;**80**:689–95.
  72. Baird DM. Telomeres. *Exp Gerontol* 2006;**41**:1223–7.
  73. Hao LY, Armanios M, Strong MA, et al. Short telomeres, even in the presence of telomerase, limit tissue renewal capacity. *Cell* 2005;**123**:1121–31.
  74. Midorikawa K, Hirakawa K, Kawanishi S. Hydroxylation of deoxyguanosine at 5' site of GG and GGG sequences in double-stranded DNA induced by carbamoyl radicals. *Free Radic Res* 2002;**36**:667–75.
  75. Chen QM, Prowse KR, Tu VC, Purdom S, Linskens MH. Uncoupling the senescent phenotype from telomere shortening in hydrogen peroxide-treated fibroblasts. *Exp Cell Res* 2001;**265**:294–303.
  76. Reddel RR. Genes involved in the control of cellular proliferative potential. *Ann N Y Acad Sci* 1998;**854**:8–19.
  77. Cerni C. Telomeres, telomerase, and myc. An update. *Mutat Res* 2000;**462**:31–47.
  78. Neumann AA, Reddel RR. Telomere maintenance and cancer – look, no telomerase. *Nat Rev Cancer* 2002;**2**:879–84.
  79. Ferenac M, Polancec D, Huzak M, Pereira-Smith OM, Rubelj I. Early-senescent human skin fibroblasts do not demonstrate accelerated telomere shortening. *J Gerontol A Biol Sci Med Sci* 2005;**60**:820–9.
  80. Jimenez R, Carracedo J, Santamaria R, et al. Replicative senescence in patients with chronic kidney failure. *Kidney Int Suppl* 2005;**S11**–5.
  81. Ramirez R, Carracedo J, Soriano S, et al. Stress-induced premature senescence in mononuclear cells from patients on long-term hemodialysis. *Am J Kidney Dis* 2005;**45**:353–9.
  82. Shao L, Wood CG, Zhang D, et al. Telomere dysfunction in peripheral lymphocytes as a potential predisposition factor for renal cancer. *J Urol* 2007;**178**:1492–6.
  83. Effros RB, Dagarag M, Spaulding C, Man J. The role of CD8+ T-cell replicative senescence in human aging. *Immunol Rev* 2005;**205**:147–57.
  84. Morla M, Busquets X, Pons J, Sauleda J, MacNee W, Agusti AG. Telomere shortening in smokers with and without COPD. *Eur Respir J* 2006;**27**:525–8.
  85. Epel ES, Blackburn EH, Lin J, et al. Accelerated telomere shortening in response to life stress. *Proc Natl Acad Sci USA* 2004;**101**:17312–5.
  86. Tsuji T, Aoshiba K, Nagai A. Alveolar cell senescence in patients with pulmonary emphysema. *Am J Respir Crit Care Med* 2006;**174**:886–93.
  87. MacNee W. Pathogenesis of chronic obstructive pulmonary disease. *Proc Am Thorac Soc* 2005;**2**:258–66. discussion 290,291.
  88. Lahmann C, Bergemann J, Harrison G, Young AR. Matrix metalloproteinase-1 and skin ageing in smokers. *Lancet* 2001;**357**:935–6.
  89. Yin L, Morita A, Tsuji T. Alterations of extracellular matrix induced by tobacco smoke extract. *Arch Dermatol Res* 2000;**292**:188–94.
  90. Kim H, Liu X, Kohyama T, et al. Cigarette smoke stimulates MMP-1 production by human lung fibroblasts through the ERK1/2 pathway. *COPD* 2004;**1**:13–23.
  91. Debacq-Chainiaux F, Borlon C, Pascal T, et al. Repeated exposure of human skin fibroblasts to UVB at subcytotoxic level triggers premature senescence through the TGF-beta1 signaling pathway. *J Cell Sci* 2005;**118**:743–58.
  92. Ning W, Dong Y, Sun J, et al. Cigarette smoke stimulates matrix metalloproteinase-2 activity via EGR-1 in human lung fibroblasts. *Am J Respir Cell Mol Biol* 2007;**36**:480–90.
  93. Martey CA, Pollock SJ, Turner CK, et al. Cigarette smoke induces cyclooxygenase-2 and microsomal prostaglandin E2 synthase in human lung fibroblasts: implications for lung inflammation and cancer. *Am J Physiol Lung Cell Mol Physiol* 2004;**287**:L981–91.
  94. Carnevali S, Nakamura Y, Mio T, et al. Cigarette smoke extract inhibits fibroblast-mediated collagen gel contraction. *Am J Physiol* 1998;**274**:L591–8.



95. Wang H, Liu X, Umino T, et al. Cigarette smoke inhibits human bronchial epithelial cell repair processes. *Am J Respir Cell Mol Biol* 2001;25:772–9.
96. Rennard SI, Togo S, Holz O. Cigarette smoke inhibits alveolar repair: a mechanism for the development of emphysema. *Proc Am Thorac Soc* 2006;3:703–8.
97. Chung HY, Sung B, Jung KJ, Zou Y, Yu BP. The molecular inflammatory process in aging. *Antioxid Redox Signal* 2006;8:572–81.
98. Tang EH, Vanhoutte PM. Gene expression changes of prostanoid synthases in endothelial cells and prostanoid receptors in vascular smooth muscle cells caused by aging and hypertension. *Physiol Genomics*; 2007.
99. Greenwald SE. Ageing of the conduit arteries. *J Pathol* 2007; 211:157–72.
100. Biondo-Simoes Mde L, Matias JE, Montibeller GR, Siqueira LC, Nunes Eda S, Grassi CA. Effect of aging on liver regeneration in rats. *Acta Cir Bras* 2006;21:197–202.
101. Jörres RA, Magnussen H. Oxidative stress in COPD. *Eur Respir Rev* 1997;7:131–5.
102. Repine JE, Bast A, Lankhorst I. Oxidative stress in chronic obstructive pulmonary disease. Oxidative Stress Study Group. *Am J Respir Crit Care Med* 1997;156:341–57.
103. Chen Q, Ames BN. Senescence-like growth arrest induced by hydrogen peroxide in human diploid fibroblast F65 cells. *Proc Natl Acad Sci USA* 1994;91:4130–4.
104. Chen JH, Stoeber K, Kingsbury S, Ozanne SE, Williams GH, Hales CN. Loss of proliferative capacity and induction of senescence in oxidatively stressed human fibroblasts. *J Biol Chem* 2004;279:49439–46.
105. Matuoka K, Chen KY. Telomerase positive human diploid fibroblasts are resistant to replicative senescence but not premature senescence induced by chemical reagents. *BioGerontology* 2002;3:365–72.
106. de Magalhaes JP. From cells to ageing: a review of models and mechanisms of cellular senescence and their impact on human ageing. *Exp Cell Res* 2004;300:1–10.
107. Molho-Pessach V, Lotem M. Ultraviolet radiation and cutaneous carcinogenesis. *Curr Probl Dermatol* 2007;35:14–27.
108. Wang JY. Cellular responses to DNA damage. *Curr Opin Cell Biol* 1998;10:240–7.
109. Mahmoudi M, Mercer J, Bennett M. DNA damage and repair in atherosclerosis. *Cardiovasc Res* 2006;71:259–68.
110. Giannakou ME, Partridge L. The interaction between FOXO and SIRT1: tipping the balance towards survival. *Trends Cell Biol* 2004;14:408–12.
111. Orrenius S, Gogvadze V, Zhivotovsky B. Mitochondrial oxidative stress: implications for cell death. *Annu Rev Pharmacol Toxicol* 2007;47:143–83.
112. Huang H, Manton KG. The role of oxidative damage in mitochondria during aging: a review. *Front Biosci* 2004;9:1100–17.
113. Chan DC. Mitochondria: dynamic organelles in disease, aging, and development. *Cell* 2006;125:1241–52.
114. Bartling B, Rehbein G, Silber RE, Simm A. Senescent fibroblasts induce moderate stress in lung epithelial cells *in vitro*. *Exp Gerontol* 2006;41:532–9.
115. Krtolica A, Parrinello S, Lockett S, Desprez PY, Campisi J. Senescent fibroblasts promote epithelial cell growth and tumorigenesis: a link between cancer and aging. *Proc Natl Acad Sci USA* 2001;98:12072–7.
116. Collins CJ, Sedivy JM. Involvement of the *INK4a/Arf* gene locus in senescence. *Aging Cell* 2003;2:145–50.
117. Stein GH, Drullinger LF, Soulard A, Dulic V. Differential roles for cyclin-dependent kinase inhibitors p21 and p16 in the mechanisms of senescence and differentiation in human fibroblasts. *Mol Cell Biol* 1999;19:2109–17.
118. Brookes S, Rowe J, Gutierrez Del Arroyo A, Bond J, Peters G. Contribution of p16(*INK4a*) to replicative senescence of human fibroblasts. *Exp Cell Res* 2004;298:549–59.
119. Haddad MM, Xu W, Schwahn DJ, Liao F, Medrano EE. Activation of a cAMP pathway and induction of melanogenesis correlate with association of p16(*INK4*) and p27(*KIP1*) to CDKs, loss of E2F-binding activity, and premature senescence of human melanocytes. *Exp Cell Res* 1999;253:561–72.
120. Harbour JW, Dean DC. The Rb/E2F pathway: expanding roles and emerging paradigms. *Genes Dev* 2000;14:2393–409.
121. Sharpless NE. *Ink4a/Arf* links senescence and aging. *Exp Gerontol* 2004;39:1751–9.
122. Satyanarayana A, Rudolph KL. p16 and ARF: activation of teenage proteins in old age. *J Clin Invest* 2004;114:1237–40.
123. Krishnamurthy J, Torrice C, Ramsey MR, et al. *Ink4a/Arf* expression is a biomarker of aging. *J Clin Invest* 2004;114:1299–307.
124. Eto I. Nutritional and chemopreventive anti-cancer agents up-regulate expression of p27*Kip1*, a cyclin-dependent kinase inhibitor, in mouse JB6 epidermal and human MCF7, MDA-MB-321 and AU565 breast cancer cells. *Cancer Cell Int* 2006;6:20.
125. Collado M, Gil J, Efeyan A, et al. Tumour biology: senescence in pre-malignant tumours. *Nature* 2005;436:642.
126. Aoshiba K, Tsuji T, Nagai A. Bleomycin induces cellular senescence in alveolar epithelial cells. *Eur Respir J* 2003;22:436–43.
127. Chkhotua A, Shohat M, Tobar A, et al. Replicative senescence in organ transplantation – mechanisms and significance. *Transpl Immunol* 2002;9:165–71.
128. Chkhotua AB, Gabusi E, Altimari A, et al. Increased expression of p16(*INK4a*) and p27(*Kip1*) cyclin-dependent kinase inhibitor genes in aging human kidney and chronic allograft nephropathy. *Am J Kidney Dis* 2003;41:1303.
129. Melk A, Schmidt BM, Vongwiwatana A, Rayner DC, Halloran PF. Increased expression of senescence-associated cell cycle inhibitor p16*INK4a* in deteriorating renal transplants and diseased native kidney. *Am J Transplant* 2005;5:1375–82.
130. Moeller A, Ask K, Warburton D, Gaudie J, Kolb M. The bleomycin animal model: a useful tool to investigate treatment options for idiopathic pulmonary fibrosis? *Int J Biochem Cell Biol*; 2007.
131. Bahar R, Hartmann CH, Rodriguez KA, et al. Increased cell-to-cell variation in gene expression in ageing mouse heart. *Nature* 2006;441:1011–4.
132. Phipps RP, Penney DP, Keng P, et al. Characterization of two major populations of lung fibroblasts: distinguishing morphology and discordant display of Thy 1 and class II MHC. *Am J Respir Cell Mol Biol* 1989;1:65–74.
133. Bagloli CJ, Bushinsky SM, Garcia TM, et al. Differential induction of apoptosis by cigarette smoke extract in primary human lung fibroblast strains: implications for emphysema. *Am J Physiol Lung Cell Mol Physiol* 2006;291:L19–29.
134. Tuder RM, Yoshida T, Fijalkowka I, Biswal S, Petrache I. Role of lung maintenance program in the heterogeneity of lung destruction in emphysema. *Proc Am Thorac Soc* 2006;3:673–9.
135. Machida K, Inoue H, Matsumoto K, et al. Activation of PI3K-Akt pathway mediates anti-apoptotic effects of beta-adrenergic agonist in airway eosinophils. *Am J Physiol Lung Cell Mol Physiol* 2005;288:L860–7.
136. Xin M, Deng X. Nicotine inactivation of the proapoptotic function of Bax through phosphorylation. *J Biol Chem* 2005;280:10781–9.
137. Chen YL, Law PY, Loh HH. Inhibition of akt/protein kinase B signaling by naltrindole in small cell lung cancer cells. *Cancer Res* 2004;64:8723–30.
138. Finan PM, Thomas MJ. PI 3-kinase inhibition: a therapeutic target for respiratory disease. *Biochem Soc Trans* 2004;32:378–82.
139. Guo J, Chu M, Abbeyquaye T, Chen CY. Persistent nicotine treatment potentiates amplification of the dihydrofolate reductase gene in rat lung epithelial cells as a consequence of Ras activation. *J Biol Chem* 2005;280:30422–31.

140. Simpson L, Parsons R. PTEN: life as a tumor suppressor. *Exp Cell Res* 2001;264:29–41.
141. Garcia Z, Kumar A, Marques M, Cortes I, Carrera AC. Phosphoinositide 3-kinase controls early and late events in mammalian cell division. *Embo J* 2006;25:655–61.
142. Matuoka K, Chen KY, Takenawa T. A positive role of phosphatidylinositol 3-kinase in aging phenotype expression in cultured human diploid fibroblasts. *Arch Gerontol Geriatr* 2003;36:203–19.
143. Wang Y, Meng A, Zhou D. Inhibition of phosphatidylinositol 3-kinase uncouples H<sub>2</sub>O<sub>2</sub>-induced senescent phenotype and cell cycle arrest in normal human diploid fibroblasts. *Exp Cell Res* 2004;298:188–96.
144. Johnston GC, Pringle JR, Hartwell LH. Coordination of growth with cell division in the yeast *Saccharomyces cerevisiae*. *Exp Cell Res* 1977;105:79–98.
145. Babar P, Adamson C, Walker GA, Walker DW, Lithgow GJ. P13-kinase inhibition induces dauer formation, thermotolerance and longevity in *C. elegans*. *Neurobiol Aging* 1999;20:513–9.
146. Collado M, Medema RH, Garcia-Cao I, et al. Inhibition of the phosphoinositide 3-kinase pathway induces a senescence-like arrest mediated by p27Kip1. *J Biol Chem* 2000;275:21960–8.
147. Tresini M, Mawal-Dewan M, Cristofalo VJ, Sell C. A phosphatidylinositol 3-kinase inhibitor induces a senescent-like growth arrest in human diploid fibroblasts. *Cancer Res* 1998;58:1–4.
148. Hanaoka N, Tanaka F, Otake Y, et al. Primary lung carcinoma arising from emphysematous bullae. *Lung Cancer* 2002;38:185–91.
149. Sen P, Mukherjee S, Ray D, Raha S. Involvement of the Akt/PKB signaling pathway with disease processes. *Mol Cell Biochem* 2003;253:241–6.
150. Baldi S, Pinna GD, Crotti P, et al. Nutritional status and airflow obstruction: two independent contributors to CO diffusing capacity impairment in COPD. *Monaldi Arch Chest Dis* 2005;63:13.
151. Cheng CL, Gao TQ, Wang Z, Li DD. Role of insulin/insulin-like growth factor 1 signaling pathway in longevity. *World J Gastroenterol* 2005;11:1891–5.
152. Barbieri M, Bonafe M, Franceschi C, Paolisso G. Insulin/IGF-I signaling pathway: an evolutionarily conserved mechanism of longevity from yeast to humans. *Am J Physiol Endocrinol Metab* 2003;285:E1064–71.
153. Nakae J, Kitamura T, Ogawa W, Kasuga M, Accili D. Insulin regulation of gene expression through the forkhead transcription factor Foxo1 (Fkhr) requires kinases distinct from Akt. *Biochemistry* 2001;40:11768–76.
154. Courtois-Cox S, Genther Williams SM, Reczek EE, et al. A negative feedback signaling network underlies oncogene-induced senescence. *Cancer Cell* 2006;10:459–72.
155. De Virgilio C, Loewith R. The TOR signalling network from yeast to man. *Int J Biochem Cell Biol* 2006;38:1476–81.
156. Gao N, Flynn DC, Zhang Z, et al. G1 cell cycle progression and the expression of G1 cyclins are regulated by PI3K/AKT/mTOR/p70S6K1 signaling in human ovarian cancer cells. *Am J Physiol Cell Physiol* 2004;287:C281–91.
157. Kim JE, Chen J. Regulation of peroxisome proliferator-activated receptor gamma activity by mammalian target of rapamycin and amino acids in adipogenesis. *Diabetes* 2004;53:2748–56.
158. Corradetti MN, Guan KL. Upstream of the mammalian target of rapamycin: do all roads pass through mTOR? *Oncogene* 2006;25:6347–60.
159. Schols AM. Pulmonary cachexia. *Int J Cardiol* 2002;85:101–10.
160. Blagosklonny MV. Aging and immortality: quasi-programmed senescence and its pharmacologic inhibition. *Cell Cycle* 2006;5:2087–102.
161. McLachlan JA, Burow M, Chiang TC, Li SF. Gene imprinting in developmental toxicology: a possible interface between physiology and pathology. *Toxicol Lett* 2001;120:161–4.
162. Benecke A. Chromatin code, local non-equilibrium dynamics, and the emergence of transcription regulatory programs. *Eur Phys J E Soft Matter* 2006;19:353–66.
163. Urnov FD. Methylation and the genome: the power of a small amendment. *J Nutr* 2002;132:2450S–6S.
164. Huschtscha LI, Reddel RR. p16(*INK4a*) and the control of cellular proliferative life span. *Carcinogenesis* 1999;20:921–6.
165. Dobosy JR, Selker EU. Emerging connections between DNA methylation and histone acetylation. *Cell Mol Life Sci* 2001;58:721–7.
166. Matheu A, Klatt P, Serrano M. Regulation of the *INK4a/ARF* locus by histone deacetylase inhibitors. *J Biol Chem* 2005;280:42433–41.
167. Nishida K, Komiyama T, Miyazawa S, et al. Histone deacetylase inhibitor suppression of autoantibody-mediated arthritis in mice via regulation of p16*INK4a* and p21(*WAF1/Cip1*) expression. *Arthritis Rheum* 2004;50:3365–76.
168. Garcia BA, Shabanowitz J, Hunt DF. Characterization of histones and their post-translational modifications by mass spectrometry. *Curr Opin Chem Biol* 2007;11:66–73.
169. Jones PA, Martienssen R. A blueprint for a Human Epigenome Project: the AACR Human Epigenome Workshop. *Cancer Res* 2005;65:11241–6.
170. Kiefer JC. Epigenetics in development. *Dev Dyn* 2007;236:1144–56.
171. Magdinier F, Wolffe AP. Selective association of the methyl-CpG binding protein MBD2 with the silent p14/p16 locus in human neoplasia. *Proc Natl Acad Sci USA* 2001;98:4990–5.
172. Ocker M, Schneider-Stock R. Histone deacetylase inhibitors: signalling towards p21*cip1/waf1*. *Int J Biochem Cell Biol* 2007;39:1367–74.
173. Li H, Wu X. Histone deacetylase inhibitor, Trichostatin A, activates p21*WAF1/CIP1* expression through downregulation of c-myc and release of the repression of c-myc from the promoter in human cervical cancer cells. *Biochem Biophys Res Commun* 2004;324:860–7.
174. Rebbaa A, Zheng X, Chu F, Mirkin BL. The role of histone acetylation versus DNA damage in drug-induced senescence and apoptosis. *Cell Death Differ* 2006;13:1960–7.
175. Sambucetti LC, Fischer DD, Zabudoff S, et al. Histone deacetylase inhibition selectively alters the activity and expression of cell cycle proteins leading to specific chromatin acetylation and antiproliferative effects. *J Biol Chem* 1999;274:34940–7.
176. Marks PA, Miller T, Richon VM. Histone deacetylases. *Curr Opin Pharmacol* 2003;3:344–51.
177. Becker PB, Horz W. ATP-dependent nucleosome remodeling. *Annu Rev Biochem* 2002;71:247–73.
178. Chai J, Charboneau AL, Betz BL, Weissman BE. Loss of the hSNF5 gene concomitantly inactivates p21*CIP1/WAF1* and p16*INK4a* activity associated with replicative senescence in A204 rhabdoid tumor cells. *Cancer Res* 2005;65:10192–8.
179. Oruetxebarria I, Venturini F, Kekalainen T, et al. P16*INK4a* is required for hSNF5 chromatin remodeler-induced cellular senescence in malignant rhabdoid tumor cells. *J Biol Chem* 2004;279:3807–16.
180. Sarma K, Reinberg D. Histone variants meet their match. *Nat Rev Mol Cell Biol* 2005;6:139–49.
181. Narita M, Nunez S, Heard E, et al. Rb-mediated heterochromatin formation and silencing of E2F target genes during cellular senescence. *Cell* 2003;113:703–16.
182. Munro J, Barr NI, Ireland H, Morrison V, Parkinson EK. Histone deacetylase inhibitors induce a senescence-like state in human cells by a p16-dependent mechanism that is independent of a mitotic clock. *Exp Cell Res* 2004;295:525–38.

183. Kronseder A, Schulze C, Imhof A, et al. Effect of cigarette smoke extract (CSE) on histone acetylation of lung fibroblasts in vitro compared to histone acetylation in patients with lung emphysema. *Eur Respir J* 2006;**28**:583s.
184. Izzotti A, Cartiglia C, Longobardi M, et al. Alterations of gene expression in skin and lung of mice exposed to light and cigarette smoke. *FASEB J* 2004;**18**:1559–61.
185. Spira A, Beane J, Shah V, et al. Effects of cigarette smoke on the human airway epithelial cell transcriptome. *Proc Natl Acad Sci USA* 2004;**101**:10143–8.
186. Szulakowski P, Crowther AJ, Jimenez LA, et al. The effect of smoking on the transcriptional regulation of lung inflammation in patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2006;**174**:41–50.
187. Ito K, Ito M, Elliott WM, et al. Decreased histone deacetylase activity in chronic obstructive pulmonary disease. *N Engl J Med* 2005;**352**:1967–76.
188. Marwick JA, Kirkham PA, Stevenson CS, et al. Cigarette smoke alters chromatin remodeling and induces proinflammatory genes in rat lungs. *Am J Respir Cell Mol Biol* 2004;**31**:633–42.
189. Bitterman KJ, Anderson RM, Cohen HY, Latorre-Esteves M, Sinclair DA. Inhibition of silencing and accelerated aging by nicotinamide, a putative negative regulator of yeast sir2 and human SIRT1. *J Biol Chem* 2002;**277**:45099–107.
190. Imai S, Armstrong CM, Kaerberlein M, Guarente L. Transcriptional silencing and longevity protein Sir2 is an NAD-dependent histone deacetylase. *Nature* 2000;**403**:795–800.
191. Buck SW, Gallo CM, Smith JS. Diversity in the Sir2 family of protein deacetylases. *J Leukoc Biol* 2004;**75**:939–50.
192. Langley E, Pearson M, Faretta M, et al. Human SIR2 deacetylates p53 and antagonizes PML/p53-induced cellular senescence. *Embo J* 2002;**21**:2383–96.
193. Rodier F, Campisi J, Bhaumik D. Two faces of p53: aging and tumor suppression. *Nucleic Acids Res*; 2007.
194. Furukawa-Hibi Y, Yoshida-Araki K, Ohta T, Ikeda K, Motoyama N. FOXO forkhead transcription factors induce G(2)-M checkpoint in response to oxidative stress. *J Biol Chem* 2002;**277**:26729–32.
195. van der Horst A, Tertoolen LG, de Vries-Smits LM, Frye RA, Medema RH, Burgering BM. FOXO4 is acetylated upon peroxide stress and deacetylated by the longevity protein hSir2(SIRT1). *J Biol Chem* 2004;**279**:28873–9.
196. Moazed D. Enzymatic activities of Sir2 and chromatin silencing. *Curr Opin Cell Biol* 2001;**13**:232–8.
197. Brunet A, Sweeney LB, Sturgill JF, et al. Stress-dependent regulation of FOXO transcription factors by the SIRT1 deacetylase. *Science* 2004;**303**:2011–5.
198. Motta MC, Divecha N, Lemieux M, et al. Mammalian SIRT1 represses forkhead transcription factors. *Cell* 2004;**116**:551–63.
199. Vogt PK, Jiang H, Aoki M. Triple layer control: phosphorylation, acetylation and ubiquitination of FOXO proteins. *Cell Cycle* 2005;**4**:908–13.
200. Yang SR, Wright J, Bauter M, Seweryniak K, Kode A, Rahman I. Sirtuin regulates cigarette smoke-induced proinflammatory mediator release via RelA/p65 NF-kappaB in macrophages in vitro and in rat lungs in vivo: implications for chronic inflammation and aging. *Am J Physiol Lung Cell Mol Physiol* 2007;**292**:L567–76.
201. Rajendrasozhan S, Yang SR, Kinnula VL, Rahman I. SIRT1, an anti-inflammatory and anti-aging protein, is decreased in lungs of patients with COPD. *Am J Respir Crit Care Med*; 2008.
202. Howitz KT, Bitterman KJ, Cohen HY, et al. Small molecule activators of sirtuins extend *Saccharomyces cerevisiae* lifespan. *Nature* 2003;**425**:191–6.
203. Agustí AG. Systemic effects of chronic obstructive pulmonary disease. *Proc Am Thorac Soc* 2005;**2**:367–70 [discussion 371,372].
204. Wolf G. Calorie restriction increases life span: a molecular mechanism. *Nutr Rev* 2006;**64**:89–92.
205. Katic M, Kahn CR. The role of insulin and IGF-1 signaling in longevity. *Cell Mol Life Sci* 2005;**62**:320–43.
206. Sinclair DA. Toward a unified theory of caloric restriction and longevity regulation. *Mech Ageing Dev* 2005;**126**:987–1002.
207. Murakami S. Stress resistance in long-lived mouse models. *Exp Gerontol* 2006;**41**:1014–9.
208. Bartke A. Mini review: role of the growth hormone/insulin-like growth factor system in mammalian aging. *Endocrinology* 2005;**146**:3718–23.
209. Pendergrass WR, Li Y, Jiang D, Fei RG, Wolf NS. Caloric restriction: conservation of cellular replicative capacity in vitro accompanies lifespan extension in mice. *Exp Cell Res* 1995;**217**:309–16.
210. Wolf NS, Penn PE, Jiang D, Fei RG, Pendergrass WR. Caloric restriction: conservation of in vivo cellular replicative capacity accompanies lifespan extension in mice. *Exp Cell Res* 1995;**217**:317–23.
211. Wolf NS, Pendergrass WR. The relationships of animal age and caloric intake to cellular replication in vivo and in vitro: a review. *J Gerontol A Biol Sci Med Sci* 1999;**54**:B502–17.
212. Sahebajami H. Effects of food deprivation on experimental emphysema in obese rats. *Exp Lung Res* 1992;**18**:675–86.
213. Massaro D, Massaro GD. Hunger disease and pulmonary alveoli. *Am J Respir Crit Care Med* 2004;**170**:723–4.
214. Cook VJ, Coxson HO, Mason AG, Bai TR. Bullae, bronchiectasis and nutritional emphysema in severe anorexia nervosa. *Can Respir J* 2001;**8**:361–5.
215. Coxson HO, Chan IH, Mayo JR, Hlynsky J, Nakano Y, Birmingham CL. Early emphysema in patients with anorexia nervosa. *Am J Respir Crit Care Med* 2004;**170**:748–52.
216. Stanescu D, Pieters T. Anorexia nervosa and emphysema. *Am J Respir Crit Care Med* 2005;**172**:398 [author reply 9].
217. Massaro D, Alexander E, Reiland K, Hoffman EP, Massaro GD, Clerch LB. Rapid onset of gene expression in lung, supportive of formation of alveolar septa, induced by refeeding mice after calorie restriction. *Am J Physiol Lung Cell Mol Physiol* 2007;**292**:L1313–26.
218. Harkema JR, Mauderly JL, Gregory RE, Pickrell JA. A comparison of starvation and elastase models of emphysema in the rat. *Am Rev Respir Dis* 1984;**129**:584–91.
219. Riley DJ, Thakker-Varia S. Effect of diet on lung structure, connective tissue metabolism and gene expression. *J Nutr* 1995;**125**:1657S–60S.
220. Hallin R, Gudmundsson G, Suppli Ulrik C, et al. Nutritional status and long-term mortality in hospitalised patients with chronic obstructive pulmonary disease (COPD). *Respir Med* 2007;**101**:1954–60.
221. Vestbo J, Prescott E, Almdal T, et al. Body mass, fat-free body mass, and prognosis in patients with chronic obstructive pulmonary disease from a random population sample: findings from the Copenhagen City Heart Study. *Am J Respir Crit Care Med* 2006;**173**:79–83.
222. Kyle UG, Morabia A, Schutz Y, Pichard C. Sedentarism affects body fat mass index and fat-free mass index in adults aged 18 to 98 years. *Nutrition* 2004;**20**:255–60.
223. Kyle UG, Genton L, Gremion G, Slosman DO, Pichard C. Aging, physical activity and height-normalized body composition parameters. *Clin Nutr* 2004;**23**:79–88.
224. Wannamethee SG, Shaper AG, Whincup PH. Body fat distribution, body composition, and respiratory function in elderly men. *Am J Clin Nutr* 2005;**82**:996–1003.
225. Balasubramanian VP, Varkey B. Chronic obstructive pulmonary disease: effects beyond the lungs. *Curr Opin Pulm Med* 2006;**12**:106–12.
226. Harik-Khan RI, Fleg JL, Wise RA. Body mass index and the risk of COPD. *Chest* 2002;**121**:370–6.

227. Hultquist CM, Meyers AW, Whelan JP, Klesges RC, Peacher-Ryan H, DeBon MW. The effect of smoking and light activity on metabolism in men. *Health Psychol* 1995;14:124–31.
228. Decramer M, De Benedetto F, Del Ponte A, Marinari S. Systemic effects of COPD. *Respir Med* 2005;99(Suppl. B):S3–10.
229. Valdes AM, Andrew T, Gardner JP, et al. Obesity, cigarette smoking, and telomere length in women. *Lancet* 2005;366:662–4.
230. Debigare R, Marquis K, Cote CH, et al. Catabolic/anabolic balance and muscle wasting in patients with COPD. *Chest* 2003;124:83–9.
231. Jankowska EA, Biel B, Majda J, et al. Anabolic deficiency in men with chronic heart failure: prevalence and detrimental impact on survival. *Circulation* 2006;114:1829–37.
232. Valenti G, Denti L, Maggio M, et al. Effect of DHEAS on skeletal muscle over the life span: the InCHIANTI study. *J Gerontol A Biol Sci Med Sci* 2004;59:466–72.
233. Gleib DA, Goldman N. Dehydroepiandrosterone sulfate (DHEAS) and risk for mortality among older Taiwanese. *Ann Epidemiol* 2006;16:510–5.
234. Soares MJ, Piers LS, O’Dea K, Collier GR. Plasma leptin concentrations, basal metabolic rates and respiratory quotients in young and older adults. *Int J Obes Relat Metab Disord* 2000;24:1592–9.
235. Russell SJ, Kahn CR. Endocrine regulation of ageing. *Nat Rev Mol Cell Biol* 2007;8:681–91.
236. Picard F, Kurtev M, Chung N, et al. Sirt1 promotes fat mobilization in white adipocytes by repressing PPAR-gamma. *Nature* 2004;429:771–6.
237. Machata B, Kronseder A, Lips K, et al. Effect of cigarette smoke extract (CSE) on the differentiation and function of adipocytes. *Eur Respir J* 2006;28:585s.
238. Barnes PJ, Adcock IM, Ito K. Histone acetylation and deacetylation: importance in inflammatory lung diseases. *Eur Respir J* 2005;25:552–63.
239. Bates JH, Davis GS, Majumdar A, Butnor KJ, Suki B. Linking parenchymal disease progression to changes in lung mechanical function by percolation. *Am J Respir Crit Care Med* 2007;176:617–23.
240. Illingworth CM. Trapped fingers and amputated finger tips in children. *J Pediatr Surg* 1974;9:853–8.
241. Gilbert KA, Rannels DE. From limbs to lungs: a Newt perspective on compensatory lung growth. *News Physiol Sci* 1999;14:260–7.
242. Solopaev BP. On the problem of regeneration of the lungs in caudate amphibia. *Bull Exp Biol Med* 1957;44:1267–71.
243. Boukamp P. Ageing mechanisms: the role of telomere loss. *Clin Exp Dermatol* 2001;26:562–5.
244. von Zglinicki T, Burkle A, Kirkwood TB. Stress, DNA damage and ageing – an integrative approach. *Exp Gerontol* 2001;36:1049–62.
245. Lou Z, Chen J. Cellular senescence and DNA repair. *Exp Cell Res* 2006;312:2641–6.
246. Fraga MF, Agrelo R, Esteller M. Cross-talk between aging and cancer: the epigenetic language. *Ann N Y Acad Sci* 2007;1100:60–74.
247. Berdyeva TK, Woodworth CD, Sokolov I. Human epithelial cells increase their rigidity with ageing in vitro: direct measurements. *Phys Med Biol* 2005;50:81–92.
248. Bailey AJ. Molecular mechanisms of ageing in connective tissues. *Mech Ageing Dev* 2001;122:735–55.
249. Friguet B. Oxidized protein degradation and repair in ageing and oxidative stress. *FEBS Lett* 2006;580:2910–6.
250. Stadtman ER. Protein oxidation and aging. *Free Radic Res* 2006;40:1250–8.
251. Frisard M, Ravussin E. Energy metabolism and oxidative stress: impact on the metabolic syndrome and the aging process. *Endocrine* 2006;29:27–32.
252. Terman A, Gustafsson B, Brunk UT. Autophagy, organelles and ageing. *J Pathol* 2007;211:134–43.