and a catalytically inactive MrpA–D sub-complex is formed in the absence of MrpE, F or G. Studies of mrpE point mutations further demonstrated the strong influence of small changes in MrpE on the antiport properties of the whole complex although MrpA and D have been hypothesized to be the actual antiporter subunits.

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S3.10 A role for uncoupling protein 1 in the formation of the mitochondrial permeability transition pore?
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The mitochondrial permeability transition pore (mPTP) is a non-specific channel that forms in the inner mitochondrial membrane in response to elevated levels of matrix calcium. The pore is generally believed to be comprised of the adenine nucleotide translocase (ANT), as well as several other mitochondrial proteins (e.g. VDAC and Cyclophilin D). Recent studies, however, indicate that the presence of ANT is not essential for permeability transition, which has led to the proposal that other members of the mitochondrial carrier protein family may be able to play a similar function to ANT in pore formation. To investigate this possibility we are studying the permeability transition properties of brown adipose tissue (BAT) mitochondria in which levels of the mitochondrial carrier protein, UCP1, can approach those of ANT. Using BAT mitochondria isolated from both wild-type and UCPTK−/− mice we assess UCP1-specific, membrane potential-independent, influences on mPTP formation by studying their swelling properties under strictly de-energised conditions. UCP1-dependent contributions to mPTP formation will, therefore, allow us to determine if the mitochondrial carrier protein involvement in permeability transition is a more general property of this family of proteins or is more likely to be restricted to specific members such as the ANT.

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S3.11 The mechanism of transport across the inner mitochondrial membrane: Clues from the PxW subfamily
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Mitochondrial carrier family members possess three conserved peptides that structural studies show to be linked by ionic bonds integral to opening the transport barrier into the mitochondrial matrix. These peptides contain the Px(D/E)x(R/K) motif across family members, but a small subset of these carriers substitute a πcation interaction. A mutation predicted to form an additional π-cation interaction with W142 and stabilize the transport barrier, eliminated MFT function. Thus, the energetics of breakage of the transport barrier are delicate and tuned to the characteristics of the substrate.

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S3.12 The ADP/ATP carrier of the apicomplexa Cryptosporidium parvum
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The eukaryotic parasite Cryptosporidium parvum has mitochondria with reduced functions. Consistent with this notion, we have identified only eight putative transport proteins of the mitochondrial carrier family (MCF). Members of the MCF support mitochondrial metabolic energy generation, macromolecular synthesis, and amino-acid metabolism by linking biochemical pathways in the mitochondrial matrix with those in the cytosol. One of the Cryptosporidium proteins has been expressed functionally in the bacterium Lactococcus lactis and has been identified by whole cell and fused vesicle uptake studies as the ADP/ATP carrier. The transporter has a similar substrate specificity and inhibitor profile as the bovine and yeast ADP/ATP carrier. By comparative modelling, the internal cavity of the ADP/ATP carrier is more akin to the bovine ADP/ATP carrier than that of the ADP/ATP carrier of Entamoeba histolytica, which is the organism that causes amoebic dysentery.

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S3.13 Determination of the dimensions and mass of membrane proteins in detergents by size exclusion chromatography
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Determining the oligomeric state of a membrane protein is an important step in understanding its mechanism. Here, we apply a new approach based on size exclusion chromatography (SEC) of membrane proteins in the alkyl-maltoside and Cymal detergent systems. The procedures will be illustrated by using the yeast mitochondrial ADP/ATP carrier, which was shown to be monomeric in detergent rather than dimeric. Several parameters of the detergent-solubilised protein were determined, such as its molecular mass, its Stokes’ radius, its radius at the midpoint of the membrane, and its excluded volume. The procedures can be used to determine the mass and dimensions of other membrane proteins when the chromatographic behaviour in SEC is determined largely by the associated detergent micelle.

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