

POSTER PRESENTATION

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Homeostatic regulation in a single neuron model from the Pre-Bötzinger Complex

Max F Oginsky^{1*}, Gennady S Cymbalyuk²

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Central Pattern Generators (CPGs) control rhythmic movements in diverse species ranging from decapods to mammals. For the same CPG, from preparation to preparation, the intrinsic membrane properties of CPG neurons are highly disparate and modulated, however the network output remains constant. This leads to

questions regarding the coregulation of currents preserving essential dynamics of a neuron. When the fast-transient potassium current, I_A , is increased in the pyloric dilator neuron of the stomatogastric ganglion, there is a compensatory increase in the hyperpolarization-activated current, $I_H[1]$. This prevents changes in the output

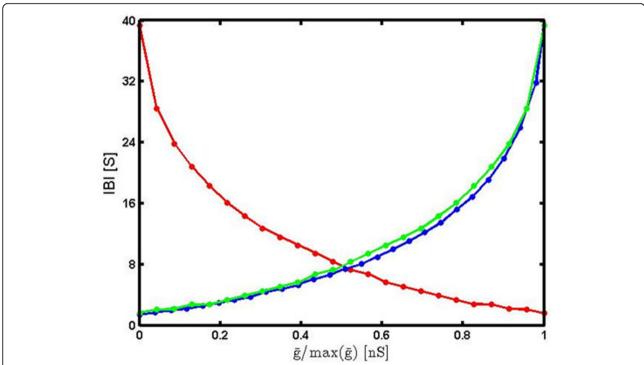


Figure 1 Dependence of interburst interval on $\tilde{\mathbf{g}}_A$ (blue) and $\tilde{\mathbf{g}}_A$ (red). Conductances were normalized to their maxima, $\max(\tilde{\mathbf{g}}_A)$ and $\max(\tilde{\mathbf{g}}_A)$.

Green curve represents the IBI dependence on $(1 - \frac{g_H}{\max(\overline{g}_H)})$

¹Department of Biology, Georgia State University, Atlanta, GA 30303, USA Full list of author information is available at the end of the article



of this well-studied CPG. Here, we test whether this coregulation would be effective if applied to the dynamics of another well-studied CPG found in mammals that controls breathing. We hypothesized that when the maximal conductance, \bar{g}_A , of I_A was increased there would be an increase in the period and interburst interval (IBI) and subsequently increasing the maximal conductance, \bar{g}_H , of I_H would provide a matching compensatory decrease. To investigate this, we modified a single neuron model of the pre-Bötzinger complex [2] by adding I_A [3] and I_H [4]. We investigated this model by systematically exploring properties of the ionic currents.

First, we increased $\bar{\mathbf{q}}_A$ from 0 to 51nS. This increased the period and IBI from 1.78s to 40.12s and 1.37s to 39.35s, respectively (Fig. 1, blue curve). At the same time, the spike frequency and spike number increased from 10.29 to 60.72Hz and 5.00 to 38.35 spikes, respectively. Then we subsequently increased \bar{g}_H from 0nS to 69nS. The period and IBI was decreased from 40.12s to 2.02s and 39.35 to 1.64s, respectively (Fig. 1, red curve). The spike frequency and spike number decreased from 60.72 to 13.93Hz and 36.50 to 2.74 spikes, respectively. In Figure 1 we presented the data for the IBIs. To place the graphs on the same scale we normalized the parameter values by the maximum values used in the study. The blue and red curves look similar, except \bar{g}_A graph grows and \bar{g}_H graph decays. To further evaluate how well the two dependences are matched, we plotted the

IBI dependence on $\left(1 - \frac{\overline{g}_H}{\max(\overline{g}_H)}\right)$ as the green curve.

One could see that the green and blue curves are well matched.

This model shows that increasing I_H opposes the effects of increasing I_A in period, IBI, frequency and spike number. This may be a common mechanism for regulating rhythmic patterns in CPGs.

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Author details

¹Department of Biology, Georgia State University, Atlanta, GA 30303, USA. ²Neuroscience Institute, Georgia State University, Atlanta, GA 30303, USA.

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