

RESEARCH ARTICLE

Oral Implant-Prostheses: New Teeth for a Brighter Brain

Vincenzo De Cicco¹, Massimo Barresi^{2*}, Maria Paola Tramonti Fantozzi¹, Enrico Cataldo³, Vincenzo Parisi⁴, Diego Manzoni¹

1 Department of Translational Research, University of Pisa, Pisa, Italy, **2** Department of Drug Sciences, University of Catania, Catania, Italy, **3** Department of Physics, University of Pisa, Pisa, Italy, **4** GB Bietti Foundation, IRCCS, Roma, Italy

* mbarresi@unict.it



OPEN ACCESS

Citation: De Cicco V, Barresi M, Tramonti Fantozzi MP, Cataldo E, Parisi V, Manzoni D (2016) Oral Implant-Prostheses: New Teeth for a Brighter Brain. PLoS ONE 11(2): e0148715. doi:10.1371/journal.pone.0148715

Editor: Mikhail A. Lebedev, Duke University, UNITED STATES

Received: August 12, 2015

Accepted: December 15, 2015

Published: February 26, 2016

Copyright: © 2016 De Cicco et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper.

Funding: The research was supported by grants of the University of Pisa, Iacer Company and Implafavourite Company. The contribution of the GB Bietti Foundation, IRCCS, was supported by Italian Ministry of Health and by Fondazione Roma. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: Products sold by Implafavourite were used in this study. The support from these

Abstract

Several studies have demonstrated that chewing can be regarded as a preventive measure for cognitive impairment, whereas masticatory deficiency, associated with soft-diet feeding, is a risk factor for the development of dementia. At present the link between orofacial sensorimotor activity and cognitive functions is unknown. In subjects with unilateral molar loss we have shown asymmetries in both pupil size and masticatory muscles electromyographic (EMG) activity during clenching: the molar less side was characterized by a lower EMG activity and a smaller pupil. Since implant-prostheses, greatly reduced both the asymmetry in EMG activity and in pupil's size, trigeminal unbalance, leading to unbalance in the activity of the Locus Coeruleus (LC), may be responsible for the pupil's asymmetry. According to the findings obtained in animal models, we propose that the different activity of the right and left LC may induce an asymmetry in brain activity, thus leading to cognitive impairment. According to this hypothesis, prostheses improved the performance in a complex sensorimotor task and increased the mydriasis associated with haptic tasks. In conclusion, the present study indicates that the implant-prosthesis therapy, which reduces the unbalance of trigeminal proprioceptive afferents and the asymmetry in pupil's size, may improve arousal, boosting performance in a complex sensorimotor task.

Introduction

Previous studies reported that mastication improves cognitive processing speed [1], alertness [2], attention [3], intelligence [4], as well as reaction time [5,6], event-related potentials latencies [7] and cerebral blood oxygen-dependent (Bold) signal [6]. It has been proposed that chewing may enhance arousal and modulate cognitive functions [7] by enhancing the activity of Ascending Reticular Activating System [8]. In addition to these short-term effects on performance, it has been suggested that the cerebral cortex activity elicited by mastication may lead to long term effects on the cerebral nervous system and be helpful in preventing degradation of brain functions [9,10,11]. Indeed, epidemiological studies have reported that tooth loss before 35 years of age was a significant risk factor for dementia or Alzheimer Disease [12,13].

companies does not alter the authors' adherence to PLOS ONE policies on sharing data and materials.

In animal experiments, it has been well documented that tooth loss, leading to long-term masticatory unbalance, decreases the number of pyramidal cells in the Hippocampal CA1 and Gyrus Dentatus [14] with impairment of spatial learning and memory in water maze tests [15]. These deficits seem to increase with aging, soft-diet feeding and time after tooth loss [16,17]. Tooth loss also increases the proliferation and the hypertrophy of hippocampal astrocytes, as it occurs following neuronal degeneration and senescence processes [16]; moreover, at hippocampal level, it decreases the number of neurons expressing c-Fos during spatial task [18], the number of dendritic spines [19] and neurogenesis [20]. It is noteworthy that the reduction in the number of c-Fos-positive cells in the hippocampal CA1 region was partially antagonized by restoring the lost molars with artificial crowns [21]. Other studies on molar less mice showed plasma glucocorticoid levels significantly greater than in molar intact control mice [22] and it is known that glucocorticoids may lead to suppression of synaptic plasticity in hippocampal neurons [23]. In addition to tooth loss, also a soft diet may affect brain structures, leading to reduced levels of brain-derived neurotrophic factor (BDNF) [24] and hippocampal neurogenesis [25]. So, there is a huge evidence that chronic masticatory dysfunction may affect brain neurobiology.

Recent studies on short-term effects of masticatory deficits on brain activity have shown that patients with temporo-mandibular disorders (TMD) show an asymmetry in both pupil size and electromyographic (EMG) activity of masticatory muscles during clenching. Moreover, the reduction of the former asymmetry by occlusal correction greatly reduce the latter, and enhances the mydriatic response associated with haptic task [26]. It is known that task-related mydriasis reflects task-associated "arousal" [27,28], "mental effort" [29,30] and, maybe, task performance (see [31]). These findings indicates that 1) trigeminal sensorimotor activity exerts a tonic effect on autonomic structures controlling pupil size and 2) its unbalance impairs cognitive performance, which is in line with the trigeminal role in long-term neurodegenerative processes shown in animal experiments and clinical studies [9–25].

However, a deeper insight is still required into the effects of occlusal unbalance and its correction upon brain activity and subject's performance. Thus the purpose of the present experiment was to investigate whether subjects with unilateral molar loss 1) show an asymmetry in the EMG activity of masticatory muscles and pupil diameter and whether replacement of the lost teeth by implant-supported prostheses 2) decrease asymmetry, 3) increases the task associated arousal estimated by the recording of mydriatic response during a haptic task, and 4) increase the sensorimotor/cognitive performance (assessed by the Spinnler-Tognoni numeric matrices [32]).

Methods

Subjects

Nine subjects (5 males and 4 female; age (mean \pm SD) 46.4 ± 7.7 years) were enrolled in the present study. They showed an unilateral loss of the first and second inferior molars, either on the left ($n = 4$) or on the right side ($n = 5$) and underwent implant of dental prostheses for restoring normal occlusal surface.

In order to evaluate the effects of simple test repetition on performance, we also studied a population of nine subjects (3 males and 6 females; age 40.2 ± 12.1 years), without occlusal alterations (controls).

Experiments consisted in routine dental care interventions, which were part of the professional practice of one of the authors (VDC) and were aimed at correcting occlusal alterations in patients and performing a preventive screening for possible deficits in normal subjects. All the subjects signed an informed consent describing the experimental design and agreed to

participate in a post-operative follow-up. They were asked to avoid caffeine and smoking for at least 2 hours before testing. None of the subjects was affected by bruxism, pain to masticatory/neck muscles, neurological, psychiatric, metabolic or endocrine diseases. None of them was under beta-blockers or corticosteroids therapy.

Surgery and implants

After radiographic bone examinations, two or three one-piece implants (3P Implafavourite, Torino, Italy), were inserted to replace the first and second molar of the mandibular arch on the left (n = 4) or the right side (n = 5). The inserted implants (n = 22) were made by a single block and screwed into a hole drilled into the bone without preliminary crest incision, piercing directly the gums. Their dimensions (diameter/length) corresponded to 4.5/10 mm (n = 10), 4.5/12 mm (n = 4) or 5.2/10 mm (n = 8). Preliminary local anaesthesia was induced by infiltration with articaine/epinephrine (Pierrel, Italy, 1/100000, 2cc). Soon after the implant placement, dental impressions were taken so to manufacture the artificial prostheses (crowns) to be mounted on the implants. The occlusal contacts of prostheses with antagonist teeth were circumscribed to dental vestibular cusps. An antimicrobial prophylaxis (Amoxycillin, Pfizer, Italy, 500 mg, twice daily) was administered for 3 days, starting 1 hour before surgery. Following the surgery, analgesic (Nimesulide, SANDOZ S.P.A, Italy, 100 mg, twice daily) was delivered for 2 days.

Experimental design

Fifteen days after surgery, temporary crowns were placed on the implants and occlusal condition was examined, in order to correct the prostheses appropriately. Then, 15 days later, before positioning of the temporary prostheses, subjects underwent evaluation of the following parameters with dental arches not touching each other (NO CONTACT condition) (see Fig 1):

1. basal pupils size evaluation while the subjects were not involved in any activity (Fig 2A);
2. pupils size evaluation during performance of a haptic task (Tan Gram) (Fig 2B),

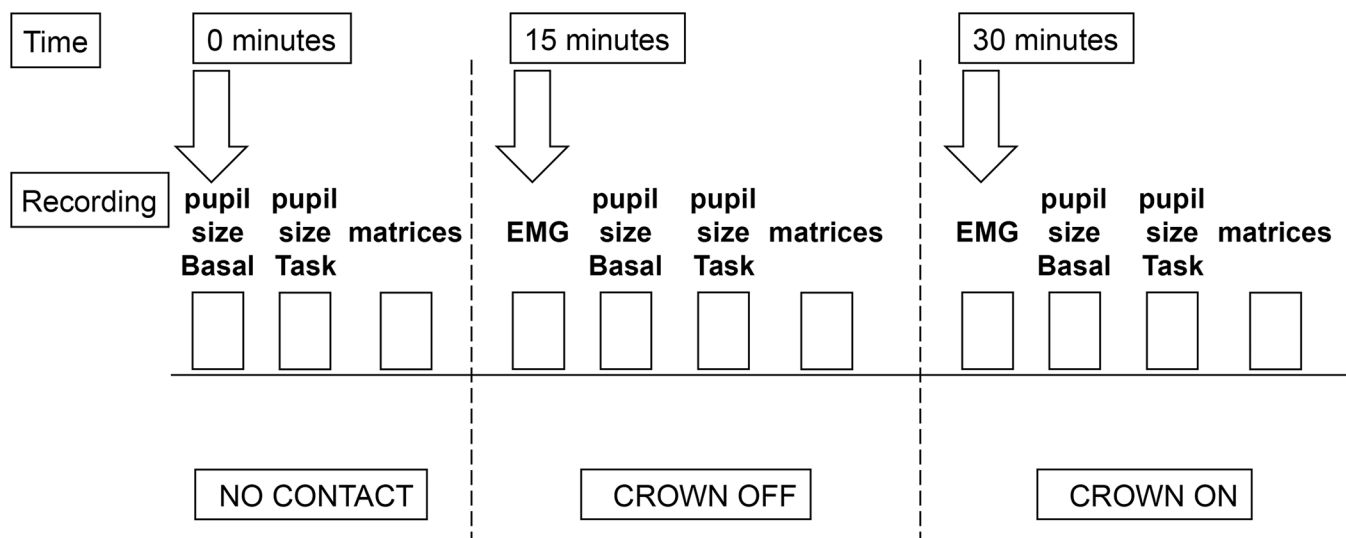


Fig 1. Experimental protocol. Flow diagram of the tests performed in all the patients at the different times. Pupil size was evaluated while the subjects were not engaged in any activity (Basal) and when they were performing a haptic task (Task). NO CONTACT: arches 1–2 mm apart. CROWN OFF: arches touching each other, no crowns on the implants. CROWN ON: arches touching each other, crowns inserted on the implants. See text for further explanation.

doi:10.1371/journal.pone.0148715.g001

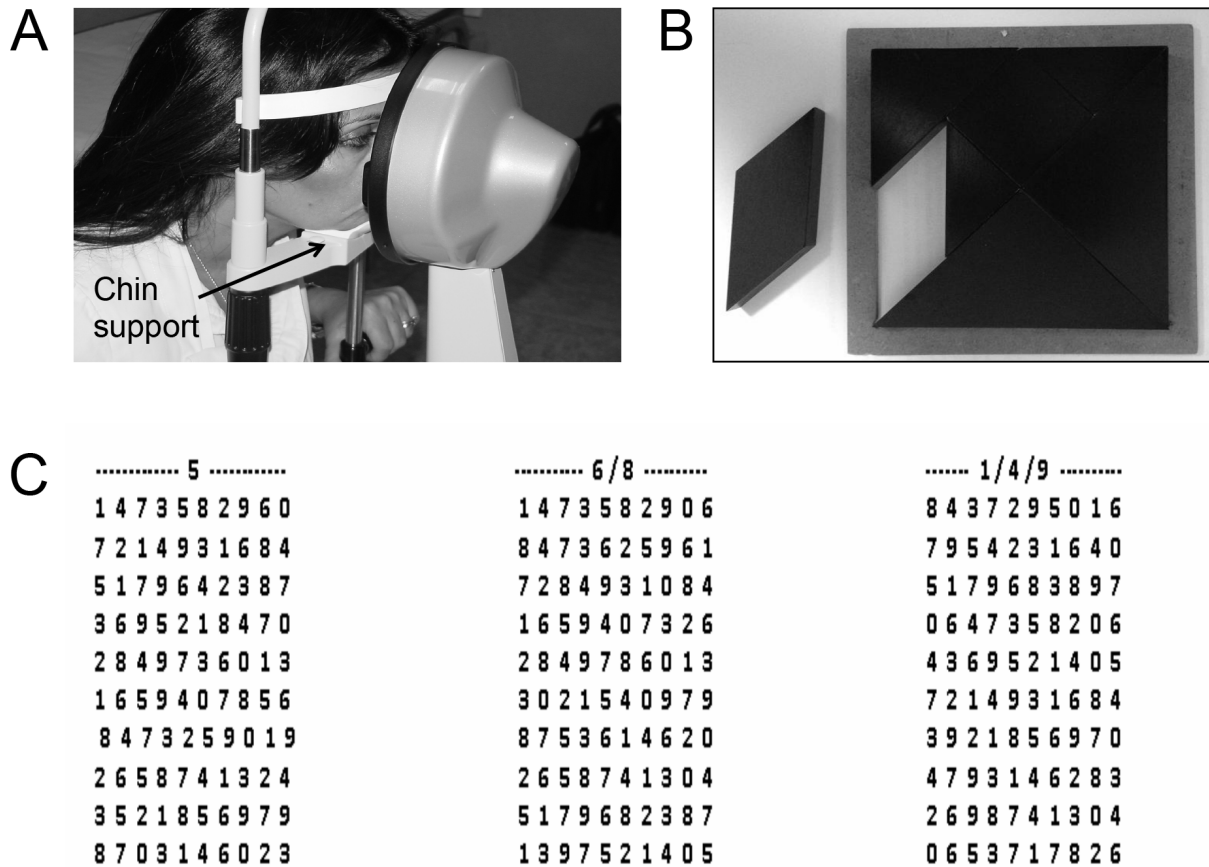


Fig 2. Pupil size recording and haptic task. A. Photograph of a representative subject with the head restrained in the pupillometric device. Note the bar for chin support that was lowered when recordings were taken with the arches 1–2 mm apart. B. Tan Gram puzzle. The parallelogram was put in the hand of the subject, while his/her head was restrained by the pupillometer as in A. The subject had to haptically reposition the piece within the puzzle. C. Example of Spinnler-Tognoni matrices. The subjects had to retrieve, from each matrix, the numbers indicated above it, underlying them with a pencil.

doi:10.1371/journal.pone.0148715.g002

- retrieval of instructed digits from Spinnler-Tognoni numeric matrices (Fig 2C). The velocity (number/sec) of number retrieved was indicated as Performance Index (PI);

Tests 1–3 were repeated with dental arches in contact (CROWN OFF condition). In this condition 4) EMG recordings of both left and right masseter activity during a clenching effort were also performed.

At this point crowns were positioned without dental cement and tests 1–4 repeated once more with dental arches in contact (CROWN ON condition).

Five of the nine subjects enrolled in the experiment (2 males and 3 females; age 47.4 ± 7.5 years) were re-tested six months following the initial session. In this second session subjects wore the final prostheses, fixed with temporary cement. Pupil size was measured and the Tan Gram and Spinnler-Tognoni numeric matrices were performed in different consecutive occlusal condition: NO CONTACT, CROWN OFF (1), CROWN ON, CROWN OFF (2). EMG evaluations were performed only in CROWN ON and CROWN OFF. With respect to the initial session, the CROWN OFF (2) condition allowed to better distinguish the effects of occlusal condition from those of test repetition.

To this aim, we performed another experiment in control of subjects showing an asymmetric EMG activity of elevator muscles during clenching, but without any occlusal alteration. They were tested for three successive times in the NO CONTACT position.

Pupil size evaluation

Pupil size measurements (mm) were performed in standard condition of artificial lighting by using a corneal topographer-pupillographer (MOD i02, with chin support, CSO, Florence, Italy) made up of a standard illuminator (halogen lamp, white light, ensuring a constant luminance level) and a camera sensor CCD1/3", with a 56 mm working distance. The operator monitored the iris image by the camera, which had an acquisition time of 33 msec. Measurements were performed for both eyes in photopic conditions, (40 lux). The arches could be 1) in contact, the chin being supported ([Fig 2A](#)) and 2) 1–2 mm apart, without chin support. Diameter values were displayed online on the computer screen. During pupil size monitoring, the subjects did not perform any clenching effort. Pupil's size was evaluated with the subject still and during the performance of a haptic task, which was practiced early only once, at the beginning of the experimental session. The task (Tan Gram), consisted of a puzzle of triangular, square and parallelogram-shaped forms. A piece of the puzzle (the parallelogram) ([Fig 2B](#)) was removed by the experimenter and put in the right hand of the subject, who had to fit it back in the original place without looking at his/her hand, keeping the head placed into the pupillometer. When the subject was at the rest, measurements were taken twice and only the second camera shot was utilised, while during task only one shot was taken, as soon as the subject began to explore the puzzle surface.

Numeric Matrix Test

In the Spinnler-Tognoni matrices test the subjects seated in front of a table, where the operator discovered a leaf containing three numerical matrices, of 10 line and 10 columns. The subjects had been previously instructed to retrieve the number 5 from the first matrix from the left, the numbers 6 and 8 from the second and the numbers 1, 4 and 9 from the third, by underlining them with a pencil (see [Fig 2C](#)). The target numbers were on the whole 60 out of the 300 included within the three matrices. The experimenter counted the numbers retrieved in a thirty seconds period and calculated the PI, i.e. the velocity of retrieval in numbers per second. The matrices presented in the different conditions analyses differed for the position of the target numbers, so that subjects could not benefit of previous spatial information for speeding up their performance.

Assessment of the occlusion

In five patients, the occlusal contact on the side of molar loss was assessed by camera shots at thirty days from the initial surgery, with and without placement of the prostheses. In this way it was possible to verify that the occlusion of the natural teeth were not modified by crown placement (see [Fig 3](#)).

EMG recordings

The EMG activity of masseter muscles was recorded by Duo-trode surface Ag/AgCl electrodes (interelectrode distance 19.5 mm, MyoTronics, Seattle, WA, USA). Electrodes were placed on the masseters belly, along an axis joining the orbit corner to the mandibular gonion, two cm far from the latter. The lead axis was parallel to the longitudinal axis of muscle fibres. Data were acquired at the sampling rate of 720 Hz by using an integrated system

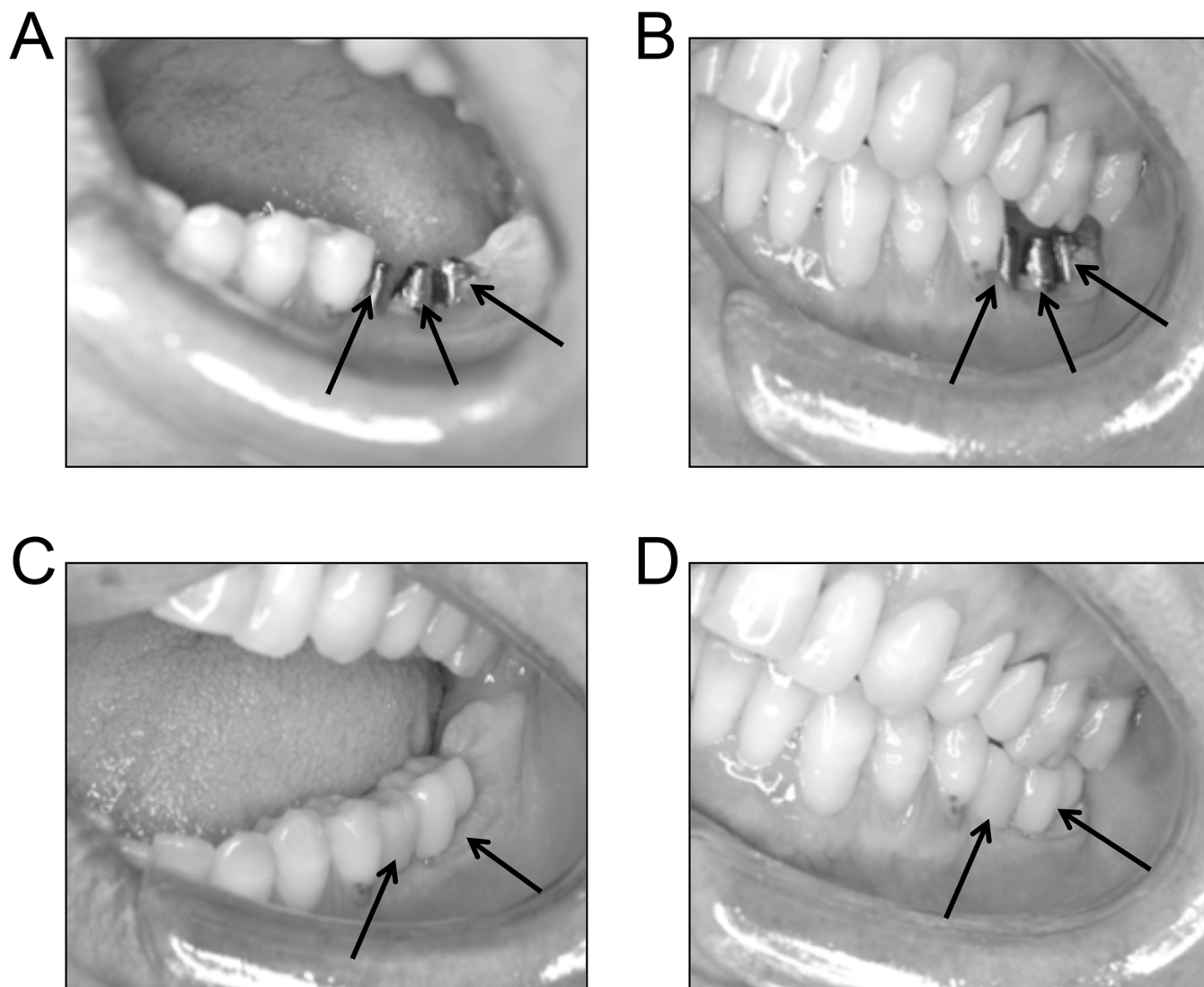


Fig 3. Evaluation of the intercuspal position. Camera shots of the arches of a subject, positioned apart (A, C) and touching each other (B, D). A-B: CROWN OFF. The arrows indicated the implants (n = 3). C-D: CROWN ON. The arrows indicated the prostheses (n = 2).

doi:10.1371/journal.pone.0148715.g003

for EMG activity and mandibular movement recording (K6-I; MyoTronics). EMG signals were acquired with a lower cutoff frequency of 15 Hz, filtered with a notch (50 Hz), full-wave rectified and displayed on the instrument monitor. The instrument provided the mean value of the rectified EMG bursts produced during clenching. Recording was allowed by the instrument software only when the resistance of the two recording leads was comparable, which allows to minimize possible bias in the asymmetry evaluation due to the different size of the EMG signal of the two sides. During evaluations of EMG asymmetries subjects were asked to develop a strong clenching effort, which abruptly raised their EMG activity (Fig 4A). At this point they were asked to raise again the effort level, leading, in general to a further increase of the EMG signal. The total time of clenching effort ranged, in different subjects, from one to three seconds.

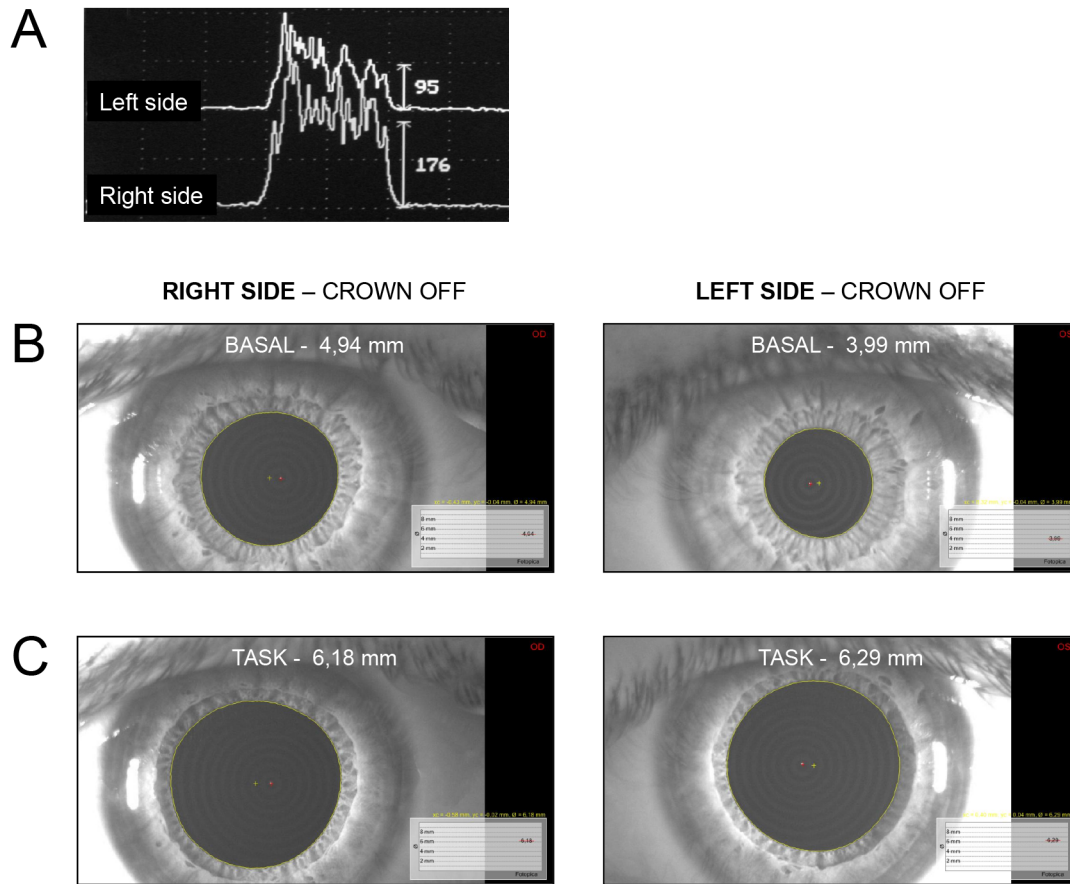


Fig 4. Examples of EMG and pupil size recordings. A. EMG activity recorded from left and right masseters in a given subject, without crown positioned and with the arches in contact (CROWN OFF). The vertical lines are calibration bars corresponding to the signal amplitude in μV . B. Recordings of pupil size in basal condition. Note that the left-right difference in pupil size is -0.95 mm. C. Recordings of pupil size during task performance (Tan Gram). Note that the left-right difference in pupil size was reduced to only 0.11 mm.

doi:10.1371/journal.pone.0148715.g004

Statistical analysis (SPSS.13)

First, in all patients we studied the left-right differences in pupil size at rest and during the haptic task as well as left-right differences in the masseter EMG activity during clenching. Positive and negative values indicated a left and right dominance, respectively. The correlation between pupil and EMG asymmetries was assessed by Pearson correlation coefficient.

The differences in size between the larger (mydriatic) and the smaller (miotic) pupil (pupil asymmetry) and the PI, were submitted to a three condition (NO CONTACT, CROWN OFF, CROWN ON) repeated measures ANOVA. The difference in the mean value of the rectified EMG burst between the hyperactive and the hypoactive side (EMG asymmetry) was analysed in a two condition (CROWN OFF, CROWN ON) repeated measures ANOVA.

Secondly, we analysed the pupils size at rest and during haptic task and the corresponding task-rest difference (mydriasis) according to a 3 condition (NO CONTACT, CROWN OFF, CROWN ON) x 2 sides (miotic/mydriatic) repeated measures ANOVA.

The EMG activity was analysed according to a two conditions (CROWN OFF, CROWN ON) x two sides (miotic/midriatic) repeated measures ANOVA. In all instances, gender was a between subjects factor.

Third, in the five patients re-tested six months following the initial session, pupil size (rest and task) and mydriasis data were processed according to a 4 condition (NO CONTACT, CROWN OFF 1, CROWN ON, CROWN OFF 2) x 2 sides (miotic/mydriatic), repeated measures ANOVA. A 4 condition (NO CONTACT, CROWN OFF 1, CROWN ON, CROWN OFF 2) design was used for pupil asymmetry and PI. A three condition (CROWN OFF 1, CROWN ON, CROWN OFF 2) design was utilized for EMG asymmetries. In these analyses gender was not considered as a between subjects factor due to the small sampling size.

Finally, the repetition effect on PI was studied in 9 control subjects showing EMG and pupil size asymmetries, but no dental loss. Data were analysed according to a 3 times (T1, T2, T3) repeated measures ANOVA, without between subjects factor.

The Greenhouse-Geisser ϵ correction was used when requested. Significance was set at $p < 0.05$.

Results

Side differences in pupil size and activity

All the subjects analysed in the CROWN OFF condition showed asymmetries in EMG activity of masseter muscles during biting (Fig 4A) and in the basal pupil size (Fig 4B). The side of the larger pupil always corresponded to the side of the higher EMG activity. Most often, the asymmetry in pupil size persisted when the subjects were involved in the haptic task. As shown in Fig 5A, a significant correlation existed between the left-right difference in basal pupil size and that in masseter EMG activity. A significant correlation was also observed between left-right pupil size differences in basal and task conditions (Fig 5B).

Crown placement greatly reduced ($F(1,7) = 42.71, P < 0.0005$) the EMG asymmetry between the mydriatic (higher EMG activity) and the miotic side (lower EMG activity), which dropped from $56.67 \pm 23.15, SD, \mu V$ (CROWN OFF) to $13.11 \pm 10.08, SD, \mu V$ (CROWN ON). A similar result (ANOVA: ($F(2,14) = 45.19, p < 0.0005$), was obtained for basal pupils asymmetry, which increased from NO CONTACT to CROWN OFF condition and dropped to the lowest value in CROWN ON condition (see Fig 6A). No significant gender effects were observed.

Similar results could be also obtained for pupil asymmetry during the haptic task ($F(2,14) = 8.30, P < 0.004$). Nonetheless, in this instance, the difference between the NO CONTACT and CROWN ON conditions was not significant (see Fig 6B). As shown in Fig 7, the left-right differences in basal and task pupil size observed in NO CONTACT and CROWN ON were strongly correlated to those observed in CROWN OFF. The same occurred for EMG differences observed in CROWN ON and CROWN OFF ($r = 0.88, P < 0.002, Y = 0.225X + 3.84$).

Performance Index

The significant condition effect observed for the PI ($F(2,14) = 248.57, P < 0.0005$), indicated that this parameter decreased from NO CONTACT to CROWN OFF condition, while increased above the NO CONTACT values in CROWN ON. As shown in Fig 8, post-hoc comparison indicated that each condition differed significantly from the others.

Pupil size, EMG activity and mydriasis on the two sides

Significant results are reported in Table 1. Table 2 reports mean \pm SD values of EMG and pupil size. These data clarify the origin of the changes in EMG and pupils asymmetries previously described. Significant condition, side and condition x side effects were found for both basal and task pupil size. Side effects were due to the fact that 1) basal sizes of larger pupils were pooled together and compared to those of the smaller ones and 2) task pupil size was in general larger

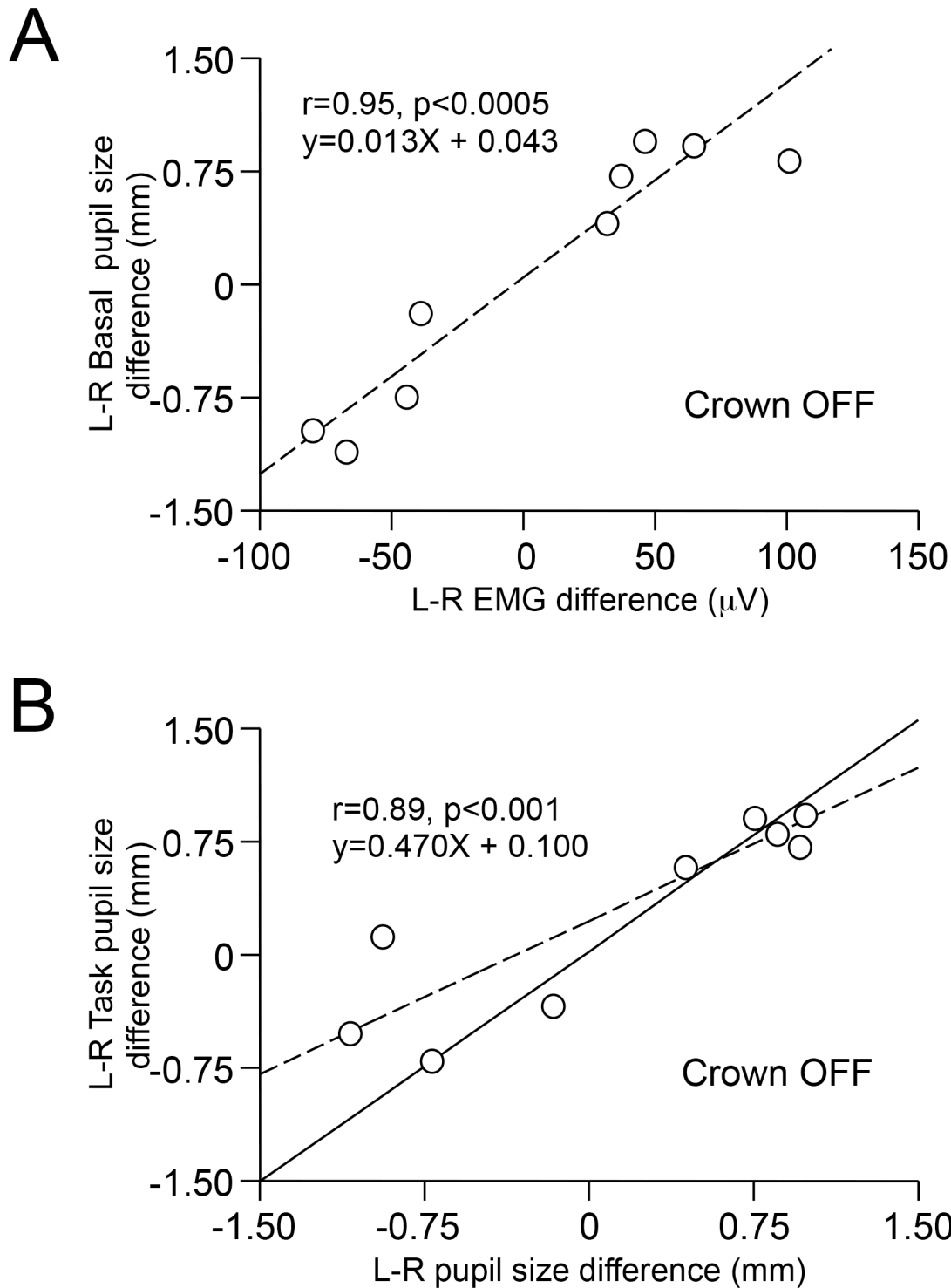
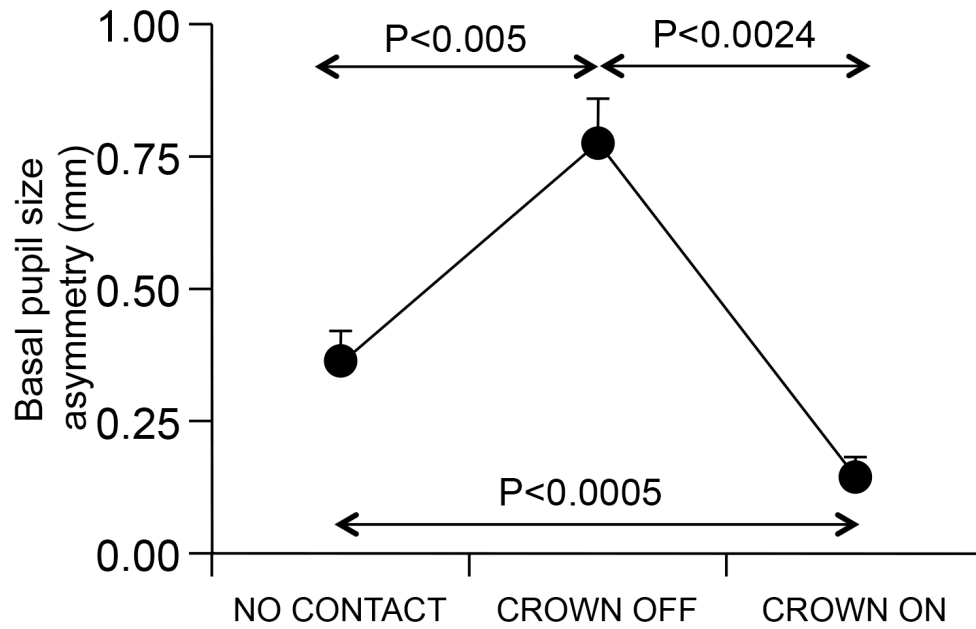


Fig 5. EMG and pupil's size asymmetries in CROWN OFF condition. A. Relation between the left-right difference in pupil size measured with the arches in contact (CROWN OFF) and the subject not attending any task (Basal) and that in masseter EMG activity observed with the arches in contact (CROWN OFF) during bite. B. Relation observed in CROWN OFF between the left-right difference in pupil size measured during the haptic task and that observed in basal condition. The continuous line represents equal values of basal and task pupil's asymmetries. In both A and B, dotted lines are the regression lines of the plotted points and their equations and coefficient of correlations are indicated in the insets.

doi:10.1371/journal.pone.0148715.g005

A



B

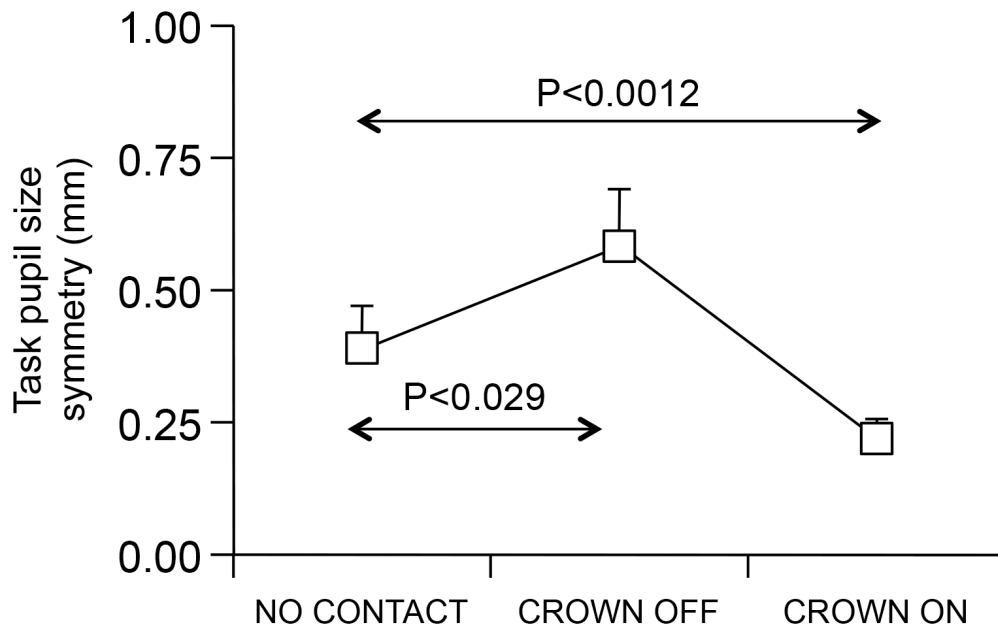


Fig 6. Pupil's size asymmetries in different conditions. The average difference in pupil size between the larger and the smaller pupil has been displayed for each of the three conditions analysed. A: asymmetries recorded while the subjects were relaxed. B: asymmetries recorded during the performance of the haptic task. In both A and B, the error bars represent standard deviations.

doi:10.1371/journal.pone.0148715.g006

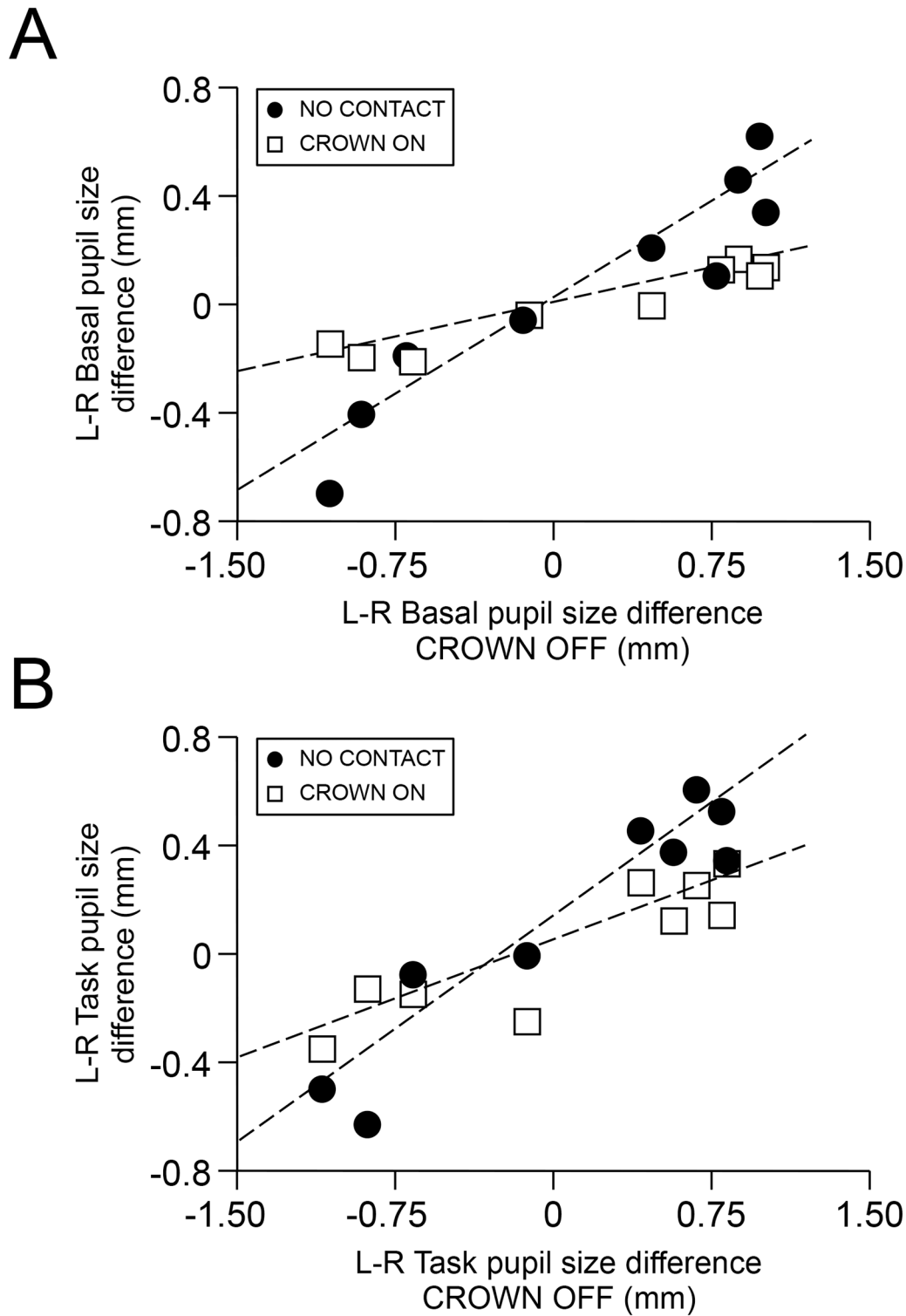


Fig 7. Relations between left-right pupil size difference in different conditions. Dots and open squares represent the left-right pupil size differences recorded in NO CONTACT and CROWN ON condition respectively, plotted as a function on the corresponding values obtained in the CROWN OFF condition. A. Basal pupil size asymmetry. B. Task pupil size asymmetry. Dotted lines are the regression lines obtained for dots and open square data, which correspond to the following equations: A. dots: $r = 0.941$, $P < 0.0005$, $Y = 0.468X + 0.18$; squares: $r = 0.958$, $P < 0.005$, $Y = 0.167X + 0.001$. B. dots: $r = 0.945$, $P < 0.0005$, $Y = 0.666X - 0.027$; squares: $r = 0.8878$, $P < 0.001$, $Y = 0.345X - 0.036$.

doi:10.1371/journal.pone.0148715.g007

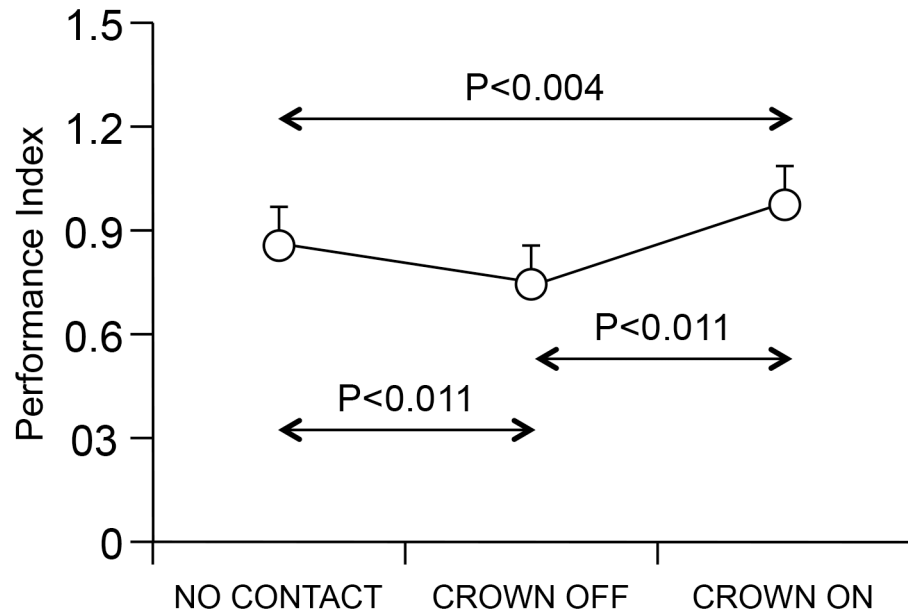


Fig 8. Performance Index in different conditions. The average values of PI have been displayed for each of the three conditions analyzed. The error bars represent standard deviations.

doi:10.1371/journal.pone.0148715.g008

in the pupil with larger basal size. Decomposition of the significant interaction (see Table 2) revealed that basal size in NO CONTACT and CROWN OFF was similar in the smaller pupil; in contrast, in the larger one, NO CONTACT values were smaller than CROWN OFF. Thus it appears that closing the arches without crowns increases the asymmetry in basal pupil size. On

Table 1. Statistically significant effects and interactions observed in the first experimental session.

Variable	Effect	p <	η ²	Post Hoc (T-Test)	p <	T
Pupil Size (Basal)	Condition F(2,14) = 14.49	0.0005	0.674	NO CONTACT < CROWN OFF	0.0005	-7.116
				CROWN ON > NO CONTACT	0.006	-3.691
	Side F(1,7) = 57.16	0.0005	0.891	miotic < mydriatic	0.0005	-7.043
Pupil Size (Task)	Condition x Side F(2,14) = 45.19	0.0005	0.866	Decomposed in Table 2		
				Condition F(2,14) = 24.78	0.0005	0.780
	Side F(1,7) = 37.41	0.0005	0.842	CROWN ON > NO CONTACT	0.001	-5.038
Mydriasis	Condition F(2,14) = 46.73	0.0005	0.870	miotic < mydriatic	0.0005	-6.559
				NO CONTACT > CROWN OFF	0.0005	8.897
	Side F(1,7) = 0.000	0.987	0.000	CROWN OFF < CROWN ON	0.0005	-8.458
EMG Activity	Condition F(2,14) = 8.30	0.004	0.543	CROWN ON > NO CONTACT	0.013	-3.174
				Condition x Side F(2,14) = 3.90	0.045	0.357
	Side F(1,7) = 52.91	0.0005	0.883	CROWN OFF < CROWN ON	0.045	-2.379
EMG Activity	Condition x Side F(1,7) = 42.71	0.0005	0.859	miotic < mydriatic	0.0005	-7.096
				Decomposed in Table 2		

Statistical significant effects observed in a three condition (NO CONTACT, CROWN OFF, CROWN ON) x 2 sides (miotic, mydriatic) repeated measures ANOVA performed on 9 patients. See text for further explanations.

doi:10.1371/journal.pone.0148715.t001

Table 2. Mean values of EMG and pupil size observed in the first experimental session.

Side	Variables	Conditions					
		1. NO CONTACT		2. CROWN OFF		3. CROWN ON	
		mean±SD	post-hoc 1–2	mean±SD	post-hoc 2–3	mean±SD	post-hoc 3–1
Smaller pupil (miotic)	basal size	3.31±0.43	NS	3.37±0.36	P<0.011	3.62±0.46	P<0.0005
	task size	4.47±0.82	NS	4.39±0.83	P<0.0005	4.89±0.67	P<0.002
	mydriasis	1.16±0.42	P<0.061	1.02±0.52	P<0.024	1.26±0.31	NS
	EMG (hypoactive)			86.8±35.4	P<0.005	130.0±54.5	
Larger pupil (mydriatic)	basal size	3.65±0.60	P<0.0005	4.12±0.54	P<0.0005	3.75±0.49	NC
	task size	4.86±0.66	NS	4.98±0.66	P<0.010	5.11±0.68	P<0.003
	mydriasis	1.21±0.33	P<0.0005	0.85±0.28	P<0.0005	1.36±0.33	P<0.011
	EMG (Hyperactive)			143.5±50.5	NS	143.1±52.5	

Average ± standard deviation values of the variables recorded in the nine subjects analyzed, submitted to a 3 condition x 2 sides repeated measures ANOVA. The EMG activity recorded on the side of the larger pupil was always higher than that recorded on the opposite side. P values refer to post-hoc analysis (see [Methods](#) for further explanations).

doi:10.1371/journal.pone.0148715.t002

the other hand, basal size of the smaller pupil increased in CROWN ON with respect to CROWN OFF, while that of the larger pupil decreased in CROWN ON: this explains why the asymmetry decreased in CROWN ON with respect to CROWN OFF. On the other hand, task size of both the smaller and larger pupils did not significantly differed between NO CONTACT and CROWN OFF. Nonetheless a slight decrease in the smaller and an increase in the larger pupil size when changing from NO CONTACT to CROWN OFF led to an enhancement of the asymmetry in CROWN OFF with respect to NO CONTACT. Finally, both pupils increased their task size in CROWN OFF with respect to CROWN ON. Since the increase was larger in the smaller pupil, the left-right side asymmetry decreased in CROWN ON. Modifications in basal and task pupil size modified the task-related mydriasis, which decreased from NO CONTACT to CROWN OFF and increased in CROWN ON (see [Fig 9](#)), with more marked changes in the larger with respect to the smaller pupil (see [Table 2](#)). As to the EMG activity, it has to be pointed out that the hypoactive muscle showed a significant increase in CROWN OFF with respect to CROWN ON, whereas the hyperactive one did not change.

Effects of test repetition in patients

In order to evaluate the effect of test repetition, five of the tested subjects with unilateral molar loss were studied once more, at six months from the initial session. In this second session, an additional final test in CROWN OFF was performed. Analysis of these data revealed condition-related changes in EMG, pupil size and mydriasis comparable to those documented in the first session (see [Tables 3](#) and [4](#)), although some differences were not significant, likely owing to the smaller number of subjects. Repetition of the condition did not change the EMG and pupil diameter, as CROWN OFF 1 and 2 were very similar. A significant condition effects ($F(3,12) = 55.02, P<0.0005$) was observed for the PI and post hoc analysis revealed smaller values in Crown OFF 1 ($0.76 \pm 0.27, P<0.001$) and CROWN OFF 2 ($0.75 \pm 0.27, P<0.002$) with respect to CROWN ON (0.96 ± 0.25). In contrast, no significant difference could be found between CROWN OFF 1 and 2. On the other hand the PI value observed in the NO CONTACT condition (0.87 ± 0.29) was significantly higher with respect to CROWN OFF (1: $P<0.001$, 2: $P<0.001$) and lower with respect to CROWN ON ($P<0.016$).

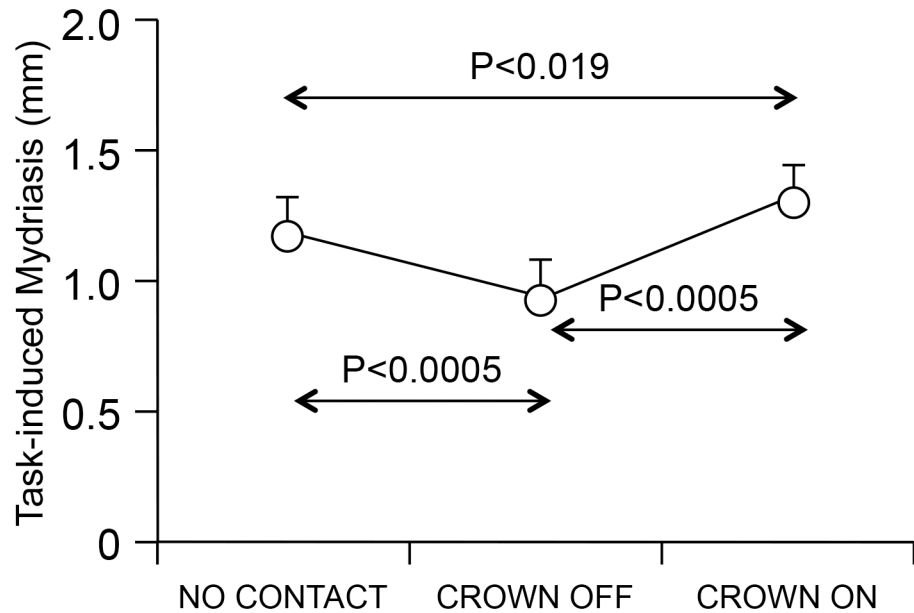


Fig 9. Task-induced mydriasis in different conditions. The average values of the task induced mydriasis have been displayed for each of the three conditions analyzed. The error bars represent standard deviations. For each subject, values relative to the left and right pupils were averaged.

doi:10.1371/journal.pone.0148715.g009

Table 3. Statistically significant effects and interactions observed in the second experimental session.

Variable	Effect	p <	η ²	Post Hoc (T-Test)	p <	t
Pupil Size (Basal)	Condition F(3,12) = 4.82	0.055	0.546	NO CONTACT < CROWN OFF1	0.013	-4.218
				NO CONTACT < CROWN OFF2	0.019	-3.820
	Side F(1,4) = 21.14	0.010	0.841	miotic < mydriatic	0.010	4.597
Pupil Size (Task)	Condition x Side F(3,12) = 27.09	0.004	0.871	Decomposed in Table 4		
				Condition F(3,12) = 13.28	0.0005	0.768
				CROWN ON > NO CONTACT	0.032	-3.231
				CROWN OFF2 < CROWN ON	0.008	4.919
	Side F(1,4) = 9.34	0.038	0.700	miotic < mydriatic	0.038	3.055
Mydriasis	Condition x Side F(3,12) = 5.80	0.066	0.592	Decomposed in Table 4		
				Condition F(3,12) = 12.54	0.009	0.758
				CROWN OFF1 < CROWN ON	0.005	-5.654
				CROWN OFF2 < CROWN ON	0.004	6.101
	Side F(1,4) = 0.22	0.664	0.052			
	Condition x Side F(3,12) = 1.87	0.240	0.319	Decomposed in Table 4		
EMG Activity	Condition F(2,8) = 11.94	0.024	0.749	CROWN OFF1 < CROWN ON	0.019	-3.816
				CROWN OFF2 < CROWN ON	0.034	3.174
	Side F(1,4) = 152.86	0.0005	0.974	miotic < mydriatic	0.0005	12.363
	Condition x Side F(2,8) = 7.40	0.050	0.649	Decomposed in Table 4		

Statistical significant effects observed in a four condition (NO CONTACT, CROWN OFF 1, CROWN ON, CROWN OFF 2) x 2 sides (miotic/mydriatic) repeated measures ANOVA performed on 5 of the 9 patients illustrated in [Table 1](#). See text for further explanations.

doi:10.1371/journal.pone.0148715.t003

Table 4. Mean values of EMG and pupil size observed in the second experimental session.

Side	Variables	Conditions									
		1. NO CONTACT		2. CROWN OFF (1)			3. CROWN ON			4. CROWN OFF (2)	
		mean±SD	post-hoc 1–2	post-hoc 1–4	mean±SD	post-hoc 2–3	post-hoc 2–4	mean±SD	post-hoc 3–1	post-hoc 3–4	mean±SD
Smaller pupil (miotic)	basal size	3.36±0.63	NS	NS	3.31±0.49	P<0.055	NS	3.60±0.49	NS	P<0.04	3.30±0.48
	Task size	4.73±0.99	NS	NS	4.51±1.07	P<0.008	NS	5.03±0.84	P<0.046	P<0.017	4.54±1.09
	Mydriasis	1.37±0.40	NS	NS	1.20±0.62	NS	NS	1.43±0.36	NS	NS	1.24±0.64
	EMG (hypoactive)				75±32.95	P<0.021	NS	133±62.45			P<0.034
Larger pupil (mydriatic)	basal size	3.45±0.63	P<0.002	P<0.003	3.93±0.65	P<0.039	NS	3.64±0.54	NS	P<0.038	3.95±0.65
	Task size	4.88±0.94	NS	NS	4.98±0.88	NS	NS	5.15±0.87	P<0.022	NS	4.98±0.85
	Mydriasis	1.43±0.35	P<0.011	P<0.014	1.05±0.26	P<0.006	NS	1.51±0.37	NS	P<0.005	1.03±0.24
	EMG (hypoactive)				116 ±42.10	P<0.022	NS	141.80 ±51.42			P<0.064

Average ± standard deviation values of the variables recorded in the five of the nine subjects shown in Table 2, s submitted to a 4 condition x 2 sides repeated measures ANOVA. The EMG activity recorded on the side of the larger pupil was always higher than that recorded on the opposite side. P values refer to post-hoc analysis (see Methods for further explanations).

doi:10.1371/journal.pone.0148715.t004

Effects of test repetition in normal subjects

Control subjects showed a significant correlation between left-right pupil size (at rest and during task) and EMG differences (basal: $r = 0.802$, $P < 0.0005$, $Y = 0.007X - 0.006$; task: $r = 0.893$, $P < 0.001$, $Y = 0.006X + 0.078$). The left-right pupil size asymmetry observed during task correlated with the corresponding value at rest ($r = 0.306$, $P < 0.424$, $Y = -0.257X + 0.168$). No significant time effect could be found for the PI (mean values: T1, 0.89 ± 0.29 ; T2, 0.92 ± 0.33 ; T3, 0.91 ± 0.32).

Discussion

The present findings indicate that, in subjects deprived unilaterally of the first and second molar, an asymmetric EMG activity of masticatory muscles develops when the arches are in contact. The side difference in EMG activity is highly correlated with the side difference observed in pupil size; both variables are smaller on the implant side. The same has been previously observed in subjects affected by TMD [26], showing asymmetry of masseter EMG activity during bite.

The observed asymmetry in pupil size was rather small, ranging from 0.18 to 0.95 mm in the different subjects (CROWN OFF, basal condition). However, it was still present, together with the correlated EMG asymmetry, six months later, which suggests it is a stable trait of subjects with unbalanced activity of elevator muscles.

In the present study, the observed EMG unbalance of masticatory muscles could depend on side differences in trigeminal sensory signals elicited during biting, which could induce asymmetric activity of trigeminal motor nuclei. In this respect it has to be pointed out that periodontal mechanoreceptors (PMRs) supply information about the forces applied to the teeth and contribute to the regulation of muscle activity generating masticatory and jaw movements [33,34]. The activation of low threshold PMRs, by the pressure exerted on the teeth can excite

at short latency masseter α and γ motoneurons [34]. So, unilateral molar loss, which reduces PMRs inputs, can contribute to the ipsilateral depression of masseter EMG activity. It is known that higher threshold PMRs give rise to the opposite effect, facilitating jaw opening [35]. This reflex component is reduced during the closing phase of chewing [36] and it is likely that the same phenomenon occurs during voluntary clenching. Crown placement did not modify the EMG activity on the hyperactive side, whereas it enhanced hypoactive side activity, possibly due to recruitment of the PMRs of the upper arch.

Another possible explanation for EMG asymmetry could be that lack of crowns on one side modifies the position of the mandible, inducing side differences in the sarcomere length of masseters and, as a consequence, in force development and motor units activity during clenching. Nonetheless, no obvious differences in the intercuspal position of patient's natural teeth could be observed when the arches were in contact with and without implanted crowns, provided that the occlusal level had been refined by milling the crown surface.

In the present study molar teeth loss-induced malocclusion seems to be the cause of the masticatory asymmetry, as the latter disappeared after crown placement. On the other hand, in some patients, malocclusion could be the consequence of prolonged, asymmetric masticatory efforts of central origin, rather than its cause. A central contribution to the masticatory asymmetry could explain why a couple of our patients showed residual EMG asymmetry following crown placement. Further investigations are required to disentangle the central and peripheral components of the muscles asymmetric activity.

As previously observed [26], the correction of EMG asymmetry was associated to a correction of pupil asymmetry. This highlights the influence of asymmetric trigeminal information on structures controlling the pupils diameter, thus emphasizing the dependence of the latter upon a trigeminal sensorimotor unbalance. The asymmetry in pupil size, which was lower when arches were in rest position (teeth 1–2 mm apart), owing to the scarce information rising from PMRs [33] and its increase by dental contact is in line with this view.

Trigeminal control of autonomic structures may develop through different pathways. Although trigeminal afferent fibres have no access to the ciliary and superior cervical ganglion [37], trigeminal input has been shown to increase the discharge of superior cervical ganglion neurons [38]. Trigeminal afferents reach autonomic structures, such as the nucleus of tractus solitarius, the ventrolateral medulla, the A5 area, the ventrolateral part of the parabrachial nucleus and the Kolliker-Fuse nucleus [39]. In addition, they reach the parvocellular reticular formation [40, 41], which mediates autonomic reflexes [42] and may influence the preganglionic parasympathetic neurons located within the Edinger-Westphal nucleus through the reticular formation and the vestibular nuclei [43,44,45]. The most important pathway is likely passing through Locus Coeruleus (LC neurons), which respond to transcutaneous electrical stimulation of the hamster's pinna [46], receive afferents from neurons localized within or near to the trigeminal Mesencephalic nucleus [47] and are electrically coupled to proprioceptive trigeminal afferents [48]. Moreover, noradrenergic LC neurons project to the preganglionic parasympathetic neurons of the Edinger-Westphal nucleus [43] and inhibits their discharge [49,50]. This inhibition is necessary to increase the pupil size, since the tonic activity of the iris constrictor would prevent pupil enlargement by dilatator pupillae [51]. The LC discharge and pupil size covary both in animals [31,52] and humans [53] and, indeed, pupil size is now considered as an indicator of LC activity [54,55,56]

Asymmetric trigeminal input to LC could be at the basis of pupils asymmetry. This hypothesis is consistent with the fact that occlusal disharmony increased the release of noradrenaline in the hypothalamic paraventricular nucleus [57] and in the frontal cortex [58].

Crown placement on dental implants also enhanced the mydriasis associated with a haptic task. Mydriasis is strictly proportional to the parallel task-related (phasic) release of

noradrenaline at cerebral cortical level [59]. Such release originates from the activation of LC, which modulates cortical arousal [50,60,61]. In contrast, LC high tonic activity reduces the phasic release of noradrenaline, decreasing task-performance [62]. So the present data suggest that reduction of the sensorimotor asymmetry by teeth substitution can reduce the asymmetry in LC discharge of the two sides and also increase the LC phasic activation, leading to an enhancement of the mydriasis associated with a cognitive task.

It is noteworthy that noradrenaline controls intracellular (Ca^{2+}) inflow in astrocytes [63], which play a key role in regulating cerebral blood flow [63,64,65], astrocytic glucose metabolism [66] as well as the cerebral synthesis and the release of BDNF [67], which is critical for long-term potentiation [68] and spatial memory [69]. This mechanism may be involved in the LC-noradrenaline system control of cognitive [70].

Thus, the improvement of the performance at the Spinnler-Tognoni matrices task induced by crown placement could be the consequence of a better phasic activation of LC neurons. Such improvement cannot be attributable to mere task repetition of the test, as the latter did not modify the PI in control subjects and the improvement elicited by crown placing in patients was completely abolished by its successive removal.

Cognitive performance may also be improved by the reduction in the left-right asymmetry of the LC tonic discharge, which may lead to a different excitability of the two hemispheres and, as a consequence, to a deterioration of cognitive performance. It has been shown, in fact, that lesion-induced unbalance in hemispheric activity may lead to severe cognitive deficits which disappears after a second, symmetric lesion on the opposite side, which doubles the extension of brain damage [71].

So, trigeminal input to the LC and, possibly, to other regions of the Ascending Reticular Activating System may help to regulate brain excitability. This hypothesis is consistent with the observation that trigeminal stimulation is useful for symptoms relief in epilepsy [72,73,74] and depression [75,76].

In this respect it has to be pointed out that also vagal stimulation may exert important effects on brain functions. In fact, vagal nerve stimulation is an approved treatment for epilepsy, depression and Alzheimer Disease [62,63] and these effects have been attributed to an activation of LC and the raphe nucleus.

Finally, it is noticeable that the masticatory activity enhances the production of BDNF and Neurotrophine-3 by muscle tissue, which are important neurotrophic factors for LC neurons and their axons [77] and this process could co-operate with the short-term influence of trigeminal afferents on LC. Thus masticatory dysfunction could impair LC neuronal functions and trigger long-term neurodegenerative processes.

In conclusion, our finding suggests that:

1. trigeminal unbalance induces asymmetric LC discharge which may exert short and long-term influences on brain excitability, leading to an asymmetric pupils size and to cognitive deficits;
2. rebalancing the activity of trigeminal afferents not only improves the masticatory activity, but also makes pupil size pupils size symmetric and improves cognitive functions.

Acknowledgments

The research was supported by grants of the University of Pisa. The contribution of the GB Bietti Foundation, IRCCS, was supported by Italian Ministry of Health and by Fondazione Roma. We thank Mr. Paolo Orsini, Francesco Montanari and Mrs Cristina Pucci for valuable technical assistance. We thank the Iacer Company for supporting Dr. Maria Paola Tramonti Fantozzi with a fellowship and the Implafavourite Company for contribution to editorial costs.

Author Contributions

Conceived and designed the experiments: VDC DM. Performed the experiments: VDC VP. Analyzed the data: VDC DM MB MPTF EC. Contributed reagents/materials/analysis tools: VDC DM. Wrote the paper: VDC DM MB MPTF. Obtained funding: DM.

References

1. Hirano Y, Obata T, Takahashi H, Tachibana A, Kuroiwa D, Takahashi T, Ikehira H, Onozuka M. Effects of chewing on cognitive processing speed. *Brain Cogn*. 2013; 81: 376–381. doi: [10.1016/j.bandc.2012.12.002](https://doi.org/10.1016/j.bandc.2012.12.002) PMID: [23375117](https://pubmed.ncbi.nlm.nih.gov/23375117/)
2. Johnson AJ, Miles C, Haddrell B, Harrison E, Osborne L, Wilson N, et al. The effects of chewing gum on physiological and self-related measures of alertness and daytime sleepiness. *Physiol Behav*. 2012; 105: 815–820. doi: [10.1016/j.physbeh.2011.10.020](https://doi.org/10.1016/j.physbeh.2011.10.020) PMID: [22061430](https://pubmed.ncbi.nlm.nih.gov/22061430/)
3. Tucha O, Mecklinger L, Maier K, Hammerl M, Lange KW. Chewing gum differentially affects aspects of attention in healthy subjects. 2004; *Appetite* 42: 327–329. PMID: [15183924](https://pubmed.ncbi.nlm.nih.gov/15183924/)
4. Smith A. Effects of chewing gum on mood, learning, memory and performance of an intelligence test. *Nutr Neurosci*. 2009; 12: 81–88. doi: [10.1179/147683009X423247](https://doi.org/10.1179/147683009X423247) PMID: [19356310](https://pubmed.ncbi.nlm.nih.gov/19356310/)
5. Allen AP, Smith AP. Effects of chewing gum and time-on-task on alertness and attention. *Nutri Neurosci*. 2012; 15: 176–185.
6. Hirano H, Onozuka M. Chewing and cognitive function. *Brain and nerve*. 2014; 66: 25–32. PMID: [24371128](https://pubmed.ncbi.nlm.nih.gov/24371128/)
7. Sakamoto K, Nakata H, Kakigi R. The effect of mastication on human cognitive processing: a study using event-related potentials. *Clin Neurophysiol*. 2009; 120: 41–50. doi: [10.1016/j.clinph.2008.10.001](https://doi.org/10.1016/j.clinph.2008.10.001) PMID: [19026594](https://pubmed.ncbi.nlm.nih.gov/19026594/)
8. Moruzzi G. and Magoun HW. Brain stem reticular formation and activation of the EEG. *Electroencephalogr Clin Neurophysiol*. 1949; 1: 455–473. PMID: [18421835](https://pubmed.ncbi.nlm.nih.gov/18421835/)
9. Ono Y, Yamamoto T, Kubo K, and Onozuka M. Occlusion and brain function: mastication as prevention of cognitive dysfunction. *J Oral Rehabil*. 2010; 37: 624–640. doi: [10.1111/j.1365-2842.2010.02079.x](https://doi.org/10.1111/j.1365-2842.2010.02079.x) PMID: [20236235](https://pubmed.ncbi.nlm.nih.gov/20236235/)
10. Okamoto N. Effect of occlusal support by implant prostheses on brain function. *J Prosthodont Res*. 2011; 55: 206–213. doi: [10.1016/j.jpor.2011.01.003](https://doi.org/10.1016/j.jpor.2011.01.003) PMID: [21333621](https://pubmed.ncbi.nlm.nih.gov/21333621/)
11. Ohkubo C, Morokuma M, Yoneyama Y, Matsuda R., Lee JS. Interactions between occlusion and human brain function activities. *J Oral Rehabil*. 2013; 40: 119–129. doi: [10.1111/j.1365-2842.2012.02316.x](https://doi.org/10.1111/j.1365-2842.2012.02316.x) PMID: [22624951](https://pubmed.ncbi.nlm.nih.gov/22624951/)
12. Weijenberg RA, Scherder EJ, Lobbezoo F. Mastication for the mind: the relationship between mastication and cognition in ageing and dementia. *Neurosci Biobehav Rev*. 2011; 35: 483–497. doi: [10.1016/j.neubiorev.2010.06.002](https://doi.org/10.1016/j.neubiorev.2010.06.002) PMID: [20547177](https://pubmed.ncbi.nlm.nih.gov/20547177/)
13. Okamoto N, Morikawa M, Tomioka K, Yanagi M, Amano N, Kurumantani N. Association between tooth loss and the development of mild memory impairment in the elderly: the Fujiwara-kyo Study. *J Alzheimers Dis*. 2015; 44: 777–786. doi: [10.3233/JAD-141665](https://doi.org/10.3233/JAD-141665) PMID: [25362033](https://pubmed.ncbi.nlm.nih.gov/25362033/)
14. Oue H, Miyamoto Y, Okada S, Koretake K, Jung C, Michikawa M, et al. Tooth loss induces memory impairment and neuronal cell loss in APP transgenic mice. *Behav Brain Res*. 2013; 252: 318–325. doi: [10.1016/j.bbr.2013.06.015](https://doi.org/10.1016/j.bbr.2013.06.015) PMID: [23773908](https://pubmed.ncbi.nlm.nih.gov/23773908/)
15. Kato T, Usami T, Noda Y, Hasegawa M, Ueda M, Nabeshima T. The effect of the loss of molar teeth on spatial memory and acetylcholine release from the parietal cortex in aged rats. *Behav Brain Res*. 1997; 83: 239–242. PMID: [9062693](https://pubmed.ncbi.nlm.nih.gov/9062693/)
16. Onozuka M, Watanabe K, Nagasaki S, Jiang Y, Ozono S, Nishiyama K, et al. Impairment of spatial memory and changes in astroglial responsiveness following loss of molar teeth in aged SAMP8 mice. *Behav Brain Res*. 2000; 108: 145–155. PMID: [10701658](https://pubmed.ncbi.nlm.nih.gov/10701658/)
17. Tsutsui K, Kaku M, Motokawa M, Tanne K. Influences of reduced masticatory sensory input from soft-diet feeding upon spatial memory/learning ability in mice. *Biomed Res*. 2007; 28: 1–7. PMID: [17379951](https://pubmed.ncbi.nlm.nih.gov/17379951/)
18. Watanabe K, Ozono S, Nishiyama K, Saito S, Tonosaki K, Fujita M, et al. The molarless condition in aged SAMP8 mice attenuates hippocampal Fos induction linked to water maze performance. *Behav Brain Res*. 2002; 128: 19–25. PMID: [11755686](https://pubmed.ncbi.nlm.nih.gov/11755686/)
19. Kubo K, Iwaku F, Watanabe K, Fujita M, Onozuka M. Molarless-induced changes of spines in hippocampal region of SAMP8 mice. *Brain Res*. 2005; 1057: 191–195. PMID: [16112090](https://pubmed.ncbi.nlm.nih.gov/16112090/)

20. Aoki H, Kimoto K, Hori N, Yamamoto Y, Onozuka M. Molarless condition suppress proliferation but not differentiation rates into neurons in the rat dentate gyrus. *Neurosci Lett*. 2010; 469: 44–48. doi: [10.1016/j.neulet.2009.11.041](https://doi.org/10.1016/j.neulet.2009.11.041) PMID: [19931591](https://pubmed.ncbi.nlm.nih.gov/19931591/)
21. Watanabe K, Tonosaki K, Karasawa N, Nagatsu I, Fujita M, Onozuka M. Evidence for involvement of dysfunctional teeth in the senile process in the hippocampus SAMP8 mice. *Exp Gerontol*. 2001; 36: 283–295. PMID: [11226743](https://pubmed.ncbi.nlm.nih.gov/11226743/)
22. Onozuka M, Watanabe K, Fujita M, Tonosaki K, Saito S. Evidence for involvement of glucocorticoid response in the hippocampal changes in molarless SAMP8 mice. *Behav Brain Res*. 2002; 131: 125–129. PMID: [11844579](https://pubmed.ncbi.nlm.nih.gov/11844579/)
23. Kim JJ, Diamond DM. The stressed hippocampus, synaptic plasticity and lost memories. *Nat Rev Neurosci*. 2002; 3: 453–462. PMID: [12042880](https://pubmed.ncbi.nlm.nih.gov/12042880/)
24. Yamamoto T, Hirayama A, Hosoe N, Furube M, Hirano S. Effects of soft-diet feeding on BDNF expression in Hippocampus of mice. *Bull Tokyo Dent Coll*. 2008; 49: 185–190. PMID: [19420879](https://pubmed.ncbi.nlm.nih.gov/19420879/)
25. Yamamoto T, Hirayama A, Hosoe N, Furube M, Hirano S. Soft diet feeding inhibits adult neurogenesis in Hippocampus of mice. *Bull Tokyo Dent Coll*. 2009; 50: 117–124. PMID: [19887754](https://pubmed.ncbi.nlm.nih.gov/19887754/)
26. De Cicco V, Cataldo E, Barresi M, Parisi V, Manzoni D. Sensorimotor trigeminal unbalance modulates pupil size. *Arch Ital Biol*. 2014; 152: 1–12. PMID: [25181592](https://pubmed.ncbi.nlm.nih.gov/25181592/)
27. Bradshaw J. Pupil size as a measure of arousal during information processing. *Nature*. 1967; 216: 515–516. PMID: [6057275](https://pubmed.ncbi.nlm.nih.gov/6057275/)
28. Bradley MM, Miccoli L, Escrig MA, Lang PJ. The pupil as a measure of emotional arousal and autonomic activation. *Psychophysiology*. 2008; 45: 602–607. doi: [10.1111/j.1469-8986.2008.00654.x](https://doi.org/10.1111/j.1469-8986.2008.00654.x) PMID: [18282202](https://pubmed.ncbi.nlm.nih.gov/18282202/)
29. Hess EH and Polt JM. Pupil size in relation to mental activity during simple problem-solving. *Science*. 1964; 143: 1190–1192. PMID: [17833905](https://pubmed.ncbi.nlm.nih.gov/17833905/)
30. Alnæs D, Sneve MH, Espeseth T, Endestad T, van de Pavert SH, Laeng B. Pupil size signals mental effort deployed during multiple object tracking and predicts brain activity in the dorsal attention network and the locus coeruleus. *J Vis*. 2014; 14: 1–20.
31. Rajkowski J, Kubiak P, Aston-Jones G (1994) Locus coeruleus activity in monkey: phasic and tonic changes are associated with altered vigilance. *Brain Res Bull* 35:607–616. PMID: [7859118](https://pubmed.ncbi.nlm.nih.gov/7859118/)
32. Spinnler H, Tognoni G. Standardizzazione e taratura Italiana di Test neuropsicologici. *Ita J Neurol Sci*. 1987; 6: 1–120.
33. Trulsson M. Sensory-motor function of human periodontal mechanoreceptors. *J Oral Rehabil*. 2006; 33: 262–273. PMID: [16629881](https://pubmed.ncbi.nlm.nih.gov/16629881/)
34. Türker KS, Sowman PF, Tuncer M, Tucker KJ, Brinkworth RS. The role of periodontal mechanoreceptors in mastication. *Arch Oral Biol*. 2007; 52: 361–364. PMID: [17222796](https://pubmed.ncbi.nlm.nih.gov/17222796/)
35. Brodin P, Türker KS, Miles TS. Mechanoreceptors around the tooth evoke inhibitory and excitatory reflexes in the human masseter muscle. *J Physiol*. 1993; 464: 711–723. PMID: [8229826](https://pubmed.ncbi.nlm.nih.gov/8229826/)
36. Lund JP, Olsson KA. The importance of reflexes and their control during jaw movement. *Trends Neurosci*. 1983; 6: 458–463.
37. Ten Tusscher MP, Klooster J, van der Want JJ, Lamers WP, Vrensen GF The allocation of nerve fibres to the anterior eye segment and peripheral ganglia of rats. I. The sensory innervation. *Brain Res*. 1989; 494: 95–104. PMID: [2475219](https://pubmed.ncbi.nlm.nih.gov/2475219/)
38. Bartsch T, Jänig W, Häbler HJ Reflex patterns in preganglionic sympathetic neurons projecting to the superior cervical ganglion in the rat. *Auton Neurosci*. 2000; 83: 66–74. PMID: [11023630](https://pubmed.ncbi.nlm.nih.gov/11023630/)
39. Panneton WM, McCulloch PF, Sun W Trigemino-autonomic connections in the muskrat: the neural substrate for the diving response. *Brain Res*. 2000; 874: 48–65. PMID: [10936223](https://pubmed.ncbi.nlm.nih.gov/10936223/)
40. Bourque MJ and Kolta A Properties and interconnections of trigeminal interneurons of the lateral pontine reticular formation in the rat. *J Neurophysiol*. 2001; 86: 2583–2596. PMID: [11698544](https://pubmed.ncbi.nlm.nih.gov/11698544/)
41. Notsu K, Tumori T, Yokota S, Semine J, Yasui Y Posterior lateral hypothalamic axon terminal are in contact with trigeminal premotor neurons in the parvicellular reticular formation of the rat medulla oblongata. *Brain Res*. 2008; 1244: 71–81. doi: [10.1016/j.brainres.2008.09.076](https://doi.org/10.1016/j.brainres.2008.09.076) PMID: [18948090](https://pubmed.ncbi.nlm.nih.gov/18948090/)
42. Esser MJ, Pronych SP, Allen GV Trigeminal-reticular connections: Possible pathways for nociception-induced cardiovascular reflex responses in the rat. *J Comp Neurol*. 1998; 391: 526–544. PMID: [9486829](https://pubmed.ncbi.nlm.nih.gov/9486829/)
43. Breen LA, Burde RM, Loewy AD Brainstem connections to the Edinger-Westphal nucleus of the cat: a retrograde tracer study. *Brain Res*. 1983; 261: 303–306. PMID: [6831211](https://pubmed.ncbi.nlm.nih.gov/6831211/)
44. Shammah-Lagnado SJ, Costa MS, Ricardo JA (1992) Afferent connections of the parvocellular reticular formation: a horseradish peroxidase study in the rat. *Neurosci*. 1992; 50: 403–425.

45. Diagne M, Valla J, Delfini C, Buisseret-Delmas C, Diagne PBTrigemino-vestibular and trigemino-spinal pathways in rats: Retrograde tracing compared with glutamic acid decarboxylase and glutamate immunohistochemistry. *J Comp Neurol*. 2006; 496: 759–772. PMID: [16628616](#)
46. Zhang J, Guan Z Pathways involved in somatosensory electrical modulation of dorsal cochlear nucleus activity. *Brain Res*. 2007; 1184: 121–131. PMID: [17964553](#)
47. Cedarbaum JM and Aghajanian GK Afferent projections to the rat locus coeruleus as determined by a retrograde tracing technique. *J Comp Neurol*. 1978; 178: 1–16. PMID: [632368](#)
48. Fujita K, Matsuo K, Yuzuriha S, Kawagishi K, Morizumi T Cell bodies of the trigeminal proprioceptive neurons that transmit reflex contraction of the levator muscle are located in the mesencephalic trigeminal nucleus in rats. *Plast Surg Hand Surg*. 2012; 46: 383–388.
49. Szabadi E and Bradshaw C Autonomic pharmacology of $\alpha 2$ -adrenoceptors. *J Physicopharmacol*. 1996; 10 (Suppl 3): s6–18.
50. Samuels ER, Szabadi E. Functional neuroanatomy of the noradrenergic locus coeruleus: its roles in the regulation of arousal and autonomic function part I: principles of functional organisation. *Curr Neuropharmacol*. 2008; 6: 235–253. doi: [10.2174/157015908785777229](#) PMID: [19506723](#)
51. Wilhelm B, Giedke H, Lütke H, Bitter E, Hofmann A., Wilhelm H Daytime variations in central nervous system activation measured by a pupillographic sleepiness test. *J Sleep Res*. 2001; 10: 1–7. PMID: [11285049](#)
52. Rajkowski J, Kubiak P, Aston-Jones G Correlations between locus coeruleus (LC) neural activity, pupil diameter and behaviour in monkey support a role of LC in attention. *Pro Soc Neurosci Abs*. 1993; 19: 974.
53. Murphy PR, O'Connell RG, O'Sullivan M, Robertson IH, Balsters JH Pupil diameter covaries with BOLD activity in human Locus Coeruleus. *Hum Brain Mapp*. 2014; 35: 4140–4154. doi: [10.1002/hbm.22466](#) PMID: [24510607](#)
54. Silvetti M, Seurinck R, van Bochove ME, Verguts T The influence of the noradrenergic system on optimal control of neural plasticity. *Front Behav Neurosci*. 2013; 7: 160. doi: [10.3389/fnbeh.2013.00160](#) PMID: [24312028](#)
55. Hoffing RC and Seitz AR Pupillometry as a glimpse into the neurochemical basis of human memory encoding. *J Cogn Neurosci*. 2015; 27: 765–774. doi: [10.1162/jocn_a_00749](#) PMID: [25390194](#)
56. Kihara K, Takeuchi T, Yoshimoto S, Kondo HM, Kawahara JI Pupillometric evidence for the locus coeruleus-noradrenaline system facilitating attentional processing of action-triggered visual stimuli. *Front Psychol*. 2015; 6: 827. doi: [10.3389/fpsyg.2015.00827](#) PMID: [26124741](#)
57. Yoshihara T, Yawaka Y Lesion of the ventral ascending noradrenergic bundles decrease the stress response to occlusal disharmony in rats. *Neurosci Lett*. 2011; 503: 43–47. doi: [10.1016/j.neulet.2011.08.004](#) PMID: [21864649](#)
58. Areso MP, Giralt MT, Sainz B, Prieto M, García-Vallejo P, Gómez FM Occlusal disharmonies modulate central catecholaminergic activity in the rat. *J Dent Res*. 1999; 78: 1204–1213. PMID: [10371243](#)
59. Gabay S, Pertzov Y, Henik A. Orienting of attention, pupil size and the norepinephrine system. *Atten Percept Psychophys*. 2011; 73: 123–129. doi: [10.3758/s13414-010-0015-4](#) PMID: [21258914](#)
60. Carter M, Yizhar O, Chikahisa S, Nguyen H, Adamantidis A, Nishino S, et al. Tuning arousal with optogenetic modulation of Locus Coeruleus neurons. *Nat Neurosci*. 2010; 13: 1526–1533. doi: [10.1038/nn.2682](#) PMID: [21037585](#)
61. Howells FM, Stein DJ, Russell VA. Synergistic tonic and phasic activity of Locus Coeruleus-Noradrenaline arousal system is required for optimal attentional performance. *Metab Brain Dis*. 2012; 27: 267–274. doi: [10.1007/s11011-012-9287-9](#) PMID: [22399276](#)
62. Gilzenrat MS, Nieuwenhuis S, Jepma M, Cohen JD. Pupil diameter tracks changes in control state predicted by the adaptive gain theory of locus coeruleus function. *Cogn Affect Behav Neurosci*. 2010; 10: 252–269.
63. Paukert M, Agarwal A, Cha J, Doze VA, Kang JU, Bergles DE. Norepinephrine controls astroglial responsiveness to local circuit activity. *Neuron*. 2014; 82: 1263–1270. doi: [10.1016/j.neuron.2014.04.038](#) PMID: [24945771](#)
64. Figley CR, Stroman PW. The role(s) of astrocytes and astrocyte activity in neurometabolism, neurovascular coupling, and the production of functional neuroimaging signals. *Eur J Neurosci*. 2011; 33: 577–588. doi: [10.1111/j.1460-9568.2010.07584.x](#) PMID: [21314846](#)
65. Girouard H, Bonev AD, Hannah RM, Meredith A, Aldrich RW, Nelson MT; Astrocytic endfoot Ca^{2+} and BK channels determine both arteriolar dilation and constriction. *Proc Natl Acad Sci U S A*. 2010; 107: 3811–3816. doi: [10.1073/pnas.0914722107](#) PMID: [20133576](#)

66. Sorg O, Magistretti PJ. Characterization of the glycogenolysis elicited by vasoactive intestinal peptide, noradrenaline and adenosine in primary cultures of mouse cerebral cortical astrocytes. *Brain Res.* 1991; 563: 227–233. PMID: [1664773](#)
67. Juric DM, Miklic S, Carman-Krzan M. Monoaminergic neuronal activity up-regulates BDNF synthesis in cultured neonatal rat astrocytes. *Brain Res.* 2006; 1108: 54–62. PMID: [16828062](#)
68. Edelmann E, Cepeda-Prado E, Franck M, Lichtenecker P, Brigadski T, Leßmann V. Theta burst firing recruits BDNF release and signaling in postsynaptic CA1 neurons in spike-timing-dependent LTP. *Neuron.* 2015; 86: 1041–1054. doi: [10.1016/j.neuron.2015.04.007](#) PMID: [25959732](#)
69. Ozawa T, Yamada K, Ichitani Y. Hippocampal BDNF treatment facilitates consolidation of spatial memory in spontaneous place recognition in rats. *Behav Brain Res.* 2014; 263: 210–216. doi: [10.1016/j.bbr.2014.01.034](#) PMID: [24503120](#)
70. Berridge CW, Waterhouse BD. The locus coeruleus-noradrenergic system: modulation of behavioral state and state-dependent cognitive processes. *Brain Res Rev.* 2003; 42: 33–84. PMID: [12668290](#)
71. Lomber SG and Payne BR Removal of two halves restores the whole: reversal of visual hemineglect during bilateral cortical or collicular inactivation in the cat. *Vis Neurosci.* 1996; 13: 1143–1156. PMID: [8961543](#)
72. DeGiorgio CM, Krahl SE. Neurostimulation for drug-resistant epilepsy. *Continuum (Minneapolis, Minn).* 2013; 19: 743–755.
73. DeGiorgio CM, Soss J, Cook IA, Markovic D, Gornbein J, Murray D, et al. Randomized controlled trial of trigeminal nerve stimulation for drug-resistant epilepsy. *Neurology.* 2013; 80: 786–791. doi: [10.1212/WNL.0b013e318285c11a](#) PMID: [23365066](#)
74. Zare M, Salehi M, Mahvari J, Najafi MR, Moradi A, Pour MH, et al. Trigeminal nerve stimulation: a new way of treatment of refractory seizures. *Adv Biomed Res.* 2014; 3: 81. doi: [10.4103/2277-9175.127994](#) PMID: [24761389](#)
75. Schrader LM, Cook IA, Miller PR, Maremont ER, DeGiorgio CM. Trigeminal nerve stimulation in major depressive disorder: first proof of concept in an open pilot trial. *Epilepsy Behav.* 2011; 22: 475–478. doi: [10.1016/j.yebeh.2011.06.026](#) PMID: [21820361](#)
76. Cook IA, Schrader LM, DeGiorgio CM, Miller PR, Maremont ER, Leuchter AF. Trigeminal nerve stimulation in major depressive disorder: acute outcomes in an open pilot study. *Epilepsy Behav.* 2013; 28: 221–226. doi: [10.1016/j.yebeh.2013.05.008](#) PMID: [23773978](#)
77. Fan G, Copray S, Huang E, Jones K, Yan Q, Walro J, et al. Formation of a full complement of cranial proprioceptors requires multiple neurotrophins. *Dev Dyn.* 2000; 218: 359–370. PMID: [10842362](#)