

1 **Kinetic and microbiological characterization of aerobic granules performing**  
2 **partial nitrification of a low-strength wastewater at 10°C**

3

4 Clara Reino<sup>1</sup>, María Eugenia Suárez-Ojeda<sup>1</sup>, Julio Pérez<sup>1,2</sup>, Julián Carrera<sup>1</sup>

5 <sup>1</sup>GENOCOV Research Group, Department of Chemical, Biological and Environmental  
6 Engineering, School of Engineering, Universitat Autònoma de Barcelona, Ed. Q-  
7 Campus UAB, 08193 Bellaterra, Barcelona, Spain

8 <sup>2</sup>Department of Biotechnology, Delft University of Technology, Julianalaan 67, Delft  
9 2628 BC, The Netherlands

10

11 \*Corresponding author: Tel. +34 935812141

12 E-mail address: [julian.carrera@uab.cat](mailto:julian.carrera@uab.cat) (J. Carrera)

13

14 Abstract

15 A granular airlift reactor enriched in ammonia oxidizing bacteria (AOB) was operated at  
16 10 °C performing stable partial nitrification in the long-term. The reactor treated a  
17 synthetic low-strength influent during 250 days with an average nitrogen loading rate of  
18  $0.63 \pm 0.06 \text{ g N L}^{-1} \text{ d}^{-1}$ . Nitrate production was barely detected, being the average  
19 concentration in the effluent of  $0.6 \pm 0.3 \text{ mg N-NO}_3 \text{ L}^{-1}$ . Furthermore, a suitable effluent  
20 for a subsequent reactor performing the anammox process was achieved. A maximum  
21 specific growth rate as high as  $0.63 \pm 0.05 \text{ d}^{-1}$  was determined by performing kinetic  
22 experiments with the granular sludge in a chemostat and fitting the results to the Monod  
23 model. Pyrosequencing analysis showed a high enrichment in AOB (41 and 65 % of the  
24 population were identified as *Nitrosomonas* genus on day 98 and 233, respectively) and  
25 an effective repression of nitrite oxidizing bacteria in the long-term. Pyrosequencing

26 analysis also identified the coexistence of nitrifying bacteria and heterotrophic  
27 psychrotolerant microorganisms in the granular sludge. Some psychrotolerant  
28 microorganisms are producers of cryoprotective extracellular polymeric substances that  
29 could explain the better survival of the whole consortia at cold temperatures.

30

31 Keywords

32 Partial nitrification; mainstream; low temperature; NOB-repression; AOB-enrichment;  
33 anammox

34

35 1. Introduction

36 Nitrogen removal is essential in urban wastewater treatment plants (WWTPs) since  
37 nitrogenous compounds are toxic to aquatic life and cause eutrophication and oxygen  
38 depletion in receiving waters. Conventional activated sludge systems are the most  
39 frequently used systems in urban WWTPs since a good removal of pollutants is  
40 guaranteed, however the costs associated to this typical biological treatment make  
41 WWTPs as very energy-demanding facilities. For the achievement of a cost-effective  
42 (energy-neutral or even energy-positive) urban WWTP, the implementation of the  
43 autotrophic biological nitrogen removal (BNR) in the mainstream has been proposed  
44 (Jetten et al., 1997; Kartal et al., 2010; Siegrist et al., 2008). Thus, aeration costs are  
45 reduced because of the lower oxygen requirements of the process compared to  
46 conventional activated sludge treatment; and furthermore, biogas production is  
47 increased since most of the organic matter will be converted to biogas in the anaerobic  
48 digestion process, with the consequent energy recovery.

49

50 Recently, many studies were focused on the implementation of autotrophic BNR in one-  
51 stage systems, such as CANON (Completely Autotrophic Nitrogen removal Over  
52 Nitrite) and OLAND (Oxygen-Limited Autotrophic Nitrification/Denitrification)  
53 technologies. Nevertheless, at low temperature and low-strength wastewaters most of  
54 these systems showed the failure of nitrification in the long-term operation, due to the  
55 growth of nitrite oxidizing bacteria (NOB) triggering the production of nitrate and the  
56 destabilization of the subsequent anammox process (De Clippeleir et al., 2013; Hu et al.,  
57 2013; Wett et al., 2013; Winkler et al., 2011). Even though Gilbert et al. (2014) reported  
58 stable operation at 10 °C with synthetic low-strength wastewater in a one-stage system,  
59 the achieved ammonium conversion rate resulted as low as 0.015 g N L<sup>-1</sup> d<sup>-1</sup>. Hence,  
60 two-stage systems appear as the alternative to overcome the destabilization problems  
61 and the low conversion rates associated to one-stage systems (Ma et al., 2011; Pérez et  
62 al., 2015; Regmi et al., 2014). Separation of the partial nitrification and the anammox  
63 process in two different reactors makes possible a more stable performance and control.  
64 In fact, stable partial nitrification at 12.5 °C with a granular sludge reactor was reported  
65 by Isanta et al. (2015) and long-term operation of an anammox reactor at temperatures  
66 between 20 and 10 °C was reported by Lotti et al. (2014). Both of these studies treated  
67 low-strength wastewater, demonstrating the feasibility of application of autotrophic  
68 BNR to mainstream conditions.

69

70 For both, one and two-stage approaches, successful implementation of autotrophic BNR  
71 at mainstream conditions relies on the stability of partial nitrification in the long-term, i.e.  
72 by achieving an effective repression of NOB activity. Previous research has shown a  
73 more sensitive temperature dependence of ammonia oxidizing bacteria (AOB)  
74 compared to that of NOB (Hunik et al., 1994; Knowles et al., 1965; Van Hulle et al.,

75 2010). Thus, different strategies have been conducted in order to favour AOB over  
76 NOB activity at mainstream conditions. On one hand, Gao et al. (2014) proposed an  
77 aeration control strategy depending on the temperature and ammonia concentration in  
78 the influent. Efficient NOB repression was obtained at room temperature (12-27 °C) but  
79 temperature fluctuated daily, being lower than 15 °C less than 10 days. On the other  
80 hand, Isanta et al. (2015) achieved stable partial nitrification for 300 days at 12.5 °C in a  
81 granular sludge system, by maintaining an adequate ratio between oxygen and  
82 ammonium concentrations in the reactor bulk liquid. However, in northern climates,  
83 temperature can easily achieve values lower than 12.5 °C during winter. In fact, average  
84 temperatures of wastewater in west European region are around 17 °C, with a minimum  
85 of 8 °C and a maximum of 29 °C (De Clippeleir et al., 2013), and thus, a temperature  
86 gradient from 20 °C in summer to 10 °C in winter was presented as representative for  
87 WWTPs in moderate climates (Gilbert et al., 2015).

88

89 In the present study we would like to demonstrate the long-term stability of partial  
90 nitrification at 10 °C for low-strength synthetic wastewater in a granular sludge reactor  
91 operated in continuous mode. Furthermore, we aim for a better understanding of the  
92 process through the in depth study of the nitrifying biomass of the granular sludge  
93 reactor. Thus, our second objective is to characterize the population developed at low  
94 temperature in the reactor from both microbiological and kinetic points of view. Finally,  
95 we correlated the special characteristics of the biomass with the nitrifying ability of the  
96 granular reactor at 10 °C.

97

98

99

100 2. Materials and methods

101 2.1. Reactor set-up and operation

102 A lab-scale airlift reactor with a total volume of 5.2 L, with a downcomer-to-separator  
103 diameter ratio of 0.36 and a total length-to-downcomer diameter ratio of 16 was used.  
104 The detailed diagram of the reactor is presented in Fig. 1. Compressed air was supplied  
105 through an air diffuser placed at the bottom of the reactor and was manually  
106 manipulated to maintain the dissolved oxygen (DO) concentration in the bulk liquid in  
107 the range 0.5-2.5 mg O<sub>2</sub> L<sup>-1</sup>. The DO concentration in the bulk liquid was measured  
108 online by means of a DO electrode (DO 60-50, Crison Instruments, Spain). The pH was  
109 measured online with a pH probe (pH 52-10, Crison Instruments, Spain) and  
110 automatically controlled at 8.0±0.1 by dosing a Na<sub>2</sub>CO<sub>3</sub> 0.5 M solution. The pH was  
111 controlled throughout the operation period to rule out any potential effects derived from  
112 pH changes. Since the effect of pH on nitrification rates is known to be reduced in the  
113 range 7.5-8, a pH set point of 8 was selected, as done in a previous study (Isanta et al.,  
114 2015). The temperature was measured and controlled at 10 °C by means of a cooling  
115 system (E100, LAUDA, Germany) and an electric heater (HBSI 0.8 m, HORST,  
116 Germany) connected to a temperature controller (BS-2400, Desin Instruments, Spain).  
117 Total ammonia nitrogen (TAN = N-NH<sub>4</sub><sup>+</sup> + N-NH<sub>3</sub>) and nitrate concentrations in the  
118 bulk liquid were measured by using an on-line probe (AN-ISE sc probe with a Cartrical  
119 cartridge plus, Hach Lange, Germany). The range of the on-line probe for TAN and  
120 nitrate concentrations was 0-1000 mg N L<sup>-1</sup> whereas the detection limit was 0.2 mg N  
121 L<sup>-1</sup> for both parameters. TAN concentration in the bulk liquid was automatically  
122 controlled by varying the inflow rate by means of a proportional controller during the  
123 whole period of operation, except between days 93-95, 144-172 and 241-245 when the

124 control was manually made based on the off-line bulk liquid TAN concentration  
125 measurement.

126

## 127 2.2. Wastewater and inoculum characteristics

128 The reactor treated a synthetic influent with an average TAN concentration of 70 mg N  
129 L<sup>-1</sup>, which mimics a pretreated municipal wastewater coming from the mixture of the  
130 effluent of a previous A-stage plus the recirculation of the reject water of the digested  
131 sludge, as in an anammox-based WWTP (Isanta et al., 2015; Kartal et al., 2010). The  
132 synthetic influent also contained: 45 mg L<sup>-1</sup> KH<sub>2</sub>PO<sub>4</sub>, 784 mg L<sup>-1</sup> NaHCO<sub>3</sub>, 80 mg L<sup>-1</sup>  
133 NaCl, 40 mg L<sup>-1</sup> CaCl<sub>2</sub>, 90 mg L<sup>-1</sup> MgCl<sub>2</sub> and 1 mL of trace elements solution per L of  
134 influent (Guerrero et al., 2011).

135 The biomass was enriched in AOB and adapted to low temperature (12.5 °C) in a  
136 reactor which was operating for more than 400 days performing stable partial nitritation  
137 (Isanta et al., 2015). Hence, the inoculum contained around 81±12 % of AOB and 1±1  
138 % of NOB as analyzed by fluorescent *in-situ* hybridization (FISH).

139

## 140 2.3. Kinetic experiments

141 Kinetic experiments were conducted in a chemostat with a working volume of 2.9 L.  
142 For each experiment, the chemostat was inoculated with nitrifying granules from the  
143 continuous airlift reactor to a final concentration of 83±3 mg VSS L<sup>-1</sup>. The same  
144 synthetic wastewater of the reactor was used as influent to carry out the kinetic  
145 experiments. DO was measured in the bulk liquid and it was maintained in excess to  
146 avoid oxygen limitations (around 9 mg O<sub>2</sub> L<sup>-1</sup>). Biomass was mixed both by mechanical  
147 stirring at 100 rpm (Stirrer type BS, VELP Scientifica, Italy) and bubbling of air to  
148 avoid mass transfer limitations. The pH was monitored and controlled at 7.5 by using an

149 ON/OFF control system by automated addition of 1 M NaOH with an automatic  
150 dispensing burette (Multi-Burette 2S-D, Crison Instruments, Spain). Temperature was  
151 maintained at 10 °C by means of a cooling system (E100, LAUDA, Germany), which  
152 provided cooled water through the jacket of the chemostat. The measurement of the  
153 particle size of the biomass of both the effluent and the reactor confirmed that biomass  
154 was not retained in the reactor and consequently the operation was as a chemostat (Fig  
155 SI-1 in Supporting Information).

156

157 Taking into account that the dilution rate is equal to the growth rate ( $\mu$ ) in a chemostat,  
158 growth rate was fixed by varying the dilution rate, and thus the inflow. The chemostat  
159 was operated continuously and experiments were finished when steady state conditions  
160 were achieved, that is, when TAN concentration in the effluent was constant. Five  
161 experiments were carried out at different growth rates ranging from 0.36 to 0.56 d<sup>-1</sup>, and  
162 a value of TAN concentration at steady state conditions was obtained for each  
163 experiment.

164

165 Kinetic parameters (maximum specific growth rate,  $\mu_{max}$ , and TAN affinity constant,  
166  $K_{S,TAN}$ ) were determined by fitting the data of the experiments to the Monod equation  
167 (Eq. 1).

168

$$169 \quad \mu = \mu_{max} \frac{[TAN]}{K_{S,TAN} + [TAN]} \quad (\text{Eq. 1})$$

170

#### 171 2.4. Analytical methods

172 Total ammonia nitrogen concentration was measured off-line with an ammonium  
173 analyzer (AMTAX sc, Hach Lange, Germany), total nitrite nitrogen (TNN = N-NO<sub>2</sub><sup>-</sup> +

174 N-HNO<sub>2</sub>) and nitrate concentrations were analyzed off-line with ionic chromatography  
175 using an ICS-2000 Integrated Reagent-Free IC system (DIONEX Corporation, USA).  
176 These measured off-line values are the ones represented in the results section. Mixed  
177 liquor total suspended solids (TSS) and mixed liquor volatile suspended solids (VSS)  
178 were analyzed according to Standard Methods (APHA, 1999). SRT was estimated by  
179 dividing the amount of VSS in the reactor by the sludge washed out with the effluent:

180

$$181 \quad SRT = \frac{[VSS]_{reactor} * V_{reactor}}{[VSS]_{effluent} * Q_{effluent}} \quad (1)$$

182

183 Where, [VSS]<sub>reactor</sub> and [VSS]<sub>effluent</sub> are the VSS concentrations in the reactor and the  
184 effluent, respectively, V<sub>reactor</sub> is the reactor volume and Q<sub>effluent</sub> is the effluent flow rate.

185

186 Average particle size was measured by a laser particle size analysis system (Malvern  
187 Mastersizer Series 2600, Malvern instruments Ltd., UK). The off-gas of the reactor was  
188 periodically collected and analyzed with gas chromatography (Agilent Technologies  
189 6890 N Network GC system, Madrid, Spain) to measure N<sub>2</sub>O emissions.

190

## 191 2.5. Fluorescence in situ hybridization (FISH) analysis

192 Abundances of AOB and NOB were analyzed by FISH coupled to confocal laser  
193 scanning microscopy (CLSM). Regarding AOB, specific probes for *Nitrosomonas* spp.  
194 and *Nitrospira* spp. were 5'-6FAM-labeled and 5'-Atto550-labeled, respectively.  
195 Regarding NOB, specific probes for *Nitrobacter* spp. and *Nitrospira* spp. were 5'-Cy3-  
196 labeled and 5'-6FAM-labeled, respectively. Hybridizations were performed with the  
197 specific and general (5'-Cy5-labeled) probes described in Table SI-1 in Supporting  
198 Information. Biomass samples were grabbed from the reactor and granules were crushed



199 by means of a mortar and a pestle in order to ease hybridization. A Leica TCS-SP5  
200 confocal laser scanning microscope (Leica Microsystem Heidelberg GmbH; Mannheim,  
201 Germany) using a Plan-Apochromatic 63x objective (NA 1.4, oil) was used to quantify  
202 biomass by analyzing 30-40 fields and following an automated image analysis  
203 procedure as described in Jubany et al. (2009).

204

## 205 2.6. Pyrosequencing analysis

206 Identification of the microbial population was performed using next-generation  
207 sequencing at day 98 and 233 of the reactor operation. DNA was extracted from  
208 biomass samples by applying the protocol of MoBio PowerBiofilm™ DNA extraction  
209 kit (MoBio Laboratories, USA). Two modifications of the manufacturer protocol were  
210 performed: 200 mL of solution BF3 were added instead of the 100 mL recommended,  
211 and 80 mL of solution BF7, instead of the 100 mL recommended. NanoDrop 1000  
212 Spectrophotometer (Thermo Fisher Scientific, USA) was used to measure the quantity  
213 and quality of extracted DNA. A minimum of 20 ng L<sup>-1</sup> of extracted DNA was  
214 guaranteed to perform pyrosequencing. Paired-end sequencing of the extracted DNA  
215 was performed on an Illumina MiSeq platform by Research and Testing Laboratory  
216 (Lubbock, Texas, USA). Bacterial 16S rRNA variable regions V2-V4 were targeted  
217 using the primer pair 341F-907R. More information about the bioinformatics applied to  
218 the sample can be found in Supporting Information.

219

## 220 2.7. Scanning electron microscopy

221 A biomass sample of 8-10 granules was fixed in 2.5 % (v/v) glutaraldehyde and 0.1 M  
222 phosphate buffer (pH 7.4) for 2 h at 4 °C, washed 4 times for 10 min each time in 0.1 M  
223 phosphate buffer, fixed in 1 % (wt/v) osmium tetroxide with 0.7 % ferrocyanide in

224 phosphate buffer, washed in water, dehydrated in an ascending ethanol series (50, 70,  
225 80, 90, and 95 % for 10 min each and twice with 100 % ethanol), and dried at critical-  
226 point with CO<sub>2</sub>. Then, the sample was metalized with Au-Pd and observed by using a  
227 scanning electron microscope (EVO MA10; Zeiss, Germany) at the following  
228 conditions: 20 kV, 100 pA, secondary electron detector (SE1).

229

### 230 3. Results and discussion

#### 231 3.1. Long-term operation at 10 °C

232 The reactor was previously operated at 12.5 °C, with an average NLR of  $0.7 \pm 0.3 \text{ g N L}^{-1}$   
233  $\text{d}^{-1}$ , for more than 400 days performing stable partial nitrification before the temperature  
234 was directly lowered to 10 °C (Isanta et al., 2015). After the decrease in temperature  
235 (day 0), the reactor was operated during 250 days with an average nitrogen loading rate  
236 (NLR) of  $0.63 \pm 0.06 \text{ g N L}^{-1} \text{ d}^{-1}$ . Stable partial nitrification was maintained in the long-  
237 term at 10 °C (Fig. 2), which was achieved by applying a ratio of DO/TAN  
238 concentrations in the bulk liquid of  $0.04 \pm 0.02 \text{ mg O}_2 \text{ mg}^{-1} \text{ N}$ . Low DO/TAN  
239 concentrations ratio was reported before to maintain stable partial nitrification in granular  
240 systems (Bartrolí et al., 2010; Isanta et al., 2015; Jemaat et al., 2013). Efficiency of the  
241 NOB repression is thought to be linked to the fact that a steep oxygen gradient is  
242 present in the granular sludge (Bartrolí et al., 2010; Isanta et al., 2015). Therefore, direct  
243 extrapolation of this strategy to other systems, such as flocculent sludge reactors in  
244 which sludge retention is assured by other means, it is not straightforward but might be  
245 object of future research. Nitrate production was barely detected, being the average  
246 concentration in the effluent of  $0.6 \pm 0.3 \text{ mg N-NO}_3^- \text{ L}^{-1}$ . Furthermore, a suitable effluent  
247 for a subsequent reactor performing the anammox process was achieved, with an  
248 average TNN/TAN concentrations ratio of  $1.1 \pm 0.2$ . The ammonium oxidation rate

249 (AOR) was maintained stable during the whole operation, with an average value of  
250  $0.34 \pm 0.06 \text{ g N L}^{-1} \text{ d}^{-1}$ . This value is considerably high compared to the one obtained in  
251 one-stage biofilm systems. Thus, Gilbert et al. (2015) reported an AOR lower than  $0.02$   
252  $\text{g N L}^{-1} \text{ d}^{-1}$  at  $10 \text{ }^\circ\text{C}$  and Hu et al. (2013) reported an AOR of  $0.03 \text{ g N L}^{-1} \text{ d}^{-1}$  at  $12 \text{ }^\circ\text{C}$ .

253

254 Particle size was maintained stable during the whole period of operation (Fig. 2A) with  
255 an average value of  $810 \pm 70 \text{ }\mu\text{m}$ . From day 50 onwards, the biomass concentration  
256 increased to an average value of  $3.6 \pm 0.1 \text{ g VSS L}^{-1}$ , as shown in Fig. 2A. In spite of the  
257 high and constant NLR and AOR achieved in the granular airlift reactor, specific rates  
258 (specific nitrogen loading rate, sNLR; specific ammonium oxidation rate, sAOR)  
259 decreased during the first 100 days at  $10 \text{ }^\circ\text{C}$  (Fig. 2C). However, sAOR remained  
260 constant from day 100 with an average value of  $0.18 \pm 0.03 \text{ g N mg}^{-1} \text{ VSS d}^{-1}$ . This fact  
261 demonstrated that biomass maintained the same activity during 150 days of operation at  
262  $10 \text{ }^\circ\text{C}$ .

263

264 Between days 93-95, 144-172 and 241-245 the ammonium control was switched from  
265 automatic to manual, but the process remained stable. Moreover, on day 98 the reactor  
266 remained without feeding for 4 hours (NLR of  $0 \text{ g N L}^{-1} \text{ d}^{-1}$ ) which resulted in an  
267 increase of DO and complete oxidation of ammonium to nitrite by AOB. Despite of the  
268 high concentration of TNN and DO, the nitrate in the bulk liquid was only  $2.2 \text{ mg L}^{-1}$   
269 and the next day the system was totally recovered. Hence, the successfully repression of  
270 NOB in the system was demonstrated and thus, the stability of this technology.

271

272 The balance of nitrogen during the operation of the reactor was fulfilled, with an  
273 average value of  $96 \pm 6 \%$  (Figure SI-2 in Supporting Information). Hence, neither

274 heterotrophic nor autotrophic (anammox process) denitrification was considered to take  
275 place in the granular airlift reactor.

276

277 Off-gas samples from days 94, 95, 241, 242 and 245 were analyzed in order to calculate  
278 the N<sub>2</sub>O emission factor of the reactor. As it is shown in Table 1, less than 0.35 % of the  
279 TAN in the influent was emitted as N-N<sub>2</sub>O. Furthermore, from the converted nitrogen  
280 the average emitted as N-N<sub>2</sub>O was 0.36±0.07 %. Thus, the N<sub>2</sub>O emissions from the  
281 granular airlift reactor performing partial nitrification of a low-strength wastewater at 10  
282 °C were very low, even at low DO concentrations (1.3±0.3 mg O<sub>2</sub> L<sup>-1</sup>) which is known  
283 to trigger high N<sub>2</sub>O emissions (Kampschreur et al., 2009). Applying the same control  
284 strategy but treating a reject water from the dewatering of digested sludge (high-strength  
285 wastewater) at 30°C, the N<sub>2</sub>O emission factor was as high as 6 % of the TAN oxidized  
286 at a DO of 1 mg O<sub>2</sub> L<sup>-1</sup> (Pijuan et al., 2014), which means more than one order of  
287 magnitude higher than the reported in this study. Hence, the temperature could be an  
288 important factor affecting N<sub>2</sub>O emissions, the lower the temperature the lower the  
289 emissions. Nevertheless further experiments are necessary to confirm this hypothesis,  
290 since other factors could cause the difference between the results of Pijuan et al. (2014)  
291 and this study.

292

### 293 3.2. Kinetics

294 As it was mentioned before, the granular airlift reactor not only achieved stable partial  
295 nitrification at 10 °C but also operated at higher NLR than other similar systems.  
296 Nitrifying capacity is related to the growth rate of AOB community and, hence, a  
297 nitrifying sludge with an unusually high maximum growth rate could explain the high  
298 activity in this airlift reactor.

299

300 The results of the five kinetic experiments carried out in a chemostat reactor are  
301 presented in Table 2. An increase in the applied growth rate caused an increase in the  
302 TAN concentration at steady state conditions, which follows satisfactorily ( $R^2 = 0.97$ )  
303 the Monod kinetic model (Figure 3). Hence, the maximum specific growth rate and  
304 TAN affinity constant were obtained by fitting the data achieved in each experiment to  
305 the Monod kinetic model. Thus  $\mu_{\max}$  and  $K_{S,TAN}$  resulted in  $0.63 \pm 0.05 \text{ d}^{-1}$  and  $2.1 \pm 0.7$   
306  $\text{mg N L}^{-1}$ , respectively.

307

308 Different values of maximum growth rate have been reported up to now, being most of  
309 them determined at high temperatures (20-30 °C) (Blackburne et al., 2007; Esquivel-  
310 Rios et al., 2014; Vadivelu et al., 2006). However, large discrepancies were found  
311 between different studies. The large variety of parameter values found in literature lies  
312 in the differences of systems evaluated, operational conditions applied, biomass growth  
313 types and the techniques used to determine the parameters themselves. Vanneck and  
314 Volcke (2015) presented a literature review on microbial characteristics of nitrifiers and  
315 reported  $\mu_{\max}$  in the range of  $0.34\text{-}3.40 \text{ d}^{-1}$  for attached growth of AOB at 30 °C and pH  
316 7.5. For suspended growth, Farges et al. (2012) used the flow cytometry technique to  
317 study the growth of *Nitrosomonas europaea* in pure cultures at 26 °C and pH 8 and  $\mu_{\max}$   
318 resulted in the range of  $0.13\text{-}0.23 \text{ d}^{-1}$ . On the other hand, Chandran et al. (2008)  
319 obtained higher values ( $\mu_{\max} = 0.24\text{-}0.74 \text{ d}^{-1}$ ) by using respirometric batch tests and  
320 substrate depletion assays in continuous reactors for an enriched nitrifying culture at 25  
321 °C and pH 7.4.

322

323 It is well known that growth rate decreases considerably with decreasing temperature.  
324 Knowles et al. (1965) reported a decrease in the  $\mu_{\max}$  from 1.5 to 0.2 d<sup>-1</sup> when  
325 temperature decreased from 27 to 8.3 °C for *Nitrosomonas* sp. in samples from Thames  
326 estuary, London, England; and afterwards, Sözen et al. (1996) determined  $\mu_{\max}$  in the  
327 range of 0.10-0.17 d<sup>-1</sup> for a nitrifying mixed culture treating real urban wastewater at 10  
328 °C. In spite of this, little has been published about kinetic parameters of nitrifying mixed  
329 cultures at low temperature, and in any case, the  $\mu_{\max}$  values reported were much lower  
330 than the one achieved in the current study ( $\mu_{\max} = 0.63 \pm 0.05$  d<sup>-1</sup> at 10 °C). In fact, to  
331 the best of the authors' knowledge, this is the highest growth rate achieved by a  
332 nitrifying sludge enriched in AOB at 10 °C. Furthermore, an estimation of the  $\mu_{\max}$  of  
333 the nitrifying sludge at higher temperatures was calculated by considering an Arrhenius-  
334 type equation ( $\mu_{1,T_1} = \mu_{2,T_2} \cdot \theta^{(T_1-T_2)}$ ) and a temperature coefficient of  $\theta = 1.13 \pm 0.03$   
335 which was determined in Isanta et al. (2015) for the inoculum of the current airlift  
336 reactor. The values obtained for  $\mu_{\max}$  were 2.1±0.2, 3.9±0.3 and 7.3±0.6 d<sup>-1</sup> at 20, 25  
337 and 30 °C, respectively. Thus, the nitrifying biomass of the granular airlift reactor  
338 presented the higher  $\mu_{\max}$  than has been reported hitherto at any temperature. A nitrifier  
339 culture with such a high  $\mu_{\max}$  could explain the high NLR and AOR achieved in the  
340 operation of the granular airlift reactor at 10 °C. A second implication is that the  
341 enrichment of an AOB population with such a high  $\mu_{\max}$  would be an advantage for  
342 NOB repression at low temperatures, since it would help to keep AOB growth rate  
343 higher than that of NOB.

344

345 Along with the high  $\mu_{\max}$  obtained, a high value for the TAN affinity constant was  
346 determined from the kinetic experiments ( $K_{S,TAN} = 2.1 \pm 0.7$  mg N L<sup>-1</sup>) compared to the  
347 previously reported by Chandran et al. (2008) at 25 °C ( $K_{S,TAN} = 0.21-0.69$  mg N L<sup>-1</sup>),

348 Knowles et al. (1965) at 8.3 °C ( $K_{S,TAN} = 0.2 \text{ mg N L}^{-1}$ ) and the proposed in the  
349 Activated Sludge Model 2d at 10 °C ( $K_{S,TAN} = 1 \text{ mg N L}^{-1}$ ; Henze et al, (2000)).

350

351 From the ecological concept, a microorganism showing quick growth on easily  
352 available substrate is defined as r-strategist microorganism (Andrews and Harris, 1986),  
353 which applied to the kinetic context represents a microorganism with high maximum  
354 specific growth rate and high substrate affinity constant (Andrews and Harris, 1986;  
355 Martín-Hernández et al., 2009). Therefore, the nitrifier population of the granular airlift  
356 reactor can be considered as r-strategist. The enrichment of a r-strategist AOB  
357 population has been reported when high residual ammonium concentrations are used  
358 (Terada et al., 2013). Hence, the high residual ammonium concentration may be also a  
359 key factor for the enrichment of r-strategist AOB population in two-stage partial  
360 nitrification/anammox reactor systems, like the one presented in this study.

361

### 362 3.3. Microbial characterization

363 FISH-CSLM was used to evaluate the enrichment in AOB and the presence of NOB in  
364 the granular sludge performing partial nitrification at 10 °C. On day 233, 92±4% of the  
365 population was quantified as AOB and less than 1±1% as NOB (specifically *Nitrobacter*  
366 spp). Since the inoculum contained 81±12% of AOB and 1±1% of *Nitrobacter* spp., a  
367 high enrichment in AOB and an effective repression of NOB was maintained in the  
368 long-term at 10 °C although NOB were always present in the biomass.

369

370 On the other hand, neither *Nitrosospira* spp. (species belonging to AOB) nor *Nitrosospira*  
371 spp. (species belonging to NOB) hybridizations were detected in the sludge. This fact  
372 was expected since they are k-strategist microorganisms and, consequently, they are not

373 favored at high TAN and TNN concentrations (Kim and Kim, 2006), such as those in  
374 the reactor of this study.

375

376 Moreover, pyrosequencing technique was used to examine the microbial community  
377 through the operation of the granular reactor at 10 °C. With that purpose, samples on  
378 days 98 and 233 were analyzed.

379

380 On sample from day 98, *Betaproteobacteria* was clearly the most abundant class of the  
381 total reads, with a relative abundance of 52 % (Fig 4). It is widely known that  
382 *Betaproteobacteria* class comprises autotrophic nitrifying microorganisms, such as  
383 AOB and NOB, and also denitrifying bacteria and organic matter decomposing bacteria.  
384 Thus, it is expected that *Betaproteobacteria* was the most abundant class in an AOB  
385 enriched sludge, such as the one of this study. *Alphaproteobacteria* was the second class  
386 in order of abundance, with a value of 23 %, following by *Actinobacteria* and  
387 *Gammaproteobacteria* representing the 7 and 5 % of total reads, respectively, among  
388 other classes of heterotrophs less abundant in the sample. These values of heterotrophic  
389 classes are in the range of the observed by Kindaichi et al. (2004), with 23 % of  
390 *Alphaproteobacteria* and 13 % of *Gammaproteobacteria* quantified in an autotrophic  
391 nitrifying biofilm system operating at 25 °C and with a high-strength synthetic  
392 wastewater.

393

394 On sample from day 233, corresponding to long-term operation of the granular reactor  
395 at 10 °C, *Betaproteobacteria* increased their relative abundance to 68%, followed by the  
396 increasing of *Cytophagia* with a 15 %; while *Alphaproteobacteria* sharply decreased to  
397 4 % (Fig. 4). There was a 10 % of reads not identified at class level, and most of them



398 comprised the phylum *Bacteroidetes*. In fact, *Bacteroidetes* abundance increased  
399 significantly compared to day 98, being the phylum with the highest increase (Figure  
400 SI-3 in Supporting Information).

401

402 At genus level, in the biomass community of day 98 (Fig. 5), *Nitrosomonas* was the  
403 most abundant genus, which was expected since the sludge was enriched in AOB and  
404 *Nitrosomonas* is the most frequently genus of AOB found in wastewater treatment  
405 systems (Wagner et al., 2002; Wang et al., 2012). Thus, *Nitrosomonas* genus counted up  
406 the 41% of the total population, indicating a majority of AOB in the sludge. Since  
407 *Nitrosomonas* genus comprises r-strategist microorganisms (Terada et al., 2013), their  
408 high abundance in the nitrifying sludge agreed with the high values of  $\mu_{\max}$  and  $K_{S,TAN}$   
409 obtained from kinetic experiments. Besides, *Nitrospira* genus was also detected in the  
410 sample with a 7 % of relative abundance, in spite of *Nitrospira* spp. were never  
411 detected by FISH technique. This may be due to the fact that FISH technique points  
412 toward the abundance of rRNA in samples, while pyrosequencing points toward the  
413 abundance of DNA (Wittebolle et al., 2005). Thus, *Nitrospira* spp. could be not  
414 detected by FISH because their probably low or null activity in the reactor, but their  
415 DNA could be still detected. Regarding NOB, 1.4 % of the total population was  
416 identified as *Nitrobacter* genus, which agrees with the production of nitrate in the  
417 reactor and with the result of the FISH analysis. Thus, *Nitrobacter* spp. were not  
418 abundant in the reactor, but active. Finally, *Nitrospira* genus was not detected in the  
419 sample, in agreement with the FISH analysis.

420

421 Other genera were detected in a low abundance, but in a more or less equal proportion  
422 between them. High bacteria richness is expected in mixed cultures and the coexistence

423 and interaction of heterotrophic bacteria and autotrophic nitrifiers were reported before  
424 (Ducey et al., 2010; Kindaichi et al., 2004; Okabe et al., 2005). In this sense, genera as  
425 *Sphingomonas* and *Dokdonella*, with a relative abundance in the sample of 8 and 4 %  
426 respectively, were reported as heterotrophic, or even autotrophic nitrifiers (Fitzgerald et  
427 al., 2015). Moreover, two other genera, *Cryobacterium* and *Flavobacterium*, with  
428 several species known to be either psychrotolerant (*Cryobacterium psychrotolerans*,  
429 Zhang et al. (2007); *Flavobacterium gelidilacus*, Van Trappen et al. (2003)) or even  
430 psychrophilic (*Cryobacterium* sp. MLB-32, Singh et al. (2015)) microorganisms, were  
431 identified in the sample with 7 and 5 % of the total reads, respectively.

432

433 On day 233 at genus level (Fig 5), enrichment in *Nitrosomonas* genus was observed to  
434 the detriment of the rest of genera. Moreover, *Nitrosomonas* genus counted the 65 % of  
435 the total reads in the sludge sample, being at day 233 the 96 % of all the  
436 *Betaproteobacteria* while at day 98 this percentage was of 78 %. Thus, the sludge was  
437 much more enriched in AOB at day 233 than at the beginning of the operation at 10 °C.  
438 *Nitrospira* genus was present in the sample in less than 0.5 % of abundance, so it was  
439 not considered for the data treatment. Since *Nitrospira* was found on day 98 (7 % of  
440 the reads), pyrosequencing analysis confirmed its wash-out from the granular airlift  
441 reactor operating at 10 °C. Furthermore, the absence of *Nitrospira* genus at day 233  
442 confirms that *Nitrospira* were not active on day 98 (no detected by FISH) and, hence,  
443 they were washed out of the reactor. Besides, since the reactor was operated at high  
444 solid retention time (80±20 days) the wash-out was slow. There may be two reasons for  
445 the wash-out of *Nitrospira* genus in the granular airlift reactor. The first one is that  
446 *Nitrospira* spp. were not favoured at the operating conditions of the reactor since they  
447 are k-strategist microorganisms (low TAN affinity constant and low specific growth

448 rate) while *Nitrosomonas* spp. are r-strategists (high TAN affinity constant and high  
449 specific growth rate). The second possible explanation is that *Nitrospira* spp. are  
450 more sensitive to temperature than *Nitrosomonas* spp (Hoang et al., 2014; Park et al.,  
451 2008).

452

453 Neither *Nitrobacter* nor *Nitrospira* genera were identified by pyrosequencing in the  
454 sample of day 233, in spite of being detected with the FISH analysis (with an abundance  
455 of  $1\pm 1$  %). Probably this could be due to poor or null amplification of a low DNA  
456 content of these bacteria in the sample. In any case, successful NOB repression in the  
457 granular airlift reactor operating at 10 °C was demonstrated.

458

459 As it is shown in Fig. 5, the second genus in order of abundance with a 15% of the total  
460 reads was unclassified at genus level (classified at order level as *Cytophagales*),  
461 however its DNA sequence was ran against BLAST and matched the one found by  
462 Larose et al. (2010) in bacteria present in snow and melt water samples from Svalbard,  
463 Norway. Therefore, the corresponding microorganism will probably be a cold-adapted  
464 microorganism (psychrotolerant or psychrophilic species) and its presence in cold  
465 waters would fit with the presence in the 10 °C system of this study.

466

467 In general terms, in addition to the enrichment in AOB, three main points can be  
468 extracted from the results obtained by pyrosequencing in this study. The first one is that  
469 the diversity of the bacterial community decreased in the long-term of operation at 10  
470 °C. The second one is that despite the fact that the influent of the reactor was devoid of  
471 an organic carbon source, a considerable part of the population in both samples was

472 composed by heterotrophic bacteria. Finally, the third one is the presence of  
473 psychrotolerant microorganisms in the sludge performing partial nitrification at 10 °C.

474

475 As shown in Table 3, there were more genera with abundances higher than 5 % on day  
476 98 than on day 233. The decrease in bacterial diversity with cold temperature was  
477 reported before (Karkman et al., 2011). Thus, not only non-adapted microorganisms to  
478 cold temperatures diminished in the long-term operation, but also diversity of  
479 psychrotolerant genera (the unclassified microorganism mentioned before appears to the  
480 detriment of *Cryobacterium* and *Flavobacterium*). Only *Nitrosomonas* and the  
481 unclassified genera (one of them corresponding to the cold-adapted microorganism  
482 mentioned before) were identified with abundance superior to 5 % on day 233.

483

484 The coexistence of nitrifying and heterotrophic bacteria in absence of organic carbon  
485 has also been reported before (Ducey et al., 2010; Hoang et al., 2014; Karkman et al.,  
486 2011). It is known that nitrifiers produce organic matter from biomass decay and  
487 substrate metabolism which is used by heterotrophs to survive. Nogueira et al. (2005)  
488 correlated the presence of heterotrophs in nitrifying biofilm reactors with the hydraulic  
489 retention time (HRT) and determined that values of HRT in the range of the one used in  
490 this study ( $2.5 \pm 0.3$  h) guarantees enough soluble microbial products (SMP) available  
491 for heterotrophic growth. There are also studies focused on the determination of these  
492 SMP derived from nitrifiers that can be used by heterotrophs (Kindaichi et al., 2004;  
493 Okabe et al., 2005).

494

495 Interaction between nitrifiers and heterotrophs is not only profitable for heterotrophic  
496 bacteria, but also for nitrifiers, becoming a synergic system as it was suggested by

497 Ducey et al. (2010) and Hoang et al. (2014). Some psychrotolerant and psychrophilic  
498 bacteria have been identified as producers of cryoprotective extracellular polymeric  
499 substances (EPS) that allow a better survival of the whole consortium at cold  
500 temperatures (Ducey et al., 2010). Therefore, this protection would affect in the same  
501 way to AOB, which could maintain the nitrification even when conditions were not ideal  
502 for their growth. Psychrotolerant microorganisms were found in both samples of the  
503 granular sludge and thus, they were present during the whole operation of the granular  
504 reactor at 10 °C. Hence, although in our system the heterotrophic population was less  
505 significant than that of nitrifiers, its presence could be essential for the maintenance of  
506 the nitrification in the granular reactor. Fig. 6 shows a SEM image of the surface of a  
507 granule where bacteria seem to be embedded in a high amount of extracellular  
508 polymeric substances, which could be the cryoprotective EPS.

509

510 This study revealed high nitrifier ability for performing partial nitrification at 10 °C:  
511 stable operation was maintained in the long-term in the granular airlift reactor and  
512 kinetic experiments showed the higher  $\mu_{\max}$  than has been hitherto determined for a  
513 nitrifying sludge at low temperature. We have two hypotheses to explain it: (i) AOB  
514 were cultivated in the long-term under low temperatures which could lead to a  
515 metabolic adjustment of the biomass and thus, to improve the ability to nitrify under this  
516 condition; (ii) granules comprised a consortium of microorganisms which included  
517 producers of cryoprotective EPS that give an adaptive advantage to AOB, protecting  
518 them from low temperatures.

519

520

521

522 4. Conclusions

523 Stable partial nitrification at 10 °C was maintained in the long-term in a granular airlift  
524 reactor operating at high NLR.

525

526 The nitrifier culture enriched in AOB presented a significantly high  $\mu_{\max}$  compared to  
527 other studies which allowed the operation at high nitrification rates, being advantageous  
528 for NOB repression.

529

530 The microbial community was dominated by AOB (specifically *Nitrosomonas* genus)  
531 throughout the whole operation of the reactor; while NOB genera were barely detected,  
532 demonstrating their effective repression from the system. Effective NOB repression was  
533 not only achieved, but also it was obtained a suitable effluent for a subsequent reactor  
534 performing the anammox process.

535

536 The operation of the granular reactor in the long-term at 10 °C with a high residual  
537 ammonium concentration decreased the microbial diversity and further enriched the  
538 granular sludge in AOB.

539

540 Partial nitrification at 10 °C can be operated with low N<sub>2</sub>O emissions since less than 0.35  
541 % of the TAN in the influent was emitted as N-N<sub>2</sub>O.

542

543 Acknowledgements

544 This study was supported by the Spanish Ministerio de Economía y Competitividad  
545 through the DESDEMONA project (CTQ2014-60495-R). The authors are members of  
546 the GENOCOV research group (2014 SGR 1255). J. Pérez acknowledges the Marie

547 Curie Intra European Fellowship (GreenN2, PIEF-GA-2012-326705) within the 7<sup>th</sup>  
548 European Community Framework Programme. The authors thank Ben A. Abbas (Dept.  
549 of Biotechnology, Technical University of Delft, The Netherlands) for helpful  
550 discussions about pyrosequencing analysis.

551

## 552 References

553 APHA, 1999. Standard Methods for the Examination of Water and Wastewater. 20th  
554 ed. American Water Works Association (AWWA) and Water Environmental Federation  
555 (WEF), Washington, DC, USA

556 Andrews, J., Harris, R., 1986. r- and K-Selection and Microbial Ecology, in: Marshall,  
557 K.C. (Ed.), *Advances in Microbial Ecology*. Springer US, pp. 99–147. doi:10.1007/978-  
558 1-4757-0611-6\_3

559 Bartrolí, A., Pérez, J., Carrera, J., 2010. Applying ratio control in a continuous granular  
560 reactor to achieve full nitrification under stable operating conditions. *Environ. Sci.*  
561 *Technol.* 44, 8930–8935.

562 Blackburne, R., Vadivelu, V.M., Yuan, Z., Keller, J., 2007. Determination of growth  
563 rate and yield of nitrifying bacteria by measuring carbon dioxide uptake rate. *Water*  
564 *Environ. Res.* 79, 2437–2445. doi:10.2175/106143007X212139

565 Chandran, K., Hu, Z., Smets, B.F., 2008. A critical comparison of extant batch  
566 respirometric and substrate depletion assays for estimation of nitrification biokinetics.  
567 *Biotechnol. Bioeng.* 101, 62–72. doi:10.1002/bit.21871

568 De Clippeleir, H., Vlaeminck, S.E., De Wilde, F., Daeninck, K., Mosquera, M., Boeckx,  
569 P., Verstraete, W., Boon, N., 2013. One-stage partial nitrification/anammox at 15 °C on  
570 pretreated sewage: feasibility demonstration at lab-scale. *Appl. Microbiol. Biotechnol.*  
571 97, 10199–210. doi:10.1007/s00253-013-4744-x

572 Ducey, T.F., Vanotti, M.B., Shriner, A.D., Szogi, A. a., Ellison, A.Q., 2010.  
573 Characterization of a microbial community capable of nitrification at cold temperature.  
574 Bioresour. Technol. 101, 491–500. doi:10.1016/j.biortech.2009.07.091  
575 Esquivel-Rios, I., Ramirez-Vargas, R., Hernandez-Martinez, G.R., Vital-Jacome, M.,  
576 Ordaz, A., Thalasso, F., 2014. A microrespirometric method for the determination of  
577 stoichiometric and kinetic parameters of heterotrophic and autotrophic cultures.  
578 Biochem. Eng. J. 83, 70–78. doi:10.1016/j.bej.2013.12.006  
579 Farges, B., Poughon, L., Roriz, D., Creuly, C., Dussap, C.G., Lasseur, C., 2012. Axenic  
580 cultures of *Nitrosomonas europaea* and *Nitrobacter winogradskyi* in autotrophic  
581 conditions: A new protocol for kinetic studies. Appl. Biochem. Biotechnol. 167, 1076–  
582 1091. doi:10.1007/s12010-012-9651-6  
583 Fitzgerald, C.M., Camejo, P., Oshlag, J.Z., Noguera, D.R., 2015. Ammonia-oxidizing  
584 microbial communities in reactors with efficient nitrification at low-dissolved oxygen.  
585 Water Res. 70, 38–51. doi:10.1016/j.watres.2014.11.041  
586 Gao, D.-W., Lu, J.-C., Liang, H., 2014. Simultaneous energy recovery and autotrophic  
587 nitrogen removal from sewage at moderately low temperatures. Appl. Microbiol.  
588 Biotechnol. 98, 2637–2645. doi:10.1007/s00253-013-5237-7  
589 Gilbert, E.M., Agrawal, S., Karst, S.M., Horn, H., Nielsen, P.H., Lackner, S., 2014.  
590 Low temperature partial nitritation/anammox in a moving bed bio fi lm reactor treating  
591 low strength wastewater. Environ. Sci. Technol. 48, 8784-8792.  
592 Gilbert, E.M., Agrawal, S., Schwartz, T., Horn, H., Lackner, S., 2015. Comparing  
593 different reactor configurations for Partial Nitritation/Anammox at low temperatures.  
594 Water Res. 81, 92–100. doi:10.1016/j.watres.2015.05.022  
595 Guerrero, J., Guisasola, A., Baeza, J.A., 2011. The nature of the carbon source rules the  
596 competition between PAO and denitrifiers in systems for simultaneous biological



597 nitrogen and phosphorus removal. *Water Res.* 45, 4793–802.  
598 doi:10.1016/j.watres.2011.06.019

599 Henze, M., Gujer, W., Mino, T., van Loosdrecht, M. C., 2000. *Activated Sludge Models*  
600 *ASM1, ASM2, ASM2d, and ASM3*. IWA Publishing, London.

601 Hoang, V., Delatolla, R., Abujamel, T., Mottawea, W., Gadbois, a., Laflamme, E.,  
602 Stintzi, a., 2014. Nitrifying moving bed biofilm reactor (MBBR) biofilm and biomass  
603 response to long term exposure to 1°C. *Water Res.* 49, 215–224.  
604 doi:10.1016/j.watres.2013.11.018

605 Hu, Z., Lotti, T., de Kreuk, M., Kleerebezem, R., van Loosdrecht, M., Kruit, J., Jetten,  
606 M.S.M., Kartal, B., 2013. Nitrogen removal by a nitrification-anammox bioreactor at low  
607 temperature. *Appl. Environ. Microbiol.* 79, 2807–12. doi:10.1128/AEM.03987-12

608 Hunik, J.H., Bos, C.G., van den Hoogen, M.P., De Gooijer, C.D., Tramper, J., 1994.  
609 Co-immobilized *Nitrosomonas europaea* and *Nitrobacter agilis* cells: validation of a  
610 dynamic model for simultaneous substrate conversion and growth in kappa-carrageenan  
611 gel beads. *Biotechnol. Bioeng.* 43, 1153–63. doi:10.1002/bit.260431121

612 Isanta, E., Reino, C., Pérez, J., Carrera, J., 2015. Stable partial nitrification for low  
613 strength wastewater at low temperature in an aerobic granular reactor. *Water Res.*  
614 doi:10.1016/j.watres.2015.04.028

615 Jemaat, Z., Bartrolí, A., Isanta, E., Carrera, J., Suárez-Ojeda, M.E., Pérez, J., 2013.  
616 Closed-loop control of ammonium concentration in nitrification: convenient for reactor  
617 operation but also for modeling. *Bioresour. Technol.* 128, 655–63.  
618 doi:10.1016/j.biortech.2012.10.045

619 Jetten, M., Horn, S., van Loosdrecht, M., 1997. Towards a more sustainable municipal  
620 wastewater treatment system. *Water Sci. Technol.* 35, 171–180. doi:10.1016/S0273-  
621 1223(97)00195-9

622 Jubany, I., Lafuente, J., Carrera, J., Baeza, J.A., 2009. Automated thresholding method  
623 (ATM) for biomass fraction determination using FISH and confocal microscopy. *J.*  
624 *Chem. Technol. Biotechnol.* 84, 1140–1145. doi:10.1002/jctb.2146

625 Kampschreur, M.J., Temmink, H., Kleerebezem, R., Jetten, M.S.M., van Loosdrecht,  
626 M.C.M., 2009. Nitrous oxide emission during wastewater treatment. *Water Res.* 43,  
627 4093–103. doi:10.1016/j.watres.2009.03.001

628 Karkman, a., Mattila, K., Tamminen, M., Virta, M., 2011. Cold temperature decreases  
629 bacterial species richness in nitrogen-removing bioreactors treating inorganic mine  
630 waters. *Biotechnol. Bioeng.* 108, 2876–2883. doi:10.1002/bit.23267

631 Kartal, B., Kuenen, J.G., van Loosdrecht, M.C.M., 2010. Engineering. Sewage  
632 treatment with anammox. *Science* 328, 702–3. doi:10.1126/science.1185941

633 Kim, D.-J., Kim, S.-H., 2006. Effect of nitrite concentration on the distribution and  
634 competition of nitrite-oxidizing bacteria in nitrification reactor systems and their kinetic  
635 characteristics. *Water Res.* 40, 887–94. doi:10.1016/j.watres.2005.12.023

636 Kindaichi, T., Ito, T., Okabe, S., 2004. Ecophysiological Interaction between Nitrifying  
637 Bacteria and Heterotrophic Bacteria in Autotrophic Nitrifying Biofilms as Determined  
638 by Microautoradiography-Fluorescence In Situ Hybridization. *Appl. Environ.*  
639 *Microbiol.* 70, 1641–1650. doi:10.1128/AEM.70.3.1641-1650.2004

640 Knowles, G., Downing, a L., Barrett, M.J., 1965. Determination of kinetic constants for  
641 nitrifying bacteria in mixed culture, with the aid of an electronic computer. *J. Gen.*  
642 *Microbiol.* 38, 263–278. doi:10.1099/00221287-38-2-263

643 Larose, C., Berger, S., Ferrari, C., Navarro, E., Dommergue, A., Schneider, D., Vogel,  
644 T.M., 2010. Microbial sequences retrieved from environmental samples from seasonal  
645 Arctic snow and meltwater from Svalbard, Norway. *Extremophiles* 14, 205–212.  
646 doi:10.1007/s00792-009-0299-2

647 Lotti, T., Kleerebezem, R., van Erp Taalman Kip, C., Hendrickx, T.L.G., Kruit, J.,  
648 Hoekstra, M., van Loosdrecht, M.C.M., 2014. Anammox growth on pretreated  
649 municipal wastewater. *Environ. Sci. Technol.* 48, 7874–80. doi:10.1021/es500632k

650 Ma, B., Zhang, S., Zhang, L., Yi, P., Wang, J., Wang, S., Peng, Y., 2011. The feasibility  
651 of using a two-stage autotrophic nitrogen removal process to treat sewage. *Bioresour.*  
652 *Technol.* 102, 8331–4. doi:10.1016/j.biortech.2011.06.017

653 Martín-Hernández, M., Carrera, J., Pérez, J., Suárez-Ojeda, M.E., 2009. Enrichment of a  
654 K-strategist microbial population able to biodegrade p-nitrophenol in a sequencing  
655 batch reactor. *Water Res.* 43, 3871–3883. doi:10.1016/j.watres.2009.06.001

656 Nogueira, R., Elenter, D., Brito, a, Melo, L.F., Wagner, M., Morgenroth, E., 2005.  
657 Evaluating heterotrophic growth in a nitrifying biofilm reactor using fluorescence in situ  
658 hybridization and mathematical modeling. *Water Sci. Technol.* 52, 135–141.

659 Okabe, S., Kindaichi, T., Ito, T., 2005. Fate of C-Labeled Microbial Products Derived  
660 from Nitrifying Bacteria in Autotrophic Nitrifying Biofilms 3987–3994.  
661 doi:10.1128/AEM.71.7.3987

662 Park, J., Byun, I., Park, S., Park, T., 2008. Nitrifying bacterial communities and its  
663 activities in aerobic biofilm reactors under different temperature conditions. *Korean J.*  
664 *Chem. Eng.* 25, 1448–1455. doi:10.1007/s11814-008-0238-4

665 Pérez, J., Isanta, E., Carrera, J., 2015. Would a two-stage N-removal be a suitable  
666 technology to implement at full scale the use of anammox for sewage treatment? *Water*  
667 *Sci. Technol.* 72, 858. doi:10.2166/wst.2015.281

668 Pijuan, M., Torà, J., Rodríguez-Caballero, A., César, E., Carrera, J., Pérez, J., 2014.  
669 Effect of process parameters and operational mode on nitrous oxide emissions from a  
670 nitrification reactor treating reject wastewater. *Water Res.* 49, 23–33.  
671 doi:10.1016/j.watres.2013.11.009

672 Regmi, P., Miller, M.W., Holgate, B., Bunce, R., Park, H., Chandran, K., Wett, B.,  
673 Murthy, S., Bott, C.B., 2014. Control of aeration, aerobic SRT and COD input for  
674 mainstream nitrification/denitrification. *Water Res.* 57, 162–71.  
675 doi:10.1016/j.watres.2014.03.035

676 Siegrist, H., Salzgeber, D., Eugster, J., Joss, A., 2008. Anammox brings WWTP closer  
677 to energy autarky due to increased biogas production and reduced aeration energy for  
678 N-removal. *Water Sci. Technol.* 57, 383–388.

679 Singh, P., Kapse, N., Arora, P., Singh, S.M., Dhakephalkar, P.K., 2015. Draft genome  
680 of *Cryobacterium* sp. MLB-32, an obligate psychrophile from glacier cryoconite holes  
681 of high Arctic. *Mar. Genomics* 21, 25–26. doi:10.1016/j.margen.2015.01.006

682 Sözen, S., Orhon, D., San, H.A., 1996. A new approach for the evaluation of the  
683 maximum specific growth rate in nitrification. *Water Res.* 30, 1661–1669.  
684 doi:10.1016/0043-1354(96)00031-0

685 Terada, A., Sugawara, S., Yamamoto, T., Zhou, S., Koba, K., Hosomi, M., 2013.  
686 Physiological characteristics of predominant ammonia-oxidizing bacteria enriched from  
687 bioreactors with different influent supply regimes. *Biochem. Eng. J.* 79, 153–161.  
688 doi:10.1016/j.bej.2013.07.012

689 Vadivelu, V.M., Keller, J., Yuan, Z., 2006. Stoichiometric and kinetic characterisation  
690 of *Nitrosomonas* sp. in mixed culture by decoupling the growth and energy generation  
691 processes. *J. Biotechnol.* 126, 342–56. doi:10.1016/j.jbiotec.2006.04.017

692 Van Hulle, S.W.H., Vandeweyer, H.J.P., Meesschaert, B.D., Vanrolleghem, P.A.,  
693 Dejana, P., Dumoulin, A., 2010. Engineering aspects and practical application of  
694 autotrophic nitrogen removal from nitrogen rich streams. *Chem. Eng. J.* 162, 1–20.  
695 doi:10.1016/j.cej.2010.05.037

696 Van Trappen, S., Mergaert, J., Swings, J., 2003. *Flavobacterium gelidilacus* sp. nov.,  
697 isolated from microbial mats in Antarctic lakes. *Int. J. Syst. Evol. Microbiol.* 53, 1241–  
698 1245. doi:10.1099/ijs.0.02583-0

699 Vannecke, T.P.W., Volcke, E.I.P., 2015. Modelling microbial competition in nitrifying  
700 biofilm reactors. *Biotechnol. Bioeng.* 9999, n/a–n/a. doi:10.1002/bit.25680

701 Wagner, M., Loy, A., Nogueira, R., Purkhold, U., Lee, N., Daims, H., 2002. Microbial  
702 community composition and function in wastewater treatment plants. *Antonie Van*  
703 *Leeuwenhoek* 81, 665–680. doi:10.1023/a:1020586312170

704 Wang, F., Liu, Y., Ma, Y., Wu, X., Yang, H., 2012. Characterization of nitrification and  
705 microbial community in a shallow moss constructed wetland at cold temperatures. *Ecol.*  
706 *Eng.* 42, 124–129. doi:10.1016/j.ecoleng.2012.01.006

707 Wett, B., Omari, a., Podmirseg, S.M., Han, M., Akintayo, O., Gómez Brandón, M.,  
708 Murthy, S., Bott, C., Hell, M., Takács, I., Nyhuis, G., O’Shaughnessy, M., 2013. Going  
709 for mainstream deammonification from bench to full scale for maximized resource  
710 efficiency. *Water Sci. Technol.* 68, 283–289. doi:10.2166/wst.2013.150

711 Winkler, M.K.H., Kleerebezem, R., Kuenen, J.G., Yang, J., Loosdrecht, M.C.M. Van,  
712 2011. Segregation of biomass in cyclic anaerobic / aerobic granular sludge allows the  
713 enrichment of anaerobic ammonium oxidizing bacteria at low temperatures. *Environ.*  
714 *Sci. Technol.* 7330–7337.

715 Wittebolle, L., Boon, N., Vanparys, B., Heylen, K., De Vos, P., Verstraete, W., 2005.  
716 Failure of the ammonia oxidation process in two pharmaceutical wastewater treatment  
717 plants is linked to shifts in the bacterial communities. *J. Appl. Microbiol.* 99, 997–1006.  
718 doi:10.1111/j.1365-2672.2005.02731.x

719 Zhang, D.C., Wang, H.X., Cui, H.L., Yang, Y., Liu, H.C., Dong, X.Z., Zhou, P.J., 2007.  
720 *Cryobacterium psychrotolerans* sp. nov., a novel psychrotolerant bacterium isolated

721 from the China No. 1 glacier. *Int. J. Syst. Evol. Microbiol.* 57, 866–869.

722 doi:10.1099/ijs.0.64750-0

723

724 Figure captions and table legends

725

726 Fig. 1 Schematic diagram of the reactor set-up showing the peripheral instrumentation

727 and control loops. DO: dissolved oxygen; TAN: total ammonia nitrogen (TAN = N-

728  $\text{NH}_4^+ + \text{N-NH}_3$ ).

729

730 Fig. 2 Continuous operation of the granular airlift reactor treating a synthetic low-

731 strength wastewater at 10 °C. (A) Biomass concentration and particle size; (B) Nitrogen

732 loading rate (NLR), and ammonium oxidation rate (AOR); (C) Specific nitrogen

733 loading rate (sNLR), specific ammonium oxidation rate (sAOR) and dissolved oxygen

734 concentration (DO); (D) Nitrogen compounds concentrations throughout the operation

735 of the granular reactor.

736

737 Fig. 3 TAN oxidation kinetics for the nitrifying granules. (○) TAN concentration at

738 steady state conditions for each specific growth rate imposed.

739

740 Fig. 4. Microbial diversity at class level. Relative abundance was calculated only

741 considering those microorganisms in which the number of 16S copies was higher than

742 0.5 % of the total copies.

743

744 Fig. 5. Microbial diversity at genus level. Relative abundance was calculated only  
745 considering those microorganisms in which the number of 16S copies was higher than  
746 0.5 % of the total copies.

747

748 Fig. 6 Scanning electron microscopy (SEM) images of a granule surface. Granule  
749 sample was taken on day 45 of operation at 10 °C. (A) 1,000x magnification; (B)  
750 10,000x magnification; (C) 20,000x magnification.

751

752 Table 1 – N<sub>2</sub>O emission factors during the operation of the granular airlift reactor  
753 treating a synthetic low-strength wastewater at 10 °C.

754

755 Table 2 – Operational parameters of the kinetic experiments in the chemostat at steady  
756 state conditions. All the experiments were conducted with the same influent of the main  
757 reactor at pH = 7.5±0.1, DO = 9.3±0.1 mg O<sub>2</sub> L<sup>-1</sup> and T = 10 °C.

758

759 Table 3 – Genera with relative abundance higher than 5 % in samples of sludge on day  
760 98 and 233.

Table 1

Day	N-N <sub>2</sub> O (% of N-influent)	N-N <sub>2</sub> O (% of N-oxidized)
94	0.13±0.01	0.23±0.02
95	0.32±0.04	0.58±0.08
241	0.23±0.04	0.40±0.08
242	0.17±0.06	0.30±0.10
245	0.14±0.02	0.28±0.04



**Table 2**

Number of Experiment	Experiment duration (days)	$\mu$ ( $\text{d}^{-1}$ )	Inflow ( $\text{L d}^{-1}$ )	$[\text{TAN}]_{\text{effluent}}$ ( $\text{mg N L}^{-1}$ )	$[\text{TNN}]_{\text{effluent}}$ ( $\text{mg N L}^{-1}$ )	$[\text{N-NO}_3^-]_{\text{effluent}}$ ( $\text{mg N L}^{-1}$ )
1	13	0.55	$1.60 \pm 0.01$	$26.3 \pm 0.7$	$49.0 \pm 2.0$	$1.9 \pm 0.1$
2	10	0.34	$1.00 \pm 0.01$	$3.4 \pm 0.2$	$30.0 \pm 4.0$	$36.0 \pm 1.0$
3	13	0.53	$1.53 \pm 0.01$	$10.2 \pm 0.6$	$47.0 \pm 2.0$	$10.8 \pm 0.4$
4	9	0.56	$1.62 \pm 0.01$	$10.0 \pm 1.0$	$56.0 \pm 2.0$	$12.0 \pm 1.0$
5	16	0.45	$1.30 \pm 0.01$	$4.3 \pm 0.4$	$34.0 \pm 9.0$	$31.0 \pm 7.0$

Table 3

Day 98		Day 233	
<i>Nitrosomonas</i>	(41%)	<i>Nitrosomonas</i>	(65%)
<i>Nitrospira</i>	(7%)	Unclassified ( <i>Cytophagales</i> Order)	(15%)
<i>Sphingomonas</i>	(8%)	Unclassified ( <i>Bacteroidetes</i> Phylum)	(8%)
<i>Sphingopyxis</i>	(5%)		
<i>Cryobacterium</i>	(7%)		
<i>Flavobacterium</i>	(5%)		
<i>Comamonas</i>	(5%)		

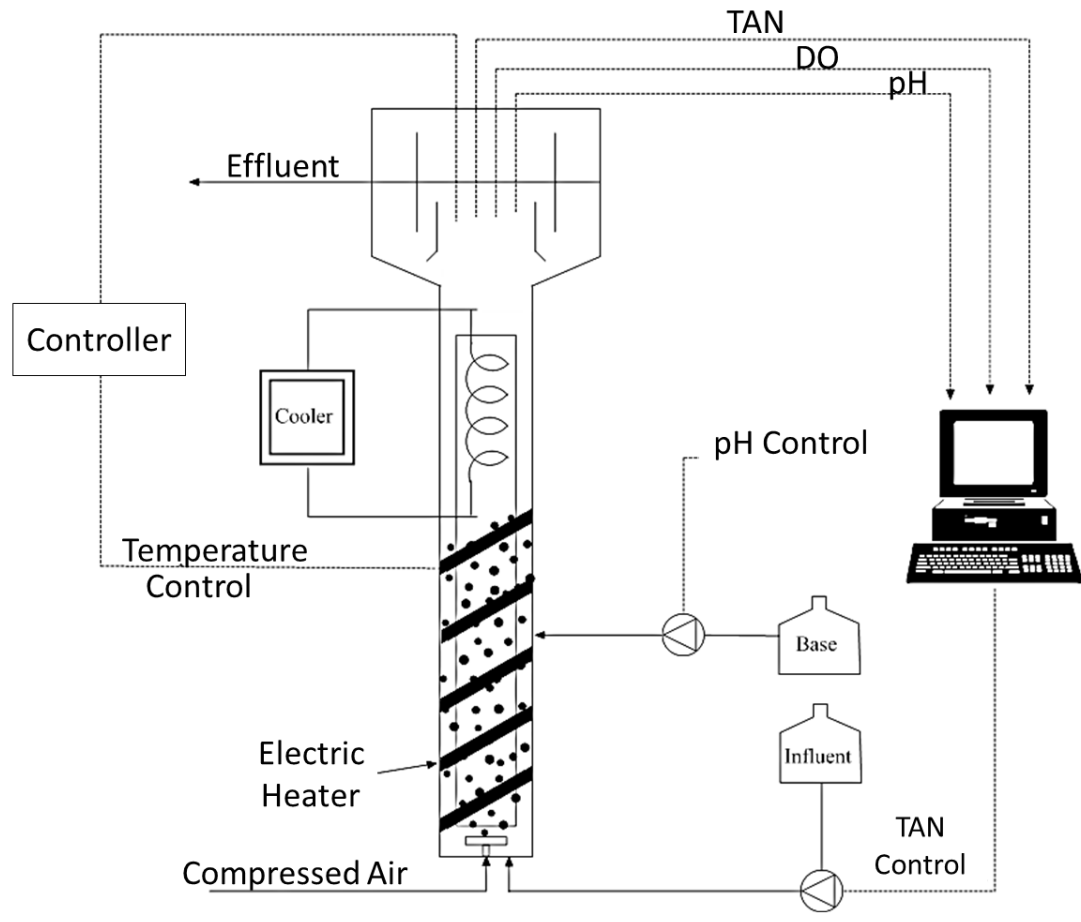


Figure 1

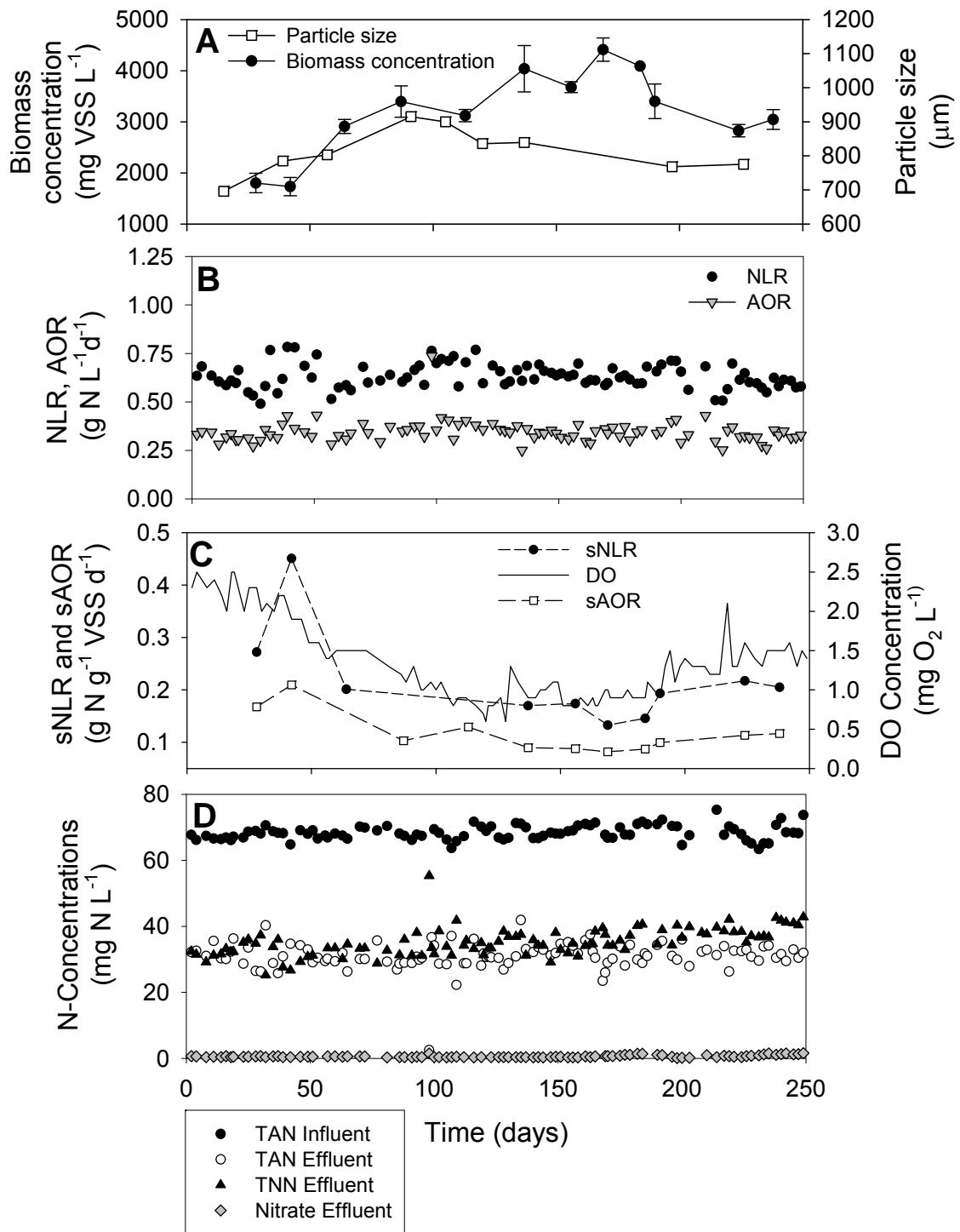


Figure 2

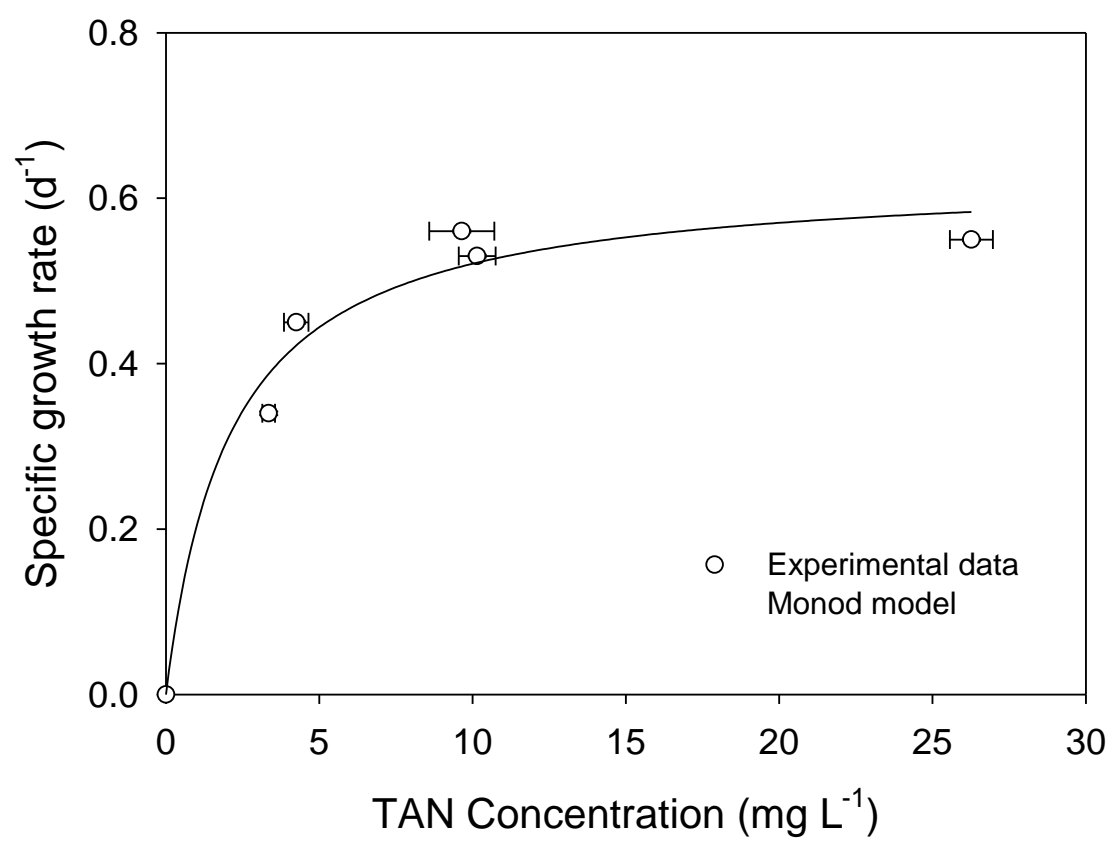


Figure 3

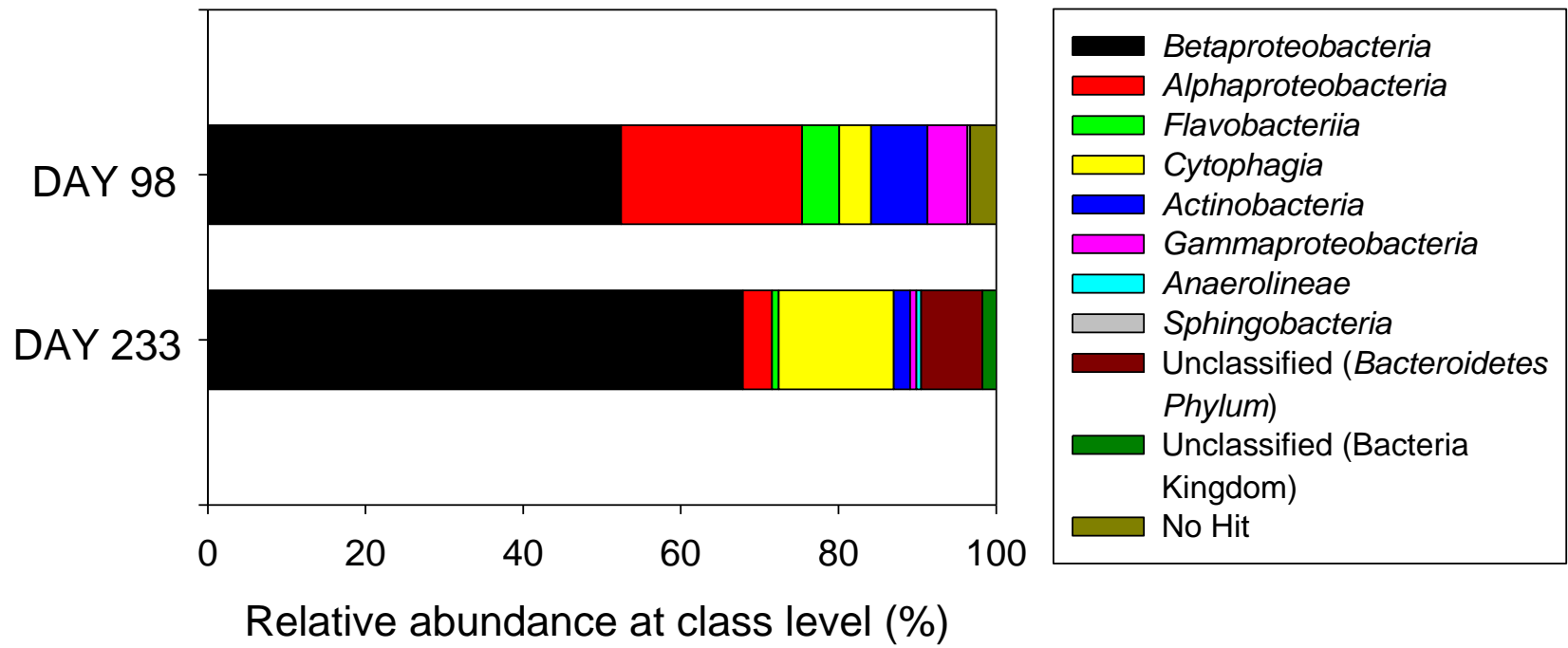


Figure 4

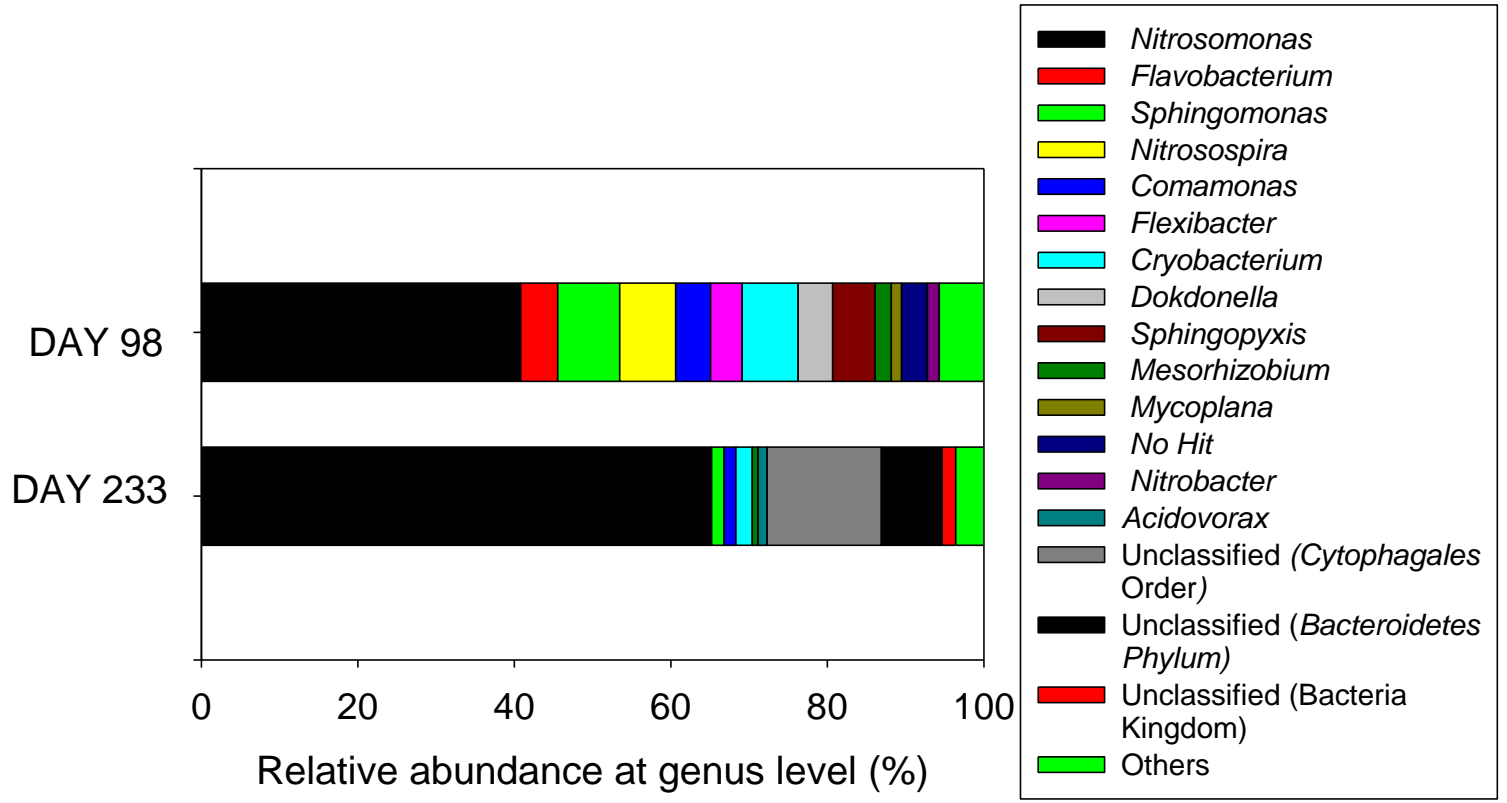


Figure 5

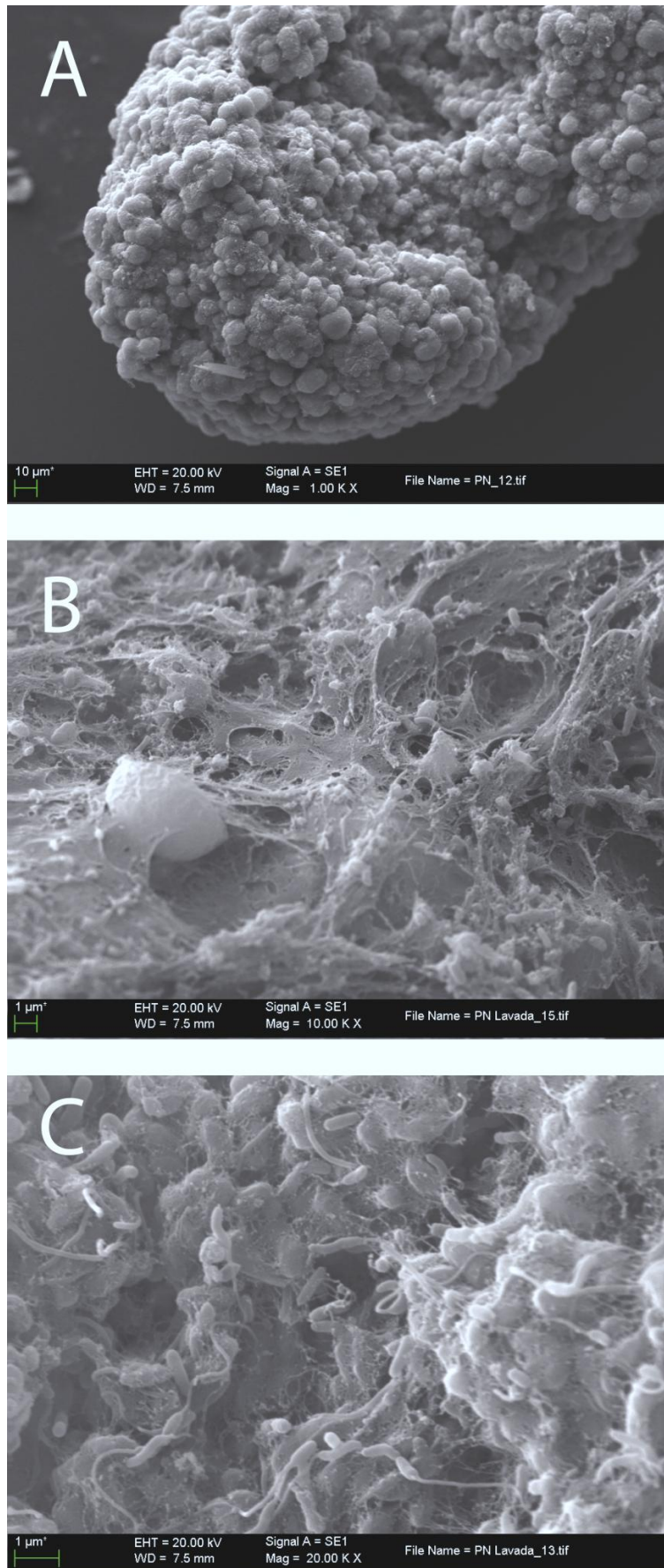


Figure 6