

REVIEW ARTICLE

Exceptionally widespread nanomachines composed of type IV pilins: the prokaryotic Swiss Army knives

Jamie-Lee Berry and Vladimir Pelicic*

MRC Centre for Molecular Bacteriology and Infection, Imperial College London, London SW7 2AZ, UK

*Corresponding author: Vladimir Pelicic, MRC Centre for Molecular Bacteriology and Infection, Imperial College London, London SW7 2AZ, UK.
Tel: +44 20 7594 2080; E-mail: v.pelicic@imperial.ac.uk

One-sentence summary: Using type IV pili as a paradigm, we review common genetic, structural and mechanistic features (many) as well as differences (few) of the exceptionally widespread and functionally versatile prokaryotic nano-machines composed of type IV pilins.

Editor: Prof. Alain Filloux

ABSTRACT

Prokaryotes have engineered sophisticated surface nanomachines that have allowed them to colonize Earth and thrive even in extreme environments. Filamentous machineries composed of type IV pilins, which are associated with an amazing array of properties ranging from motility to electric conductance, are arguably the most widespread since distinctive proteins dedicated to their biogenesis are found in most known species of prokaryotes. Several decades of investigations, starting with type IV pili and then a variety of related systems both in bacteria and archaea, have outlined common molecular and structural bases for these nanomachines. Using type IV pili as a paradigm, we will highlight in this review common aspects and key biological differences of this group of filamentous structures.

Keywords: type IV pilus; type II secretion system; archaellum; class III signal peptide; prepilin peptidase

INTRODUCTION

Since the inception of cellular life, prokaryotes (bacteria and archaea) have been faced with a conundrum crucial for their survival: How to efficiently interact with their environment through the formidable barriers that define their very existence? These unicellular organisms have therefore engineered a variety of macromolecular nanomachines on their surface, which are assembled across highly impermeable membrane(s) and/or thick cell walls, and play important and diverse roles in microbial biology. One exceptionally widespread and multipurpose group of nanomachines uses filaments composed of subunits with a characteristic N-terminal sequence motif named class III signal peptide (Szabó *et al.*, 2007), generically named type IV pilins. Because they were the first to be identified and have been extensively studied ever since, the paradigm of this class are the surface-exposed organelles known as type IV pili (Tfp) (Pelicic 2008). Tfp—also known as bundle-forming pili (Bfp) in enteropathogenic *Escherichia coli* (EPEC) (Donnenberg, Zhang

and Stone 1997), toxin co-regulated pili (Tcp) in *Vibrio cholerae* (Manning 1997), fimbrial low-molecular-weight protein (Flp) pili in *Aggregatibacter actinomycetemcomitans* (Tomich, Planet and Figurski 2007), etc.—are long surface-exposed filaments composed of type IV pilins, whose biogenesis depends on a set of distinctive proteins. Studies in numerous species of bacteria and archaea have later revealed that several other systems with widely different morphological features are evolutionarily related to Tfp (Hobbs and Mattick 1993; Jarrell, Bayley and Kostyukova 1996) because they are also composed of type IV pilins and assembled by similar sets of proteins. These structures have names as diverse as secretion (Pugsley 1993b), archaellum (Jarrell and Albers 2012) or bindosome (Zolghadr *et al.*, 2007).

In this review, we will provide an overview of the complex biology of the machineries composed of type IV pilins whose biogenesis depends on a conserved set of proteins, for which we would like to introduce the unifying name type IV filaments (Tff). We will briefly list known Tff systems, the wide array of

Received: 8 August 2014; Accepted: 30 September 2014

© FEMS 2015. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

functions they mediate and their astonishing distribution in two out of the three domains of life. With an emphasis on the best characterized Tfp, we will then discuss in detail molecular and structural aspects of Tff and their complex biogenesis, underlining the many important commonalities, as well as a few significant differences.

TFF: AN AMAZING VARIETY OF MORPHOLOGIES AND ASSOCIATED FUNCTIONS

Unlike any other type of prokaryotic surface nanomachines, Tff come in a variety of shapes and promote a vast array of seemingly unrelated properties such as adhesion, motility, protein secretion and DNA uptake. Hence, these functionally versatile nanomachines can be viewed as the prokaryotic equivalents of the world-famous multitool pocket knife, which has inspired the title of this review. In this section, we will list the different types of Tff and the variety of functions they have been associated with.

Bacterial Tfp: the Tff paradigm

Bacterial Tfp conspicuous morphological features (Fig. 1)—i.e. they are surface-exposed filaments displaying a pronounced flexibility, a propensity to interact laterally to form bundles and are up to 1000 times longer (up to several μm) than they are wide (usually 60–80 Å)—were used in early electron microscopy (EM) studies to define them as a distinct type of pili (Ottow 1975), hence their Tfp moniker. Tfp have since been found and studied in many species of Gram-negative (Mattick 2002; Pelicic 2008) and Gram-positive bacteria (Melville and Craig 2013). Unlike other types of pili, Tfp are capable of retracting and generating forces ranging from 100 pN per single filament (Maier et al., 2002), to several nN for bundles (Biais et al., 2008). Retraction,

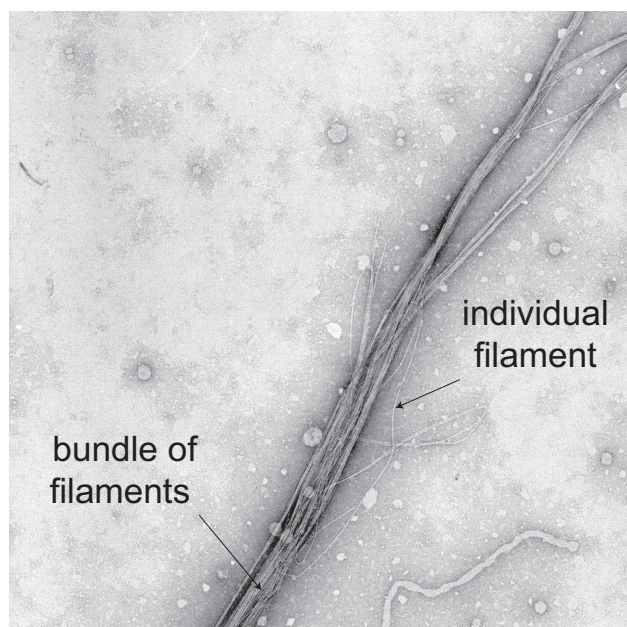


Figure 1. Conspicuous morphological features of bacterial Tfp, i.e. several μm length, 6–8 nm width, flexibility and propensity to interact laterally to form bundles. Bundle of filaments produced by *N. meningitidis* (individual filaments are 60 Å wide) were visualized by EM after negative staining with phosphotungstic acid.

which occurs through rapid depolymerization of pilin subunits, has so far been directly demonstrated only for one sub-class of Tfp (Tfpa see below). This is consistent with the fact that the PilT ATPase powering this process (Merz, So and Sheetz 2000) is restricted to the corresponding bacterial species. It remains a burning question whether other sub-classes of Tfp and/or other Tff can also retract as there is at best only indirect evidence so far (Bieber et al., 1998; Zahavi et al., 2011). This huge force generation makes Tfp the most potent linear molecular motor described to date. Critically, it endows Tfp with properties not commonly associated with other pili and can even influence their morphology, i.e. when under tension *Neisseria gonorrhoeae* Tfp reversibly transition into a 40% narrower elongated conformation (Biais et al., 2010).

Tfp are 'sticky' organelles

Like other types of bacterial pili, Tfp mediate attachment to and colonization of a wide variety of surfaces, both abiotic (plastic, glass, metal, etc.) and biotic (host cells and extracellular matrix in commensals and pathogens). Attachment to biotic surfaces makes Tfp key virulence factors in several human pathogens responsible for infections leading to dramatic morbidity and/or mortality worldwide—cholera, diarrhoea, meningitis and gonorrhoea, to cite but a few—and is the main reason Tfp have been a hot topic for research for decades. Tfp mediate adhesion in several ways in contrast to other types of pili that most often harbour a minor (low-abundance) subunit with intrinsic adhesive properties at their tip, e.g. type I pili (Lillington, Geibel and Waksman 2014). For example, the major Tfp subunit itself can have adhesive properties, such as the Bfp subunit that is a lectin with affinity for N-acetyllactosamine receptors on human host cells (Hyland et al., 2008), or PilA1 that mediates adhesion of the Gram-positive *Ruminococcus albus* to cellulose in the gastrointestinal tract of ruminants (Pegden et al., 1998; Rakotoarivonina et al., 2002). Alternatively, minor pilins can play important roles in attachment, such as PilV in the pathogenic *Neisseria* species *N. meningitidis* and *N. gonorrhoeae* (Winther-Larsen et al., 2001; Brown et al., 2010). PilV first binds CD147, mediating tight adhesion to host cells (Bernard et al., 2014), and then it binds β -adrenergic receptors (Coureuil et al., 2010) triggering the formation of membrane protrusions around adhering meningococci and enhancing their ability to resist mechanical forces generated by the blood flow (Mikaty et al., 2009). Non-pilin proteins associated with Tfp can also play key roles in adhesion as shown for PilC/PilY1 in multiple species (Rudel et al., 1992; Nassif et al., 1994; Kehl-Fie, Miller and St Geme 2008; Heiniger et al., 2010). Recently, an Arg-Gly-Asp motif in the N-terminal domain (NTD) of *Pseudomonas aeruginosa* PilY1 was found to bind integrin (Johnson et al., 2011), providing evidence that it is a *bona fide* adhesin. Since the NTD of PilC/PilY1 is species-specific, it is likely that orthologues in different species mediate adhesion to different receptors. Finally, Tfp retraction can influence their adhesive properties. For example, *N. gonorrhoeae* promotes colonization by mechanically stimulating pathways in host cells upon pilus retraction (Howie, Glogauer and So 2005).

Tfp promote interbacterial contacts

Another very common property of Tfp that influences attachment to surfaces is their ability to promote interaction between neighbouring bacteria via pilus–pilus contacts. These contacts lead to the formation of aggregates or micro-colonies (Kirm et al., 2000), which can even become biofilms if embedded within a matrix of extracellular polymeric substance

(O'Toole and Kolter 1998). Although they both involve pilus-pilus contacts, it should be pointed out that formation of micro-colonies and pilus bundling are distinct properties (Kirm et al., 2000). Formation of micro-colonies can be promoted by the major pilus subunit as in *V. cholerae* Tcp (Chiang et al., 1995), or by a minor pilin such as PilX in *N. meningitidis* (Helaine et al., 2005). PilX subunits in the filaments of interacting meningococci are thought to brace against each other upon pilus retraction through surface-exposed 'hooks' and thereby stabilize micro-colonies in face of pilus retraction (Helaine et al., 2007), although an alternative hypothesis has recently been proposed (Imhaus and Dumenil 2014). The formation of micro-colonies, which can also be seen in liquid culture, is however not limited to enhancing surface colonization. As shown for the R64 plasmid thin pilus of *E. coli* (Yoshida et al., 1998), Tfp-mediated aggregation promotes subsequent exchange of DNA between cells.

Tfp power twitching motility

Almost all bacterial species with retractile Tfp undergo surface-associated motility known as twitching motility (or social motility in *Myxococcus xanthus*) because cells exhibit jerky movements (Henrichsen 1972). Bacteria use Tfp as 'grappling hooks' and upon PilT-mediated pilus retraction pull themselves towards the site where the pilus is attached (Mattick 2002). The force generated by a single filament retraction (Maier et al., 2002) allows the bacterium to move 10 000 times its own body weight (Baker, Biais and Tama 2013), which results in rapid movement (Merz, So and Sheetz 2000). The irregular motion, abrupt turns and changes of direction, characteristic of twitching, are due to the release of single filaments, while others are still under tension, so that the bacterium rapidly 'slingshots' to a new orientation (Jin et al., 2011). Twitching motility has probably evolved to allow surface exploration that can be random or directed, e.g. towards light sources in the cyanobacterium *Synechocystis* sp. (Bhaya et al., 2000), and can even be optimized for 2D exploration when bacteria stand upright and 'walk' (Gibiansky et al., 2010).

Tfp promote DNA uptake during natural transformation

In contrast to the widespread nature of twitching motility, only a subset of bacterial species with retractile Tfp use these to promote the earliest step of natural transformation, i.e. import (or uptake) of free extracellular DNA across the outer membrane and/or thick layer of peptidoglycan (PG) (Chen and Dubnau 2004). In these species, DNA uptake is directly powered by pilus retraction and is abolished in a *pilT* mutant (Wolfgang et al., 1998a). Imported DNA is used to generate genetic diversity, as a template for the repair of damaged DNA, or as a source of food (Chen and Dubnau 2004). Tfp bind extracellular DNA (van Schaik et al., 2005) most likely through a major or minor pilin, as confirmed by the recent discovery that the ComP minor pilin in the Neisseriaceae family has intrinsic DNA-binding ability (Berry et al., 2013; Cehovin et al., 2013). Furthermore, ComP binds better to short and specific sequence motifs hyperabundant in these species genomes, explaining how they manage to preferentially take up their own DNA.

Uncommon and/or indirect Tfp properties

Another property of bacterial Tfp, which further extends the versatility of this class of filaments, might be viewed as 'exotic' or as a mere curiosity. In *Geobacter* species, Tfp have been found to be electrically conductive 'nanowires' transferring electrons from the cells to extracellular electron acceptors (Reguera et al., 2005). Although it has been proposed that conductivity might result from electrons 'hopping' between cytochromes at-

tached to *Geobacter* Tfp (Boesen and Nielsen 2013), filaments are likely to have intrinsic metal-like electron-conductive properties through stacking of aromatic residues of the major pilin PilA. Accordingly, filaments composed of *pilA* mutants lacking these residues have reduced conductive properties (Vargas et al., 2013).

It is worth mentioning here that retractile Tfp have also been hijacked by bacterial viruses. Some bacteriophages bind to the side or tip of Tfp, and are brought in contact with a cell surface-associated receptor upon pilus retraction (Skerker and Shapiro 2000). Historically, this is an important property since it is the observation by EM that shortening of *P. aeruginosa* pili occurred after phage attachment and was necessary for phage infection that led Bradley to propose that Tfp are capable of retraction (Bradley 1972). Furthermore, the finding that phage-resistant mutants that were unable to retract their pili were also defective for twitching allowed the same author to link Tfp retraction with twitching motility for the first time (Bradley 1980).

Other widespread bacterial Tff

Competence (pseudo)pili

In most naturally competent bacterial species, DNA uptake is not mediated by Tfp but rather by elusive Tff structures known as competence 'pseudopili' formed of a major 'pseudopilin', which cannot be directly visualized by EM because they are too short (Chen and Dubnau 2004). Key evidence for their existence is the recent discovery of extended competence organelles in *Streptococcus pneumoniae* (Laurenceau et al., 2013; Balaban et al., 2014), which remained unnoticed for decades in this species where competence has been intensively investigated (Johnston et al., 2014). Although no PilT is associated with competence pseudopili, a study in *Bacillus subtilis* showed that these are nevertheless force-generating motors exerting forces in excess of 40 pN on free DNA, and transporting it into the cell in a linear fashion (Maier et al., 2004). Transport is powered by proton motive force but the underlying molecular mechanism for force generation remains to be determined.

Type II secretion systems

Rather than for DNA import, pseudopili are used by many Gram-negative species as part of the secretion machinery to 'push' or 'lift' fully folded periplasmic proteins or protein complexes into the extracellular milieu across a dedicated channel in the outer membrane (Douzi, Filloux and Voulhoux 2012; Korotkov, Sandkvist and Hol 2012; Nivaskumar and Francetic 2014). Perhaps misleadingly, this process is known as type II secretion, whereas type IV secretion is mediated by a machinery unrelated to Tfp (Low et al., 2014). However, these unfortunate differences in nomenclature predate the discovery that type II secretion systems (T2SS) are evolutionarily related to Tfp, and further support our proposal of a unifying Tff name. As for competence pseudopili, T2SS pseudopili have never been directly visualized, probably because they are too short, barely spanning the periplasmic space (Douzi, Filloux and Voulhoux 2012; Nivaskumar and Francetic 2014). However, a key piece of evidence for their existence is the fact that surface-exposed filaments similar to Tfp, named 'hyper-pseudopili', can be seen when T2SS-harboring bacteria are genetically engineered to overexpress the major pseudopilin (Sauvonnnet et al., 2000; Durand et al., 2003; Vignon et al., 2003). T2SS-exported proteins, often enzymes, play important roles in lifestyles of the secreting bacteria by providing them with essential nutrients or by having toxic effects on host cells in pathogens (Nivaskumar and Francetic 2014). Interestingly, protein secretion can also be

mediated by some Tfp machineries as in *V. cholerae* and *Dichelobacter nodosus* (Kirn, Bose and Taylor 2003; Han et al., 2007). This blurs the lines between the different Tff and strengthens the notion that they are a homogeneous class of nanomachines.

Archaeal Tff

It is now clear that Tff are also widespread in a second domain of life, archaea. Although they often have different morphological features from bacterial Tfp—notably diameters up to 140 Å (Jarrell et al., 2013)—*bona fide* Tfp (long surface-exposed organelles composed of type IV pilins whose biogenesis depends on similar sets of proteins) have been identified in many archaeal species. Two main groups of archaeal filaments are distinguished based on the functions they have been associated with. Those filaments involved in attachment to surface and/or promotion of interarchaeal contacts are generically called pili. As in bacteria, Tfp-mediated aggregation can promote subsequent exchange of DNA between archaeal cells as shown for UV-induced pili of *Sulfolobus* (Fröls et al., 2008). Archaeal filaments that power swimming such as in *Sulfolobus acidocaldarius* are defined as flagella. Indeed, these Tff drive swimming by rotating (Shahapure et al., 2014) and are thus functional analogues of bacterial flagella (Jarrell et al., 2013). To underline these differences, a new name ‘archaellum’ has been coined for these archaeal flagella (Jarrell and

Albers 2012), but it is not universally accepted. Intriguingly, it has been shown that the FlaI ATPase powers both archaellum assembly and rotation (Reindl et al., 2013). The fact that a protein universally energizing Tff assembly (see below) can power filament rotation in archaea might have biological implications for this whole class of filaments, as will be discussed later. Among the many reasons why archaella might be the only Tff powering swimming are a higher rigidity, the possibility that they are Archimedes’ ‘screws’, the presence of additional machinery components, etc.

Possibly the most peculiar Tff machinery is the elusive *S. solfataricus* structure named bindosome that facilitates sugar uptake, which consists of several surface-exposed sugar-binding proteins that harbour class III signal peptides (Zolghadr et al., 2007). The bindosome has never been visualized and evidence that it produces (or not) even short filaments is still to be obtained. However, the bindosome surface localization depends on a canonical Tff named Bas.

TFF BIOGENESIS RELIES ON A LARGE AND CONSERVED SET OF DEDICATED PROTEINS

Tff are not only formed of subunits sharing a well-defined and conserved N-terminal motif but their biogenesis relies on conserved multiprotein machineries (Fig. 2), which is in support of their evolutionary relationship.

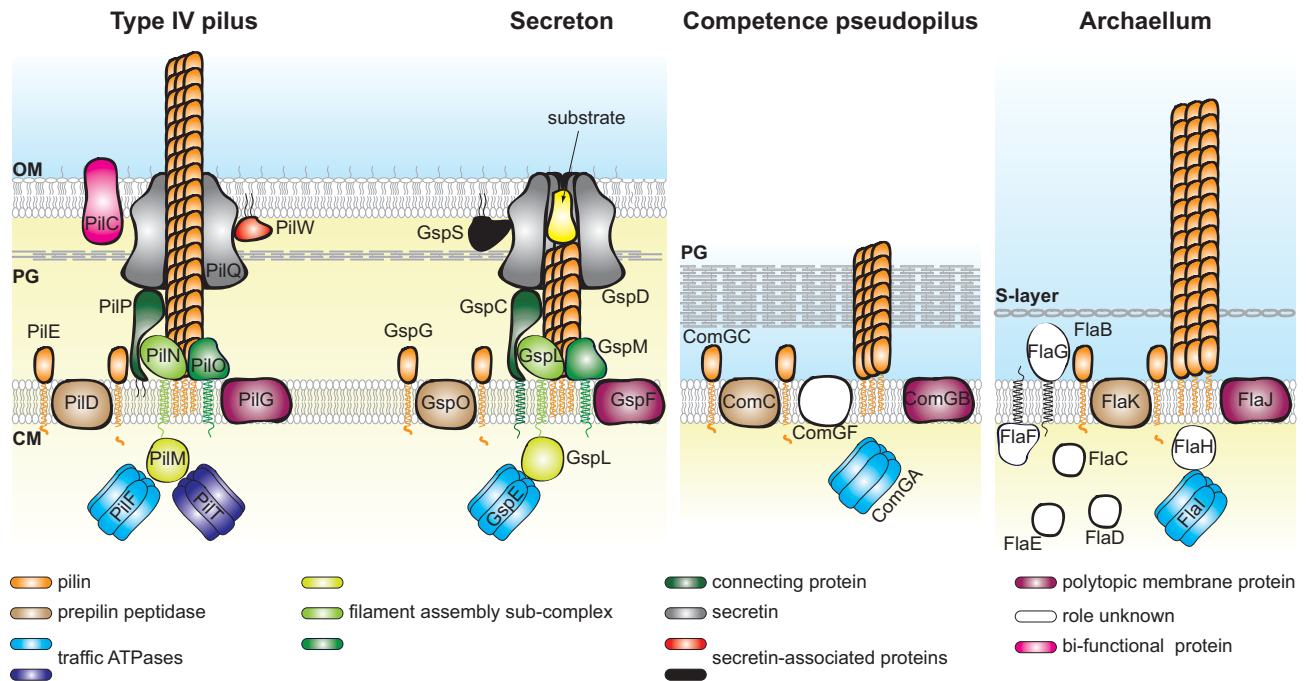


Figure 2. Schematic representation of different Tff nanomachines: Tfp, T2SS, competence pseudopili and archaella. For Tfp, the well-characterized Tfp_a system in *N. meningitidis* is shown. It should be noted that the protein nomenclature varies widely in other Tfp_a-expressing species, although all the proteins are highly conserved. However, a major difference in Gram-positive pilated species is the absence of the proteins forming the outer membrane sub-complex (PilC, PilP, PilQ and PilW). For T2SS, the system exporting pullulanase in *K. oxytoca* is shown, but the proteins have been given their unifying Gsp names. For competence pseudopili, *B. subtilis* has been chosen. For archaeal flagella, we have chosen the representative *M. maripaludis* archaellum. All these systems are evolutionarily related as they are composed of proteins that show sequence and/or structural similarity and perform the same functions. To facilitate comparisons, proteins of similar function have identical colour. In brief, major (pseudo)pilins are processed by a dedicated prepilin peptidase, which removes a short hydrophilic leader peptide. For the sake of clarity, minor (pseudo)pilins that also undergo this processing are not shown. Traffic ATPases power filament extension from the inner membrane through ATP hydrolysis. The PilT ATPase which powers filament retraction has so far been identified only in Tfp_a. The energy generated by ATP hydrolysis is translocated across the membrane by a multiprotein sub-complex, although the polytopic protein showing universal sequence conservation (purple) has also been proposed to play this role. In Gram-negative species, the inner membrane sub-complex is linked via a connecting protein to an outer membrane sub-complex centred on a multimeric channel known as the secretin. Several other proteins important for secretin stability/function are also part of this sub-complex. To facilitate visualization, the secretin dodecamer is shown as a vertical cross-section. OM, outer membrane; PG, peptidoglycan; CM, cytoplasmic membrane.

Type IV pilins

Tff subunits, which are synthesized as precursors, share an N-terminal class III signal peptide (Fig. 3) that is well defined and distinctive (Dalrymple and Mattick 1987; Szabó et al., 2007). These subunits will be generically referred to as type IV pilins, although this might be a misnomer because not all of them are involved in the biogenesis of pili. The class III signal peptide starts with a leader peptide containing a majority of hydrophilic and neutral residues, whose length varies between 6 and 26 amino acids (aa) in *P. aeruginosa* PilA

(Paranchych et al., 1979) and *A. actinomycetemcomitans* Flp1 (Kachlany et al., 2001), respectively. Although there are many exceptions in archaea, this leader peptide invariably ends with a conserved Gly, and is followed by a tract of 21 predominantly hydrophobic residues (Fig. 3). A negatively charged Glu₅ residue almost invariably interrupts this stretch, although it is often absent in archaeal pilins (Szabó et al., 2007; Jarrell et al., 2013). Importantly, these features distinguish class III signal peptides, which are processed by a dedicated prepilin peptidase after the conserved Gly on the cytoplasmic side of the

Major (pseudo)pilins

Tfpa

PilE *Nme*

PilA *Pae*

PilA1* *Cdi*

PilE1* *Ssa*

Tfpb

BfpA EPEC

TcpA *Vch*

PilS *Eco* (R64 Pil)

Flp1 *Aac* (Tad)

PilA *Ccr* (Cpa)

T2SS

PulG *Kox*

XcpT *Pae*

Archaeal Tfp

Aap *Sac*

MMP1685 *Mma*

IGNI0670 *Iho*

Archaeallins

FlaB *Sso*

FlaB1* *Mma*

Minor (pseudo)pilins

Tfpa (conserved)

PilH *Nme*

PilI *Nme*

PilJ *Nme*

PilK *Nme*

Tfpa (species-specific)

ComP *Nme*

PilV *Nme*

PilX *Nme*

T2SS (conserved)

PulH *Kox*

Pull *Kox*

PulJ *Kox*

PulK *Kox*

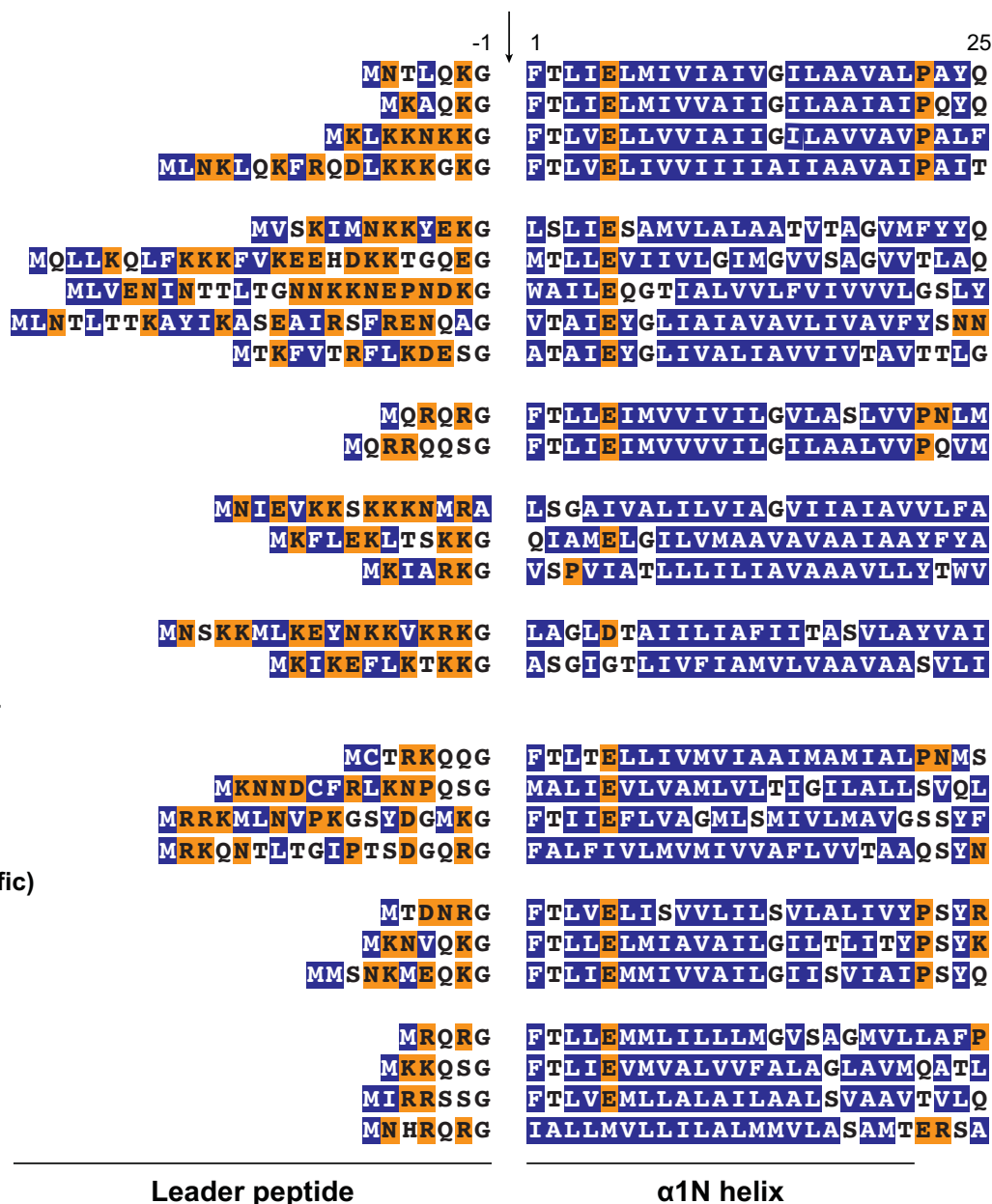


Figure 3. Conserved N-terminal sequence motif defining pilin subunits in a variety of Tff. This motif, known as class III signal peptide (Szabó et al., 2007), is composed of a hydrophilic leader peptide followed by a stretch of hydrophobic residues (except for a negatively charged Glu₅). The 6–26 aa leader peptide contains a majority of hydrophilic (shaded in orange) and neutral (no shading) residues, and invariably ends with a Gly (except in archaea). The following tract of 21 predominantly hydrophobic residues (shaded in blue) forms an extended α -helix that is the main assembly interface of subunits within Tff. This class III signal peptide is recognized by a dedicated prepilin peptidase and cleaved (indicated by a vertical arrow) after the conserved Gly⁻¹. *Filaments in these species might be composed of more than one major pilin. *Nme*, *N. meningitidis*; *Pae*, *P. aeruginosa*; *Cdi*, *C. difficile*; *Ssa*, *Streptococcus sanguinis*; *Vch*, *V. cholerae*; *Eco*, *E. coli*; *Aac*, *A. actinomycetemcomitans*; *Ccr*, *C. crescentus*; *Kox*, *K. oxytoca*; *Sac*, *S. acidocaldarius*; *Mma*, *M. maripaludis*; *Iho*, *Ignicoccus hospitalis*; *Sso*, *S. solfataricus*.

membrane (Nunn and Lory 1991) (Fig. 2), from standard (class I) and lipoprotein (class II) signal peptides, which are processed by different peptidases on the periplasmic side (Pugsley 1993b).

Historically, differences in length of prepilins and their leader peptides in the extensively studied Tfp systems were used to define two sub-classes of Tfp, Tfp_a and Tfp_b (Giltner, Nguyen and Burrows 2012). Prepilins of Tfp_a are shorter (140–170 aa) with shorter leader peptides (less than 10 residues), while Tfp_b prepilins are longer (180–200 aa) with longer leader peptides (about 15–30 aa) (Fig. 3). This distinction is consistent with important differences between the machineries involved in their biogenesis (Pelicic 2008). The discovery and study of many other Tff in more varied species means that these traditional length parameters are not valid anymore. Nevertheless, phylogenetic analysis of bacterial prepilins showed that two families consistent with the original classification can be distinguished (Kachlany et al., 2001). Type IV_a prepilins, some of which have leader peptides as long as 18 residues, share substantial sequence homology in their N-terminal hydrophobic stretch (Fig. 3) and they are by far the most widespread, being found both in pili and pseudopili (Giltner, Nguyen and Burrows 2012). Type IV_b prepilins, which can be as short as 44 residues in *Caulobacter crescentus* (Skerker and Shapiro 2000), also group together although they show less sequence homology, except for the Flp sub-family (Tomich, Planet and Figurski 2007). Most archaeal prepilins do not readily fit in the above two families and are likely to form one or more families of their own (Desmond, Brochier-Armanet and Gribaldo 2007; Szabó et al., 2007), which could tentatively be named type IV_c, type IV_d, etc.

Tff biogenesis machineries

The complete sets of proteins dedicated to Tff biology have been defined by systematic genetic studies in several model systems: Tfp_a in *P. aeruginosa* and *N. meningitidis* (Alm and Mattick 1997; Carbonnelle et al., 2006), Tfp_b in EPEC, *V. cholerae* and *E. coli* (Yoshida, Kim and Komano 1999; Kirn, Bose and Taylor 2003) and T2SS in multiple species such as *Klebsiella oxytoca*, *Erwinia chrysanthemi* and *P. aeruginosa* (Douzi, Filloux and Voulhoux 2012; Nivaskumar and Francetic 2014). This revealed that although Tff are primarily polymers of a single protein, their biogenesis requires complex machineries of 10–18 proteins. In most systems—Tfp_b, T2SS, archaeal Tff—the corresponding genes cluster together but they are scattered throughout the genome for Tfp_a, except in the few Gram-positive species in which these are found (Pelicic 2008; Douzi, Filloux and Voulhoux 2012; Jarrell and Albers 2012; Melville and Craig 2013; Nivaskumar and Francetic 2014). Unlike for Tfp_b that are more heterogeneous—Flp that are assembled by the conserved tight adherence (Tad) locus are an exception (Tomich, Planet and Figurski 2007)—genes are conserved ‘en bloc’ for Tfp_a or T2SS with similar genetic organizations. In Tfp_a for example, there are virtually no differences between *P. aeruginosa* and *N. meningitidis* that are distant γ and β Proteobacteria. The only major difference might be the pilY2 gene essential for piliation, which is restricted to *P. aeruginosa* (Alm et al., 1996b). It should be noted here that additional ‘accessory’ proteins are often key for Tff function(s) while being dispensable for filament biogenesis, which makes Tff biology even more complex. For example, in *N. meningitidis*, there are seven accessory proteins—three of which (PilT, PilU and PilZ) are widely conserved—that modulate one or several Tfp_a functions (Brown et al., 2010): three minor pilins (ComP, PilV and PilX), three ATPases (PilT, PilT2 and PilU) and PilZ.

The most important finding in the above studies was that Tff are unequivocally a homogeneous class of filaments since many proteins involved in their biogenesis show sequence homology, several of which are even universally conserved (Fig. 2). These conserved proteins are often designated ‘core’ but recent structural findings (see below) show that this distinction is misleading and should probably be abandoned. This set of conserved proteins consists of at least one protein (but more often several) with a class III signal peptide, a dedicated prepilin peptidase for prepilin processing, a ‘traffic’ ATPase to power filament assembly and a polytopic cytoplasmic membrane protein of unknown function. Intriguingly, many of the other non-core proteins important for Tff biogenesis exhibit no obvious sequence conservation between the different systems, which led to questioning whether a unique mechanism was used for Tff biogenesis (Pelicic 2008). This is nevertheless likely to be the case as suggested by recent structural data showing that non-core proteins exhibiting no sequence homology have strikingly similar 3D structures. For example, the structure of the N-terminal part of BfpC that is involved in Tfp_b biogenesis (Yamagata et al., 2012) is similar to the structures of PilM involved in Tfp_a biogenesis (Karuppiah and Derrick 2011) and GspL from T2SS (Abendroth et al., 2004a). Unfortunately, although a unified Gsp (general secretory pathway) nomenclature has been proposed for T2SS (Pugsley 1993b), these proteins have different names in different species or systems, which makes comparisons difficult. To help the reader, proteins performing the same function in different systems have the same colour in Fig. 2. It should also be noted that, unless stated otherwise, the *N. meningitidis* nomenclature will be privileged in the rest of this review.

TFF ARE VIRTUALLY UNIVERSAL IN PROKARYOTES

Various types of Tff nanomachines have been identified and experimentally studied in many prokaryotic species. However, their global distribution remains an open question. This can be addressed by mining the ever increasing amount of sequence data in the databanks in search of signature motifs found uniquely in proteins dedicated to Tff biogenesis. Using BioMart (Guberman et al., 2011), we have therefore performed a global search of the InterPro database (Hunter et al., 2012) for the distribution of such motifs across all the species sequenced so far. This analysis shows that Tff are one order of magnitude more widespread than previously anticipated (Pelicic, 2008), and are actually virtually universal in prokaryotes (Fig. 4). This is likely to be a consequence of their extreme functional versatility and their ancient nature since a Tff, whose function can only be guessed, was already present in a common ancestor to bacteria and archaea that have diverged more than 3 billion years ago.

The most obvious sequence signature to start this bioinformatic search was the one found in Tff subunits: the distinctive class III signal peptide motif (Fig. 3) (Dalrymple and Mattick, 1987; Szabó et al., 2007). A simple query using InterPro domains absolutely specific for bacterial (IPR012912 and/or IPR007047) and archaeal type IV prepilins (IPR002774) suggests that Tff machineries are encoded in the genomes of approx. 1800 different species. These species span 4/5 phyla of archaea (Korarchaeota are apparently the only exception) and all 26 phyla or group of phyla of bacteria (Fig. 4). Strikingly, this estimate is even likely to be conservative since simple queries using domains specific for other components invariably found in Tff machineries, namely the prepilin peptidase (IPR010627 and/or IPR000045), the

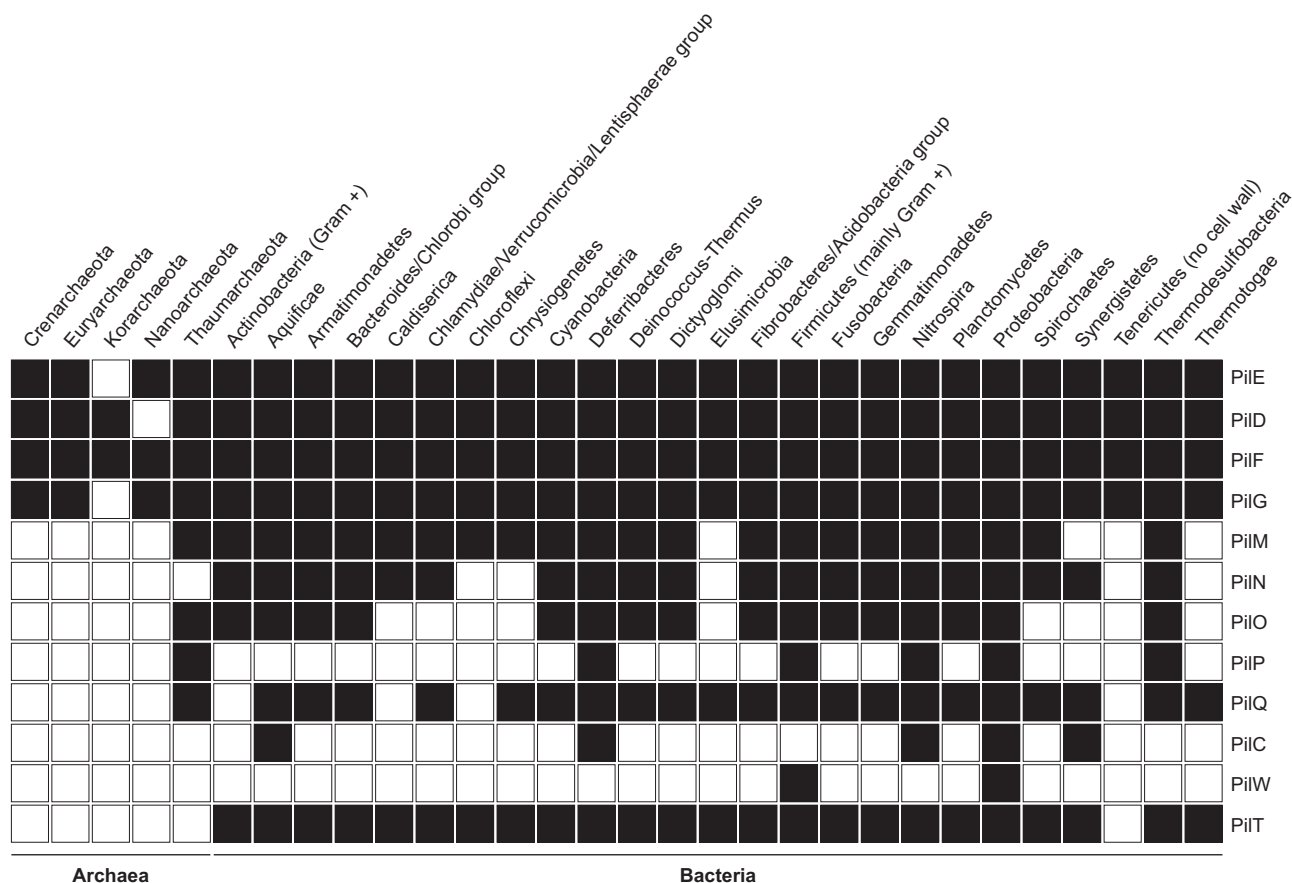


Figure 4. Phylogenetic distribution of the proteins involved in Tff biogenesis in archaea and bacteria. Using BioMart (Guberman et al., 2011), we have performed a global search of the InterPro database (Hunter et al., 2012) for signature motifs found only in proteins dedicated to Tff biogenesis. A black box indicates that the corresponding protein is found in the analysed phylum, while a white box indicates that it is absent. PilE, IPR007047, IPR002774 or IPR012912; PilD, IPR010627 or IPR000045; PilF, IPR007831 or IPR001482; PilG, IPR018076; PilM, IPR005883; PilN, IPR007813; PilO, IPR007445; PilP, IPR007446; PilQ, IPR001775 or IPR013355; PilC, IPR008707; PilW, IPR013360; PilT, IPR006321. In Firmicutes, the outer membrane sub-complex proteins (PilP, PilQ and PilW) are found only in the very few Gram-negative species in this phylum.

traffic ATPase (IPR007831 and/or IPR001482) and the PilG polytopic membrane protein (IPR018076), each returned more than 2000 different species, including some in which no prepilins could be found. The possibility that prepilins not readily identified using the above pilin signatures might be present in those 'extra' species is supported by the PilFind algorithm that predicts many non-canonical type IV prepilins (Imam et al., 2011), including a new class that we are currently characterizing (Pelicic, unpublished). This new class of prepilins corresponds to a well-defined domain of so far unknown function that is widespread in Actinobacteria (275 different species).

These findings are strengthened by a multiquery using all the above domains. This search revealed that the genomes of 1656 different species, in which type IV prepilins are found, encode simultaneously a prepilin peptidase, a traffic ATPase and the polytopic cytoplasmic membrane protein (Fig. 4). These species span all the phyla of archaea and bacteria listed above, except Nanoarchaeota in which no prepilin peptidase could be detected. Several other interesting observations arose from these queries. Firstly, prepilin peptidases often consist of two domains, one of which (IPR000045) always corresponds to the peptidase domain. In bacteria, the most common second domain (IPR010627) is almost certainly responsible for the well-known N-methylation of the first residue of many mature pilins (Strom et al., 1993). In archaea, where this N-methylation is not observed

(Jarrell et al., 2013), IPR010627 is replaced by IPR009655 whose function is unknown. This suggests that archaeal pilins might undergo another sort of N-terminal modification. Interestingly, in *Methanococcus maripaludis* Tfp, the N-terminal Gln of the major pilin is modified into a pyroglutamate (Ng et al., 2011). Secondly, almost 1000 species encode the above four proteins as well as PilT retraction ATPase (IPR006321), and are likely to express Tff capable of retraction. Of these 1000 species, 500 also encode the PilM, PilN, PilO proteins (IPR005883, IPR007813 and IPR007445, respectively) and are likely to express *bona fide* Tfp (Pelicic, 2008). Strikingly, the entire set of 15 proteins involved in Tfp biogenesis in *N. meningitidis* is found in as many as 270 different species of Proteobacteria.

TFF: COMMON STRUCTURAL ASPECTS

Tff have been intensively studied from a structural point of view. This has confirmed that they are a homogeneous group of filamentous nanomachines in which pilin subunits share a distinctive fold and are assembled in a similar way (Fig. 5).

Type IV pilins: a universally conserved structural fold

Once their leader peptide is processed, Tff subunits share an N-terminal tract of highly hydrophobic residues with predicted

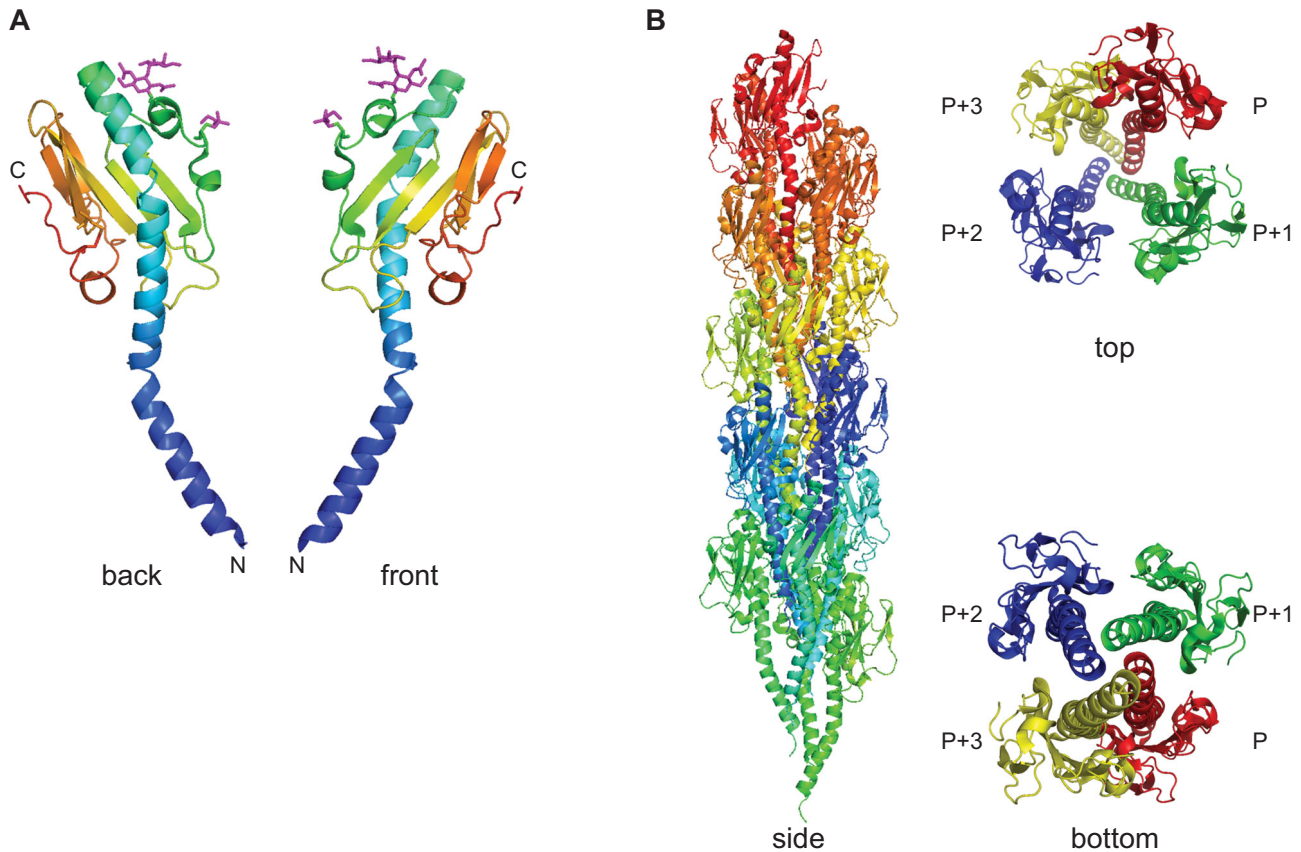


Figure 5: Common structural features of Tff and their subunits illustrated with *N. gonorrhoeae* Tfp (Craig et al., 2006). (A) Structure of the full-length PilE from *N. gonorrhoeae* (PDB entry 2HI2) showing the conserved lollipop shape with a protruding α 1N-helix (back view on the left, front view on the right). Post-translational modifications on the $\alpha\beta$ -loop, carbohydrate at Ser₆₃ and phosphate at Ser₆₈, are shown in magenta. (B) Structural model of gonococcal Tfp (PDB entry 2HIL) obtained by combining X-ray crystallography and cryo-EM (side view on the left, top view on the upper right, bottom view on the lower right). In the 60 Å diameter right-handed 1-start helical assembly, the pilins are arranged in an 'ear of wheat' fashion (side view). The α 1N-helices provide the main polymerization interface and are buried within the filament core parallel to its long axis (top and bottom views). Figures were generated using PyMOL (<http://www.pymol.org>).

propensity to form an extended α -helix. There is little, if any, sequence homology in the rest of these proteins. Notwithstanding, structural studies have revealed that type IV pilins share a common 3D architecture (Craig, Pique and Tainer 2004). Full-length structures of mature Tfp-forming type IVa pilins from *N. gonorrhoeae* (Parge et al., 1995), *P. aeruginosa* (Craig et al., 2003) and *D. nodosus* (Hartung et al., 2011), all resemble 'lollipops', with a 'stick' and a globular head (Fig. 5A). The stick corresponds to the N-terminus of the pilins and consists of a ~50 residue α -helix (α 1). The N-terminal half of α 1 (α 1N) protrudes from the C-terminal globular head and corresponds primarily to the stretch of 21 highly hydrophobic residues that is part of the class III signal peptide (Fig. 3). The C-terminal half of α 1 (α 1C) is packed against the globular head that consists mainly of a β -meander motif of four antiparallel β -strands. Most of the structural diversity between pilins lies in the two regions flanking the β -meander, the $\alpha\beta$ -loop that connects α 1 and the first β -strand of the β -meander, and the C-terminus known as the D-region because it is usually stapled to the last β -strand of the β -meander by a disulphide bond. D-region is, however, a misnomer since these C-terminal Cys are not always conserved and the disulphide bond can be replaced by a network of hydrogen bonds as in the *D. nodosus* pilin (Hartung et al., 2011). Recently, the full-length structure of a much shorter (66 aa) type IVa pilin subunit of *Geobacter sulfurreducens* nanowires has been determined in detergent micelles (Reardon and Mueller 2013). This structure,

which almost exclusively consists of α 1 (i.e. there is no globular head), confirmed that the first 21 residues are inserted within the membrane prior filament assembly and showed that an extended N-terminal α -helix might be viewed as the minimal common feature of type IV pilins.

The many other structures available for type IVa and type IVb major and minor (pseudo)pilins lack the protruding and highly hydrophobic α 1N—for a review see Giltner, Nguyen and Burrows (2012). Truncation makes the recombinant proteins soluble and more readily amenable to purification and structural characterization. It has (i) minimal structural impact as shown with the *P. aeruginosa* PAK pilin for which full-length and truncated structures are essentially identical (Craig et al., 2003), and (ii) no consequence on the utility of these structures since the missing hydrophobic region can reliably be modelled as an extended α -helix. All these structures confirm that the overall architecture of type IV pilins is conserved, i.e. α 1C packs against a β -meander motif that is composed of three to seven antiparallel β -strands depending on the size of the protein. As mentioned above, the structural diversity, which accounts for the functional differences between corresponding proteins and/or filaments, lies in the $\alpha\beta$ -loop and D-region. For example, the D-region of the minor pilin PilX that is key for the formation of bacterial aggregates in *N. meningitidis* consists of a short 'pigtail' α -helix and a hook that protrudes from the surface of the filaments (Helaine et al., 2007). Interestingly, major pilins undergo

post-translational modifications in these regions (Fig. 5A), most often addition of glycans, although phosphorylcholine, phosphoethanolamine and phosphoglycerol, have also been described (Forest *et al.*, 1999; Hegge *et al.*, 2004; Gault *et al.*, 2013). The functional significance of these modifications is unclear and varies from system to system. In bacteria, they are usually dispensable for Tfp assembly and have little effect on Tfp-mediated functions (Marceau *et al.*, 1998; Smedley *et al.*, 2005), while in archaea non-glycosylated subunits cannot be assembled in filaments (Chaban *et al.*, 2006; VanDyke *et al.*, 2008).

Until very recently, only structures of pilins from Gram-negative bacteria were available and it could only be speculated that the above architecture was universally conserved. However, the recently determined structure of the minor pilin PilJ from the Gram-positive *Clostridium difficile* (Piepenbrink *et al.*, 2014) not only displays the typical pilin fold but is, curiously, the only type IV pilin with a dual pilin fold likely to have resulted from a duplication or a fusion event (Piepenbrink *et al.*, 2014). This finding is important because it strengthens the notion that the classical fold is likely to be universal in type IV pilins. However, it would be interesting to determine the structures of more distant members, notably pilins from archaea (Szabó *et al.*, 2007) which display important sequence differences (Fig. 3) or the very short pilins from the Flp sub-family (Kachlany *et al.*, 2001). The latter ones are puzzling because they are even shorter than the *G. sulfurreducens* pilin (Reardon and Mueller 2013) and possibly consist only of the α 1N-helix, which might thus be viewed as the minimal universally conserved structural fold defining type IV pilins.

Similar Tff models

Consistent with this universally conserved fold, all models predicting how type IV pilins are arranged within filaments agree that Tff are helical polymers in which α 1N-helices provide the principal polymerization interface and are buried within the filament core parallel to the filament axis, in an ‘ear of wheat’ fashion (Fig. 5B). This is true even in archaeal species producing ‘unorthodox’ Tff (Wang *et al.*, 2008; Yu *et al.*, 2012). Several different helical models have been proposed using a variety of methods. The prototype model for Tfp_a, which has been determined for *N. gonorrhoeae* filaments (Craig *et al.*, 2006), is marked by high ridges (corresponding to the $\alpha\beta$ -loop and D-region) and deep grooves that wind around the filament axis (Fig. 5B). The α 1-helices are within the core of the filament, almost parallel to its long axis. In the 60 Å diameter right-handed 1-start helix, consecutive subunits are separated by a rise of 10.5 Å and show 101° azimuthal rotation. In this pseudo-atomic resolution model, charges in α 1 are neutralized by inter-subunit salt bridges, such as the one between Glu₅ of one protomer (P) and the N-terminal amide of Phe₁ of the next (P+1) (Craig *et al.*, 2006). A 3-start mechanism for Tfp_a assembly was proposed (Craig *et al.*, 2006), in which three pilin subunits are added simultaneously around the filament circumference.

Although the corresponding pilin subunit and the filament are significantly larger, a similar approach (which was refined by comparing solvent accessibility of the pilin residues in monomer versus in filaments) yielded a similar architecture for Tfp_b from *V. cholerae* (Li *et al.*, 2008; Li, Egelman and Craig 2012). In this model, consecutive subunits in a 88 Å diameter right-handed 1-start helix are separated by a rise of 8.4 Å and show 97° azimuthal rotation. Unlike in the Tfp_a model in which they are exposed on the surface, the $\alpha\beta$ -loops are implicated in holding the subunits together and are buried within the filament, whereas a sizeable segment of α 1N is curiously more exposed to the sol-

vent (Li *et al.*, 2008). This looser packing might explain, at least in part, the reduced resistance of Tfp_b to denaturing agents when compared to Tfp_a (Li, Egelman and Craig 2012).

A different approach has been used to model the T2SS hyper-pseudopilus of *K. oxytoca* (Campos *et al.*, 2010). In this computational approach, a multistage minimization and molecular dynamics modelling procedure used the structure of the PulG subunit and symmetry parameters of the helix obtained from classical EM studies (Campos, Francetic and Nilges 2011). This procedure allows the protomers to ‘explore’ conformations that would be missed in the above ‘rigid’ approaches. It led to one major model that closely resembles that of *N. gonorrhoeae* Tfp_a, i.e. a right-handed 1-start helix with an axial rise of 10.4 Å between consecutive subunits. The salt bridges and hydrophobic contacts predicted between α 1N-helices have been elegantly validated by single and complementary charge inversions and/or double Cys substitutions and cross-linking (Campos *et al.*, 2010). Using this strategy, reliable models consistent with the above ones could also be obtained for Tfp_a and Tfp_b (Campos, Francetic and Nilges 2011). Interestingly, in this flexible modelling approach, the predicted salt bridge between Glu₅ (P) and Phe₁ (P+1) in Tfp_a (Craig *et al.*, 2006) and Tfp_b (Li, Egelman and Craig 2012) was found only in a minority of models, since in the most stable conformation charged residues in α 1C of P formed a salt bridge with Glu₅ from P+3. This suggested that several conformational states might coexist, which is supported by EM in which a continuum of structures with different azimuthal rotations between subunits could be observed (Nivaskumar *et al.*, 2014). It was proposed that three main conformational groups might be consecutive transitions during filament assembly, which led to a model for hyper-pseudopilus assembly. In this model, a rotation-driven mechanism was proposed (Nivaskumar *et al.*, 2014) in which filament assembly is initiated by P–P+1 contacts between pilins still localized in the membrane (docking step). Then, the force generated by the traffic ATPase spools a newly docked protomer into the fibre, leading to its partial extraction from the membrane and its rotation by an average 84°. Two subsequent rounds of elongation/rotation lead to full extraction of this protomer from the membrane, and the overall 252° rotation allows P–P+3 contacts that consolidate the filament. The finding that archaea are rotating Tfp (Shahapure *et al.*, 2014) gives further support to this rotation-coupled assembly as a common mechanism for all Tff (although angles will be different in different systems).

MOLECULAR MECHANISM OF TFF BIOGENESIS

Molecular mechanisms of Tff biogenesis are still poorly understood and we cannot definitely answer the fundamental question: How are Tff assembled? However, much progress has been made with the identification of several discrete steps in Tff biogenesis, much better knowledge of the different protein sub-complexes involved (Fig. 2) and (at least partial) 3D structural information for all the key players. Important parallels between different systems can now be drawn, but it would be imprudent to make generalized conclusions at this stage.

Prepilin transport and processing

The translocation of prepilins across the cytoplasmic membrane and subsequent processing by the prepilin peptidase is the first and best understood stage of Tff biogenesis. As shown in *E. coli* in the absence of any other component of the Tff machinery (Strom and Lory 1987; Dupuy *et al.*, 1991), the conserved

N-terminal motif of prepilins is sufficient to promote their insertion in the cytoplasmic membrane. Membrane topology is determined by the conserved sequence features of the class III signal peptide. Following the 'positive-inside' rule (von Heijne and Gavel 1988), the hydrophilic leader peptide remains in the cytoplasm, the hydrophobic α 1N-helix behaves as a transmembrane domain, while the rest of the prepilin is exposed to the periplasmic space (Strom and Lory 1987; Dupuy et al., 1991). Insertion of prepilins in the membrane relies on a universal machinery composed of the signal recognition particle (SRP) and the Sec translocon (Arts et al., 2007b; Francetic et al., 2007). The SRP binds the conserved N-terminal motif when the nascent prepilin polypeptide emerges from the ribosome and feeds it to the Sec translocon, which translocates it across the cytoplasmic membrane and integrates it into this lipid bilayer where it adopts its typical 3D structure.

Since class III signal peptides lack the specific cleavage residues found after the hydrophobic tract in class I and class II signal peptides (Pugsley 1993b), prepilins are catalytically processed by dedicated prepilin peptidases (Kaufman, Seyer and Taylor 1991; Nunn and Lory 1991), which in many (but not all) bacteria also often methylate the first residue of mature pilins (Strom, Nunn and Lory 1993). Prepilin peptidases form a new superfamily of polytopic membrane aspartic acid proteases lacking the canonical cleavage site and low pH requirements of classical aspartate proteases (LaPointe and Taylor 2000). Cleavage of the leader peptide occurs after the conserved Gly on the cytoplasmic side of the membrane (Fig. 2) and effectively leaves the mature pilin as a membrane protein with no remaining domain in the cytoplasm (Lemkul and Bevan 2011). Prepilin processing and N-methylation by the prepilin peptidase do not require any other component of the Tff biogenesis machinery, since they can be observed upon co-synthesis of these two proteins in a cell-free translation system (Aly et al., 2013). It is very likely (although it remains to be formally demonstrated) that the IPR010627 domain catalyzes the N-methylation activity since zinc-binding via a CysXXCys-X_n-CysXXCys motif was found to be important (Aly et al., 2013). It remains to be determined whether IPR009655 in archaeal prepilin peptidases catalyzes an alternative N-terminal modification, perhaps the cyclization into a lactam ring of the N-terminal-free amino group as identified in *M. maripaludis* pilin (Ng et al., 2011). Importantly, although there are exceptions (de Bentzmann et al., 2006; Szabó et al., 2007), many prepilin peptidases show substrate promiscuity and can process proteins harbouring (even non-canonical) class III signal peptides (Winther-Larsen et al., 2005) from other Tff systems in the studied species (Nunn and Lory 1993), or even from pilins coming from other species (Nunn and Lory 1991).

Many site-directed mutagenesis studies have highlighted the importance of two conserved Asp residues in prepilin peptidases for efficient prepilin processing (LaPointe and Taylor 2000; Bardy and Jarrell 2003; Akahane et al., 2005; Tomich, Fine and Figurski 2006). Critically, the recent crystal structure of the *M. maripaludis* FlaK prepilin peptidase (Hu et al., 2011) provided a framework for understanding the mechanism of catalysis. FlaK, which is 'tilted' in the membrane, consists of a membrane-spanning central domain composed of six transmembrane helices and a soluble domain with four anti-parallel β -strands protruding into the cytoplasm, which constitutes the bulk of the archaea-specific IPR009655 motif. Strikingly, the crystal structure indicated that FlaK must undergo significant conformational change to bring the two catalytic Asp residues—separated by as much as 12 Å—close enough for catalysis to occur. This change in conformation is likely to occur upon loading of the prepilin substrate.

Accordingly, interfering with this conformational shift abolished the peptidase proteolytic activity (Hu et al., 2011).

Site-directed mutagenesis studies have also been performed on prepilins and revealed common principles (Strom and Lory 1991; Pugsley 1993a; Horiuchi and Komano 1998; Thomas, Chao and Jarrell 2001). The leader peptide is necessary for efficient processing—but it can in some systems be shortened without adverse effects (Horiuchi and Komano 1998; Ng et al., 2009)—and the last residue (the conserved Gly) is absolutely critical. In contrast, the conserved Glu₅ in the hydrophobic stretch of bacterial (pseudo)pilins, which appears to be important for methylation in Tfp (Pasloske and Paranchych 1988; Strom and Lory 1991) but not in T2SS (Pugsley 1993a), is always dispensable for processing. Glu₅ pilin mutants cannot be assembled into homopolymeric Tfp (Pasloske, Scraba and Paranchych 1989; Strom and Lory 1991; Horiuchi and Komano 1998), but they can be efficiently incorporated together with wild-type subunits into compound filaments (Pasloske, Scraba and Paranchych 1989; Aas et al., 2007). The lack of homopolymerization of Glu₅ mutants is unlikely to be due to their lack of methylation since other mutants that are not methylated can be efficiently assembled into functional Tfp (Strom and Lory 1991). Therefore, the functional role of N-terminal methylation of some bacterial pilins remains enigmatic.

Traffic ATPases power Tff (dis)assembly

Once they have been processed, mature pilins remain membrane proteins (Strom and Lory 1987; Dupuy et al., 1991) and must be actively extruded from the lipid bilayer in order to be polymerized into the base of growing filaments. This process is powered by cytoplasmic proteins from the superfamily of traffic ATPases (Planet et al., 2001; Peabody et al., 2003), which are thought to invariably function as oligomers (Sakai, Horiuchi and Komano 2001; Rose et al., 2011). These are associated with the membrane in a non-covalent fashion, via interaction with the membrane proteins involved in Tff assembly (Fig. 2) (Sandkvist et al., 1995; Tripathi and Taylor 2007). Oligomerization occurs once the traffic ATPase binds ATP, which leads to its association with a membrane partner and stimulates ATPase activity (Shiue et al., 2006). Consistent with membrane localization of this complex, ATPase activity is dramatically increased in the presence of phospholipids (Camberg et al., 2007). In Tfp, the extension ATPase PilF also interacts strongly with the cytoplasmic PilZ protein (Guzzo et al., 2009; Georgiadou et al., 2012), but the functional significance of this is unclear (Alm et al., 1996a; Brown et al., 2010). As shown by site-directed mutagenesis, the highly conserved Walker motifs in traffic ATPases are essential for their enzymatic activity (Turner et al., 1993; Possot and Pugsley 1994; Jakovljevic et al., 2008; Patrick et al., 2011). These properties are shared by the retraction ATPase PilT (Brossay et al., 1994; Herdendorf, McCaslin and Forest 2002; Chiang, Habash and Burrows 2005). PilT powers disassembly of pilin subunits from the base of filaments, which form a pool in the cytoplasmic membrane ready to be polymerized again (Morand et al., 2004). These different traffic ATPases have distinctive sequence features which probably account for their different functional roles. For example, the C-terminal domain (CTD) of PilT contains a helical AIRNLIRE motif, which is critical for pilus retraction but is not required for ATPase activity or oligomerization (Aukema et al., 2005). Two-hybrid studies in *N. meningitidis* have recently identified an intricate interaction network between the four-traffic ATPases found in this species (Georgiadou et al., 2012). This introduced the notion that the situation might be more complex than alternative switching of two

steadfast antagonistic motors at the base of the Tff, and that hetero-hexamers or higher order complex motors might exist. Therefore, the highly dynamic nature of Tfp_a would be under the dependence of the composition of these 'hybrid' motors, which remains to be formally demonstrated.

Several crystallographic studies of extension motors (Yamagata and Tainer 2007; Lu et al., 2013), the PilT retraction motor (Satyshur et al., 2007; Masic, Satyshur and Forest 2010) and the archaeum FlaI ATPase (Reindl et al., 2013), have provided insight into how chemical energy resulting from ATP hydrolysis might be transformed into mechanical energy. All of these structures describe a hexameric ring arrangement of bilobed monomers, in which distinct NTD and CTD are connected by flexible linkers. In each case, the NTD adopts a PAS-like fold usually comprising six anti-parallel β -strands flanked on one side by two or three α -helices. The CTD is topologically similar to RecA-like proteins with four signature ATPase motifs (Walker A and B motifs, and Asp and His boxes) which surround the nucleotide-binding pocket and form the catalytic centre. A striking feature of these hexameric rings is their dynamic nature, as they undergo dramatic domain movements and consequently adopt very different conformations upon ATP binding and hydrolysis. In brief, binding of the ATP involving conserved Arg residues forming a 'basic clamp' brings NTD and CTD from one subunit closer together (a 'closure' stage later released upon ATP hydrolysis), driving a 'swinging' motion of the neighbouring subunit CTD with some residues moving tens of Å (Satyshur et al., 2007; Reindl et al., 2013).

Assembling Tff: the last frontier

How the mechanical energy generated by domain motion within traffic ATPases is translated to pilins during Tff (dis)assembly remains the main mystery in the field. Since mature pilins do not have a cytoplasmic domain (Lemkul and Bevan 2011) and Tff assembly occurs on the periplasmic side of the membrane, this mechanical energy must be transduced to pilins across the membrane, via one or several membrane proteins that together form a Tff assembly sub-complex (Fig. 2). Although there is widespread agreement on this point, discrepant findings have been reported and two different models coexist.

A seminal finding concerning this step comes from Tfp_a studies showing that not all proteins essential for pilus biogenesis are involved in filament assembly *per se*. This was first established in *N. gonorrhoeae* by showing that the piliation defect in a pilC mutant could be suppressed by a second mutation in pilT (Wolfgang et al., 1998b). PilC is thus dispensable for Tfp assembly in the absence of pilus retraction, suggesting that PilC exerts its role in Tfp biogenesis at a late stage (see below). Importantly, piliation is also restored in a pilY1/T double mutant in the phylogenetically distant *P. aeruginosa*, where PilY1 is the orthologue of meningococcal PilC (Heiniger et al., 2010). An important caveat is that when piliation is not restored in a double mutant, this is (indirect) evidence that the corresponding protein might be involved in pilus assembly. This approach has been used in a systematic fashion in *N. meningitidis* and revealed that, out of the 15 proteins required for piliation, a surprisingly small number might be involved in filament assembly (PilD, PilE, PilF, PilM, PilN, PilO and PilP) since piliation could be restored in all the other double mutants (Carbonnelle et al., 2006). Many of these findings are consistent with those in the closely related *N. gonorrhoeae* (Wolfgang et al., 2000; Winther-Larsen et al., 2005).

Therefore, since the roles of PilD, PilE and PilF are known, these results suggested that it is the PilMNOP sub-complex that

uses the energy generated by PilF in the cytoplasm and translocates it across the membrane to mature pilins while polymerizing them into helical filaments (Fig. 2). Although they share little, if any, sequence homology, structural data have clearly shown that structural homologues of PilMNOP are found in other well-studied Tff systems (Korotkov, Sandkvist and Hol 2012). The structure of PilM (Karuppiyah and Derrick 2011) is similar to the cytoplasmic domains of BfpC and GspL that are involved in Tfp_b biogenesis and T2SS, respectively (Abendroth et al., 2004a; Yamagata et al., 2012). The structures of PilN and PilO (Sampaleanu et al., 2009; Karuppiyah et al., 2013), which exhibit similar circular permutations of the ferredoxin fold, are similar to the periplasmic domain of GspL and to GspM, respectively (Abendroth et al., 2004b; Abendroth, Kreger and Hol 2009a). Finally, PilP is a structural homologue of the homology region (HR) domain that is found in all GspC proteins (Golovanov et al., 2006; Korotkov et al., 2011b; Tammam et al., 2011; Gu et al., 2012). There is now a wealth of functional information confirming the existence of a PilMNOP sub-complex at the cytoplasmic membrane. This was obtained in different bacterial species and systems, using a wide variety of approaches (stability assays in which the absence of one of these proteins negatively impacts the stability of the others, two-hybrid studies, co-immunoprecipitation, co-purification and/or co-crystallization). In brief, PilM interacts with the short cytoplasmic portion of PilN (Karuppiyah and Derrick 2011; Georgiadou et al., 2012), making the PilMN complex an orthologue of GspL (Fig. 2). PilN interacts with PilO (Ayers et al., 2009; Sampaleanu et al., 2009; Georgiadou et al., 2012), which is also well documented for their GspL and GspM counterparts (Sandkvist et al., 1999; Py, Loiseau and Barras 2001), forming together a PilMNO complex (Karuppiyah et al., 2013). PilP is also part of this complex, but it is unclear whether it interacts with PilN, PilO or both (Georgiadou et al., 2012; Tammam et al., 2013). Critically, consistent with a role in Tff assembly, this complex interacts with the pilin substrate and the traffic ATPase. Interaction with pilin subunits (Karuppiyah et al., 2013; Tammam et al., 2013) occurs either via PilN (or the equivalent periplasmic portion of GspL), PilO, or both (Gray et al., 2011; Georgiadou et al., 2012). There is extensive experimental evidence in T2SS that the traffic ATPase GspE interacts with GspL (Sandkvist et al., 1995; Py, Loiseau and Barras 1999; Possot et al., 2000; Robert, Filloux and Michel 2005; Shiue et al., 2006), including the 3D structure of a complex between the cytoplasmic domain of GspL (GspL_{cyto}) and an N-terminal fragment of GspE (Abendroth et al., 2005). In the absence of GspL, GspE mislocalizes to the cytoplasm (Sandkvist et al., 1995), which is a strong indication that GspL anchors traffic ATPases to the membrane where they power Tff (dis)assembly. The very recent crystal structure of the full length GspE-GspL_{cyto} complex (Lu, Korotkov and Hol 2014b) identified linear 'arrays' of GspL_{cyto}, which could be the driving force for the formation of the assembly sub-complex. The subsequent swinging motion of traffic ATPase domains within a hexamer would thus 'drag' interacting GspL together with GspM, thereby powering pilin assembly into Tff. The possibility that PilM links PilF (and PilT) with the rest of the (dis)assembly sub-complex remains to be formally demonstrated, but it is very likely since the patch of residues that mediate the interaction of GspL with GspE is conserved in PilM (Abendroth et al., 2005; Karuppiyah and Derrick 2011). Importantly, this might also be the norm in archaea as suggested by the finding that FlaH (that is an ATP-binding protein like PilM) interacts with the FlaI traffic ATPase (Banerjee et al., 2013).

Recently, however, when the above pilT suppressor assay was used in *P. aeruginosa*, important discrepancies with the above

model emerged. It was reported that *pilM/T*, *pilN/T*, *pilO/T* and *pilP/T* mutants exhibit piliation (although at low levels when compared to the wild type), while the *pilC/T* remains non-piliated (PilC is the orthologue of meningococcal PilG) (Takhar et al., 2013). In contrast, the *pilG/T* meningococcal mutant is heavily piliated and even exhibits a partial restoration of Tfp-linked phenotypes (Carbonnelle et al., 2006). The main experimental difference between the two studies is that piliation was assessed by immunofluorescence microscopy in *N. meningitidis*, which is a direct assay of Tfp production, while in the *P. aeruginosa* study it was assessed indirectly via immunodetection of the pilin subunit in sheared supernatant fractions. Their results led Takhar et al., to propose that the energy generated by the traffic ATPase in the cytoplasm is translocated to pilins mainly through the PilG 'platform' protein, and that owing to the interaction between PilP and the secretin (see below) PilMNOP would represent a 'secretin dynamic-associated' sub-complex (Ayers, Howell and Burrows 2010) or a 'connecting module' between inner and outer membrane sub-complexes (Nivaskumar and Francetic 2014). A strong indication against such a merely connecting role is the finding that PilM, PilN and PilO orthologues are also found in Gram-positive bacteria expressing Tfp (Melville and Craig 2013). Nevertheless, the belief that PilG must play a key role in Tff assembly is longstanding (Hobbs and Mattick 1993) and mainly derives from the fact that it is universal in Tff systems (Fig. 4). Unfortunately, although PilG has been one of the first Tff proteins to be identified (Nunn, Bergman and Lory 1990), its exact function remains unknown. What is clear is that PilG is a polytopic cytoplasmic membrane protein with three transmembrane domains leaving the N-terminus and a large loop in the distal part of the protein exposed in the cytoplasm, while only a very small loop and few residues of the C-terminus are located in the periplasm (Thomas, Reeves and Salmond 1997; Arts et al., 2007a). A different topology has also been reported but was judged to be uncommon (Blank and Donnenberg 2001). 3D structures of the N-terminal cytoplasmic portion of this protein (Abendroth et al., 2009b; Karupiah et al., 2010; Kolappan and Craig 2013), which revealed a six-helix bundle, strongly suggested that it functions by interacting with other cytoplasmic proteins but offered little clue to its exact functional role. Accordingly, and although it is much less extensive than above, there is evidence that PilG interacts with the traffic ATPase (Py, Loiseau and Barras 2001; Chiang, Habash and Burrows 2005; Arts et al., 2007a; Takhar et al., 2013), with the pilin subunit (Georgiadou et al., 2012), and also the above PilMNOP sub-complex (Py, Loiseau and Barras 2001; Georgiadou et al., 2012), all of which are compatible with a role in Tff assembly.

Surmising that Tfp assembly is even less likely to be different in two Tffa-producing species such as *N. meningitidis* and *P. aeruginosa* that have essentially identical sets of Tfp biogenesis proteins, than it is in different Tff systems, more direct evidence will be necessary to confirm and/or refute one, the other or both of the above models. Furthermore, how the corresponding sub-complex might then polymerize mature pilins into Tff remains to be determined.

Crossing the outer membrane

In Gram-negative bacteria, there is an additional sub-complex in the outer membrane centred on the secretin PilQ, which forms a gated pore allowing translocation across this second permeability barrier of the Tff themselves as in the case of Tfp, or Tff-secreted substrates as in T2SS (Fig. 2). It is the finding in pathogenic *Neisseria* species that filaments in a *pilQ/T*

mutant remain trapped in the periplasm that provided the most compelling evidence that Tfp emerge on the surface through the secretin pore and that this sub-complex is not involved in pilus assembly *per se* (Wolfgang et al., 2000; Carbonnelle et al., 2006). Secretins are a vast group of outer membrane proteins found in different bacterial secretion systems (Tff, T3SS and filamentous phages), and have therefore been intensively studied (Korotkov, Gonen and Hol 2011a). They generally share a high level of homology at their C-terminus, which is necessary for oligomerization within the outer membrane to generate multimers of usually 12–14 subunits, often heat- and SDS-resistant (Kazmierczak et al., 1994; Hardie, Lory and Pugsley 1996a; Drake, Sandstedt and Koomey 1997). Towards their N-terminus there is decreasing homology (Korotkov, Gonen and Hol 2011a; Berry et al., 2012). This part of the protein is constituted by a number of discrete and more flexible globular domains, which extend deeply into the periplasm, possibly up to the cytoplasmic membrane. The other member of the secretin sub-complex in most systems is PilP/GspC (Fig. 2), an inner membrane lipoprotein/bitopic protein, that interacts with the secretin and connects the inner and outer membrane sub-complexes (Possot, Gerard-Vincent and Pugsley 1999; Balasingham et al., 2007; Tammam et al., 2013). As revealed by their very similar 3D structures (Golovanov et al., 2006; Tammam et al., 2011; Gu et al., 2012), PilP and the HR domain of GspC are functionally equivalent with a long disordered N-terminal 'arm' and a folded C-terminal globular β -sandwich domain. The globular domain of PilP interacts with the N0 domain of the secretin, thus bridging the different sub-complexes involved in Tff biogenesis (Korotkov et al., 2011b; Berry et al., 2012; Tammam et al., 2013). An 'outside-in' assembly model, whereby the secretin sub-complex would form first and then recruit and stabilize the other sub-complexes, has been proposed in some species (Lybarger et al., 2009; Friedrich, Bulyha and Sogaard-Andersen 2014). However, it is unclear whether this is a general feature since in *Neisseria* species intra-periplasmic Tfp can be assembled in a *pilQ/T* double mutant (Wolfgang et al., 2000; Carbonnelle et al., 2006), suggesting that the other sub-complexes are assembled and functional in the absence of the secretin sub-complex. Another important observation consistent with the role of the PilPQ sub-complex is the absence of both proteins in Gram-positive species expressing Tfp (Melville and Craig 2013). This suggests that PilP is unlikely to play a role in Tff assembly, which was inferred from the absence of Tfp in a meningococcal *pilP/T* mutant (Carbonnelle et al., 2006). It is possible that the lack of piliation in this mutant was merely a consequence of the dramatic instability of PilN and PilO in the absence of PilP (Ayers et al., 2009; Georgiadou et al., 2012).

There is an abundance of structural information concerning secretins that has outlined common features. Early visualization of PilQ multimers by EM revealed ring-like dodecameric structures (Linderoth, Simon and Russel 1997; Nouwen et al., 1999). Increasingly high-resolution 3D cryo-EM reconstructions showed cylindrical dodecamers spanning the periplasm consisting of a series of rings defining outer membrane and periplasmic domains with a closed central cavity (the so-called 'periplasmic vestibule'), the size of which is compatible with the passage of Tff (Opalka et al., 2003; Collins et al., 2004; Chami et al., 2005; Burkhardt et al., 2012; Tosi et al., 2014). It has been demonstrated *in vitro* that *N. meningitidis* PilQ multimers are able to physically accommodate purified Tfp while undergoing important conformational changes (Collins et al., 2005). Atomic resolution 3D structure of a full-length secretin is still elusive, but structures for several of the globular domains of the

periplasmic vestibule have been obtained. The N0 and N1 domains that are found in all secretins form compact and globular α/β folds (Korotkov et al., 2009; Berry et al., 2012). An important and unique feature in most Tfp-dependent secretins (*T. thermophilus* appears to be an exception) is the presence of one or two β -sandwich domains (B1/2) at their extreme N-terminus (Berry et al., 2012).

The secretin sub-complex often contains (sometimes transiently) other components that are necessary for the pore function and/or stability (Koo, Burrows and Howell 2012). Following the seminal studies with GspS from *K. oxytoca* (Hardie, Lory and Pugsley 1996a), most of these proteins have been (mis)named pilotins. GspS, which is a small outer membrane lipoprotein (D'Enfert and Pugsley 1989), binds the CTD of GspD monomers (Daefler et al., 1997), protects them from degradation and is necessary for their correct localization in the outer membrane (Hardie, Lory and Pugsley 1996a; Hardie et al., 1996b). In the absence of GspS, or its mislocalization, GspD multimers mislocalize to the cytoplasmic membrane (Guilvout et al., 2006). These findings, which were confirmed in many different species/systems (Shevchik, Robert-Baudouy and Condemine 1997; Schuch and Maurelli 2001), showed that the secretin does not require GspS for multimerization or membrane insertion, but rather that GspS is a chaperone that 'pilots' the secretin monomers to the outer membrane, which it reaches via the Lol pathway (hence its pilotin moniker). Strikingly, pilotins in different secretion systems are extremely diverse, showing no sequence or structural homology (Lario et al., 2005; Tosi et al., 2011). Many, possibly most, Tff systems do not need pilotins either because secretins are lipoproteins themselves (Schmidt et al., 2001; Viarre et al., 2009) or use general pathways for protein targeting to the outer membrane (Voulhoux et al., 2003). Another protein that is often part of secretin sub-complexes is PilW (Fig. 2). This outer membrane lipoprotein (Carbonnelle et al., 2005), which is almost exclusively found in Proteobacteria and most likely restricted to Tfpa (Fig. 4), has been found to interact with PilQ (Koo et al., 2013) and is key for the stability of PilQ multimers. Indeed, in PilW absence PilQ multimers could not be detected (Carbonnelle et al., 2005; Nudleman, Wall and Kaiser 2006; Koo et al., 2008). Unlike what has been observed for GspS (Hardie et al., 1996b), PilW mislocalization does not prevent PilQ localization to the outer membrane and/or piliation (Koo et al., 2008; Szeto et al., 2011) and it is therefore not a *bona fide* pilotin. Two PG-binding proteins, FimV and TsaP, have also been found as part of the secretin sub-complex in some Tfpa-expressing species, but it is unclear if this is conserved in other Tff. While FimV has been shown in *P. aeruginosa* to be important for PilQ stability and Tfp-linked functions (Wehbi et al., 2011), TsaP was identified as a protein interacting peripherally with secretin rings in *N. gonorrhoeae* (Siewering et al., 2014). Although lack of TsaP did not affect PilQ multimerization, the secretin channel was apparently inactive since a *tsaP* mutant exhibited intra-periplasmic fibres, reminiscent of those in a *pilQ/T* mutant (Wolfgang et al., 2000; Carbonnelle et al., 2006). These PG-binding proteins were suggested to anchor the secretin multimers to the cell wall, enabling them to withstand dramatic forces generated during pilus (dis)assembly, which warrants further investigation.

Late stages in Tff biogenesis

As mentioned above, most proteins essential for Tfpa biogenesis are actually dispensable for filament assembly *per se* since the piliation defect in the corresponding genes could be suppressed by a second mutation in *pilT*. Except for *pilQ/T*, all the

other double mutants display surface-exposed Tfp (Carbonnelle et al., 2006), suggesting that the corresponding proteins (PilC, PilG, PilH, PilI, PilJ, PilK and PilW) act later than PilQ. Their putative role is to shift the Tfp dynamics towards assembly and recent publications have shed light on the function of the pilin-like proteins (PilH, PilI, PilJ and PilK) and PilC/PilY1.

In each Tff system, there are almost invariably additional genes beside the major (pseudo)pilin that encode proteins with class III signal peptides, are cleaved by the prepilin peptidase and play key roles in Tff biology. PilH, PilI, PilJ and PilK are thus required for Tfpa biogenesis (Winther-Larsen et al., 2005; Carbonnelle et al., 2006), and their GspH, GspI, GspJ and GspK counterparts are essential for T2SS (Lu, Motley and Lory 1997). An idea of the role of these minor (pseudo)pilins came from the 3D structure of a GspIJK hetero-trimer that revealed a Tff-like architecture with an axial rise of 10 Å between neighbouring subunits (Korotkov and Hol 2008). The fourth pseudopilin GspH was found to bind to GspJ at the base of this trimer (Douzi et al., 2009) and the T2SS substrate was found to interact with its tip (Douzi et al., 2011). Importantly, GspK that caps this complex has a large domain positioned in such a way that no additional subunit can be added above it (Korotkov and Hol 2008), which would be consistent with a localization at the tip of pseudopili and a possible role in priming filament assembly. However, in *P. aeruginosa* Tfp, the corresponding proteins have not been detected at the tip but rather distributed throughout the filaments (Giltner, Habash and Burrows 2010). Nevertheless, in accord with this scenario, hyper-pseudopilus abundance and/or length was found to be affected in mutants in *gspI*, *gspJ* and *gspK* (but not *gspH*) and was abolished in the concurrent absence of all these proteins (Durand et al., 2005; Cisneros et al., 2012a). Cysteine cross-linking experiments and molecular dynamics simulations outlined a model in which GspI and GspJ form a staggered complex which recruits GspK, partially extracting it from the membrane by 10 Å (Cisneros et al., 2012a). This was proposed to 'kick-start' initiation of pseudopilus assembly, possibly by activating the assembly ATPase in the cytoplasm (although this remains purely speculative). The finding that the *E. coli* Tfp equivalents of these pseudopilins (from the PpdD pilus) can complement the hyper-pseudopiliation defect in a *gspHIJK* polymutant is strong evidence that the role of these proteins in Tff assembly might be conserved (Cisneros et al., 2012b). In Tfp, however, the finding in *N. gonorrhoeae* that wild-type levels of piliation can be restored in the concurrent absence of PilH, PilI, PilJ and PilK when *pilT* is also mutated (Winther-Larsen et al., 2005) seems difficult to reconcile with a role of these proteins in initiating filament assembly.

PilC/PilY1 is a protein found predominantly in Tfpa produced by Proteobacteria (Fig. 4), first identified in *N. gonorrhoeae* (Jonsson, Nyberg and Normark 1991). Orthologues in different species have diverse NTD, while their C-terminus is highly conserved. The protein is predicted to be associated with the outer membrane (Rahman et al., 1997; Carbonnelle et al., 2005), although a pilus localization has also been proposed (Rudel, Scheuerpflug and Meyer 1995). A series of studies in different species have shown that PilC is a bifunctional protein involved in Tfp-mediated adhesion (already discussed) and pilus biogenesis. The conserved CTD is responsible for PilC role in Tfp biogenesis (Orans et al., 2010). The 3D structure of the corresponding domain in PilY1 revealed a modified β -propeller fold with a distinct and highly conserved EF-hand-like calcium-binding site (Orans et al., 2010). Calcium was proposed to control PilT-mediated pilus retraction, consistent with the phenotype of a *pilC/T* mutant. Calcium-bound PilY1 inhibits PilT-mediated pilus retraction, while in a calcium-free state it is unable to do so,

resulting in a non-piliated phenotype. However, although later studies agree that calcium binding by PilC is key for Tff biology in other pilated proteobacterial species as well, no major effect on piliation was seen in calcium-free mutants (Cheng et al., 2013; Porsch et al., 2013).

CONCLUDING REMARKS

Despite considerable progress, our understanding of Tff biology remains incomplete and there are important gaps in knowledge to be filled. Obviously, the main challenge consists in improving our understanding of the molecular mechanisms of Tff assembly. Perhaps the most spectacular advances in recent years came from structural studies showing that proteins in diverse Tff systems that have diverged to the point that no sequence homology is discernible do exhibit highly similar 3D structures. This is a strong argument in favour of the notion that common molecular principles govern Tff biology. Additional structural information has the potential to further improve our understanding of these principles. For example, cryo-EM combined with direct electron detectors (Lu et al., 2014a) could be used to obtain much higher, possibly atomic, resolution reconstruction of various Tff, including Flp pili that are formed of a 'minimal' pilin (Tomich, Planet and Figurski 2007). The 3D structure of more 'orphan' Tff proteins from less well characterized systems might reveal that they are structural homologues of bacterial proteins to which they bear no sequence homology (e.g. is FlaH an orthologue of PilM?), fuelling the emergence of one unifying mechanism for Tff assembly. Atomic-resolution structures of a secretin channel or a bacterial prepilin peptidase in complex with a prepilin substrate would shed more light on these key steps in Tff biology. Finally, although this promises to be utterly complex, it would be worth trying to determine the structure of the various sub-complexes discussed here (full-length proteins rather than soluble domains) because this would dramatically improve our understanding of Tff biology and bring it a little closer to the exquisite understanding of pilus assembly by the chaperone-usher pathway (Phan et al., 2011). Such advances will be instrumental in the rational design of specific inhibitors of Tff assembly, analogues of 'pilicides' interfering with pilus assembly by the chaperone-usher pathway (Pinkner et al., 2006) or curli biogenesis (Cegelski et al., 2009), which would represent exceptionally broad-spectrum anti-microbial compounds (Fig. 4).

Structural biology is obviously not the only research avenue that should be privileged, and further genetic, biochemical and dynamic modelling studies of different Tff properties and/or proteins are warranted. As confirmed by the wealth of interesting data coming from archaeal Tff (Jarrell et al., 2013), future research should not only favour the 'historic' models (although there is still plenty to be learned in T2SS, Bfp, Tcp or *Neisseria* and *P. aeruginosa* Tfp) but should be extended to new models, more distant from an evolutionary point of view. An in-depth study of Gram-positive Tfp would be for example of great interest as such bacteria are inherently 'simpler' because of the absence of an outer membrane (Melville and Craig 2013). This could be key in understanding how Tff cross the PG. Another attractive 'reductionist' approach would be to determine which proteins are necessary and sufficient for filament assembly by creating a minimal system, which could be done in several ways. Similar to what was done for Tfpb or T2SS with the transfer of the entire corresponding operons (D'Enfert, Ryter and Pugsley 1987; Sohler et al., 1996; Stone et al., 1996), a surrogate non-piliated organism could be used to determine which subset of genes is sufficient

to promote Tff assembly. The *pilT* suppressor assay pioneered in *Neisseria* species (Wolfgang et al., 1998b) could be extended by creating polymutants in a *pilT* mutant background in which all the genes not involved in Tff assembly would be deleted. Ultimately, a minimal Tff assembly system could be reconstituted from proteins expressed in a cell-free system (Aly et al., 2013) or even purified, as has been achieved for type I pili (Nishiyama et al., 2008). All of these studies, and many more, will undoubtedly continue to 'feed' the Tff field with interesting results and exciting concepts in the years to come.

ACKNOWLEDGEMENTS

We would like to thank Chiara Recchi and Vivianne Goosens (Imperial College London) for critical reading of this manuscript. Work in our laboratory is currently supported by the Biotechnology and Biological Sciences Research Council and Medical Research Council.

Conflict of interest statement. None declared.

REFERENCES

- Aas FE, Winther-Larsen HC, Wolfgang M, et al., Substitutions in the N-terminal alpha helical spine of *Neisseria gonorrhoeae* pilin affect type IV pilus assembly, dynamics and associated functions. *Mol Microbiol* 2007;**63**:69–85.
- Abendroth J, Bagdasarian M, Sandkvist M, et al., The structure of the cytoplasmic domain of EpsL, an inner membrane component of the type II secretion system of *Vibrio cholerae*: an unusual member of the actin-like ATPase superfamily. *J Mol Biol* 2004a;**344**:619–33.
- Abendroth J, Kreger AC, Hol WG. The dimer formed by the periplasmic domain of EpsL from the type 2 secretion system of *Vibrio parahaemolyticus*. *J Struct Biol* 2009a;**168**:313–22.
- Abendroth J, Mitchell DD, Korotkov KV, et al., The three-dimensional structure of the cytoplasmic domains of EpsF from the type 2 secretion system of *Vibrio cholerae*. *J Struct Biol* 2009b;**166**:303–15.
- Abendroth J, Murphy P, Sandkvist M, et al., The X-ray structure of the type II secretion system complex formed by the N-terminal domain of EpsE and the cytoplasmic domain of EpsL of *Vibrio cholerae*. *J Mol Biol* 2005;**348**:845–55.
- Abendroth J, Rice AE, McLuskey K, et al., The crystal structure of the periplasmic domain of the type II secretion system protein EpsM from *Vibrio cholerae*: the simplest version of the ferredoxin fold. *J Mol Biol* 2004b;**338**:585–96.
- Akahane K, Sakai D, Furuya N, et al., Analysis of the *pilU* gene for the prepilin peptidase involved in the biogenesis of type IV pili encoded by plasmid R64. *Mol Genet Genomics* 2005;**273**:350–9.
- Alm RA, Boder AJ, Free PD, et al., Identification of a novel gene, *pilZ*, essential for type 4 fimbrial biogenesis in *Pseudomonas aeruginosa*. *J Bacteriol* 1996a;**178**:46–53.
- Alm RA, Hallinan JP, Watson AA, et al., Fimbrial biogenesis genes of *Pseudomonas aeruginosa*: *pilW* and *pilX* increase the similarity of type 4 fimbriae to the GSP protein-secretion systems and *pilY1* encodes a gonococcal PilC homologue. *Mol Microbiol* 1996b;**22**:161–73.
- Alm RA, Mattick JS. Genes involved in the biogenesis and function of type 4 fimbriae in *Pseudomonas aeruginosa*. *Gene* 1997;**192**:89–98.
- Aly KA, Beebe ET, Chan CH, et al., Cell-free production of integral membrane aspartic acid proteases reveals zinc-dependent

- methyltransferase activity of the *Pseudomonas aeruginosa* prepilin peptidase PilD. *Microbiologyopen* 2013;2:94–104.
- Arts J, de Groot A, Ball G, et al., Interaction domains in the *Pseudomonas aeruginosa* type II secretory apparatus component XcpS (GspF). *Microbiology* 2007a;153:1582–92.
- Arts J, van Boxtel R, Filloux A, et al., Export of the pseudopilin XcpT of the *Pseudomonas aeruginosa* type II secretion system via the signal recognition particle-Sec pathway. *J Bacteriol* 2007b;189:2069–76.
- Aukema KG, Kron EM, Herdendorf TJ, et al., Functional dissection of a conserved motif within the pilus retraction protein PilT. *J Bacteriol* 2005;187:611–8.
- Ayers M, Howell PL, Burrows LL. Architecture of the type II secretion and type IV pilus machineries. *Future Microbiol* 2010;5:1203–18.
- Ayers M, Sampaleanu LM, Tammam S, et al., PilM/N/O/P proteins form an inner membrane complex that affects the stability of the *Pseudomonas aeruginosa* type IV pilus secretin. *J Mol Biol* 2009;394:128–42.
- Baker JL, Biaisi N, Tama F. Steered molecular dynamics simulations of a type IV pilus probe initial stages of a force-induced conformational transition. *PLoS Comput Biol* 2013;9:e1003032.
- Balaban M, Battig P, Muschiol S, et al., Secretion of a pneumococcal type II secretion system pilus correlates with DNA uptake during transformation. *Proc Natl Acad Sci USA* 2014;111:E758–65.
- Balasingham SV, Collins RF, Assalkhou R, et al., Interactions between the lipoprotein PilP and the secretin PilQ in *Neisseria meningitidis*. *J Bacteriol* 2007;189:5716–27.
- Banerjee A, Neiner T, Tripp P, et al., Insights into subunit interactions in the *Sulfolobus acidocaldarius* archaeum cytoplasmic complex. *FEBS J* 2013;280:6141–9.
- Bardy SL, Jarrell KF. Cleavage of preflagellins by an aspartic acid signal peptidase is essential for flagellation in the archaeon *Methanococcus voltae*. *Mol Microbiol* 2003;50:1339–47.
- Bernard SC, Simpson N, Join-Lambert O, et al., Pathogenic *Neisseria meningitidis* utilizes CD147 for vascular colonization. *Nat Med* 2014;20:725–31.
- Berry JL, Cehovin A, McDowell MA, et al., Functional analysis of the interdependence between DNA uptake sequence and its cognate ComP receptor during natural transformation in *Neisseria* species. *PLoS Genet* 2013;9:e1004014.
- Berry JL, Phelan MM, Collins RF, et al., Structure and assembly of a trans-periplasmic channel for type IV pili in *Neisseria meningitidis*. *PLoS Pathog* 2012;8:e1002923.
- Bhaya D, Bianco NR, Bryant D, et al., Type IV pilus biogenesis and motility in the cyanobacterium *Synechocystis* sp. PCC6803. *Mol Microbiol* 2000;37:941–51.
- Biaisi N, Higashi DL, Brujic J, et al., Force-dependent polymorphism in type IV pili reveals hidden epitopes. *Proc Natl Acad Sci USA* 2010;107:11358–63.
- Biaisi N, Ladoux B, Higashi D, et al., Cooperative retraction of bundled type IV pili enables nanonewton force generation. *PLoS Biol* 2008;6:e87.
- Bieber D, Ramer SW, Wu C, et al., Type IV pili, transient bacterial aggregates, and virulence of enteropathogenic *Escherichia coli*. *Science* 1998;280:2114–8.
- Blank TE, Donnenberg MS. Novel topology of BfpE, a cytoplasmic membrane protein required for type IV fimbrial biogenesis in enteropathogenic *Escherichia coli*. *J Bacteriol* 2001;183:4435–50.
- Boesen T, Nielsen LP. Molecular dissection of bacterial nanowires. *mBio* 2013;4:e00270–13.
- Bradley DE. Shortening of *Pseudomonas aeruginosa* pili after RNA-phage adsorption. *J Gen Microbiol* 1972;72:303–19.
- Bradley DE. A function of *Pseudomonas aeruginosa* PAO polar pili: twitching motility. *Can J Microbiol* 1980;26:146–54.
- Brossay L, Paradis G, Fox R, et al., Identification, localization, and distribution of the PilT protein in *Neisseria gonorrhoeae*. *Infect Immun* 1994;62:2302–8.
- Brown D, Helaine S, Carbonnelle E, et al., Systematic functional analysis reveals that a set of 7 genes is involved in fine tuning of the multiple functions mediated by type IV pili in *Neisseria meningitidis*. *Infect Immun* 2010;78:3053–63.
- Burkhardt J, Vonck J, Langer JD, et al., Unusual N-terminal $\alpha\alpha\beta\beta\beta$ fold of PilQ from *Thermus thermophilus* mediates ring formation and is essential for piliation. *J Biol Chem* 2012;287:8484–94.
- Camberg JL, Johnson TL, Patrick M, et al., Synergistic stimulation of EpsE ATP hydrolysis by EpsL and acidic phospholipids. *EMBO J* 2007;26:19–27.
- Campos M, Francetic O, Nilges M. Modeling pilus structures from sparse data. *J Struct Biol* 2011;173:436–44.
- Campos M, Nilges M, Cisneros DA, et al., Detailed structural and assembly model of the type II secretion pilus from sparse data. *Proc Natl Acad Sci USA* 2010;107:13081–6.
- Carbonnelle E, Helaine S, Nassif X, et al., A systematic genetic analysis in *Neisseria meningitidis* defines the Pil proteins required for assembly, functionality, stabilization and export of type IV pili. *Mol Microbiol* 2006;61:1510–22.
- Carbonnelle E, Helaine S, Prouvensier L, et al., Type IV pilus biogenesis in *Neisseria meningitidis*: PilW is involved in a step occurring after pilus assembly, essential for fiber stability and function. *Mol Microbiol* 2005;55:54–64.
- Cegelski L, Pinkner JS, Hammer ND, et al., Small-molecule inhibitors target *Escherichia coli* amyloid biogenesis and biofilm formation. *Nat Chem Biol* 2009;5:913–9.
- Cehovin A, Simpson PJ, McDowell MA, et al., Specific DNA recognition mediated by a type IV pilin. *Proc Natl Acad Sci USA* 2013;110:3065–70.
- Chaban B, Voisin S, Kelly J, et al., Identification of genes involved in the biosynthesis and attachment of *Methanococcus voltae* N-linked glycans: insight into N-linked glycosylation pathways in Archaea. *Mol Microbiol* 2006;61:259–68.
- Chami M, Guilvout I, Gregorini M, et al., Structural insights into the secretin PulD and its trypsin-resistant core. *J Biol Chem* 2005;280:37732–41.
- Chen I, Dubnau D. DNA uptake during bacterial transformation. *Nat Rev Microbiol* 2004;2:241–9.
- Cheng Y, Johnson MD, Burillo-Kirch C, et al., Mutation of the conserved calcium-binding motif in *Neisseria gonorrhoeae* PilC1 impacts adhesion but not piliation. *Infect Immun* 2013;81:4280–9.
- Chiang P, Habash M, Burrows LL. Disparate subcellular localization patterns of *Pseudomonas aeruginosa* type IV pilus ATPases involved in twitching motility. *J Bacteriol* 2005;187:829–39.
- Chiang SL, Taylor RK, Koomey M, et al., Single amino acid substitutions in the N-terminus of *Vibrio cholerae* TcpA affect colonization, autoagglutination, and serum resistance. *Mol Microbiol* 1995;17:1133–42.
- Cisneros DA, Bond PJ, Pugsley AP, et al., Minor pseudopilin self-assembly primes type II secretion pseudopilus elongation. *EMBO J* 2012a;31:1041–53.
- Cisneros DA, Pehau-Arnaudet G, Francetic O. Heterologous assembly of type IV pili by a type II secretion system reveals the role of minor pilins in assembly initiation. *Mol Microbiol* 2012b;86:805–18.

- Collins RF, Frye SA, Balasingham S, et al., Interaction with type IV pili induces structural changes in the bacterial outer membrane secretin PilQ. *J Biol Chem* 2005;**280**:18923–30.
- Collins RF, Frye SA, Kitmitto A, et al., Structure of the *Neisseria meningitidis* outer membrane PilQ secretin complex at 12 Å resolution. *J Biol Chem* 2004;**279**:39750–6.
- Coureuil M, Lecuyer H, Scott MG, et al., Meningococcus hijacks a β 2-adrenoceptor/ β -arrestin pathway to cross brain microvasculature endothelium. *Cell* 2010;**143**:1149–60.
- Craig L, Pique ME, Tainer JA. Type IV pilus structure and bacterial pathogenicity. *Nat Rev Microbiol* 2004;**2**:363–78.
- Craig L, Taylor RK, Pique ME, et al., Type IV pilin structure and assembly. X-ray and EM analyses of *Vibrio cholerae* toxin-coregulated pilus and *Pseudomonas aeruginosa* PAK pilin. *Mol Cell* 2003;**11**:1139–50.
- Craig L, Volkmann N, Arvai AS, et al., Type IV pilus structure by cryo-electron microscopy and crystallography: implications for pilus assembly and functions. *Mol Cell* 2006;**23**:651–62.
- D'Enfert C, Pugsley AP. *Klebsiella pneumoniae* pulS gene encodes an outer membrane lipoprotein required for pullulanase secretion. *J Bacteriol* 1989;**171**:3673–9.
- D'Enfert C, Ryter A, Pugsley AP. Cloning and expression in *Escherichia coli* of the *Klebsiella pneumoniae* genes for production, surface localization and secretion of the lipoprotein pullulanase. *EMBO J* 1987;**6**:3531–8.
- Daefler S, Guilvout I, Hardie KR, et al., The C-terminal domain of the secretin PulD contains the binding site for its cognate chaperone, PulS, and confers PulS dependence on pIVf1 function. *Mol Microbiol* 1997;**24**:465–75.
- Dalrymple B, Mattick JS. An analysis of the organization and evolution of type 4 fimbrial (MePhe) subunit proteins. *J Mol Evol* 1987;**25**:261–9.
- de Bentzmann S, Aurouze M, Ball G, et al., FppA, a novel *Pseudomonas aeruginosa* prepilin peptidase involved in assembly of type IVb pili. *J Bacteriol* 2006;**188**:4851–60.
- Desmond E, Brochier-Armanet C, Gribaldo S. Phylogenomics of the archaeal flagellum: rare horizontal gene transfer in a unique motility structure. *BMC Evol Biol* 2007;**7**:106.
- Donnenberg MS, Zhang HZ, Stone KD. Biogenesis of the bundle-forming pilus of enteropathogenic *Escherichia coli*: reconstitution of fimbriae in recombinant *E. coli* and role of DsbA in pilin stability—a review. *Gene* 1997;**192**:33–8.
- Douzi B, Ball G, Cambillau C, et al., Deciphering the Xcp *Pseudomonas aeruginosa* type II secretion machinery through multiple interactions with substrates. *J Biol Chem* 2011;**286**:40792–801.
- Douzi B, Durand E, Bernard C, et al., The XcpV/GspI pseudopilin has a central role in the assembly of a quaternary complex within the T2SS pseudopilus. *J Biol Chem* 2009;**284**:34580–9.
- Douzi B, Filloux A, Voulhoux R. On the path to uncover the bacterial type II secretion system. *Philos T Roy Soc B* 2012;**367**:1059–72.
- Drake SL, Sandstedt SA, Koomey M. PilP, a pilus biogenesis lipoprotein in *Neisseria gonorrhoeae*, affects expression of PilQ as a high-molecular-mass multimer. *Mol Microbiol* 1997;**23**:657–68.
- Dupuy B, Taha MK, Pugsley AP, et al., *Neisseria gonorrhoeae* prepilin export studied in *Escherichia coli*. *J Bacteriol* 1991;**173**:7589–98.
- Durand E, Bernadac A, Ball G, et al., Type II protein secretion in *Pseudomonas aeruginosa*: the pseudopilus is a multifibrillar and adhesive structure. *J Bacteriol* 2003;**185**:2749–58.
- Durand E, Michel G, Voulhoux R, et al., XcpX controls biogenesis of the *Pseudomonas aeruginosa* XcpT-containing pseudopilus. *J Biol Chem* 2005;**280**:31378–89.
- Forest KT, Dunham SA, Koomey M, et al., Crystallographic structure reveals phosphorylated pilin from *Neisseria*: phosphoserine sites modify type IV pilus surface chemistry and fibre morphology. *Mol Microbiol* 1999;**31**:743–52.
- Francetic O, Buddelmeijer N, Lewenza S, et al., Signal recognition particle-dependent inner membrane targeting of the PulG pseudopilin component of a type II secretion system. *J Bacteriol* 2007;**189**:1783–93.
- Friedrich C, Bulyha I, Sogaard-Andersen L. Outside-in assembly pathway of the type IV pilus system in *Myxococcus xanthus*. *J Bacteriol* 2014;**196**:378–90.
- Fröls S, Ajon M, Wagner M, et al., UV-inducible cellular aggregation of the hyperthermophilic archaeon *Sulfolobus solfataricus* is mediated by pili formation. *Mol Microbiol* 2008;**70**:938–52.
- Gault J, Malosse C, Dumenil G, et al., A combined mass spectrometry strategy for complete posttranslational modification mapping of *Neisseria meningitidis* major pilin. *J Mass Spectrom* 2013;**48**:1199–206.
- Georgiadou M, Castagnini M, Karimova G, et al., Large-scale study of the interactions between proteins involved in type IV pilus biology in *Neisseria meningitidis*: characterization of a subcomplex involved in pilus assembly. *Mol Microbiol* 2012;**84**:857–73.
- Gibiansky ML, Conrad JC, Jin F, et al., Bacteria use type IV pili to walk upright and detach from surfaces. *Science* 2010;**330**:197.
- Giltner CL, Habash M, Burrows LL. *Pseudomonas aeruginosa* minor pilins are incorporated into type IV pili. *J Mol Biol* 2010;**398**:444–61.
- Giltner CL, Nguyen Y, Burrows LL. Type IV pilin proteins: versatile molecular modules. *Microbiol Mol Biol R* 2012;**76**:740–72.
- Golovanov AP, Balasingham S, Tzitzilonis C, et al., The solution structure of a domain from the *Neisseria meningitidis* lipoprotein PilP reveals a new β -sandwich fold. *J Mol Biol* 2006;**364**:186–95.
- Gray MD, Bagdasarjan M, Hol WG, et al., In vivo cross-linking of EpsG to EpsL suggests a role for EpsL as an ATPase-pseudopilin coupling protein in the type II secretion system of *Vibrio cholerae*. *Mol Microbiol* 2011;**79**:786–98.
- Gu S, Kelly G, Wang X, et al., Solution structure of homology region (HR) domain of type II secretion system. *J Biol Chem* 2012;**287**:9072–80.
- Guberman JM, Ai J, Arnaiz O, et al., BioMart Central Portal: an open database network for the biological community. *Database (Oxford)* 2011;bar041.
- Guilvout I, Chami M, Engel A, et al., Bacterial outer membrane secretin PulD assembles and inserts into the inner membrane in the absence of its pilotin. *EMBO J* 2006;**25**:5241–9.
- Guzzo CR, Salinas RK, Andrade MO, et al., PilZ protein structure and interactions with PilB and the FimX EAL domain: implications for control of type IV pilus biogenesis. *J Mol Biol* 2009;**393**:848–66.
- Han X, Kennan RM, Parker D, et al., Type IV fimbrial biogenesis is required for protease secretion and natural transformation in *Dichelobacter nodosus*. *J Bacteriol* 2007;**189**:5022–33.
- Hardie KR, Lory S, Pugsley AP. Insertion of an outer membrane protein in *Escherichia coli* requires a chaperone-like protein. *EMBO J* 1996a;**15**:978–88.
- Hardie KR, Seydel A, Guilvout I, et al., The secretin-specific, chaperone-like protein of the general secretory pathway: separation of proteolytic protection and piloting functions. *Mol Microbiol* 1996b;**22**:967–76.

- Hartung S, Arvai AS, Wood T, et al., Ultrahigh resolution and full-length pilin structures with insights for filament assembly, pathogenic functions, and vaccine potential. *J Biol Chem* 2011;**286**: 44254–65.
- Hegge FT, Hitchen PG, Aas FE, et al., Unique modifications with phosphocholine and phosphoethanolamine define alternate antigenic forms of *Neisseria gonorrhoeae* type IV pili. *Proc Natl Acad Sci USA* 2004;**101**:10798–803.
- Heiniger RW, Winther-Larsen HC, Pickles RJ, et al., Infection of human mucosal tissue by *Pseudomonas aeruginosa* requires sequential and mutually dependent virulence factors and a novel pilus-associated adhesin. *Cell Microbiol* 2010;**12**:1158–73.
- Helaine S, Carbonnelle E, Prouvensier L, et al., PilX, a pilus-associated protein essential for bacterial aggregation, is a key to pilus-facilitated attachment of *Neisseria meningitidis* to human cells. *Mol Microbiol* 2005;**55**:65–77.
- Helaine S, Dyer DH, Nassif X, et al., 3D structure/function analysis of PilX reveals how minor pilins can modulate the virulence properties of type IV pili. *Proc Natl Acad Sci USA* 2007;**104**:15888–93.
- Henrichsen J. Bacterial surface translocation: a survey and a classification. *Bacteriol Rev* 1972;**36**:478–503.
- Herdendorf TJ, McCaslin DR, Forest KT. *Aquifex aeolicus* PilT, homologue of a surface motility protein, is a thermostable oligomeric NTPase. *J Bacteriol* 2002;**184**:6465–71.
- Hobbs M, Mattick JS. Common components in the assembly of type 4 fimbriae, DNA transfer systems, filamentous phage and protein-secretion apparatus: a general system for the formation of surface-associated protein complexes. *Mol Microbiol* 1993;**10**:233–43.
- Horiuchi T, Komano T. Mutational analysis of plasmid R64 thin pilus prepilin: the entire prepilin sequence is required for processing by type IV prepilin peptidase. *J Bacteriol* 1998;**180**:4613–20.
- Howie HL, Glogauer M, So M. The *N. gonorrhoeae* type IV pilus stimulates mechanosensitive pathways and cytoprotection through a PilT-dependent mechanism. *PLoS Biol* 2005;**3**:e100.
- Hu J, Xue Y, Lee S, et al., The crystal structure of GXGD membrane protease FlaK. *Nature* 2011;**475**:528–31.
- Hunter S, Jones P, Mitchell A, et al., InterPro in 2011: new developments in the family and domain prediction database. *Nucleic Acids Res* 2012;**40**:D306–12.
- Hyland RM, Sun J, Griener TP, et al., The bundlin pilin protein of enteropathogenic *Escherichia coli* is an N-acetylglucosamine-specific lectin. *Cell Microbiol* 2008;**10**:177–87.
- Imam S, Chen Z, Roos DS, et al., Identification of surprisingly diverse type IV pili, across a broad range of Gram-positive bacteria. *PLoS One* 2011;**6**:e28919s.
- Imhaus AF, Dumenil G. The number of *Neisseria meningitidis* type IV pili determines host cell interaction. *EMBO J* 2014;**33**:1767–83.
- Jakovljevic V, Leonardy S, Hoppert M, et al., PilB and PilT are ATPases acting antagonistically in type IV pilus function in *Myxococcus xanthus*. *J Bacteriol* 2008;**190**:2411–21.
- Jarrell KF, Albers SV. The archaeellum: an old motility structure with a new name. *Trends Microbiol* 2012;**20**:307–12.
- Jarrell KF, Bayley DP, Kostyukova AS. The archaeal flagellum: a unique motility structure. *J Bacteriol* 1996;**178**:5057–64.
- Jarrell KF, Ding Y, Nair DB, et al., Surface appendages of archaea: structure, function, genetics and assembly. *Life* 2013;**3**:86–117.
- Jin F, Conrad JC, Gibiansky ML, et al., Bacteria use type-IV pili to slingshot on surfaces. *Proc Natl Acad Sci USA* 2011;**108**:12617–22.
- Johnson MD, Garrett CK, Bond JE, et al., *Pseudomonas aeruginosa* PilY1 binds integrin in an RGD- and calcium-dependent manner. *PLoS One* 2011;**6**:e29629.
- Johnston C, Campo N, Berge MJ, et al., *Streptococcus pneumoniae*, le transformiste. *Trends Microbiol* 2014;**22**:113–9.
- Jonsson AB, Nyberg G, Normark S. Phase variation of gonococcal pili by frameshift mutation in pilC, a novel gene for pilus assembly. *EMBO J* 1991;**10**:477–88.
- Kachlany SC, Planet PJ, DeSalle R, et al., flp-1, the first representative of a new pilin gene subfamily, is required for non-specific adherence of *Actinobacillus actinomycetemcomitans*. *Mol Microbiol* 2001;**40**:542–54.
- Karuppiiah V, Collins RF, Thistlethwaite A, et al., Structure and assembly of an inner membrane platform for initiation of type IV pilus biogenesis. *Proc Natl Acad Sci USA* 2013;**110**:E4638–47.
- Karuppiiah V, Derrick JP. Structure of the PilM-PilN inner membrane type IV pilus biogenesis complex from *Thermus thermophilus*. *J Biol Chem* 2011;**286**: 24434–42.
- Karuppiiah V, Hassan D, Saleem M, et al., Structure and oligomerization of the PilC type IV pilus biogenesis protein from *Thermus thermophilus*. *Proteins* 2010;**78**:2049–57.
- Kaufman MR, Seyer JM, Taylor RK. Processing of TCP pilin by TcpJ typifies a common step intrinsic to a newly recognized pathway of extracellular protein secretion by Gram-negative bacteria. *Gene Dev* 1991;**5**:1834–46.
- Kazmierczak BI, Mielke DL, Russel M, et al., pIV, a filamentous phage protein that mediates phage export across the bacterial cell envelope, forms a multimer. *J Mol Biol* 1994;**238**:187–98.
- Kehl-Fie TE, Miller SE, St Geme JW, III. *Kingella kingae* expresses type IV pili that mediate adherence to respiratory epithelial and synovial cells. *J Bacteriol* 2008;**190**:7157–63.
- Kirn TJ, Bose N, Taylor RK. Secretion of a soluble colonization factor by the TCP type 4 pilus biogenesis pathway in *Vibrio cholerae*. *Mol Microbiol* 2003;**49**:81–92.
- Kirn TJ, Lafferty MJ, Sandoe CMP, et al., Delineation of pilin domains required for bacterial association into microcolonies and intestinal colonization by *Vibrio cholerae*. *Mol Microbiol* 2000;**35**:896–910.
- Kolappan S, Craig L. Structure of the cytoplasmic domain of TcpE, the inner membrane core protein required for assembly of the *Vibrio cholerae* toxin-coregulated pilus. *Acta Crystallogr D* 2013;**69**:513–9.
- Koo J, Burrows LL, Howell PL. Decoding the roles of pilotins and accessory proteins in secretin escort services. *FEMS Microbiol Lett* 2012;**328**:1–12.
- Koo J, Tammam S, Ku SY, et al., PilF is an outer membrane lipoprotein required for multimerization and localization of the *Pseudomonas aeruginosa* type IV pilus secretin. *J Bacteriol* 2008;**190**:6961–9.
- Koo J, Tang T, Harvey H, et al., Functional mapping of PilF and PilQ in the *Pseudomonas aeruginosa* type IV pilus system. *Biochemistry* 2013;**52**:2914–23.
- Korotkov KV, Gonen T, Hol WG. Secretins: dynamic channels for protein transport across membranes. *Trends Biochem Sci* 2011a;**36**:433–43.
- Korotkov KV, Hol WG. Structure of the GspK-GspI-GspJ complex from the enterotoxigenic *Escherichia coli* type 2 secretion system. *Nat Struct Mol Biol* 2008;**15**:462–8.

- Korotkov KV, Johnson TL, Jobling MG, et al., Structural and functional studies on the interaction of GspC and GspD in the type II secretion system. *PLoS Pathog* 2011b;7:e1002228.
- Korotkov KV, Pardon E, Steyaert J, et al., Crystal structure of the N-terminal domain of the secretin GspD from ETEC determined with the assistance of a nanobody. *Structure* 2009;17:255–65.
- Korotkov KV, Sandkvist M, Hol WG. The type II secretion system: biogenesis, molecular architecture and mechanism. *Nat Rev Microbiol* 2012;10:336–51.
- LaPointe CF, Taylor RK. The type 4 prepilin peptidases comprise a novel family of aspartic acid proteases. *J Biol Chem* 2000;275:1502–10.
- Lario PI, Pfuetzner RA, Frey EA, et al., Structure and biochemical analysis of a secretin pilot protein. *EMBO J* 2005;24:1111–21.
- Laurenceau R, Pehau-Arnaudet G, Baconnais S, et al., A type IV pilus mediates DNA binding during natural transformation in *Streptococcus pneumoniae*. *PLoS Pathog* 2013;9:e1003473.
- Lemkul JA, Bevan DR. Characterization of interactions between PilA from *Pseudomonas aeruginosa* strain K and a model membrane. *J Phys Chem B* 2011;115:8004–8.
- Li J, Egelman EH, Craig L. Structure of the *Vibrio cholerae* type IVb pilus and stability comparison with the *Neisseria gonorrhoeae* type IVa pilus. *J Mol Biol* 2012;418:47–64.
- Li J, Lim MS, Li S, et al., *Vibrio cholerae* toxin-coregulated pilus structure analyzed by hydrogen/deuterium exchange mass spectrometry. *Structure* 2008;16:137–48.
- Lillington J, Geibel S, Waksman G. Biogenesis and adhesion of type 1 and P pili. *Biochim Biophys Acta* 2014;1840:2783–93.
- Linderoth NA, Simon MN, Russel M. The filamentous phage pIV multimer visualized by scanning transmission electron microscopy. *Science* 1997;278:1635–8.
- Low HH, Gubellini F, Rivera-Calzada A, et al., Structure of a type IV secretion system. *Nature* 2014;508:550–3.
- Lu A, Magupalli VG, Ruan J, et al., Unified polymerization mechanism for the assembly of ASC-dependent inflammasomes. *Cell* 2014a;156:1193–206.
- Lu C, Korotkov KV, Hol WG. Crystal structure of the full-length ATPase GspE from the *Vibrio vulnificus* type II secretion system in complex with the cytoplasmic domain of GspL. *J Struct Biol* 2014b;187:223–35.
- Lu C, Turley S, Marionni ST, et al., Hexamers of the type II secretion ATPase GspE from *Vibrio cholerae* with increased ATPase activity. *Structure* 2013;21:1707–17.
- Lu HM, Motley ST, Lory S. Interactions of the components of the general secretion pathway: role of *Pseudomonas aeruginosa* type IV pilin subunits in complex formation and extracellular protein secretion. *Mol Microbiol* 1997;25:247–59.
- Lybarger SR, Johnson TL, Gray MD, et al., Docking and assembly of the type II secretion complex of *Vibrio cholerae*. *J Bacteriol* 2009;191:3149–61.
- Maier B, Chen I, Dubnau D, et al., DNA transport into *Bacillus subtilis* requires proton motive force to generate large molecular forces. *Nat Struct Mol Biol* 2004;11:643–9.
- Maier B, Potter L, So M, et al., Single pilus motor forces exceed 100 pN. *Proc Natl Acad Sci USA* 2002;99:16012–7.
- Manning PA. The *tcp* gene cluster of *Vibrio cholerae*. *Gene* 1997;192:63–70.
- Marceau M, Forest K, Beretti J-L, et al., Consequences of the loss of O-linked glycosylation of meningococcal type IV pilin on piliation and pilus-mediated adhesion. *Mol Microbiol* 1998;27:705–15.
- Mattick JS. Type IV pili and twitching motility. *Annu Rev Microbiol* 2002;56:289–314.
- Melville S, Craig L. Type IV pili in Gram-positive bacteria. *Microbiol Mol Biol R* 2013;77:323–41.
- Merz AJ, So M, Sheetz MP. Pilus retraction powers bacterial twitching motility. *Nature* 2000;407:98–102.
- Mikaty G, Soyer M, Mairey E, et al., Extracellular bacterial pathogen induces host cell surface reorganization to resist shear stress. *PLoS Pathog* 2009;5:e1000314.
- Misic AM, Satyshur KA, Forest KT. *P. aeruginosa* PilT structures with and without nucleotide reveal a dynamic type IV pilus retraction motor. *J Mol Biol* 2010;400:1011–21.
- Morand PC, Bille E, Morelle S, et al., Type IV pilus retraction in pathogenic *Neisseria* is regulated by the PilC proteins. *EMBO J* 2004;23:2009–17.
- Nassif X, Beretti J-L, Lowy J, et al., Roles of pilin and PilC in adhesion of *Neisseria meningitidis* to human epithelial and endothelial cells. *Proc Natl Acad Sci USA* 1994;91:3769–73.
- Ng SY, VanDyke DJ, Chaban B, et al., Different minimal signal peptide lengths recognized by the archaeal prepilin-like peptidases FlaK and PibD. *J Bacteriol* 2009;191:6732–40.
- Ng SY, Wu J, Nair DB, et al., Genetic and mass spectrometry analyses of the unusual type IV-like pili of the archaeon *Methanococcus maripaludis*. *J Bacteriol* 2011;193:804–14.
- Nishiyama M, Ishikawa T, Rechsteiner H, et al., Reconstitution of pilus assembly reveals a bacterial outer membrane catalyst. *Science* 2008;320:376–9.
- Nivaskumar M, Bouvier G, Campos M, et al., Distinct docking and stabilization steps of the pseudopilus conformational transition path suggest rotational assembly of type IV pilus-like fibers. *Structure* 2014;22:685–96.
- Nivaskumar M, Francetic O. Type II secretion system: a magic beanstalk or a protein escalator. *Biochim Biophys Acta* 2014;1843:1568–77.
- Nouwen N, Ranson N, Saibil H, et al., Secretin PulD: association with pilot PulS, structure, and ion-conducting channel formation. *Proc Natl Acad Sci USA* 1999;96:8173–7.
- Nudleman E, Wall D, Kaiser D. Polar assembly of the type IV pilus secretin in *Myxococcus xanthus*. *Mol Microbiol* 2006;60:16–29.
- Nunn D, Bergman S, Lory S. Products of three accessory genes, *pilB*, *pilC*, and *pilD*, are required for biogenesis of *Pseudomonas aeruginosa* pili. *J Bacteriol* 1990;172:2911–9.
- Nunn DN, Lory S. Product of the *Pseudomonas aeruginosa* gene *pilD* is a prepilin leader peptidase. *Proc Natl Acad Sci USA* 1991;88:3281–5.
- Nunn DN, Lory S. Cleavage, methylation, and localization of the *Pseudomonas aeruginosa* export proteins XcpT, -U, -V, and -W. *J Bacteriol* 1993;175:4375–82.
- O'Toole G, Kolter R. Flagellar and twitching motility are necessary for *Pseudomonas aeruginosa* biofilm development. *Mol Microbiol* 1998;30:295–304.
- Opalka N, Beckmann R, Boisset N, et al., Structure of the filamentous phage pIV multimer by cryo-electron microscopy. *J Mol Biol* 2003;325:461–70.
- Orans J, Johnson MD, Coggan KA, et al., Crystal structure analysis reveals *Pseudomonas* PilY1 as an essential calcium-dependent regulator of bacterial surface motility. *Proc Natl Acad Sci USA* 2010;107:1065–70.
- Ottow JC. Ecology, physiology, and genetics of fimbriae and pili. *Annu Rev Microbiol* 1975;29:79–108.
- Paranchych W, Sastry PA, Frost LS, et al., Biochemical studies on pili isolated from *Pseudomonas aeruginosa* strain PAO. *Can J Microbiol* 1979;25:1175–81.

- Parge HE, Forest KT, Hickey MJ, et al., Structure of the fibre-forming protein pilin at 2.6 Å resolution. *Nature* 1995;**378**:32–8.
- Pasloske BL, Paranchych W. The expression of mutant pilins in *Pseudomonas aeruginosa*: fifth position glutamate affects pilin methylation. *Mol Microbiol* 1988;**2**:489–95.
- Pasloske BL, Scraba DG, Paranchych W. Assembly of mutant pilins in *Pseudomonas aeruginosa*: formation of pili composed of heterologous subunits. *J Bacteriol* 1989;**171**:2142–7.
- Patrick M, Korotkov KV, Hol WG, et al., Oligomerization of EpsE coordinates residues from multiple subunits to facilitate ATPase activity. *J Biol Chem* 2011;**286**:10378–86.
- Peabody CR, Chung YJ, Yen MR, et al., Type II protein secretion and its relationship to bacterial type IV pili and archaeal flagella. *Microbiology* 2003;**149**:3051–72.
- Pegden RS, Larson MA, Grant RJ, et al., Adherence of the Gram-positive bacterium *Ruminococcus albus* to cellulose and identification of a novel form of cellulose-binding protein which belongs to the Pil family of proteins. *J Bacteriol* 1998;**180**:5921–7.
- Pellic V. Type IV pili: *e pluribus unum*? *Mol Microbiol* 2008;**68**:827–37.
- Phan G, Remaut H, Wang T, et al., Crystal structure of the FimD usher bound to its cognate FimC-FimH substrate. *Nature* 2011;**474**:49–53.
- Piepenbrink KH, Maldarelli GA, de la Pena CF, et al., Structure of *Clostridium difficile* PilJ exhibits unprecedented divergence from known type IV pilins. *J Biol Chem* 2014;**289**:4334–45.
- Pinkner JS, Remaut H, Buelens F, et al., Rationally designed small compounds inhibit pilus biogenesis in uropathogenic bacteria. *Proc Natl Acad Sci USA* 2006;**103**:17897–902.
- Planet PJ, Kachlany SC, DeSalle R, et al., Phylogeny of genes for secretion NTPases: identification of the widespread *tadA* subfamily and development of a diagnostic key for gene classification. *Proc Natl Acad Sci USA* 2001;**98**:2503–8.
- Porsch EA, Johnson MD, Broadnax AD, et al., Calcium binding properties of the *Kingella kingae* PilC1 and PilC2 proteins have differential effects on type IV pilus-mediated adherence and twitching motility. *J Bacteriol* 2013;**195**:886–95.
- Possot O, Pugsley AP. Molecular characterization of PulE, a protein required for pullulanase secretion. *Mol Microbiol* 1994;**12**:287–99.
- Possot OM, Gerard-Vincent M, Pugsley AP. Membrane association and multimerization of secretion component PulC. *J Bacteriol* 1999;**181**:4004–11.
- Possot OM, Vignon G, Bomchil N, et al., Multiple interactions between pullulanase secretion components involved in stabilization and cytoplasmic membrane association of PulE. *J Bacteriol* 2000;**182**:2142–52.
- Pugsley AP. Processing and methylation of PulG, a pilin-like component of the general secretory pathway of *Klebsiella oxytoca*. *Mol Microbiol* 1993a;**9**:295–308.
- Pugsley AP. The complete general secretory pathway in Gram-negative bacteria. *Microbiol Rev* 1993b;**57**:50–108.
- Py B, Loiseau L, Barras F. Assembly of the type II secretion machinery of *Erwinia chrysanthemi*: direct interaction and associated conformational change between OutE, the putative ATP-binding component and the membrane protein OutL. *J Mol Biol* 1999;**289**:659–70.
- Py B, Loiseau L, Barras F. An inner membrane platform in the type II secretion machinery of Gram-negative bacteria. *EMBO Rep* 2001;**2**:244–8.
- Rahman M, Källström H, Normark S, et al., PilC of pathogenic *Neisseria* is associated with the bacterial cell surface. *Mol Microbiol* 1997;**25**:11–25.
- Rakotoarivonina H, Jubelin G, Hebraud M, et al., Adhesion to cellulose of the Gram-positive bacterium *Ruminococcus albus* involves type IV pili. *Microbiology* 2002;**148**:1871–80.
- Reardon PN, Mueller KT. Structure of the type IVa major pilin from the electrically conductive bacterial nanowires of *Geobacter sulfurreducens*. *J Biol Chem* 2013;**288**:29260–6.
- Reguera G, McCarthy KD, Mehta T, et al., Extracellular electron transfer via microbial nanowires. *Nature* 2005;**435**:1098–101.
- Reindl S, Ghosh A, Williams GJ, et al., Insights into Flal functions in archaeal motor assembly and motility from structures, conformations, and genetics. *Mol Cell* 2013;**49**:1069–82.
- Robert V, Filloux A, Michel GP. Subcomplexes from the Xcp secretion system of *Pseudomonas aeruginosa*. *FEMS Microbiol Lett* 2005;**252**:43–50.
- Rose I, Biukovic G, Aderhold P, et al., Identification and characterization of a unique, zinc-containing transport ATPase essential for natural transformation in *Thermus thermophilus* HB27. *Extremophiles* 2011;**15**:191–202.
- Rudel T, Scheuerpflug I, Meyer TF. *Neisseria* PilC protein identified as a type-4 pilus-tip located adhesin. *Nature* 1995;**373**:357–9.
- Rudel T, vanPutten JPM, Gibbs CP, et al., Interaction of two variable proteins (PilE and PilC) required for pilus-mediated adherence of *Neisseria gonorrhoeae* to human epithelial cells. *Mol Microbiol* 1992;**6**:3439–50.
- Sakai D, Horiuchi T, Komano T. ATPase activity and multimer formation of PilQ protein are required for thin pilus biogenesis in plasmid R64. *J Biol Chem* 2001;**276**:17968–75.
- Sampaleanu LM, Bonanno JB, Ayers M, et al., Periplasmic domains of *Pseudomonas aeruginosa* PilN and PilO form a stable heterodimeric complex. *J Mol Biol* 2009;**394**:143–59.
- Sandkvist M, Bagdasarian M, Howard SP, et al., Interaction between the autokinase EpsE and EpsL in the cytoplasmic membrane is required for extracellular secretion in *Vibrio cholerae*. *EMBO J* 1995;**14**:1664–73.
- Sandkvist M, Hough LP, Bagdasarian MM, et al., Direct interaction of the EpsL and EpsM proteins of the general secretion apparatus in *Vibrio cholerae*. *J Bacteriol* 1999;**181**:3129–35.
- Satyshur KA, Worzalla GA, Meyer LS, et al., Crystal structures of the pilus retraction motor PilT suggest large domain movements and subunit cooperation drive motility. *Structure* 2007;**15**:363–76.
- Sauvonnet N, Vignon G, Pugsley AP, et al., Pilus formation and protein secretion by the same machinery in *Escherichia coli*. *EMBO J* 2000;**19**:2221–8.
- Schmidt SA, Bieber D, Ramer SW, et al., Structure-function analysis of BfpB, a secretin-like protein encoded by the bundle-forming-pilus operon of enteropathogenic *Escherichia coli*. *J Bacteriol* 2001;**183**:4848–59.
- Schuch R, Maurelli AT. MxiM and MxiJ, base elements of the Mxi-Spa type III secretion system of *Shigella*, interact with and stabilize the MxiD secretin in the cell envelope. *J Bacteriol* 2001;**183**:6991–8.
- Shahapure R, Driessen RP, Haurat MF, et al., The archaeum: a rotating type IV pilus. *Mol Microbiol* 2014;**91**:716–23.
- Shevchik VE, Robert-Baudouy J, Condemine G. Specific interaction between OutD, an *Erwinia chrysanthemi* outer membrane protein of the general secretory pathway, and secreted proteins. *EMBO J* 1997;**16**:3007–16.

- Shiue SJ, Kao KM, Leu WM, et al., XpsE oligomerization triggered by ATP binding, not hydrolysis, leads to its association with XpsL. *EMBO J* 2006;**25**:1426–35.
- Siewering K, Jain S, Friedrich C, et al. Peptidoglycan-binding protein TsaP functions in surface assembly of type IV pili. *Proc Natl Acad Sci USA* 2014;**111**:E953–61.
- Skerker JM, Shapiro L. Identification and cell cycle control of a novel pilus system in *Caulobacter crescentus*. *EMBO J* 2000;**19**:3223–34.
- Smedley JG, III, Jewell E, Roguskie J, et al., Influence of pilin glycosylation on *Pseudomonas aeruginosa* 1244 pilus function. *Infect Immun* 2005;**73**:7922–31.
- Sohel I, Puente JL, Ramer SW, et al., Enteropathogenic *Escherichia coli*: identification of a gene cluster coding for bundle-forming pilus morphogenesis. *J Bacteriol* 1996;**178**:2613–28.
- Stone KD, Zhang HZ, Carlson LK, et al., A cluster of fourteen genes from enteropathogenic *Escherichia coli* is sufficient for the biogenesis of a type IV pilus. *Mol Microbiol* 1996;**20**:325–37.
- Strom MS, Lory S. Mapping of export signals of *Pseudomonas aeruginosa* pilin with alkaline phosphatase fusions. *J Bacteriol* 1987;**169**:3181–8.
- Strom MS, Lory S. Amino acid substitutions in pilin of *Pseudomonas aeruginosa*. Effect on leader peptide cleavage, aminoterminal methylation, and pilus assembly. *J Biol Chem* 1991;**266**:1656–64.
- Strom MS, Nunn DN, Lory S. A single bifunctional enzyme, PilD, catalyzes cleavage and N-methylation of proteins belonging to the type IV pilin family. *Proc Natl Acad Sci USA* 1993;**90**:2404–8.
- Szabó Z, Stahl AO, Albers SV, et al., Identification of diverse archaeal proteins with class III signal peptides cleaved by distinct archaeal prepilin peptidases. *J Bacteriol* 2007;**189**:772–8.
- Szeto TH, Dessen A, Pelicic V. Structure/function analysis of *Neisseria meningitidis* PilW, a conserved protein playing multiple roles in type IV pilus biology. *Infect Immun* 2011;**79**:3028–35.
- Takhar HK, Kemp K, Kim M, et al., The platform protein is essential for type IV pilus biogenesis. *J Biol Chem* 2013;**288**:9721–8.
- Tammam S, Sampaleanu LM, Koo J, et al., Characterization of the PilN, PilO and PilP type IVa pilus subcomplex. *Mol Microbiol* 2011;**82**:1496–514.
- Tammam S, Sampaleanu LM, Koo J, et al., PilMNOPQ from the *Pseudomonas aeruginosa* type IV pilus system form a transenvelope protein interaction network that interacts with PilA. *J Bacteriol* 2013;**195**:2126–35.
- Thomas JD, Reeves PJ, Salmond GP. The general secretion pathway of *Erwinia carotovora* subsp. *carotovora*: analysis of the membrane topology of OutC and OutF. *Microbiology* 1997;**143**:713–20.
- Thomas NA, Chao ED, Jarrell KF. Identification of amino acids in the leader peptide of *Methanococcus voltae* preflagellin that are important in posttranslational processing. *Arch Microbiol* 2001;**175**:263–9.
- Tomich M, Fine DH, Figurski DH. The TadV protein of *Actinobacillus actinomycetemcomitans* is a novel aspartic acid prepilin peptidase required for maturation of the Flp1 pilin and TadE and TadF pseudopilins. *J Bacteriol* 2006;**188**:6899–914.
- Tomich M, Planet PJ, Figurski DH. The *tad* locus: postcards from the widespread colonization island. *Nat Rev Microbiol* 2007;**5**:363–75.
- Tosi T, Estrozi LF, Job V, et al., Structural similarity of secretins from type II and type III secretion systems. *Structure* 2014;**22**:1348–55.
- Tosi T, Nickerson NN, Mollica L, et al., Pilotin-secretin recognition in the type II secretion system of *Klebsiella oxytoca*. *Mol Microbiol* 2011;**82**:1422–32.
- Tripathi SA, Taylor RK. Membrane association and multimerization of TcpT, the cognate ATPase ortholog of the *Vibrio cholerae* toxin co-regulated pilus biogenesis apparatus. *J Bacteriol* 2007;**189**:4401–9.
- Turner LR, Lara JC, Nunn DN, et al., Mutations in the consensus ATP-binding sites of XcpR and PilB eliminate extracellular protein secretion and pilus biogenesis in *Pseudomonas aeruginosa*. *J Bacteriol* 1993;**175**:4962–9.
- van Schaik EJ, Giltner CL, Audette GF, et al., DNA binding: a novel function of *Pseudomonas aeruginosa* type IV pili. *J Bacteriol* 2005;**187**:1455–64.
- VanDyke DJ, Wu J, Ng SY, et al., Identification of a putative acetyltransferase gene, MMP0350, which affects proper assembly of both flagella and pili in the archaeon *Methanococcus maripaludis*. *J Bacteriol* 2008;**190**:5300–7.
- Vargas M, Malvankar NS, Tremblay PL, et al., Aromatic amino acids required for pili conductivity and long-range extracellular electron transport in *Geobacter sulfurreducens*. *mBio* 2013;**4**:e00105–13.
- Viarre V, Cascales E, Ball G, et al., HxcQ liposecretin is self-piloted to the outer membrane by its N-terminal lipid anchor. *J Biol Chem* 2009;**284**:33815–23.
- Vignon G, Kohler R, Larquet E, et al., Type IV-like pili formed by the type II secretin: specificity, composition, bundling, polar localization, and surface presentation of peptides. *J Bacteriol* 2003;**185**:3416–28.
- von Heijne G, Gavel Y. Topogenic signals in integral membrane proteins. *Eur J Biochem* 1988;**174**:671–8.
- Voulhoux R, Bos MP, Geurtsen J, et al., Role of a highly conserved bacterial protein in outer membrane protein assembly. *Science* 2003;**299**:262–5.
- Wang YA, Yu X, Ng SY, et al., The structure of an archaeal pilus. *J Mol Biol* 2008;**381**:456–66.
- Wehbi H, Portillo E, Harvey H, et al., The peptidoglycan-binding protein FimV promotes assembly of the *Pseudomonas aeruginosa* type IV pilus secretin. *J Bacteriol* 2011;**193**:540–50.
- Winther-Larsen HC, Hegge FT, Wolfgang M, et al., *Neisseria gonorrhoeae* PilV, a type IV pilus-associated protein essential to human epithelial cell adherence. *Proc Natl Acad Sci USA* 2001;**98**:15276–81.
- Winther-Larsen HC, Wolfgang M, Dunham S, et al., A conserved set of pilin-like molecules controls type IV pilus dynamics and organelle-associated functions in *Neisseria gonorrhoeae*. *Mol Microbiol* 2005;**56**:903–17.
- Wolfgang M, Lauer P, Park HS, et al., PilT mutations lead to simultaneous defects in competence for natural transformation and twitching motility in piliated *Neisseria gonorrhoeae*. *Mol Microbiol* 1998a;**29**:321–30.
- Wolfgang M, Park HS, Hayes SF, et al., Suppression of an absolute defect in type IV pilus biogenesis by loss-of-function mutations in *pilT*, a twitching motility gene in *Neisseria gonorrhoeae*. *Proc Natl Acad Sci USA* 1998b;**95**:14973–8.
- Wolfgang M, van Putten JP, Hayes SF, et al., Components and dynamics of fiber formation define a ubiquitous biogenesis pathway for bacterial pili. *EMBO J* 2000;**19**:6408–18.
- Yamagata A, Milgotina E, Scanlon K, et al., Structure of an essential type IV pilus biogenesis protein provides insights into

- pilus and type II secretion systems. *J Mol Biol* 2012;**419**:110–24.
- Yamagata A, Tainer JA. Hexameric structures of the archaeal secretion ATPase GspE and implications for a universal secretion mechanism. *EMBO J* 2007;**26**:878–90.
- Yoshida T, Furuya N, Ishikura M, et al., Purification and characterization of thin pili of IncI1 plasmids Collb-P9 and R64: formation of PilV-specific cell aggregates by type IV pili. *J Bacteriol* 1998;**180**:2842–8.
- Yoshida T, Kim SR, Komano T. Twelve *pil* genes are required for biogenesis of the R64 thin pilus. *J Bacteriol* 1999;**181**:2038–43.
- Yu X, Goforth C, Meyer C, et al., Filaments from *Ignicoccus hospitalis* show diversity of packing in proteins containing N-terminal type IV pilin helices. *J Mol Biol* 2012;**422**:274–81.
- Zahavi EE, Lieberman JA, Donnenberg MS, et al., Bundle-forming pilus retraction enhances enteropathogenic *Escherichia coli* infectivity. *Mol Biol Cell* 2011;**22**:2436–47.
- Zolghadr B, Weber S, Szabo Z, et al., Identification of a system required for the functional surface localization of sugar binding proteins with class III signal peptides in *Sulfolobus solfataricus*. *Mol Microbiol* 2007;**64**:795–806.