Volume 236, number 1, 119–122

FEB 06177

August 1988

Effect of truncated glucagon-like peptide 1 on cAMP in rat gastric glands and HGT-1 human gastric cancer cells

A.B. Hansen⁺, C.P. Gespach, G.E. Rosselin and J.J. Holst⁺

Unité INSERM U55, Hôpital Saint-Antoine, 184, rue du Faubourg Saint-Antoine, 75571 Paris Cédex 12, France and +Institute of Medical Physiology C, The Panum Institute, Blegdamsvej 3c, DK-2200 Copenhagen N, Denmark

Received 6 June 1988

We tested the truncated 7-37 glucagon-like peptide 1 (TGLP-1), a naturally occurring porcine intestinal peptide, and other members of the glucagon family, including pancreatic glucagon (G-29), GLP-1 and GLP-2 for their ability to activate the cAMP generating system in rat gastric glands and HGT-1 human gastric cancer cells. In rat fundic glands, TGLP-1 was about 100 times more potent ($EC_{50}=2.8 \times 10^{-9}$ M) than GLP-1 of G-29, and 10 times more potent than G-29 in the HGT-1 cell line. Our results support the notion that TGLP-1 plays a direct role in the regulation of acid secretion in rat and human gastric mucosa.

Enteroglucagon; Proglucagon 78-107; cyclic AMP; (Gastric mucosa, Fundic gland)

1. INTRODUCTION

The mammalian glucagon precursor is a 180 amino acid peptide [1,2]. Besides pancreatic glucagon (G-29), glicentin (G-69) and oxyntomodulin (G-37), it contains two glucagon-like sequences, GLP-1 and GLP-2, which share 50% homology with G-29. A truncated form of GLP-1, lacking the N-terminal 6-amino acid sequence, has been isolated from the porcine intestinal mucosa [3]. This peptide (TGLP-1) is strongly insulinotropic [3,4], as opposed to GLP-1, and specific receptors for TGLP-1 have recently been demonstrated on rat insulinoma cells [5]. In man, G-29 is a potent inhibitor of meal- and pentagastrin-induced gastric acid secretion [6] and stimulates mucus secretion [7]. GLP-1 and TGLP-1 both inhibit pentagastrin-induced gastric acid secretion in man [8]. Consistent with these physiological data, G-29 has been shown to activate membrane receptors in rat gastric glands [9] and in the human gastric cancer cell line HGT-1

Correspondence address: J.J. Holst, Institute of Medical Physiology C, The Panum Institute, Blegdamsvej 3c, DK-2200 Copenhagen N, Denmark [10]. We therefore studied the effects of G-29, GLP-1, TGLP-1 and GLP-2 on the receptorcAMP systems previously evidenced in these two models.

2. MATERIALS AND METHODS

2.1. Peptides

TGLP-1, GLP-1, GLP-2 and oxyntomodulin were from Peninsula Europe (St. Helens, Merseyside, England). Crystalline, pure porcine pancreatic glucagon (G-29) was from Novo Research Institute (lot 42306, Bagsvaerd, Denmark). Synthetic porcine secretin was prepared by Professor E. Wünsch (Max-Planck-Institut für Peptidchemie, Martinsried, FRG).

2.2. Tissues

Adult male Wistar rats (200–250 g) were from our own colony. Mucosal glands were isolated from the rat fundus and antrum, using EDTA as chelator of divalent cations [11]. The HGT-1 cell line was routinely cultured, as described [12].

2.3. cAMP assay

In a standard cAMP assay, $150 \,\mu$ l from the HGT-1 cell suspension (1-2 × 10⁶ cells/ml) or from the rat gastric gland preparation (50-150 μ g cell protein/ml) was preincubated at 20°C for 10 min in 250 μ l KRP buffer containing 1% bovine serum albumin (BSA, fraction V) and IBMX as a cAMP phosphodiesterase inhibitor [11,13]. Cyclic AMP was determined by our radioimmunoassay method [14].

2.4. Calculations

The apparent EC_{50} was the concentration required to produce 50% of the maximal stimulation produced by peptides. The significance of the differences observed was assessed using Student's *t*-test.

3. RESULTS

As shown in fig.1, TGLP-1 $(10^{-10} - 10^{-8} \text{ M})$ increased cAMP production in rat fundic glands 80-fold with a potency $EC_{50} = 2.8 \pm 0.7 \times 10^{-9} M$ (n = 6). The glucagon-related peptides GLP-1 and G-29 produced similar and parallel dose-response curves at a much lower potency: $EC_{50} = 3.4 \pm$ 1.5×10^{-7} M and $2.3 \pm 0.4 \times 10^{-7}$ M, respectively (n = 6). In contrast, GLP-2 at concentrations as high as 4×10^{-7} M was ineffective in increasing basal cAMP levels $(3.5 \pm 0.8 \text{ pmol cAMP/mg pro-}$ tein, n = 6). We verified that the kinetics of cAMP generation induced by either 10^{-7} M G-29 or 10⁻⁸ M TGLP-1 were similar in rat gastric glands (not shown). The three peptides also increased cAMP generation in rat antral glands, from 4.0 \pm 0.4 to 97 \pm 10 pmol cAMP/mg protein in the presence of 10⁻⁷ M TGLP-1 (20-fold increase), to 58 ± 10 and 59 ± 14 pmol cAMP/mg protein in the presence of 10⁻⁷ M G-29 and GLP-1, respectively (n = 3-5 experiments). At maximally effective doses, secretin and TGLP-1 produced additive stimulations in rat fundic glands (fig.2). In contrast, combinations of G-29 or oxyntomodulin with TGLP-1 did not elevate significantly the cAMP values measured in the presence of TGLP-1 alone.

In HGT-1 human gastric cancer cells, TGLP-1 increased basal cAMP levels 3.7-fold, from 4.3 \pm 0.6 to 16 \pm 2.3 pmol/10⁶ cells, with EC₅₀ = 1.8 \times 10⁻⁹ M (fig.3). Pancreatic glucagon was much less potent (EC₅₀ = 1.6 \times 10⁻⁸ M) but more efficient than TGLP-1 in the system (7.5-fold increase over basal). The intact peptide GLP-1 and GLP-2 were ineffective.

4. DISCUSSION

We have shown here that TGLP-1 is the most potent glucagon-like peptide so far examined in stimulating cAMP production in rat fundic glands and the HGT-1 cell line. These peptides included G-29 [9,10], oxyntomodulin [15,16], GLP-1 and GLP-2 in the present study. TGLP-1 exhibits the same potency in the two systems, being 10 times more potent than G-29 in HGT-1 cells and 100 times more potent than intact GLP-1 and G-29 in rat fundic glands. According to its high potency in these two models, the present data suggest that TGLP-1 is the specific ligand for the glucagon-like receptor in gastric mucosa. No additive effect was



Fig.1. Effects of the glucagon-like peptides, TGLP-1 (●), G-29
(○), GLP-1 (▲) and GLP-2 (△), on cAMP generation in rat fundic glands. Cyclic AMP was determined after 1 h incubation at 20°C, in the presence of 0.5 mM IBMX [9]. The results are from one experiment, typical of 5 others. Data are means of duplicate determinations of cAMP production.



Fig.2. Effects of TGLP-1, G-29, oxyntomodulin and secretin alone (□) or in combination with TGLP-1 (S) on cAMP generation in rat fundic glands. The peptides were tested at the following concentrations: 10⁻⁸ M TGLP-1, 10⁻⁶ M G-29, 10⁻⁷ M oxyntomodulin (OX) or 10⁻⁷ M secretin (SEC). Values shown are the means ± SE of 6 experiments. Each determination was performed in duplicate.



Fig.3. Effects of the glucagon-like peptides, TGLP-1 (●), G-29
(○), GLP-1 (▲) and GLP-2 (△), on cAMP generation in HGT-1 cells. Cyclic AMP was determined after 15 min incubation at 20°C in the presence of 1 mM IBMX [13]. Values shown are the means ± SE of 5 experiments. Each determination was performed in duplicate.

found when TGLP-1 was combined with the other natural glucagon analogs, G-29 and oxyntomodulin. Our data are compatible with the hypothesis that TGLP-1 acts on the acid-secreting parietal cells in rat fundic glands: (i) the actions of TGLP-1 and secretin are additive; (ii) somatostatin selectively inhibits cAMP generation induced by TGLP-1 and histamine [9,17]. Also consistent with these results, is the observation that TGLP-1 is a potent inhibtor of in vivo pentagastrin-induced gastric acid secretion in man [8]. Truncated GLP-1 might also exert an effect on mucus secretion since glucagon-like peptides elevate cAMP generation in rat antral glands and stimulate mucus secretion from the surface epithelial cells in the fundic part of the human stomach [7]. Similar to its effect on insulin secretion [3,4], the truncated GLP-1 peptide showed a higher potency than the intact GLP-1 in rat gastric glands. Thus, from the cleavage of active peptides by proteolytic enzymes (arginine vasopressin, pancreatic glucagon and GLP-1) truncated derivatives arise which have potent and new biological activities [3,4,18,19].

The present data support the notion that TGLP-1 plays an important role in acid and mucus secretions in gastric mucosa. In this connection, it is interesting that glucagon containing A-like cells are present in the oxyntic part of the rat stomach [9], suggesting a paracrine action of glucagons, together with endocrine secretion from the distal gut [3,20]. We and other authors have observed that both histamine and glucagon-like peptides increase cAMP levels in rat parietal cells [9,21]. Since histamine and glucagons exert an opposite effect on acid secretion [6,8,15,21-23], and regarding the central role of cAMP in acid secretion [23], it is therefore likely that other intracellular messengers are involved in the inhibition of gastric acid secretion by TGLP-1. In the liver, pancreatic glucagon has been shown to produce its biological effects on two different receptor types [24]. At low concentrations, the GR-1 receptor stimulates the production of inositol phosphates $(0.25 \times$ 10^{-9} M), at higher concentrations, the GR-2 receptor activates the adenylate cyclase system ($K_a = 6.3$ $\times 10^{-9}$ M). Further studies are therefore required to delineate the actual contribution of cAMP and inositides in the biological actions of the glucagonlike peptides on gastric secretion.

REFERENCES

- Bell, G.I., Santerre, R.F. and Mullenbach, G.T. (1983) Nature 302, 716-718.
- [2] Heinrich, G., Gros, P., Lund, P.K. and Habener, J.F. (1984) J. Biol. Chem. 259, 14082–14087.
- [3] Holst, J.J., Ørskov, C., Nielsen, O.V. and Schwartz, T.W. (1987) FEBS Lett. 211, 169-174.
- [4] Moisov, S., Weir, G.C. and Habener, J.F. (1987) J. Clin. Invest. 79, 616–619.
- [5] Ørskov, C. and Nielsen, J.H. (1988) FEBS Lett. 229, 175-178.
- [6] Christiansen, J., Holst, J.J. and Kalaja, E. (1976) Gastroenterology 70, 688-692.
- [7] Stachura, J., Tarnawski, A., Bogdal, J., Krause, W. and Ivey, K. (1981) Gastroenterology 80, 474-481.
- [8] Scholdager, B., Mortensen, P.E., Christiansen, J., Ørskov, C. and Holst, J.J. (1988) Dig. Dis. Sci., in press.
- [9] Gespach, C., Bataille, D., Dutrillaux, M.C. and Rosselin, G. (1982) Biochim. Biophys. Acta 720, 7-16.
- [12] Emami, S., Chastre, E., Bodéré, H., Gespach, C., Bataille, D. and Rosselin, G. (1986) Peptides 7, 121–127.
- [11] Gespach, C., Bataille, D., Dupont, C., Rosselin, G., Wünsch, E. and Jeager, E. (1980) Biochim. Biophys. Acta 630, 433-441.
- [12] Laboisse, C.L., Augeron, C., Couturier-Turpin, M.-H., Gespach, C., Cheret, A.M. and Potet, F. (1982) Cancer Res. 42, 1541-1548.
- [13] Emami, S., Gespach, C., Forgue-Lafitte, M.E., Broer, Y. and Rosselin, G. (1983) Life Sci. 33, 415-423.
- [14] Gespach, C., Hui Bon Hoa, D. and Rosselin, G. (1983) Endocrinology 112, 1597-1606.

Volume 236, number 1

- [15] Bataille, D., Gespach, C., Tatemoto, K., Marie, J.C., Coudray, A.M., Rosselin, G. and Mutt, V. (1981) Peptides 2, 41-44.
- [16] Bataille, D., Gespach, C., Coudray, A.M. and Rosselin, G. (1981) Biosci. Rep. 1, 151-155.
- [17] Gespach, C., Hansen, A., Fagot, D., Holst, J. and Rosselin, G. (1988) Agents Actions, in press.
- [18] Burbach, J.P.H., Kovacs, G.L., De Wied, D., Van Nispen, J.W. and Greven, H.M. (1983) Science 221, 1310-1312.
- [19] Mallat, A., Pavoine, C., Dufour, M., Lotersztajn, S., Bataille, D. and Pecker, F. (1987) Nature 325, 620-622.
- [20] Ørskov, C., Holst, J.J., Knuthsen, S., Baldissera, F.G.A., Poulsen, S.S. and Nielsen, O.V. (1986) Endocrinology 119, 1467-1475.
- [21] Schepp, W. and Rouff, H.-J. (1984) Eur. J. Clin. Pharmacol. 98, 9-18.
- [22] Kirkegaard, A., Moody, A.J., Holst, J.J., Loud, F.B., Skov Olsen, P. and Christiansen, J. (1982) Nature 297, 156-157.
- [23] Gespach, C. and Emami, S. (1985) Adv. Biosci. 51, 265-273.
- [24] Wakelam, M.J., Murphy, G.J., Hruby, V.J. and Houslay, M.D. (1986) Nature 323, 68-70.