

offer a new perspective on (TTAGGG)_n maximum length and on the relative stability of this telomeric hexamer, both in vitro and in vivo.

Acknowledgments

We thank Drs E. Fritz, S. Meyn, F. Calafell, S. Baserga, G. Isaya, and T. Ashley for helpful discussion and advice, and we thank B. King for her gift of DNA samples from mice. This work was supported in part by National Institutes of Health grants MH44876, MH39239, and MH50390.

GIORGIO SIRUGO AND KENNETH K. KIDD

Department of Genetics
Yale University School of Medicine
New Haven

Electronic-Database Information

Accession numbers and URLs for data in this article are as follows:

Epicentre Forum, http://www.epicentre.com/f2_3/f2_3al.html

References

- Allshire RC, Dempster M, Hastie ND (1989) Human telomeres contain at least three types of G-rich repeat distributed non-randomly. *Nucleic Acids Res* 17:4611–4627
- Ashley CJ, Warren ST (1995) Trinucleotide repeat expansion and human disease. *Annu Rev Genet* 29:703–704
- Barnes WM (1994) PCR amplification of up to 35-kb DNA with high fidelity and high yield from λ bacteriophage templates. *Proc Natl Acad Sci USA* 91:2216–2220
- Baskaran N, Kandpal RP, Bhargava AK, Glynn MW, Bale A, Weissman SM (1996) Uniform amplification of a mixture of deoxyribonucleic acids with varying GC content. *Genome Res* 6:633–638
- Blasco MA, Lee HW, Hande MP, Samper E, Lansdorp PM, Depinho RA, Greider CW (1997) Telomere shortening and tumour formation by mouse cells lacking telomerase RNA. *Cell* 91:25–34
- Brown WR, MacKinnon PJ, Villasante A, Spurr N, Buckle VJ, Dobson MJ (1990) Structure and polymorphism of human telomere-associated DNA. *Cell* 63:119–132
- Bryan TM, Englezou A, Dalla-Pozza L, Dunham MA, Reddel RR (1997) Evidence for an alternative mechanism for maintaining telomere length in human tumors and tumor-derived cell lines. *Nat Med* 3:1271–1274
- Bryan TM, Englezou A, Gupta J, Bacchetti S, Reddel RR (1995) Telomere elongation in immortal human cells without detectable telomerase activity. *EMBO J* 14:4240–4248
- de Lange T (1995) Telomere dynamics and genome instability in human cancer. In: Blackburn EH, Greider CW (eds) *Telomeres*. Cold Spring Harbor Laboratory Press, New York, NY, pp 265–293
- de Lange T, Shiu L, Myers RM, Cox DR, Naylor SL, Killery AM, Varmus H (1990) Structure and variability of human chromosome ends. *Mol Cell Biol* 10:518–527
- Gordenin DA, Kunkel TA, Resnik M (1997) Repeat expansion—all in a flap? *Nat Genet* 16:116–118
- Hanish JP, Yanowitz JL, de Lange T (1994) Stringent sequence requirements for the formation of human telomeres. *Proc Natl Acad Sci USA* 91:8861–8865
- Kipling D (1995) *The telomere*. Oxford University Press, New York
- Martens UM, Zijlmans JMJM, Poon SSS, Dragowska W, Yui J, Chavez E, Ward RK, et al (1998) Short telomeres on human chromosome 17p. *Nat Genet* 18:76–80
- Meyerson M, Counter CM, Eaton EN, Ellisen LW, Steiner P, Caddle SD, Ziaugra L, et al (1997) hEST2, the putative human telomerase catalytic subunit gene, is up-regulated in tumor cells and during immortalization. *Cell* 90:785–795
- Nakamura TM, Morin GB, Chapman KB, Weinrich SL, Andrews WH, Lingner J, Harley CB, et al (1997) Telomerase catalytic subunit homologs from fission yeast and human. *Science* 277:955–959
- Notaro R, Cimmino A, Tabarini D, Rotoli B, Luzzatto L (1997) *In vivo* telomere dynamics of human hematopoietic stem cells. *Proc Natl Acad Sci USA* 94:13782–13785
- Schalling M, Hudson TJ, Buetow KH, Housman DE (1993) Direct detection of novel expanded trinucleotide repeats in the human genome. *Nat Genet* 4:135–139
- Sirugo G, Kidd KK (1995) Repeat expansion detection using ampligase thermostable DNA ligase. *Epicentre Forum* 2:1–3
- Sirugo G, Deinard AS, Kidd JR, Kidd KK (1997) Survey of maximum CTG/CAG repeat lengths in humans and non-human primates: total genome scan in populations using the repeat expansion detection method. *Hum Mol Genet* 6:403–408
- Van Steensel B, de Lange T (1997) Control of telomere length by the human telomeric protein TRF1. *Nature* 385:740–743
- Vaziri H, Schachter F, Uchida I, Wei L, Zhu X, Effros R, Cohen D, et al (1993) Loss of telomeric DNA during aging of normal and trisomy 21 human lymphocytes. *Am J Hum Genet* 52:661–667
- Zijlmans JMJM, Martens U, Poon SSS, Raap AK, Tanke HJ, Ward RK, Lansdorp PM (1997) Telomeres in the mouse have large inter-chromosomal variations in the number of T₂AG₃ repeats. *Proc Natl Acad Sci USA* 94:7423–7428

Address for correspondence and reprints: Dr. G. Sirugo, Department of Genetics, Yale University School of Medicine, New Haven, CT 06520-8005. E-mail: Sirugo@biomed.med.yale.edu

© 1998 by The American Society of Human Genetics. All rights reserved. 0002-9297/98/6302-0040\$02.00

Am. J. Hum. Genet. 63:651–654, 1998

A retGC-1 Mutation in Autosomal Dominant Cone-Rod Dystrophy

To the Editor:

Choroidretinal dystrophies represent a clinically and genetically heterogeneous group of disorders that in-

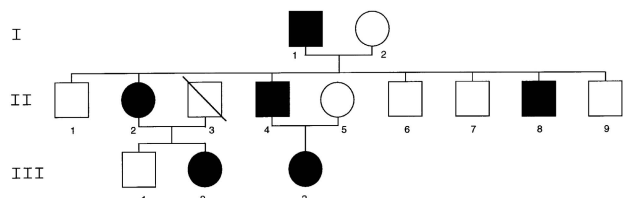


Figure 1 Pedigree of family segregating autosomal dominant cone-rod dystrophy (CORD6).

cludes retinitis pigmentosa (RP). On the other hand, cone-rod dystrophies (CRDs) long have been regarded as inverse RP and are characterized clinically by an initial cone dysfunction followed by a progressive peripheral disease (Rabb et al. 1986; Heckenlively 1987). The main symptoms at onset of the disease are a decrease of visual acuity, with loss of color discrimination and photophobia. As the disease progresses, nyctalopia, progressive peripheral visual field deficit, and decreasing scotopic electroretinogram (ERG) amplitudes are observed (Moore 1995). Autosomal dominant, autosomal recessive, and X-linked recessive patterns of inheritance have been observed (Bird 1995), and five CRD loci have been mapped: CORD1 to 18q21.1 (Warburg et al. 1991), CORD2 to 19q13 (Evans et al. 1994), CORD3 to Xp22.13-p22.11 (McGuire et al. 1995), the peripherin/retinal degeneration slow (RDS) gene to 6p21.2-cen (Travis et al. 1991), and CORD6 to 17p12-p13 (Kelsell et al. 1997). Yet, only two disease-causing genes have

been identified for CRD—namely, the peripherin/RDS gene (Nakazawa et al. 1994, 1996; Kohl et al. 1997) and the photoreceptor-specific homeobox gene, CRX, corresponding to CORD2 (Freund et al. 1997).

Since CORD6 maps to the genetic interval encompassing the retinal-specific guanylate cyclase gene (*retGC-1*) and especially since *retGC-1* mutations have been reported elsewhere for Leber congenital amaurosis (LCA1) (Perrault et al. 1996), we screened *retGC-1* for mutations in a large CRD pedigree consistent with linkage to CORD6. In addition, very recently a large deletion of the GC1 gene, the avian orthologue of *retGC-1*, had been reported in the rd/rd chicken affected with a congenital retinal degeneration similar to LCA (Semple-Rowland et al. 1997).

All affected individuals displayed an early cone dysfunction characterized by decreased vision acuity, with severe color dyschromatopsia and photophobia, during the 1st decade of life. At this stage, ophthalmoscopy examinations were not specific. By contrast, electrophysiological testing revealed marked loss of photopic function, with scotopic function relatively well preserved, and the visual field showed a consistent central scotoma. During the 2d and 3d decades, visual acuity decreased dramatically, and the color-vision defect was confined to achromatopsia, hampering normal schooling and professional insertion. After 40 years, peripheral visual field loss and progressive night blindness were observed, and the ERG became unrecordable (individual I-1; fig. 1).

For PCR-based genotypic analyses, genomic DNA

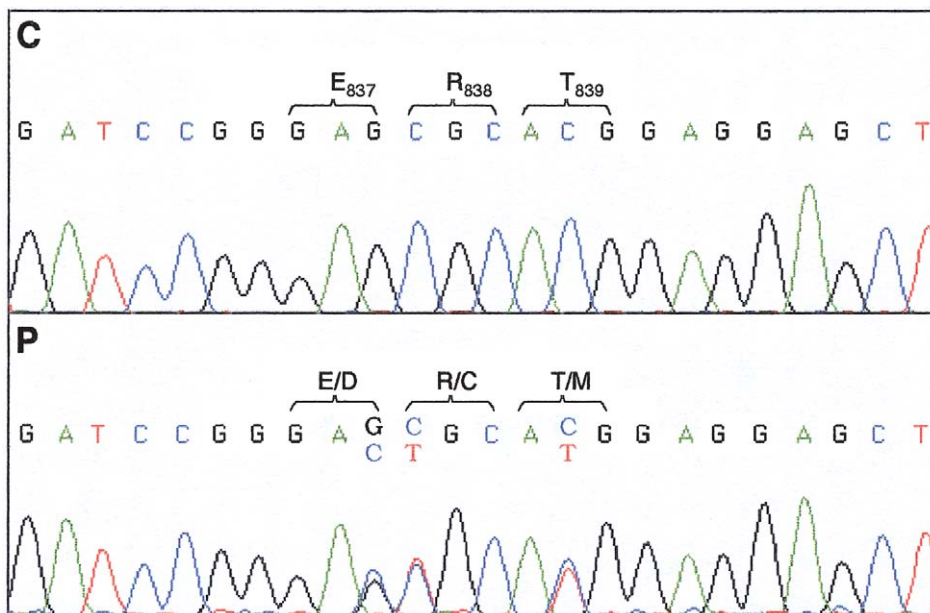


Figure 2 Identification of a heterozygote mutant genotype of the *retGC-1* gene in CORD6. C, Control. P, Patient.

(200 ng) was tested as described elsewhere (Perrault et al. 1998), and linkage analyses were performed by use of the MLINK and LINKMAP options of the LINKAGE program, version 5.1 (Lathrop et al. 1985). For mutation screening of the 18 coding exons of the retGC-1 gene, genomic DNA (200 ng) was PCR amplified by use of 1 μ M of the intronic primers, as described elsewhere (Perrault et al. 1996). Amplification products were loaded onto a 1% agarose low-melting-temperature gel, were purified by phenol-chloroform extraction, and were recovered by ethanol precipitation. Purified fragments were sequenced directly, by use of primers specific for the cDNA sequence and the Big Dye Terminator Cycle Sequencing kit (Perkin Elmer), on an automatic fluorometric DNA sequencer (Applied Biosystems).

Positive LOD-score values were obtained with polymorphic markers flanking retGC-1 at loci D17S1796 and D17S1881 (maximum LOD score of 2.71 at recombination fraction 0, for both markers). The coding sequence of the retGC-1 gene was screened for point mutations or minute changes, by direct-sequencing analysis of genomic DNA. The proband was heterozygous for a complex mutational event including three consecutive missense mutations in exon 13: (1) a G→C transversion at nucleotide 2584, changing a glutamate to an aspartate at codon 837 (E837D); (2) a C→T transition at nucleotide 2585, changing an arginine to a cysteine at codon 838 (R838C); and (3) a C→T transition at nucleotide 2589, changing a threonine to a methionine at codon 839 (T839M) (see fig. 2). This mutational event was found in all affected individuals and was absent in all healthy members of the family. No base change was found in the remaining exons.

retGC-1 mutations previously had been shown to account for LCA1. Interestingly, none of the 17 retGC-1 mutations identified in 20 unrelated LCA1 families involved the putative dimerization domain encoded by exons 11–13 (Laura et al. 1996). Conversely, no visual impairment was present in individuals heterozygous for the LCA1 mutations. We speculate that mutations at these codons led to the production of a mutant cyclase that interfered with normal protein dimerization, thereby limiting the production of cGMP in the retina, via a dominant negative effect of the mutant protein on the wild-type gene product.

In conclusion, it appears that the same gene—namely, retGC-1—can result in either an autosomal dominant cone-rod dystrophy or an autosomal recessive retinal degeneration (Leber disease), depending on the location of the mutation in the gene. The wide clinical spectrum of retGC-1 mutations gives additional support to the relevance of visual-transduction-cascade genes in a variety of retinal diseases.

Acknowledgments

This study was supported by the Association Retina France.

ISABELLE PERRAULT,¹ JEAN-MICHEL ROZET,¹
SYLVIE GERBER,¹ ROSEMARY E. KELSELL,² ERIC SOUÏED,¹
ANNICK CABOT,¹ DAVID M. HUNT,²
ARNOLD MUNNICH,¹ AND JOSSELINE KAPLAN¹

¹Unité de Recherches sur les Handicaps Génétiques de l'Enfant, Institut National de la Santé et de la Recherche Médicale U393, Hôpital des Enfants Malades, Paris; and
²Department of Molecular Genetics, Institute of Ophthalmology, University College London, London

References

- Bird AC (1995) Retinal photoreceptor dystrophies LI. Edward Jackson Memorial Lecture. *Am J Ophthalmol* 119:543–562
- Evans K, Fryer A, Inglehearn C, Duvall-Young J, Wittaker JL, Gregory CY, Butler R, et al (1994) Genetic linkage of cone-rod retinal dystrophy to chromosome 19q and evidence for segregation distortion. *Nat Genet* 6:210–213
- Freund CL, Gregory-Evans CY, Furukawa T, Papaioannou M, Looser J, Ploder L, Bellingham J, et al (1997) Cone-rod dystrophy due to mutations in a novel photoreceptor-specific homeobox gene (CRX) essential for maintenance of the photoreceptor. *Cell* 91:543–553
- Heckenlively JR (1987) RP cone-rod degeneration. *Trans Am Ophthalmol Soc* 85:438–470
- Kelsell RE, Evans K, Gregory CY, Moore AT, Bird AC, Hunt DM (1997) Localisation of a gene for dominant cone-rod dystrophy (CORD6) to chromosome 17p. *Hum Mol Genet* 6:597–600
- Kohl S, Christ-Adler M, Apfelstedt-Stylla E, Kellner U, Eckstein A, Zrenner E, Wissinger B (1997) RDS/peripherin gene mutations are frequent causes of central retinal dystrophies. *J Med Genet* 34:620–626
- Lathrop GM, Lalouel JM, Julier C, Ott J (1985) Multilocus linkage analysis in humans: detection of linkage and estimation of recombination. *Am J Hum Genet* 37:482–498
- Laura RP, Dizhoor A, Hurley JB (1996) The membrane guanylyl cyclase, retinal guanylyl cyclase-1, is activated through its intracellular domain. *J Biol Chem* 271:11646–11651
- McGuire RE, Sullivan LS, Blanton SH, Church MW, Heckenlively JR, Daiger SP (1995) X-linked dominant cone-rod degeneration: linkage mapping of a new locus for retinitis pigmentosa (RP15) to Xp22.13-p22.11. *Am J Hum Genet* 57:87–94
- Moore AT (1992) Cone and cone-rod dystrophies. *J Med Genet* 29:289–290
- Nakazawa M, Kikawa E, Chida Y, Shiono T, Tamai M (1996) Autosomal dominant cone-rod dystrophy associated with mutations in codon 244 (Asn244His) and codon 184 (Tyr184Ser) of the peripherin/RDS gene. *Arch Ophthalmol* 114:72–78
- Nakazawa M, Kikawa E, Chida Y, Tamai M (1994) Asn244His mutation of peripherin/RDS gene causing autosomal dominant cone-rod degeneration. *Hum Mol Genet* 3:1195–1196
- Perrault I, Châtelain S, Nancy V, Rozet JM, Gerber S, Ghazi I,

- Souied E, et al (1998) Exclusion of five subunits of cGMP phosphodiesterase in Leber's congenital amaurosis. *Hum Genet* 102:322–326
- Perrault I, Rozet JM, Calvas P, Gerber S, Camuzat A, Dollfus H, Châtelain S, et al (1996) Retinal-specific guanylate cyclase gene mutations in Leber's congenital amaurosis. *Nat Genet* 14:461–464
- Rabb MF, Tso MO, Fishman GA (1986) Cone-rod dystrophy: a clinical and histopathologic report. *Ophthalmology* 93: 1443–1451
- Semple-Rowland SL, Lee NR, Van Hooser JP, Palczewski K, Baehr W (1998) A null mutation in the photoreceptor guanylate cyclase gene causes the retinal degeneration chicken phenotype. *Proc Natl Acad Sci USA* 95:1271–1276
- Travis GH, Christerson L, Danielson PE, Klisak I, Sparkes RS, Hahn LB, Dryja TP, et al (1991) The human retinal degeneration slow rds gene: chromosome assignment and structure of the mRNA. *Genomics* 10:733–739
- Warburg M, Sjo O, Fledelius HC (1991) Deletion mapping of a retinal cone-rod dystrophy: assignment to 18q21.1. *Am J Med Genet* 39:288–293

Address for correspondence and reprints: Dr. Josseline Kaplan, Unité de Recherches sur les Handicaps Génétiques de l'Enfant, INSERM U393, Hôpital des Enfants Malades, 149 rue de Sèvres, 75743 Paris Cedex 15, France. E-mail: munnich3@citi2.fr

© 1998 by The American Society of Human Genetics. All rights reserved. 0002-9297/98/6302-0041\$02.00

Am. J. Hum. Genet. 63:654–656, 1998

Mapping Genes by Drift-Generated Linkage Disequilibrium

To the Editor:

In human populations that have remained of small and constant size, high levels of linkage disequilibrium (LD) are generated by genetic drift (Slatkin 1994; Laan and Pääbo 1997). Theoretical considerations suggest that such LD can be used to identify chromosomal regions involved in diseases or other traits, by “drift mapping” (Terwilliger et al. 1998). This concept relies on the assumption that when “cases” and “controls” are compared within a population in which extensive LD exists,

disequilibrium will be observed between the trait and marker loci close to the gene(s) that contributes to the trait. Furthermore, genetic differentiation between the cases and controls will be observed in genomic regions contributing to the trait, whereas no differentiation will be seen in other parts of the genome. Computer simulations indicate that, under reasonable assumptions with regard to population size, population age, and marker heterozygosity (Terwilliger et al. 1998), it might be possible to map genes by use of this approach.

To empirically evaluate this idea, we have studied polymorphic loci in and around the gene that encodes the renin-binding protein (RnBP), a component of the renin-angiotensin system involved in the regulation of blood pressure. The RnBP gene is located on Xq28 and contains a point mutation, T61C, that occurs with a frequency of .18 in Germans (Knöll et al. 1997). We scored this polymorphism in males from the Saami and the Finns, two populations that differ radically in their demographic history. Whereas the Saami have not expanded during historical times and show no indication of expansion in tests based on DNA sequence variability (von Haeseler et al. 1996), the Finns are thought to have expanded drastically during the past few thousand years, on the basis of both epidemiological (Peltonen et al. 1995) and genetic evidence (Sajantila et al. 1996). The frequencies of the C allele were found to be .21 and .19 in the Saami and the Finns, respectively. The fact that the C allele occurs at appreciable frequencies in three European populations indicates that it is older than these populations. It is therefore a useful model of alleles involved in complex traits, since such alleles are expected to be both frequent in the population and of old age.

Four microsatellites located ~1.0–7.8 cM from the RnBP gene (fig. 1), as well as the T61C polymorphism, were typed in 53 Saami and 80 Finns. In addition, 10 microsatellite loci on Xp22 and Xq13, which had numbers of alleles comparable to the numbers of those around the RnBP gene, were typed in the same individuals (Laan and Pääbo 1997; authors' unpublished data), to assess the extent to which loci situated far from the RnBP gene might yield spurious associations with the T61C polymorphism. When the RnBP polymorphism and the microsatellite loci were analyzed for allelic as-

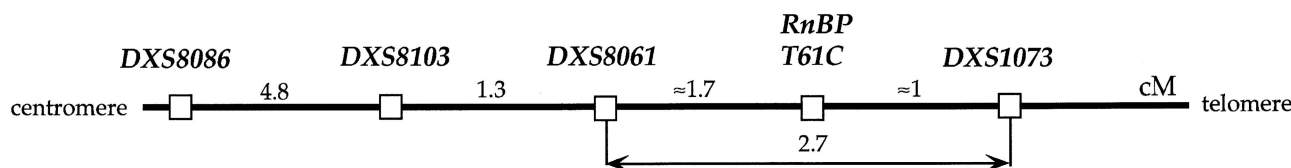


Figure 1 Genetic map (Nelson et al. 1995; Dib et al. 1996; Esposito et al. 1997; Nagaraja et al. 1997) of studied microsatellite loci around the RnBP gene.