

## BRIEF REPORT

# Characterization of Fibroblast Growth Factor Receptor 1 in Small-Cell Lung Cancer

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**Introduction:** There remains a significant therapeutic need for small-cell lung cancer (SCLC). We and others have reported high frequency of copy number gains in cytogenetic bands encoding fibroblast growth factor receptor 1 (FGFR1) in SCLC tumors and cell lines.

**Methods:** Thirteen SCLC cell lines and 68 SCLC patient tumor samples were studied for FGFR1 amplification. Growth inhibition assays were performed using PD173074, a pan-FGFR inhibitor to determine the correlation between FGFR1 expression and drug sensitivity.

**Results:** We did not detect *FGFR1* mutations in SCLC cell lines. Focal amplification of *FGFR1* gene was found in five tumor samples (7%), with high-level focal amplification in only one tumor sample (1%). Amplification owing to polysomy of chromosome 8, where FGFR1 locates, was observed in 22 tumor samples (32%). There was no correlation between *FGFR1* gene copy number and messenger RNA expression or protein expression in SCLC cells. *FGFR* inhibitor sensitivity correlated with *FGFR1* copy number determined by real-time polymerase chain reaction assay ( $r = -0.79$ ;  $p = 0.01$ ).

**Conclusion:** *FGFR1* gene mutations and focal amplification are rare in SCLC, but polysomy of chromosome 8 is relatively common. *FGFR1* copy number gain predicts sensitivity to FGFR inhibition, and FGFR expression correlates inversely with chemosensitivity.

**Key Words:** Small-cell lung cancer, FGFR1, Focal amplification, Copy number gain.

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Small-cell lung cancer (SCLC) comprises 13% of all lung cancers.<sup>1</sup> Despite extensive research over the last two decades, treatment for SCLC has not improved significantly and prognosis remains poor.<sup>2</sup> Genetic characterization of driver mutations could potentially offer effective drug targets and novel therapies in SCLC. Although previous studies have reported recurrent genetic alterations in SCLC,<sup>3–5</sup> therapeutically tractable targets have so far been elusive.

Fibroblast growth factors and fibroblast growth factor receptors (FGFR) play essential roles in regulation of normal cell proliferation, survival, migration, and differentiation.<sup>6</sup> The FGFR tyrosine kinase family comprises four kinases: FGFR1, FGFR2, FGFR3, and FGFR4. *FGFR1* amplification has been reported in 22% of squamous cell lung carcinoma and may predict sensitivity to FGFR inhibition.<sup>7</sup> The fibroblast growth factor signaling pathway may influence growth of SCLC. FGFR inhibition impairs SCLC growth in vitro and in vivo, reduces intratumor proliferation, and increases apoptosis.<sup>8</sup> We and others have reported high copy number gains (CNGs) in cytogenetic bands encoding FGFR1 in SCLC cell lines and tumors.<sup>4,5</sup> High CNGs in some of the samples suggest a role for FGFR1 as a potential therapeutic target in a subgroup of SCLC. In this study, we sought to further characterize the role of FGFR1 in SCLC.

## PATIENTS AND METHODS

### Tumor Samples, Tissue Microarray, and Cancer Cell Lines

Thirteen SCLC cell lines and 68 SCLC tumor samples were studied. Use of human samples was approved by the National Cancer Institute (NCI) Institutional Review Board. Details of cells and tumors are described in the Supplementary Materials and Methods (Supplementary Digital Content 1, <http://links.lww.com/JTO/A534>).

### Array Comparative Genomic Hybridization Analysis

We performed array comparative genomic hybridization (aCGH) analysis on five patient samples from the NCI cohort. aCGH was performed and analyzed as described previously.<sup>5</sup> A reference genomic DNA was used for hybridization. Data analysis was performed using Nexus 4.0 software (Biodiscovery Inc., Hawthorne, CA). CNG and loss of the *FGFR1* gene were defined

if the log<sub>2</sub> ratio values of sample DNA/reference genomic DNA were more than 0.2 and less than -0.2, respectively.

### Sequencing

Exons 2 to 18 of *FGFR1* gene were sequenced in 13 SCLC cell lines as described in the Supplementary Materials and Methods (Supplementary Digital Content 1, <http://links.lww.com/JTO/A534>).

### Real-Time Polymerase Chain Reaction

The messenger RNA (mRNA) expression of *FGFR1* gene and the copy numbers of the *FGFR1* gene in SCLC cells were evaluated using the Taqman gene expression assay and Taqman copy number assay, respectively (Applied Biosystems, Foster City, CA), following manufacturer's instruction. Details of the method are described in the Supplementary Materials and Methods (Supplementary Digital Content 1, <http://links.lww.com/JTO/A534>).

### Western Blot

Western blotting was performed as previously described.<sup>9</sup> Anti-FGFR1 antibody was obtained from Abcam (Cambridge, United Kingdom), anti-p-FGFR antibody was from Cell Signaling Technology (Danvers, MA), and anti-Actin antibody was from Sigma-Aldrich (St. Louis, MO). Semiquantification of signals of Western blot was analyzed by Image J software (National Institutes of Health, Bethesda, MD). FGFR1 protein expression was normalized by actin

protein expression and then calibrated by the expression level of the H69 cells before further analysis.

### Growth Inhibition Assays

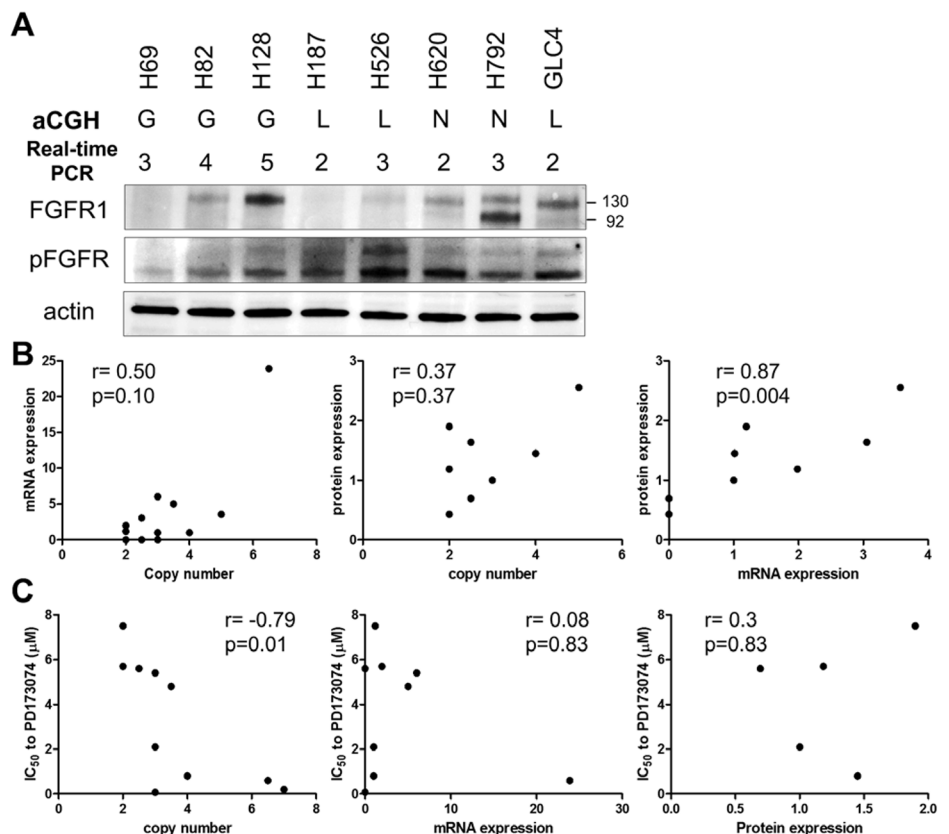
The concentration that inhibits 50% to drugs in SCLC cells was determined as described in the Supplementary Materials and Methods (Supplementary Digital Content 1, <http://links.lww.com/JTO/A534>).

### Fluorescence In Situ Hybridization for the *FGFR1* Gene

Fluorescence in situ hybridization (FISH) assays were performed on 5 μm formalin-fixed paraffin-embedded tumor sections using a laboratory standardized protocol with slight modifications.<sup>10</sup> Details of the method are described in the Supplementary Materials and Methods (Supplementary Digital Content 1, <http://links.lww.com/JTO/A534>). CNG was defined as red FGFR1 signals with the corresponding control chromosome enumeration probes (CEP8) more than or equal to four copies per cell in more than 20% of cells. Focal amplification was defined if the ratio of the *FGFR1* gene to the CEP8 was more than 2 (low-level amplification 2-4; high-level amplification >4).

### Statistical Analysis

Correlation between variables was analyzed using Spearman's method, and *p* values less than 0.05 were regarded as significant.



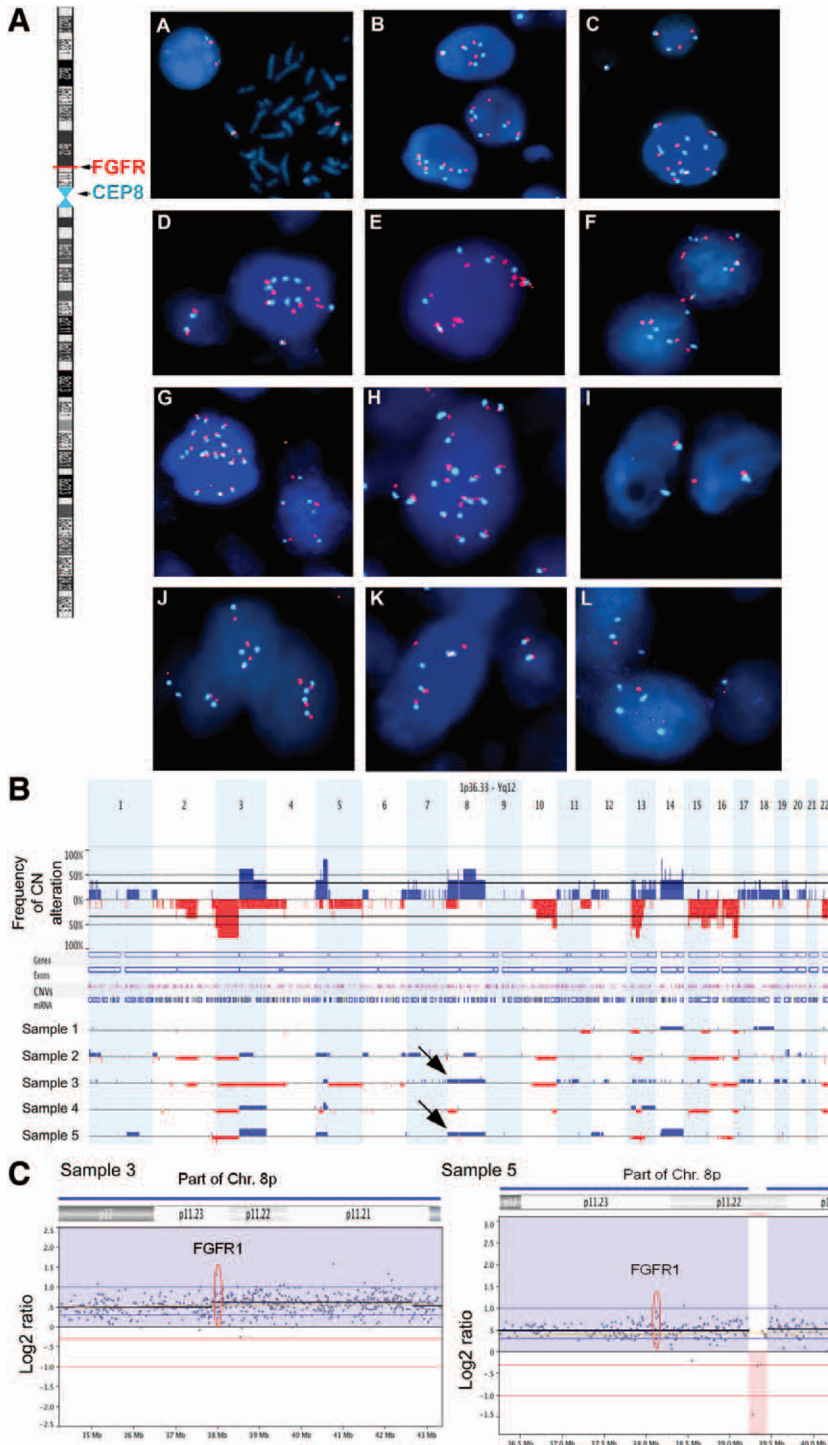
**FIGURE 1.** FGFR1 in SCLC cells. *A*, FGFR1 protein expression in SCLC cells. G, copy number gain; N, copy number normal; L, copy number loss. The numbers indicate the copy number of the *FGFR1* gene determined by the real-time PCR assay. *B*, Correlation of *FGFR1* copy number assessed by real-time PCR and mRNA expression and protein expression. *C*, Correlation of concentration that inhibits 50% to PD173074 and *FGFR1* copy number assessed by real-time PCR assay, *FGFR1* mRNA expression, or *FGFR1* protein expression. FGFR1, fibroblast growth factor receptor 1; SCLC, small-cell lung cancer; PCR, polymerase chain reaction; mRNA, messenger RNA.

## RESULTS

Expression of *FGFR1* Is Unrelated to Gene Copy Number

We previously reported genetic alterations of 13 SCLC cell lines using aCGH assay.<sup>5</sup> We further determined the copy number of the *FGFR1* gene using real-time polymerase chain reaction assay and evaluated whether the copy

number alteration of the *FGFR1* gene is associated with its protein expression (Fig. 1A). Whereas the protein expression of the *FGFR1* gene was related to its mRNA expression, protein or mRNA expression of the *FGFR1* gene in SCLC cells was unrelated to its gene copy number (Fig. 1B). As the anti-p-FGFR antibody is not *FGFR1* specific, we failed to observe correlation between expression of p-FGFR and copy number of the *FGFR1* gene ( $r = -0.19$ ,  $p = 0.66$ ), *FGFR1*



**FIGURE 2.** *FGFR1* in SCLC tumors. *A*, Dual-color fluorescence in situ hybridization showing *FGFR1* (red fluorescence signal) gene copy number variations along with the reference chromosome 8 centromeric probe (aqua fluorescence signal) in SCLC cell lines and primary tumors. Left: Ideogram of chromosome 8 with *FGFR1* gene mapped to 8p12-p11.23 chromosomal region. Normal peripheral blood lymphocytes showing two copies of *FGFR1* gene on interphase and metaphase cells. *B*, Genome-wide frequency of copy number alterations. Gains and losses are shown in blue and red, respectively. *C*, High-resolution analysis showing copy number alterations in chromosome 8p where *FGFR1* is located of patient samples 3 and 5 which had chromosome 8p polysomy. *FGFR1*, fibroblast growth factor receptor 1; SCLC, small-cell lung cancer.



mRNA expression ( $r = -0.31$ ,  $p = 0.45$ ), and FGFR1 protein expression ( $r = -0.34$ ,  $p = 0.42$ ). We sequenced the exons 2 to 18 of the *FGFR1* gene in 13 SCLC cell lines, and no mutation was detected.

### FGFR1 Inhibitor Sensitivity Is Related to CNG of the Gene

SCLC cells with higher copies of the *FGFR1* gene, detected by real-time polymerase chain reaction assay, were more sensitive to PD173074, a pan-FGFR inhibitor ( $r = -0.79$ ;  $p = 0.01$ ) (Fig. 1C). Nevertheless, we failed to observe a correlation between the sensitivity to PD173074 and FGFR1 mRNA expression ( $r = 0.45$ ;  $p = 0.13$ ) or FGFR1 protein expression (Fig. 1C). The expression of p-FGFR was unrelated to the sensitivity to PD173074 ( $r = 0.31$ ,  $p = 0.61$ ). We studied the correlation between *FGFR1* copy number (Supplementary Fig. S1A and B, Supplementary Digital Content 1, <http://links.lww.com/JTO/A534>) and FGFR1 mRNA expression (Supplementary Fig. S1C and D, Supplementary Digital Content 1, <http://links.lww.com/JTO/A534>) and sensitivity to the cytotoxic agents cisplatin and etoposide, both of which are standard therapies for SCLC. We observed that SCLC cells expressing higher FGFR1 mRNA were more resistant to etoposide ( $r = 0.68$ ;  $p = 0.01$ ) (Supplementary Fig. S1D, Supplementary Digital Content 1, <http://links.lww.com/JTO/A534>). Nevertheless, the protein expression of the *FGFR1* gene was unrelated to the sensitivities to etoposide ( $r = 0.43$ ,  $p = 0.29$ ).

### Focal Amplification of *FGFR1* Gene Was Infrequent, But CNG Was Common

We studied, using FISH, whether the CNG was due to focal amplification or to polysomy of chromosome 8. Focal amplification was observed in only five of 68 tumor samples (7%) and among those high-level focal amplification was detected only in 1% (Fig. 2A).

Nevertheless, 22 of the 68 tumor samples (32%) displayed moderate amplification ranging from 4 to 9 *FGFR1* gene copy numbers per cell, which includes the entire chromosome 8 (Table 1). The same pattern, that is, polysomy of chromosome 8, was seen in the SCLC cell lines H678 and DMS114. We performed aCGH in five samples from the NCI cohort and observed CNG of the whole chromosome 8p, implying polysomy in two samples (Fig. 2B and C). The results were consistent with what was observed by using FISH.

### DISCUSSION

Recent large-scale sequencing efforts have identified FGFR1 as a potentially therapeutically tractable target in a subgroup of SCLC. Consistent with previous reports, we found no mutations in *FGFR1* exons.<sup>11</sup> Peifer et al.<sup>4</sup> detected focal amplification of *FGFR1* locus in SCLC by copy number analysis (4 of 63, 6%) and validated the results by FISH in an independent cohort (3 of 51, 6%). Nevertheless, no amplification of the *FGFR1* locus was identified in a similar analysis of 56 cases.<sup>3</sup> Among 68

**TABLE 1.** FGFR1 Fluorescence In Situ Hybridization in the Small-Cell Lung Cancer Tumor Samples and TMA Specimens

Tumor Source	No. of Patients	FGFR1 Copy Number Gain (%)	FGFR1 Focal Amplification (%)
National Cancer Institute cohort	20	12 (60)	1 (5)
TMA	14	5 (36)	0
Multitumor block	34	5 (15)	4 (12)

FGFR1, Fibroblast growth factor receptor 1; TMA, tissue microarray.

SCLC tumor specimens, we found focal *FGFR1* amplification in 7% of the samples and true high-level amplification in only 1% of the sample. Our findings are comparable to results of large-scale integrated genomics studies where the frequency of *FGFR1* amplification ranged from 0% to 6%,<sup>3,4</sup> and mutation was absent.

We found no correlation between *FGFR1* copy number and mRNA expression and between *FGFR1* copy number and protein expression. A similar lack of correlation between FGFR1 mRNA expression and copy number of the gene was also demonstrated in squamous cell carcinomas of lung.<sup>12</sup> In SCLC cells, the lack of correlation between *FGFR1* copy number, mRNA expression, and protein expression suggests that chromosome 8 polysomy may have little impact on FGFR1 expression. Despite SCLC cells with higher copies of the *FGFR1* gene were more sensitive to PD173074, a non-isoform-specific FGFR inhibitor, we cannot exclude the possibility that other genes locating in the chromosome 8 contribute to the sensitivity to PD173074 in SCLC cells.

In conclusion, mutations or focal amplification of *FGFR1* gene are rare in SCLC. Nevertheless, *FGFR1* gene CNG is common and is related to sensitivity to FGFR inhibition. FGFR1 inhibitors should be explored in patients with SCLC with high copy numbers of the gene.

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