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Study on antibacterial activity of silver nanoparticles synthesized by gamma irradiation method using different stabilizers

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Abstract

Colloidal solutions of silver nanoparticles (AgNPs) were synthesized by gamma Co-60 irradiation using different stabilizers, namely polyvinyl pyrrolidone (PVP), polyvinyl alcohol (PVA), alginate, and sericin. The particle size measured from TEM images was 4.3, 6.1, 7.6, and 10.2 nm for AgNPs/PVP, AgNPs/PVA, AgNPs/alginate, and AgNPs/sericin, respectively. The influence of different stabilizers on the antibacterial activity of AgNPs was investigated. Results showed that AgNPs/alginate exhibited the highest antibacterial activity against *Escherichia coli* (*E. coli*) among the as-synthesized AgNPs. Handwash solution has been prepared using Na lauryl sulfate as surfactant, hydroxyethyl cellulose as binder, and 15 mg/L of AgNPs/alginate as antimicrobial agent. The obtained results on the antibacterial test of handwash for the dilution to 3 mg AgNPs/L showed that the antibacterial efficiency against *E. coli* was of 74.6%, 89.8%, and 99.0% for the contacted time of 1, 3, and 5 min, respectively. Thus, due to the biocompatibility of alginate extracted from seaweed and highly antimicrobial agent in biomedicine, cosmetic, and in other fields.

Keywords: Silver nanoparticles; Gamma irradiation; Antibacterial; E. coli; Handwash

Background

The common stabilizers used in bottom-up approach of metallic nanoparticle synthesis (Au, Pt, Ag, etc.) are polymers and surfactants such as sodium dodecyl sulfate (SDS) and Tween 80 [1]. Furthermore, citrate which is a typical electro-static stabilizer has been popularly used [2]. Followed by citrate, the synthetic polymers have also been widely used as stabilizers, typically polyvinyl alcohol (PVA) [3-5] and polyvinyl pyrrolidone (PVP) [5-9]. On the other hand, among the natural polysaccharide stabilizers, chitosan [10-14] and alginate [15,16] are commonly used. In addition, the derivatives of cellulose such as carboxylmethyl cellulose [1,17], hydroxypropyl cellulose [18], and gelatin [7,19] were also used as stabilizers for the synthesis of metallic nanoparticles.

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The study results of Kvítek et al. showed that the antibacterial activity of silver nanoparticles (AgNPs) was significantly enhanced by using the suitable stabilizers such as the following surfactants: SDS, Tween 80, and PVP K90 [1]. Moreover, the study results of El Badawy et al. also showed that AgNPs/PVP had the higher antibacterial activity than AgNPs/citrate and especially AgNPs capped by branched polyethyleneimine (BPEI) had better antibacterial activity than the other ones [20]. They also claimed that the mechanism of AgNP toxicity may involve a combination of both physical and chemical interactions. There was a direct correlation between the toxicity of AgNPs and their surface charge. The more negative the zeta value, the less toxic are the AgNPs to bacillus species. The zeta potential of AgNPs/citrate was -38 mV, whereas the zeta potential of AgNPs/PVP and AgNPs/BPEI were -10 and +40 mV, respectively [20]. Therefore, the various stabilizers for AgNPs affect not only on the stability but also on the antibacterial activity of AgNP colloid [1,14,20,21].

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In this study, we prepared four colloidal AgNP solutions at a concentration of 1-mM Ag in different stabilizers, namely PVP, PVA, alginate, and sericin with the same concentration of 0.5% (w/v). Subsequently, the antibacterial activity of these colloidal AgNP solutions was investigated. To further demonstrate the effect of AgNPs on antibacterial activity and apply the development in practice, the AgNPs were added into a handwash solution, and the antibacterial activity was also tested.

Methods

Material

Pure-grade AgNO₃ was purchased from Shanghai Chemical Reagent Co., Shanghai, China The pharmaceutical grade PVP K90 was a product from Merck, Darmstadt, Germany. PVA 217 was a product of Kuraray, Tokyo, Japan. Alginate was a product of Hayashi Pure Chemical Industries, Osaka, Japan, and sericin was purchased from Sigma, St. Louis, MO, USA. Distilled water was used throughout the preparation of colloidal AgNP solutions. The strain of *Escherichia coli* ATCC 6538 was provided by the University of Medical Pharmacy, Ho Chi Minh City. The Luria-Bertani (LB) medium purchased form Himedia, Mumbai, India contains 10 g triptone, 5 g yeast extract, 10 g sodium chloride, and 1 L distilled water.

Synthesis of AgNPs

Four colloidal solution samples of 1-mM AgNPs stabilized in 0.5% (w/v) stabilizers of PVP, PVA, alginate, and sericin were prepared by gamma Co-60 irradiation method as described in our previous papers [9,13]. Briefly, the stabilizers were dissolved in water to reach a concentration of 0.5%. AgNO₃ was then dissolved in the above prepared solution to obtain a final concentration of 1-mM Ag⁺. The mixture was poured into glass bottles with plastic caps. The irradiation of these solutions at dose of 6 kGy for the synthesis of AgNPs was carried out on a Co-60 irradiator with a dose rate of approximately 1.2 kGy/h at VINA-GAMMA Center, Ho Chi Minh City. Absorption spectra of the irradiated AgNP solutions with dilution by water to 0.1-mM AgNPs were taken on an UV-vis spectrophotometer, Jasco V-630 (Easton, MD, USA). The AgNP sizes were measured using a transmission electron microscope (TEM; JEM 1010, JEOL, Tokyo, Japan).

Antibacterial activity of AgNPs

A 99-mL sterilized LB medium and AgNPs in different stabilizers as prepared for the final concentration of 1-mg/L AgNPs were added into conical flasks (250 mL), and the control sample just contained a 99-mL LB medium. A 1-ml *E. coli* suspension (approximately 10^7 CFU/mL) was added to each flask. The cultures were shaken at 150 rpm, and the bacterial growth curves were determined by measuring optical density (OD) at 600 nm on a UV-vis Jasco V-630 with 30-min interval [11,22,23].

Bactericidal activity of handwash containing AgNPs

A handwash solution was prepared using Na lauryl sulfate (Na-LS) as surfactant, hydroxyethyl cellulose (HEC) as binder, and 15 mg/L of AgNPs/alginate as antimicrobial agent. The bactericidal activity assay of the handwash against *E. coli* was carried out by culture medium toxicity method [11,13] as follows: the handwash samples (with and without AgNPs) were put into 99-mL LB medium for the final concentration of 3-mg/L AgNPs, whereas the control sample just contains 99-mL LB. Subsequently, 1-mL *E. coli* suspension of 10^7 CFU/mL was injected to each sample. The samples were shaken at 150 rpm at room temperature for 1, 3, and 5 min.





After that, the number of bacteria in each mixture was quantified by spread plate technique on LB agar plates.

Results and discussion

The successful synthesis of AgNPs stabilized in different polymer solutions was first revealed by the specific colors that the colloidal AgNP solution displays (Figure 1). A UV-vis spectrum with a maximum wavelength (λ_{max}) of 413 nm, TEM image with quasi-spherical particles, and narrow size distribution of AgNPs stabilized by alginate were typically described in Figure 2. It is clear that the resulting colloidal solutions exhibited the characteristic surface plasmon resonance (SPR) band of AgNPs with λ_{max} at 410 to 420 nm (see Table 1) [4,11].

The results in Table 1 also indicated that the AgNP average diameters were 6.1, 4.3, 10.2, and 7.6 nm for PVA, PVP, sericin, and alginate stabilizer, respectively. It is obvious that the stabilizers affected the size of AgNPs synthesized by the gamma Co-60 irradiation method. In addition, the stabilizers were also found to influence the stability and antibacterial activity of the AgNPs [1,21,24]. According to Zhang et al., the stabilizers was in the following sequence: AgNPs/PVP > AgNPs/casein > AgNPs/dextrin [24]. Furthermore, the results of Liu et al. [15] and Lan et al. [16] also confirmed the good stability of AgNPs synthesized by gamma Co-60 irradiation method using alginate as the stabilizer.

The gamma Co-60 irradiation method is fairly suitable to create the smaller AgNPs compared to chemical reduction

Table 1 The λ_{max} , OD, and average size (d) of the colloidal AgNP solution in different stabilizers

Stabilizers	λ _{max} (nm)	OD	<i>d</i> (nm)
PVA	411	0.80	6.1 ± 0.2
PVP	407	0.65	4.3 ± 0.4
Sericin	418	0.25	10.2 ± 1.1
Alginate	413	0.76	7.6 ± 0.5

method [8]. It is generally admitted that the smaller the AgNPs, the stronger the antibacterial effect. AgNPs have been currently applied as disinfecting agents in general practice due to their antibacterial effects (www. nanotechproject.org/inventories/consumer/analysis_draft/). Therefore, antibacterial activity of the resulted AgNP solutions, namely AgNPs/PVA, AgNPs/PVP, AgNPs/sericin, and AgNPs/alginate was tested.

Figure 3 displayed the dynamics of bacterial growth in liquid LB medium supplemented with 10^7 *E. coli* cells/ 100 mL and 1-mg/L AgNPs in different stabilizers. OD_o and OD_t (Figure 3) are the optical density values of the studied sample solutions at the beginning and at the different contacting time, respectively. In all AgNP-treated samples, the AgNPs caused a growth delay of *E. coli* compared with the control sample, and the growth delay effect was different in the following sequence: AgNPs/al-ginate (7.6 nm) > AgNPs/PVA (6.1 nm) > AgNPs/PVP (4.3 nm) > AgNPs/sericin (10.2 nm). The obtained results also proved that the antibacterial effect of AgNPs





depends not only on the size but also on the stabilizer used.

In addition, Sondi and Salopek-Sondi [25] and Tiwari et al. [22] reported that the concentration of AgNPs is mainly responsible for the antibacterial effect along with treatment time. Moreover, the results of El Badawy et al. have also confirmed that the stabilizers of the AgNPs were one of the most important determinants of the antibacterial activity of AgNPs [20]. For that reason, upon each application purpose, the appropriate stabilizer should be chosen for capping AgNPs, especially for applying AgNPs as antibacterial agents. Therefore, in this study, an antibacterial handwash solution was prepared using Na-LS as surfactant, HEC as binder, and 15 mg/L of AgNPs/alginate as antimicrobial agent.

Photographs of handwash solutions and bactericidal activity were showed in Figure 4. The handwash without AgNPs (HW) was almost non-antibacterial against *E. coli*; the η value reached approximately 6.2% only. The bactericidal efficiency with only 3-mg/L AgNPs diluted from the handwash solution against *E. coli* with a bioburden of approximately 10⁷ CFU/100 ml (*E. coli* infection is much higher in comparison with real conditions) was 74.6%, 89.8%, and 99.0% for 1, 3, and 5 min of contacting time, respectively (Table 2).

Wei et al. also reported the high bactericidal effect of AgNPs with sizes of 6 to 8 nm against *E. coli*, particularly

Table 2 The bactericidal efficiency (η) of handwash/ AgNPs with contacting time

Time	E. coli (CFU/mL)	η (%)
Control (LB)	33.9×10^{5}	-
Control (HW)	31.8×10^{5}	6.2
1 min	86.0×10^{4}	74.6
3 min	34.6×10^{4}	89.8
5 min	3.3×10^{4}	99.0

the η value of 10-mg/L AgNPs which was approximately 99.9% for 2 min of contacting time [11]. Our obtained results revealed the high bactericidal activity of the asprepared handwash that can be recommended to communities to apply for daily sanitary handwash for prevention of infectious diseases, such as diarrhea and enterovirus infection. Furthermore, Petica et al. reported that using Na-LS as co-stabilizer was highly effective for obtaining stable colloidal AgNP solution with very good antimicrobial and antifungal properties [26]. Concerning the environmental impact of AgNPs, it is also worth to note that the AgNPs in wastewater is almost completely transformed into Ag₂S that has extremely low solubility and exhibits a much lower toxicity than other forms of silver [27,28]. Therefore, the asprepared handwash/AgNP solution is expected to be stable for a longer duration and to maintain a bactericidal activity due to the presence of Na-LS as co-stabilizer. In addition, AgNPs eliminated from the handwash after use into wastewater will be transformed into Ag₂S that is considered to have no significant impact to the environment [27].

Conclusions

The colloidal AgNP solutions stabilized by PVA, PVP, sericin, and alginate were successfully synthesized by gamma Co-60 irradiation method. Results on antibacterial activity test demonstrated that AgNPs/alginate with an average size of 7.6 nm exhibited the highest antibacterial activity among the as-synthesized AgNP solutions. The as-prepared handwash with 15-mg/L AgNPs/alginate showed a high antibacterial efficiency with rather short contacting time. Thus, handwash/AgNPs can be potentially used as a daily sanitary handwash to prevent transmission of infectious diseases.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

NQH came up with the idea. DVP and LAQ designed and set up the experimental procedure. NND, LQL, and BDD planed the experiments and

agreed to the publication of the paper. NTKL conducted the size measurement of the as-prepared silver nanoparticles by TEM. NND, LQL, and LAQ performed the UV-vis measurement of the AgNP solutions stabilized by different polymers and evaluated the antibacterial efficiency of AgNP solutions and handwash solution containing AgNPs. NQH, LQL, and DVP analyzed the data, drafted the manuscript, revised the manuscript critically, and made a few changes. All authors read and approved the final manuscript.

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References

- Kvítek L, Panáček A, Soukupová J, Kolář M, Večeřová R, Prucek R, Holecová M, Zbořil R: Effect of surfactants and polymers on stability and antibacterial activity of silver nanoparticles (NPs). J Phys Chem C 2008, 112:5825–5834.
- Henglein A, Giersig M: Formation of colloidal silver nanoparticles: capping action of citrate. J Phys Chem B 1999, 103:9533–9539.
- 3. Temgire MK, Joshi SS: **Optical and structural studies of silver** nanoparticles. *Rad Phys Chem* 2004, **71:**1039–1044.
- Bogle KA, Dhole SD, Bhoraskar VN: Silver nanoparticles: synthesis and size control by electron irradiation. Nanotechnology 2006, 17:3204–3208.
- Patakfalvi R, Papp S, Dékány I: The kinetics of homogenous nucleation of silver nanoparticles stabilized by polymers. J Nanopart Res 2007, 9:353–364.
- Zhang Z, Zhao B, Hu L: PVP protective mechanism of ultrafine silver power synthesized by chemical reduction processes. J Solid State Chem 1996, 121:105–110.
- Kapoor S: Preparation, characterization, and surface modification of silver particles. Langmuir 1998, 14:1021–1025.
- Li T, Park HG, Choi SH: γ-irradiation-induced preparation of Ag and Au nanoparticles and their characterizations. *Mater Chem Phys* 2007, 105:325–330.
- Du BD, Phu DV, Duy NN, Lan NTK, Lang VTK, Thanh NVK, Phong NTP, Hien NQ: Preparation of colloidal silver nanoparticles in poly(*N*-vinylpyrrolidone) by y-irradiation. J Exper Nanosci 2008, 3:207–213.
- Sanpui P, Murugadoss A, Prasad PVD, Ghosh SS, Chattopadhyay A: The antibacterial properties of a novel chitosan-Ag-nanoparticle composite. *Inter J Food Microbiol* 2008, 124:142–146.
- Wei D, Sun W, Qian W, Ye Y, Ma X: The synthesis of chitosan-based silver nanoparticles and their antimicrobial activity. *Carbohydr Res* 2009, 344:2375–2382.
- Huang NM, Radiman S, Lim HN, Khiew PS, Chiu WS, Lee KH, Syahida A, Hashim R, Chia CH: γ-ray assisted synthesis of silver nanoparticles in chitosan solution and the antimicrobial properties. *Chem Engin J* 2009, 155:499–507.
- Phu DV, Lang VTK, Lan NTK, Duy NN, Chau ND, Du BD, Cam BD, Hien NQ: Synthesis and antimicrobial effects of colloidal silver nanoparticles in chitosan by γ-irradiation. J Exper Nanosci 2010, 5:169–179.
- Potara M, Jakab E, Damert A, Popescu O, Canpean V, Astilean S: Synergistic antibacterial activity of chitosan-silver nanocomposites on Staphylococcus aureus. Nanotechnology 2011, 22:135101.
- Liu Y, Chen S, Zhong L, Wu G: Preparation of high-stable silver nanoparticle dispersion by using sodium alginate as a stabilizer under gamma radiation. *Rad Phys Chem* 2009, 78:251–255.
- 16. Lan NTK, Phu DV, Lang VTK, Duy NN, Hanh TT, Anh NT, Hien NQ: Study on preparation of silver nanoparticles by gamma Co-60 irradiation using

- Hebeish AA, El-Rafie MH, Abdel-Mohdy FA, Abdel-Halim ES, Emam HE: Carboxymethyl cellulose for green synthesis and stabilization of silver nanoparticles. Carbohydr Polym 2010, 82:933–941.
- Abdel-Halim ES, Al-Deyab SS: Utilization of hydroxypropyl cellulose for green and efficient synthesis of silver nanoparticles. *Carbohydr Polym* 2011, 82:1615–1622.
- Darroudi M, Zak AK, Muhamad MR, Huang NM, Hakimi M: Green synthesis of colloidal silver nanoparticles by sonochemical method. *Mater Lett* 2012, 66:117–120.
- El Badawy AM, Silva RG, Morris B, Scheckel KG, Tolaymat TM: Surface charge-dependent toxicity of silver nanoparticles. *Environ Sci Technol* 2011, 45:283–287.
- El Badawy AM, Scheckel KG, Suidan M, Tolaymat T: The impact of stabilization mechanism on the aggregation kinetics of silver nanoparticles. Sci Total Environ 2012, 429:325–331.
- 22. Tiwari DK, Behari J, Sen P: Time and dose-dependent antimicrobial potential of Ag nanoparticles synthesized by top-down approach. *Curr Sci* 2008, **95**:647–655.
- Li WR, Xie XB, Shi QS, Zeng HY, Ou-Yang YS, Chen YB: Antibacterial activity and mechanism of silver nanoparticles on *Escherichia coli*. *Appl Microbiol Biotechnol* 2010, 85:1115–1122.
- Zhang H, Smith JA, Oyanedel-Craver V: The effect of natural water condition on the antibacterial performance and stability of silver nanoparticles capped with different polymers. *Water Res* 2012, 46:691–699.
- Sondi I, Salopek-Sondi B: Silver nanoparticles as antimicrobial agent: a case study on *E. coli* as a model for Gram-negative bacteria. *J Colloid Interf Sci* 2004, 275:177–182.
- Petica A, Gavriliu S, Lungu M, Buruntea N, Panzaru C: Colloidal silver solutions with antimicrobial properties. *Mater Sci Engin B* 2008, 152:22–27.
- Kaegi R, Voegelin A, Sinnet B, Zuleeg S, Hagendorfer H, Burkhardt M, Siegrist H: Behavior of metallic silver nanoparticles in a pilot wastewater treatment plant. Environ Sci Technol 2011, 45:3902–3908.
- 28. Ratte HT: Bioaccumulation and toxicity of silver compounds: a review. *Environ Toxicol Chem* 1999, **18**:89–108.

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