

Neuroendocrine Control of Female Puberty: Glial and Neuronal Interactions

Ying J. Ma and Sergio R. Ojeda

Division of Neuroscience, Oregon Regional Primate Research Center, Beaverton, Oregon, U.S.A.

Emerging evidence suggests that, in addition to neuronal inputs, growth factors of glial origin are also important in the control of mammalian puberty via a cell-cell interaction that ultimately affects the neurons that release gonadotropin-releasing hormone (GnRH), a neurohormone controlling sexual development. Among these growth factors, transforming growth factor- α (TGF α) appears to be one of the physiologic components that controls the onset of female puberty by affecting GnRH neuronal activity in a glia-mediated autocrine/paracrine manner. Specifically, TGF α induces glia to produce bioactive substances, such as prostaglandin E₂ (PGE₂). In turn, PGE₂ directly acts on GnRH neurons to stimulate the

release of GnRH. Furthermore, the neuregulin of glial origin neu differentiation factor (NDF) was found to facilitate the action of TGF α , suggesting that other growth factors may exert their biologic effects on GnRH neuronal function via a glia/neuron interaction. Another indication that glial cells may be involved in the regulation of neuroendocrine function is the presence of estrogen receptors on hypothalamic astrocytes. Thus, region-specific glial cells appear to play an integral role in the regulation of neuroendocrine function. Key words: astrocytes/growth factors/receptors/GnRH. *Journal of Investigative Dermatology Symposium Proceedings 2:19-22, 1997*

The onset of mammalian female puberty depends on the enhanced secretory activity of a specialized group of neurons that produce gonadotropin-releasing hormone (GnRH), a key neurohormone for sexual maturation and reproductive function. The regulation of GnRH neuronal function has been a focal issue for understanding the cellular and molecular mechanisms controlling sexual development and reproductive biology. It is believed that neuron to neuron transsynaptic inputs, both excitatory (Brann and Mahesh, 1994; Crowley *et al* 1995) and inhibitory (Terasawa, 1995), contribute to the regulation of GnRH secretory activity. A combination of a decrease in inhibitory tone and an increase in excitatory input has been generally considered to bring about the initiation of puberty. The main excitatory inputs include neuronal systems that utilize norepinephrine (Ramirez *et al* 1984), neuropeptide Y (Crowley *et al* 1995), and excitatory amino acids (Ojeda, 1991; Brann and Mahesh, 1994; Plant, 1994). The inhibitory components, on the other hand, are γ -aminobutyric acid (Masotto and Negro-Vilar, 1987; Terasawa, 1995) and opioid peptides (Bhanot and Wilkinson, 1983).

Although this neuronal regulation of the GnRH neuronal network is well-documented, recent studies have raised the intriguing possibility that glial cells may also play a role in the control of GnRH neuronal activity (Ojeda and Ma, 1995). Anatomical studies revealed that the glial ensheathment of GnRH cell bodies in primates is prominent, as glial processes surround the GnRH cells (Witkin *et al* 1991). Such a structural relationship suggests a direct glial influence on GnRH neuronal activity. This hypothesis is

further supported by the evidence that glial cells produce a variety of bioactive molecules and growth factors able to affect the activity of GnRH neurons. Furthermore, glial apposition to GnRH nerve terminals in the median eminence of the hypothalamus is yet another specialized structural feature in both rats and primates. The predominant cells in the median eminence are astrocytes and modified ependymal glia known as tanyocytes (Kobayashi and Matsui, 1969). Interestingly, these specialized glial cells that line the wall of the third ventricle send their processes toward the endothelial wall of the portal blood vasculature, neighboring the GnRH nerve terminals (Kobayashi and Matsui, 1969; Kozlowski and Coates, 1985). More intriguingly, sex steroids appear to modulate this structural arrangement as demonstrated by the finding that the space between the GnRH-nerve terminals and the basal lamina of the portal vasculature decreases after ovariectomy, presumably due to retraction of the glial cells. These structural changes may lead to an increase in the access of GnRH terminals to the portal vessels. The evidence that about 50% of glial cells in the median eminence have estrogen receptors (Langub and Watson, 1992) further supports the notion that a direct action of estrogen on glial cells is region-specific (Ma *et al*, 1994a). Because glial-neuronal synaptoid specializations are common structural features in the median eminence (Kobayashi and Matsui, 1969), the concept of a direct glial involvement in the control of GnRH release via cell-cell interactions with GnRH nerve terminals was proposed. In this article, we will discuss some recent results obtained by our laboratory that underscore the emerging role of glial cells in the regulation of GnRH neuronal function.

We demonstrated that glia-derived transforming growth factor alpha (TGF α), a mitogenic polypeptide member of the epidermal growth factor (EGF) family (Derynck, 1988; Carpenter and Cohen, 1990; Massague, 1990), plays an important role in the glia-neuron-mediated control of GnRH secretory activity (Ma *et al* 1992, 1994a,

Reprint requests to: Dr. Ying J. Ma, Division of Neuroscience, Oregon Regional Primate Research Center, 505 N.W. 185th Avenue, Beaverton, OR 97006.

Abbreviations: EGFR, epidermal growth factor receptor.

The Control of GnRH Neurons

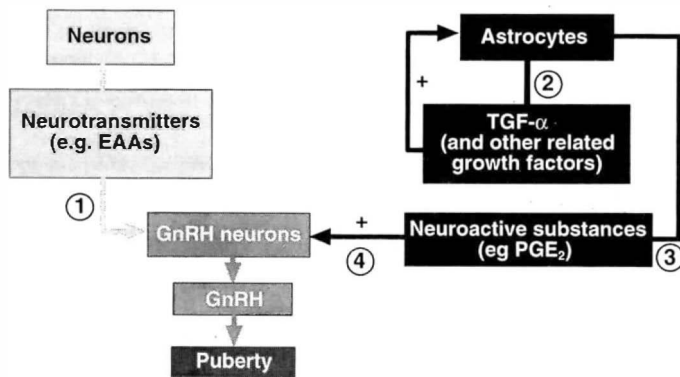


Figure 1. A simplified schematic illustration of the postulated control of GnRH neuronal function. (1) Transsynaptic inputs regulate GnRH release. For detailed description, see text. (2)–(3) Glial-derived TGF α and other related growth factors stimulate the secretion of neuroactive substances (i.e., PGE $_2$) in a paracrine/autocrine fashion. (4) Neuroactive substances act on GnRH neurons to enhance secretion. See text for further description.

1994b, 1994c, 1994d; Ojeda, 1994) (see Fig 1). Other distant members of the EGF family, neuregulins of glial origin—including heregulin (Holmes *et al*, 1992) and neu differentiation factor (Wen *et al*, 1992), the alternatively spliced products from a single gene (Wen *et al*, 1992; Marchionni *et al*, 1993;)—appear to be also involved in the regulation of GnRH release via a mechanism by which neu differentiation factor facilitates the action of TGF α (YJ Ma, DF Hill, M Costa, SR Ojeda, unpublished observation). Like other members of the EGF family, TGF α is synthesized as a transmembrane precursor. It is important to note that this precursor is able to interact with neighboring cells endowed with its receptors, without the need of releasing the mature peptide. This type of cellular action is designated “juxtacrine” (Massague, 1990) and represents an additional cellular signaling mechanism of membrane-anchored growth factors in the central nervous system.

The biologic actions of TGF α , like other members of the EGF family, are initiated by binding to a family of membrane-spanning receptors with protein kinase activity for cellular signaling (Schechter *et al*, 1985; Carpenter, 1987; Kraus *et al*, 1989; Plowman *et al*, 1990, 1993). The receptor for TGF α (as well as EGF, amphiregulins, and others) is a 170-kDa glycosylated protein known as the EGF receptor (EGFR) that has an extracellular domain containing the amino acid motif essential for specific ligand recognition, a single transmembrane domain, and an intracellular enzyme catalytic element containing the tyrosine kinase activity. Other EGFR-related members (encoded by different proto-oncogenes) including HER-2 (neu) receptor (Karunakaran *et al*, 1996), HER-3 (named ERBB-3) (Kraus *et al*, 1989; Plowman *et al*, 1990), and HER-4 (Plowman *et al*, 1993) also have similar structural features. These EGFR congeners, however, do not directly bind to TGF α , EGF, and amphiregulin, but instead recognize other members of the EGF family. Recent studies have suggested that the heterodimers (HER-2/HER-3 or HER-2/HER-4) and homodimer (HER-4/HER-4) are the functional receptor units required for the signal transduction of neuregulins (for review see Lemke, 1996). Furthermore, HER-2 can be transmodulated by the activation of EGFR (Stern and Kamps, 1988), suggesting interactions between EGFR and HER-2. This view is supported by the observation that HER-2 is able to function as an auxiliary component that enhances the affinity of EGFR by stabilizing dimerization of the receptor (Karunakaran *et al*, 1996).

In our studies of both rats (Ma *et al*, 1992) and nonhuman primates (Ma *et al*, 1994b), we found that the TGF α gene and its protein product are predominantly expressed in astroglial cells of

certain hypothalamic nuclei involved in the control of GnRH neuronal function (Ma *et al*, 1992), such as the suprachiasmatic and arcuate nucleus. Both TGF α mRNA and protein levels are also abundant in tanyocytes and glial cells of the median eminence, and the levels are significantly elevated around the time of puberty (Ma *et al*, 1992, 1994b). Furthermore, EGFR in the median eminence is colocalized with TGF α (Ma *et al*, 1994d), and—at least in monkeys (Ma *et al*, 1994b)—its developmental expression profile parallels that of TGF α , indicating the involvement of the glia-derived growth factor and its receptor in neuroendocrine regulation. That TGF α may be involved in the control of GnRH neuronal activity was demonstrated by the finding that TGF α is able to stimulate the release of GnRH from median eminence fragments in a dose-response-related manner (Ojeda *et al*, 1990). Subsequent studies further supported the notion that a TGF α /EGFR system contributes, at least in part, to the normal initiation of mammalian female puberty because overexpression of TGF α in transgenic mice induced precocious puberty (Ma *et al*, 1994c). In contrast, blockade of EGFR tyrosine kinase activity by RG-50864 (Yaish *et al*, 1988) targeted to the median eminence resulted in a delayed onset of puberty (Ma *et al*, 1992), suggesting that activation of the EGFR-mediated signaling pathway is critical for the initiation of reproductive competence. Our most recent evidence (YJ Ma, DF Hill, M Costa, SR Ojeda, unpublished data) show that glia-derived neuro differentiation factor is able to enhance the stimulatory action of TGF α on GnRH release via activation of an HER-2/HER-4 receptor complex, indicating that other members of the EGF family may also be involved in the process of sexual development. This view is further supported by the evidence (YJ Ma, DF Hill, M Costa, SR Ojeda, unpublished observation) that disruption of HER-2 synthesis via an anti-sense oligonucleotide attenuated the stimulatory effect of TGF α on GnRH release.

The effect of TGF α on GnRH secretion does not appear to be direct, as GnRH neurons do not express EGFR. The GnRH terminals in the median eminence, however, are in close association with glial cells that express TGF α and EGFR (Ma *et al*, 1992, 1994d), suggesting that these glial cells mediate the effect of TGF α . This view is supported by the recent findings (Ma *et al*, 1997) that isolated hypothalamic astrocytes respond to TGF α with the release of prostaglandin E $_2$ (PGE $_2$), a bioactive substance that stimulates GnRH release. That PGE $_2$ (although one cannot rule out other bioactive substances), in turn, acts on GnRH neurons to stimulate the release of GnRH is indicated by the ability of conditioned medium from hypothalamic astrocytes treated with TGF α to enhance GnRH release from the GnRH-producing cell line GT1-1. These cells express no EGFR and are unresponsive to TGF α itself (Voigt *et al*, 1996). Moreover, blockade of PGE $_2$ action by immunoneutralization in the conditioned medium abolished the effect of the conditioned medium on GnRH release (Ma *et al*, 1997).

The biologic actions of PGE $_2$ are initiated via the activation of its receptors, including at least four subtypes known as EP1, EP2, EP3, and EP4 (Sugimoto *et al*, 1992; Honda *et al*, 1993; Watabe *et al*, 1993). Activation of EP1 leads to increased calcium flux, whereas EP2 enhances and EP3 attenuates adenylate cyclase activity, suggesting that the diversity of actions of PGE $_2$ in different circumstances may depend on the type of PGE $_2$ receptors expressed. By ribonuclease protection assay, we observed the presence of mRNAs encoding EP1 and EP3 in the immortalized GnRH neuronal GT1-1 cell line. Utilizing *in situ* hybridization combined with immunocytochemistry, we further observed that EP1 mRNA is expressed by GnRH neurons in mouse hypothalamus (F Rage, BJ Lee, YJ Ma, SR Ojeda, unpublished data). These studies provided cellular and molecular evidence supporting the view that GnRH secretion is regulated by biomolecules produced by glial cells in response to the action of TGF α .

The importance of glial-neuronal interactions in the regulation of GnRH neuronal activity is further indicated by the finding that glutamic acid, a major excitatory amino acid neurotransmitter in the neuroendocrine brain (van den Pol *et al*, 1990), is released from astrocytes and affects neuronal activity (Parpura *et al*, 1994). We

found that glutamate increases TGF α mRNA expression in hypothalamic astroglia via activation of metabotropic glutamate receptors located on the astrocytes.¹ Thus, specific metabotropic glutamate receptor subtypes may be involved in the process by which excitatory amino acids control glial TGF α production and, hence, facilitate GnRH release. Other excitatory neurotransmitters were also shown to influence glial function (Chiu and Kriegler, 1994; Glowinski *et al.*, 1994).

It is well-documented that estradiol is important in regulating GnRH neuronal function (for review, see Ojeda and Urbanski, 1994; Kalra, 1986). The action of estradiol on GnRH release, however, is indirect, as GnRH neurons themselves do not contain detectable estrogen receptors (Shivers *et al.*, 1983), indicating that other cell types expressing the receptors mediate this effect. That hypothalamic astrocytes, in addition to other neurons, may contribute to mediating the action of estradiol is indicated by the finding that estrogen receptor mRNA was detected in hypothalamic astrocytes (Ma *et al.*, 1994a). Estradiol is able to enhance TGF α gene expression in hypothalamic, but not cerebellar (i.e., non-neuroendocrine), astrocytes (Ma *et al.*, 1994a). Moreover, the culture medium of astrocytes exposed to estradiol increases PGE₂ receptor EP1 gene expression in GT1-1 cells. No change in EP1 mRNA levels was observed, however, in GT1-1 cells directly treated with estradiol (F Rage, BJ Lee, YJ Ma, SR Ojeda, unpublished data). These findings suggest that astrocytes of the neuroendocrine brain are indeed able to mediate the effect of gonadal steroids on LHRH release.

In summary, mammalian puberty is initiated within the brain by multiple but well-coordinated events that ultimately affect the activity of the GnRH neuronal network. These events include: (i) an increase in transsynaptic excitatory inputs such as excitatory amino acids, norepinephrine, and neuropeptide Y; (ii) a decrease in the transsynaptic γ -aminobutyric acid-inhibitory regulation, and (iii) increased glial production of growth factors able to stimulate the production of neuroactive substances, such as prostaglandins. It appears that members of the EGF family, such as TGF α and its distant relatives neuregulins, are factors contributing to this glial involvement. Finally, it is of great interest to determine whether these glial influences are also essential for the regulation and maintenance of normal reproductive function during adulthood. Future experiments are needed to address this important issue.

This work was supported by National Institutes of Health Grants HD25123 and RR00163.

REFERENCES

- Bhanot R, Wilkinson M: Opiatergic control of gonadotropin secretion during puberty in the rat. *Endocrinology* 113:596-602, 1983
- Brann DW, Mahesh VB: Excitatory amino acids: function and significance in reproduction and neuroendocrine regulation. *Front Neuroendocrinol* 15:3-49, 1994
- Carpenter G: Receptors for epidermal growth factor and other polypeptide mitogens. *Annu Rev Biochem* 56:881-914, 1987
- Carpenter G, Cohen S: Epidermal growth factor. *J Biol Chem* 265:7709-7712, 1990
- Chiu SY, Kriegler S: Neurotransmitter-mediated signaling between axons and glial cells. *Glia* 11:191-200, 1994
- Crowley WR, Parker SL, Sahu A, Kalra SP: Interacting transmembrane signals regulating GnRH and LH secretion. In: Plant TM, Lee PA (eds.). *The Neurobiology of Puberty*. Journal of Endocrinology Ltd, Bristol, U.K., 1995, pp 41-54
- Derynck R: Transforming growth factor α . *Cell* 54:593-595, 1988
- Glowinski J, Marin P, Tence M, Stella N, Giaume C, Premont J: Glial receptors and their intervention in astrocyte-astrocytic and astrocyte-neuronal interactions. *Glia* 11:201-208, 1994
- Holmes WE, Sliwkowski MX, Akita RW, Henzel WJ, Lee J, Park JW, Yansura D, Abadi N, Raab H, Lewis GD, Shepard HM, Kuang W, Wood WI, Goeddel DV, Vandlen RL: Identification of heregulin, a specific activator of p185^{erbB2}. *Science* 256:1205-1210, 1992
- Honda A, Sugimoto Y, Namba T, Watabe A, Irie A, Negishi M, Narumiya S, Ichikawa A: Cloning and expression of a cDNA for mouse prostaglandin E receptor EP₂ subtype. *J Biol Chem* 268:7759-7762, 1993
- Kalra SP: Neural circuitry involved in the control of LHRH secretion: a model for preovulatory LH release. In: Ganong WF, Martini L (eds.). *Frontiers in Neuroendocrinology*. Vol. 9. Raven Press, New York, 1986, pp 31-75
- Karunakaran D, Tsahar E, Beerli RR, Chen X, Graus-Porta D, Ratzkin BJ, Seger R, Hynes NE, Yarden Y: ErbB-2 is a common auxiliary subunit of NDF and EGF receptors: implications for breast cancer. *EMBO J* 15:254-264, 1996
- Kobayashi H, Matsui T: Fine structure of the median eminence and its functional significance. In: Ganong WF, Martini L (eds.). *Frontiers in Neuroendocrinology*. Oxford University Press, New York, 1969, pp 3-46
- Kozlowski GP, Coates PW: Ependymoneuronal specializations between LHRH fibers and cells of the cerebroventricular system. *Cell Tissue Res* 242:301-311, 1985
- Kraus MH, Issing W, Miki T, Popescu NC, Aaronson SA: Isolation and characterization of ERBB3, a third member of the ERBB/epidermal growth factor receptor family: evidence for overexpression in a subset of human mammary tumors. *Proc Natl Acad Sci USA* 86:9193-9197, 1989
- Langub MC, Jr, Watson RE, Jr: Estrogen receptor-immunoreactive glia, endothelia, and ependyma in guinea pig preoptic area and median eminence: electron microscopy. *Endocrinology* 130:364-372, 1992
- Lemke G: Neuregulins in development. *Mol Cell Neurosci* 7:247-262, 1996
- Ma YJ, Junier M, Costa ME, Ojeda SR: Transforming growth factor alpha (TGF α) gene expression in the hypothalamus is developmentally regulated and linked to sexual maturation. *Neuron* 9:657-670, 1992
- Ma YJ, Berg-von der Emde K, Moholt-Siebert M, Hill DF, Ojeda SR: Region-specific regulation of transforming growth factor α (TGF α) gene expression in astrocytes of the neuroendocrine brain. *J Neurosci* 14:5644-5651, 1994a
- Ma YJ, Costa ME, Ojeda SR: Developmental expression of the genes encoding transforming growth factor alpha (TGF α) and its receptor in the hypothalamus of female rhesus macaques. *Neuroendocrinology* 60:346-359, 1994b
- Ma YJ, Dissen GA, Merlino G, Coquelin A, Ojeda SR: Overexpression of a human transforming growth factor alpha (TGF α) transgene reveals a dual antagonistic role of TGF α in female sexual development. *Endocrinology* 135:1392-1400, 1994c
- Ma YJ, Hill DF, Junier M, Costa ME, Felder SE, Ojeda SR: Expression of epidermal growth factor receptor changes in the hypothalamus during the onset of female puberty. *Mol Cell Neurosci* 5:246-262, 1994d
- Ma YJ, Berg-von der Emde K, Rage F, Wetsel WC, Ojeda SR: Hypothalamic astrocytes respond to transforming growth factor alpha with secretion of neuroactive substances that stimulate the release of luteinizing hormone-releasing hormone. *Endocrinology* 138:19-25, 1997
- Marchionni MA, Goodearl ADJ, Chen MS, Birmingham-McDonogh O, Kirk C, Hendricks M, Danehy F, Misumi D, Sudhalter J, Kobayashi K, Wroblewski D, Lynch C, Baldassare M, Hiles I, Davis JB, Hsuan JJ, Totty NF, Otsu M, McBurney RN, Waterfield MD, Stroobant P, Gwynne D: Glial growth factors are alternatively spliced erbB2 ligands expressed in the nervous system. *Nature* 362:312-318, 1993
- Masotto C, Negro-Vilar A: Activation of gamma-amino butyric acid B-receptors abolishes naloxone-stimulated luteinizing hormone release. *Endocrinology* 121:2251-2255, 1987
- Massague J: Transforming growth factor- α . A model for membrane-anchored growth factors. *J Biol Chem* 265:21393-21396, 1990
- Ojeda SR, Urbanski HF, Costa ME, Hill DF, Moholt-Siebert M: Involvement of transforming growth factor α in the release of luteinizing-hormone releasing hormone from the developing female hypothalamus. *Proc Natl Acad Sci USA* 87:9698-9702, 1990
- Ojeda SR: The mystery of mammalian puberty: how much more do we know?. *Perspect Biol Med* 34:365-383, 1991
- Ojeda SR: The neurobiology of mammalian puberty: has the contribution of glial cells been underestimated?. *J NIH Res* 6:51-56, 1994
- Ojeda SR, Ma YJ: Neuroendocrine control of female puberty: the role of brain growth factors. In: Sarkar DK, Barnes CD (eds.). *Reproductive Neuroendocrinology of Aging and Drug Abuse*. CRC Press, Boca Raton, FL, 1995, pp 41-58
- Ojeda SR, Urbanski HF: Puberty in the rat. In: Knobil E, Neill JD (eds.). *The Physiology of Reproduction*. 2nd ed. Vol 2. Raven Press, New York, 1994, pp 363-409
- Parpura V, Basarsky TA, Liu F, Jęftinija K, Jęftinija S, Haydon PG: Glutamate-mediated astrocyte-neuron signalling. *Nature* 369:744-747, 1994
- Plant TM: Puberty in primates. In: Knobil E, Neill J (eds.). *The Physiology of Reproduction*. 2nd ed. Vol 2. Raven Press, New York, 1994, pp 453-485
- Plowman GD, Whitney GS, Neubauer MG, Green JM, McDonald VL, Todaro GJ, Shoyab M: Molecular cloning and expression of an additional epidermal growth factor receptor-related gene. *Proc Natl Acad Sci USA* 87:4905-4909, 1990
- Plowman GD, Culouscou J, Whitney GS, Green JM, Carlton GW, Foy L, Neubauer MG, Shoyab M: Ligand-specific activation of HER4/p180^{erbB4}, a fourth member of the epidermal growth factor receptor family. *Proc Natl Acad Sci USA* 90:1746-1750, 1993
- Ramirez VD, Feder HH, Sawyer CH: The role of brain catecholamines in the regulation of LH secretion: a critical inquiry. In: Martini L, Ganong WF (eds.). *Frontiers in Neuroendocrinology*. Vol. 8. Raven Press, New York, 1984, pp 27-84
- Schechter AL, Hung M, Vaidyanathan L, Weinberg RA, Yang-Feng RL, Francke U, Ullrich A, Coussens L: The *neu* gene: an *erb-B*-homologous gene distinct from and unlinked to the gene encoding the EGF receptor. *Science* 229:976-978, 1985
- Shivers BD, Harlan RE, Morrell JI, Pfaff DW: Absence of oestradiol concentration in cell nuclei of LHRH-immunoreactive neurones. *Nature* 304:345-347, 1983
- Stern DF, Kamps MP: EGF-stimulated tyrosine phosphorylation of p185^{neu}: a potential model for receptor interactions. *EMBO J* 7:995-1001, 1988

¹ Ma YJ, Rage F, Ojeda SR: Regulation of transforming growth factor alpha (TGF α) gene expression in hypothalamic astrocytes by metabotropic glutamate receptors. *Soc Neurosci Abstr* 20:1070, 1994 (abstr).

- Sugimoto Y, Namba T, Honda A, Hayashi Y, Negishi M, Ichikawa A, Narumiya S: Cloning and expression of a cDNA for mouse prostaglandin E receptor EP₃ subtype. *J Biol Chem* 267:6463-6466, 1992
- Terasawa E: Mechanisms controlling the onset of puberty in primates: the role of GABAergic neurons. In: Plant TM, Lee PA (eds.). *The Neurobiology of Puberty*. Journal of Endocrinology Ltd, Bristol, U.K., 1995, pp 139-151
- van den Pol AN, Wuarin J, Dudek FE: Glutamate, the dominant excitatory transmitter in neuroendocrine regulation. *Science* 250:1276-1278, 1990
- Voigt P, Ma YJ, Gonzalez D, Fahrenbach WH, Wetsel WC, Berg-von der Emde K, Hill DF, Taylor KG, Costa ME, Seidah NG, Ojeda SR: Neural and glial-mediated effects of growth factors acting via tyrosine kinase receptors on LHRH neurons. *Endocrinology* 137:2593-2605, 1996
- Watabe A, Sugimoto Y, Honda A, Irie A, Namba T, Negishi M, Ito S, Narumiya S, Ichikawa A: Cloning and expression of cDNA for a mouse EP₁ subtype of prostaglandin E receptor. *J Biol Chem* 268:20175-20178, 1993
- Wen D, Peles E, Cupples R, Suggs SV, Bacus SS, Luo Y, Trail G, Hu S, Silbiger SM, Ben Levy R, Koski RA, Lu HS, Yarden Y: Neu differentiation factor: a transmembrane glycoprotein containing an EGF domain and an immunoglobulin homology unit. *Cell* 69:559-572, 1992
- Witkin JW, Ferin M, Popilskis SJ, Silverman A: Effects of gonadal steroids on the ultrastructure of GnRH neurons in the rhesus monkey: synaptic input and glial apposition. *Endocrinology* 129:1083-1092, 1991
- Yaish P, Gazit A, Gilon C, Levitzki A: Blocking of EGF-dependent cell proliferation by EGF receptor kinase inhibitors. *Science* 242:933-935, 1988